

# Dorsal and ventral horn atrophy is associated with clinical outcome after spinal cord injury

Eveline Huber, MSc, Gergely David, MSc, Alan J. Thompson, MD, Nikolaus Weiskopf, PhD, Siawoosh Mohammadi, PhD, and Patrick Freund, MD, PhD

*Neurology*® 2018;90:e1510-e1522. doi:10.1212/WNL.0000000000005361

## Correspondence

Dr. Freund  
patrick.freund@balgrist.ch

## Abstract

### Objective

To investigate whether gray matter pathology above the level of injury, alongside white matter changes, also contributes to sensorimotor impairments after spinal cord injury.

### Methods

A 3T MRI protocol was acquired in 17 tetraplegic patients and 21 controls. A sagittal T2-weighted sequence was used to characterize lesion severity. At the C2-3 level, a high-resolution T2\*-weighted sequence was used to assess cross-sectional areas of gray and white matter, including their subcompartments; a diffusion-weighted sequence was used to compute voxel-based diffusion indices. Regression models determined associations between lesion severity and tissue-specific neurodegeneration and associations between the latter with neurophysiologic and clinical outcome.

### Results

Neurodegeneration was evident within the dorsal and ventral horns and white matter above the level of injury. Tract-specific neurodegeneration was associated with prolonged conduction of appropriate electrophysiologic recordings. Dorsal horn atrophy was associated with sensory outcome, while ventral horn atrophy was associated with motor outcome. White matter integrity of dorsal columns and corticospinal tracts was associated with daily-life independence.

### Conclusion

Our results suggest that, next to anterograde and retrograde degeneration of white matter tracts, neuronal circuits within the spinal cord far above the level of injury undergo trans-synaptic neurodegeneration, resulting in specific gray matter changes. Such improved understanding of tissue-specific cord pathology offers potential biomarkers with more efficient targeting and monitoring of neuroregenerative (i.e., white matter) and neuroprotective (i.e., gray matter) agents.

## CME Course

[NPublic.org/cmelist](https://npublic.org/cmelist)

From the Spinal Cord Injury Center (E.H., G.D., P.F.), Balgrist University Hospital, Zurich, Switzerland; Department of Brain Repair and Rehabilitation (A.J.T., P.F.) and Wellcome Trust Centre for Neuroimaging (N.W., S.M., P.F.), UCL Institute of Neurology, University College London, UK; Department of Neurophysics (N.W., P.F.), Max Planck Institute for Human Cognitive and Brain Sciences, Leipzig, Germany; and Department of Systems Neuroscience (S.M.), University Medical Center Hamburg-Eppendorf, Germany.

Go to [Neurology.org/N](https://Neurology.org/N) for full disclosures. Funding information and disclosures deemed relevant by the authors, if any, are provided at the end of the article.

The Article Processing Charge was funded by Wellcome Trust.

This is an open access article distributed under the terms of the Creative Commons Attribution License 4.0 (CC BY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## Glossary

**AD** = axonal diffusivity; **CHEP** = contact heat evoked potential; **CI** = confidence interval; **DCA** = dorsal column area; **DHA** = dorsal horn area; **DTI** = diffusion tensor imaging; **FA** = fractional anisotropy; **FOV** = field of view; **GMA** = gray matter area; **GRASSP** = Graded Redefined Assessment of Strength, Sensibility and Prehension; **MD** = mean diffusivity; **RD** = radial diffusivity; **SCA** = spinal cord area; **SCI** = spinal cord injury; **SCIM** = Spinal Cord Independence Measure; **SSEP** = somatosensory evoked potential; **VHA** = ventral horn area; **WISCI** = Walking Index for Spinal Cord Injury; **WMA** = white matter area.

Spinal cord injury (SCI) usually leads to sensorimotor dysfunction resulting from damage at the level of injury. However, a complex cascade of secondary neurodegenerative processes occur across the spinal cord and brain.<sup>1</sup> In chronic SCI, cervical cord atrophy of up to 30% has been reported above the level of injury; its magnitude relates to the degree of clinical impairment.<sup>2</sup> Recent improvements in diffusion-weighted imaging and anatomic sequences with higher in-plane resolution,<sup>3</sup> combined with advanced postprocessing techniques,<sup>4,5</sup> now allow the assessment of gray and white matter changes in the cervical spinal cord occurring after SCI.

Although white matter pathology within the spinal cord contributes to sensorimotor impairments, the functional effects of gray matter pathology above the level of injury are uncertain. Improved understanding of tissue-specific cord pathology may allow more efficient targeting and monitoring of neuroregenerative and neuroprotective agents. This study therefore addresses to what extent cord atrophy above the level of injury is driven by pathophysiologic processes occurring in gray and white matter, whether lesion severity is associated with the magnitude of neurodegeneration above the level of injury, and whether the tissue-specific neurodegeneration is associated with neurophysiologic and clinical outcome.

Using structural and diffusion MRI data, we assessed tissue-specific cord pathology above the level of injury in patients with chronic SCI compared to healthy controls. These measures included the assessment of dorsal horn area (DHA) and ventral horn area (VHA),<sup>6</sup> diffusivity changes within the major spinal pathways, and associations between lesion severity,<sup>7</sup> tissue-specific pathology, and neurophysiologic changes.

## Methods

### Standard protocol approvals, registrations, and patient consents

Our study protocol was designed in accordance with the Declaration of Helsinki and was approved by the local ethics committee of Zurich (KEK-ZH-Nr. 2012-0343, PB\_2016-00623). All participants gave their written informed consent before participation.

### Participants

We recruited 17 patients with SCI (mean age  $48.7 \pm 14.1$  years, 3 female patients) between November 2014 and May

2016 who were previously admitted to the University Hospital Balgrist (Zurich, Switzerland). Twenty-one healthy controls (mean age  $41.7 \pm 11.3$  years, 7 female controls) from the local neighborhood served as a control dataset that was acquired and used in a previous study.<sup>8</sup>

Inclusion criteria for patients with SCI were traumatic cervical SCI, no other neurologic or mental disorders affecting clinical outcome, age between 18 and 70 years, MRI compatible, and no pregnancy.

### Clinical assessments

All patients were examined with comprehensive clinical protocols to assess neurologic and functional impairment. These included the International Standards for Neurological Classification of Spinal Cord Injury protocol for motor, light-touch, and pinprick score and completeness of injury<sup>9</sup>; the Spinal Cord Independence Measure (SCIM) to measure daily life independence<sup>10</sup>; the Graded Redefined Assessment of Strength, Sensibility and Prehension (GRASSP) for assessing upper limb function<sup>11</sup>; and the Walking Index for Spinal Cord Injury (WISCI).<sup>12</sup> All patients completed the full protocol, except GRASSP score was not available for 1 patient.

### Neurophysiologic assessments

Contact heat evoked potentials (CHEPs) and somatosensory evoked potentials (SSEPs) were acquired bilaterally in patients at the dermatomes C4, C6, and C8 to measure the integrity of the spinothalamic tract (i.e., CHEPs) and the dorsal column (i.e., SSEPs). For the acquisition of CHEPs<sup>13</sup> and SSEPs,<sup>14</sup> the same protocols were applied as previously described.

### Contact heat evoked potentials

A contact heat stimulator (PATHWAY Pain & Sensory Evaluation System, Medoc, Ramat Yishay, Israel) was used to deliver contact heat stimuli from a baseline temperature of 35°C to a peak temperature of 52°C with a heating rate of 70°C/s and a cooling rate of 40°C/s. For each dermatome, we first assessed heat perception and pain thresholds within 2 consecutive trials. For the CHEPs recording, scalp recording sites were prepared with Nuprep (D.O. Weaver & Co, Aurora, CO) and alcohol. Three 9-mm Ag/AgCl surface disk electrodes were positioned according to the international 10-20 system with the active electrode at the Cz position and referenced to linked earlobes (A1-A2);

impedances were kept  $<5$  k $\Omega$ . Ten to 15 contact heat stimuli were applied (interstimulus interval 8–12 seconds). Two seconds after each stimulus, an audio cue appeared, and patients rated their perceived intensity according to a numeric rating scale. All signals were sampled from 100 milliseconds before the trigger to 1,500 milliseconds after the trigger at a sampling rate of 2,000 Hz with a preamplifier (20,000 $\times$  bandpass filter = 0.25–300 Hz; ALEA Solutions, Switzerland). Data were recorded in a LabView-based program (V1.43 CHEP; ALEA Solutions, Zurich, Switzerland) with a 100-millisecond period before the trigger and 1-second posttrigger period. Raw data were bandpass filtered from 0.5 to 30 Hz.

### Somatosensory evoked potentials

For dermatomal SSEPs, Key Point (Medtronic, Mississauga, ON, Canada) was used to record and deliver electric stimulation of 3 Hz. Stimuli were elicited by single 0.2-millisecond, repetitive, square-wave electric stimulation. We first assessed electric perception and pain thresholds for each dermatome (not exceeding 40 mA) within 2 consecutive trials. For the recording of SSEPs, surface gel electrodes (10 mm) were used on each dermatome after the skin was prepared with Nuprep (D.O. Weaver & Co) and alcohol. Disposable needle electrodes (Spes Medica, Srl, Genova, Italy) were positioned according to the international 10-20 system with the active electrode positioned at the contralateral side for the stimulated dermatome (C3-4) referenced to Fz; impedances were kept  $<5$  k $\Omega$ . The stimulation intensity was individually set as 3-fold electric perception threshold. Averages of 2 traces of 300 cortical responses were obtained for each dermatome. Raw data were bandpass filtered from 2 to 2,000 Hz.

### Neurophysiologic classification

We determined amplitudes and latencies of each dermatome for each patient after averaging all single-trial waveforms for CHEPs (i.e., N2P2, N2, P2) and SSEPs (i.e., N1P1, N1, P1).

Furthermore, CHEPs and SSEPs were classified as normal (onset latency  $\leq 2$  SDs from control dermatome recording), pathologic (onset latency  $>2$  SDs from control dermatome recording), or absent (not recordable).<sup>14</sup> The CHEPs protocol was acquired fully in 14 patients and partially in 1 patient. For SSEPs, 12 patients received the full protocol and 2 patients participated in part of the protocol.

### Image acquisition

All imaging was performed on a clinical 3T Skyra<sup>Fit</sup> scanner (Siemens Healthcare, Erlangen, Germany) equipped with a 16-channel radiofrequency receive-only head and neck coil and a radiofrequency body transmit coil. A stiff neck (Laerdal Medicals, Stavanger, Norway) was used in all participants to minimize motion artifacts. As a result of motion artifacts, 1 patient was excluded from macrostructural analysis, and 3 patients had to be excluded from microstructural analysis.

At the lesion epicenter, a sagittal T1-weighted (repetition time 600 milliseconds, echo time 9.9 milliseconds, flip angle 150°, in-plane resolution 0.57  $\times$  0.57 mm, slice thickness 3.3 mm), a sagittal T2-weighted (repetition time 3,500 milliseconds, echo time 84 milliseconds, flip angle 160°, in-plane resolution 0.34  $\times$  0.34 mm, slice thickness 2.75 mm), and an axial T2-weighted image (repetition time 5,510 milliseconds, echo time 93 milliseconds, flip angle 150°, in-plane resolution 0.5  $\times$  0.5 mm, slice thickness 3.6 mm) were acquired to assess the lesion size.

At the cervical cord above the level of injury (centered at C2-3), 5 volumes were acquired with a T2\*-weighted 3-dimensional multiecho gradient recall echo sequence (multiple echo data image combination<sup>15</sup>) in the oblique axial plane (i.e., perpendicular to the cord) to assess gray and white matter atrophy. Each of the 5 volumes acquired consisted of 20 partitions with a resolution of 0.5  $\times$  0.5 mm<sup>2</sup> (field of view 192  $\times$  162 mm<sup>2</sup>, slice thickness 2.50 mm [10% gap], repetition time 44 milliseconds, echo time 19 milliseconds, flip angle 11°, readout bandwidth 260 Hz/pixel). Each volume took 2.13 minutes to acquire. Application of zero-filling interpolation doubled the nominal in-plane resolution (0.25  $\times$  0.25 mm<sup>2</sup>).

At the identical level, a high-resolution diffusion tensor imaging (DTI) dataset was acquired with a cardiac-gated reduced-FOV single-shot spin-echo echo planar imaging sequence with outer volume suppression<sup>16</sup> to assess microstructural changes of the whole spinal cord. Four measurements of  $b = 0$  (T2-weighted) and 30  $b = 500$  s/mm<sup>2</sup> volumes were acquired, resulting in 144 images per participant and a nominal acquisition time of 6.17 minutes. The following parameters were applied: repetition time of 350 milliseconds; echo time of 71 milliseconds; slice thickness of 5 mm (10% interslice gap); resolution of 0.76  $\times$  0.76 mm<sup>2</sup>; FOV of 133  $\times$  30 mm<sup>2</sup>; phase oversampling of 50%; 5/8 partial-Fourier imaging in the phase-encoding direction; cardiac trigger delay of 200 milliseconds; and minimal time between triggers of 1,800 milliseconds. After acquisition, zero-filling interpolation was used to double the in-plane resolution (0.38  $\times$  0.38 mm<sup>2</sup>).

### Image processing

#### Lesion segmentation

With the use of the Jim 6.0 software (Xinapse Systems, Aldwincle, UK), the lesion was segmented on the midsagittal T2-weighted images, being visible as a high-signal-intensity area within the spinal cord, as previously described.<sup>7</sup> The following parameters were quantified: midsagittal anterior-posterior lesion width (equal to the maximal anterior-posterior width of the lesion), midsagittal rostrocaudal lesion length (equal to the maximal caudocranial extent of the lesion), total midsagittal lesion area, and midsagittal thickness of midsagittal ventral and dorsal tissue bridges at the widest point of the lesion, which was summed up to the total amount of midsagittal tissue bridges.

## Processing of high-resolution macrostructural data above the level of injury

We used serial longitudinal registration<sup>17</sup> embedded within SPM12 to average the five 3-dimensional MEDIC volumes, accounting for intraparticipant motion. To further increase the signal-to-noise ratio, the average volume was resampled at a double slice thickness. We then used the Jim 6.0 software to measure cross-sectional spinal cord area (SCA) of 3 slices. After the midpoint of the spinal cord was marked manually in each slice, the SCA was calculated automatically with the semiautomatic 3-dimensional active-surface model.<sup>18</sup> Gray matter area (GMA), dorsal column area (DCA), VHA (approximately lamina VI–IX), and DHA (approximately lamina I–V) were extracted manually.<sup>6</sup> White matter area (WMA) was calculated by subtracting the GMA from the SCA. The mean interobserver reliability and intraobserver reliability for these measures were previously shown.<sup>8,16</sup>

## Preprocessing and estimation of DTI data

All processing of the DTI data was carried out with a modified version of the MatLab-based ACID toolbox optimized for the spinal cord. First, we reduced the in-plane FOV to  $24 \times 24 \text{ mm}^2$  to exclude much of the non-spinal cord tissue in each participant. Then, DTI volumes were slice-wise linearly registered with 3 *df* (translation in the frequency- and phase-encoding direction, scaling in the phase-encoding direction) to correct for intraparticipant motion and eddy-current artifacts.<sup>19</sup> A diffusion tensor was fitted by use of a robust tensor fitting algorithm that accounts for outlier volumes due to motion and physiologic artifacts.<sup>20</sup> The following DTI index maps were extracted: fractional anisotropy (FA) and mean, axial, and radial diffusivity (MD, AD, and RD).

These DTI index maps were then spatially normalized to a self-constructed mean diffusivity template residing in the spinal Montreal Neurological Institute space.<sup>21</sup> To further refine the accuracy of the registration, a manual slice-by-slice registration (in-plane translation and scaling) was performed. Finally, all DTI index maps were smoothed with a full width at half-maximum gaussian kernel with  $0.5 \times 0.5 \times 5 \text{ mm}^3$ .

## Statistical analysis

Statistical analysis of all macrostructural MRI, neurophysiologic, and clinical data was performed with Stata13 (StataCorp LP, College Station, TX). The mean age was not statistically different between healthy controls and patients (Mann-Whitney *U* test:  $z = -1.61$ ,  $p = 0.10$ ). All images were visually inspected for artifacts, and the analysis was conducted on 3 slices from each modality at the same level.

First, we assessed the morphometric differences in SCA, GMA, WMA, DCA, VHA, and DHA between patients and healthy controls by means of analysis of covariance, adjusted for age. For microstructural differences between patients and healthy controls, we used SPM12 for voxel-based analysis of the different DTI indexes (FA, MD, AD, RD) by means of analysis of covariance, adjusted for age. All statistical

parametric maps were initially thresholded with a cluster-defining threshold of  $p < 0.01$  (uncorrected) and clusters surpassing a cluster threshold of  $p < 0.05$  (family-wise error corrected) are reported. Next, we used linear regression analysis to investigate the relationship between changes at the lesion site (midsagittal lesion area, length and width, and size of midsagittal tissue bridges) and remote cord macrostructural and microstructural changes. We then determined associations between macrostructural (SCA, GMA, WMA, DCA, VHA, and DHA) and microstructural (DTI indexes within lateral corticospinal tract, dorsal column, and spinal lemniscus) parameters and tract-specific clinical measures (motor, pinprick, and light-touch score, GRASSP, SCIM) using linear regression models, adjusted for age and lesion area. Finally, we investigated associations between macrostructural and microstructural MRI indexes and neurophysiologic outcome measures using linear regression models, adjusted for age and lesion area. Note that only patients with both recordable electrophysiologic potentials and available MRI data entered this regression analysis, resulting in a total number of 8 patients. For all microstructural associations, we extracted mean values of DTI indexes within anatomic regions of interest (lateral corticospinal tract, dorsal column, and spinal lemniscus [containing spinothalamic and spinoreticular tracts]) as embedded in the Spinal Cord Toolbox.<sup>22</sup> The level of significance was set to  $p < 0.05$ .

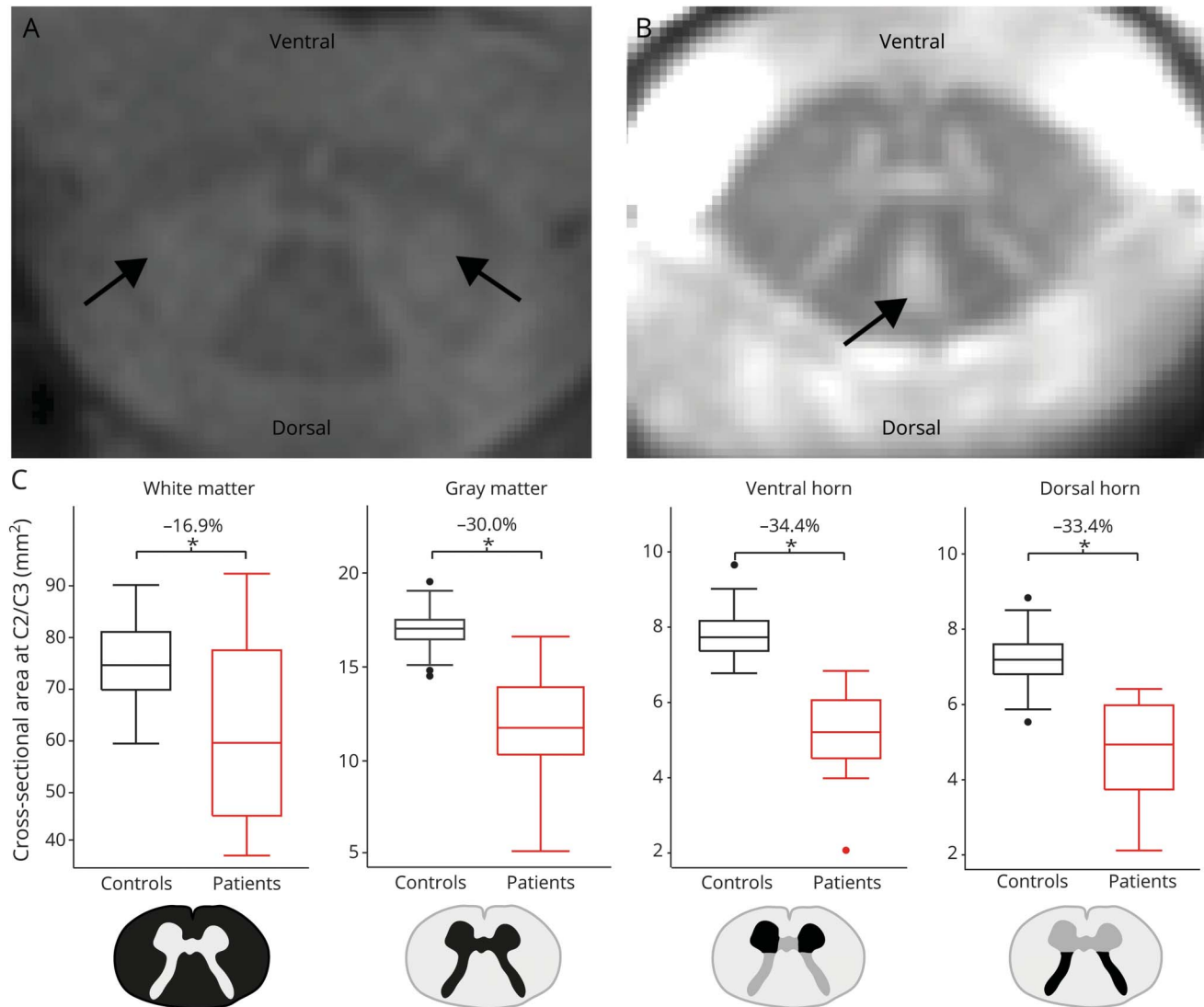
## Results

### Radiologic, clinical, and neurophysiologic characteristics

Patients were scanned  $6.7 \pm 7.8$  years after injury. An area of hyperintense signal was visible on the T2-weighted sagittal images in 16 patients (figure 1, A and B); 13 patients showed hyperintensities in their dorsal column, covering on average  $41.4 \pm 21.0\%$  of the whole dorsal column, and 2 patients showed hyperintensities in the dorsolateral funiculus (e.g., corticospinal tract). The radiologic level of injury (hyperintense T2-weighted signal) covered the vertebral level C3-5 in 1 patient, C3 in 1 patient, C4 in 2 patients, C4-5 in 1 patient, C5 in 2 patients, C6 in 2 patients, C6-7 in 4 patients, and C7 in 2 patients. Two patients showed no signal alteration within the cord. The average lesion area was  $45.4 \pm 66.6 \text{ mm}^2$  with a lesion length of  $11.3 \pm 9.4 \text{ mm}$  and a lesion width of  $4.3 \pm 3.5 \text{ mm}$ . In 2 patients, the lesion occupied the full cord area, and no midsagittal tissue bridges could be identified. In the remaining 15 patients, the midsagittal tissue bridges had an average width of  $2.9 \pm 1.9 \text{ mm}$ . No magnetic resonance abnormalities were identified in the control group.

Two patients were motor and sensory complete; 2 patients were motor complete and sensory incomplete; and the remaining 13 patients were motor and sensory incomplete. The motor score (maximum 100) was  $68.1 \pm 30.4$ ; the light-touch score was (mean  $\pm$  SD)  $66.3 \pm 32.7$  (maximum 112); and the pinprick score (maximum 112) was  $52.7 \pm$

**Figure 1** Macrostructural changes above the level of injury



Hyperintense regions most likely indicating (A) retrograde degeneration in the corticospinal tract and (B) anterograde degeneration in the dorsal column. Arrows indicate the corresponding locations. (C) Differences between the cross-sectional white matter area, cross-sectional gray matter area, cross-sectional ventral horn area, and cross-sectional dorsal horn area in patients compared to healthy controls.

35.0. Manual dexterity was impaired as assessed by the GRASSP score ( $149.8 \pm 66.3$  [maximum 232]), and functional independence was impaired as assessed by the SCIM score ( $[63.1 \pm 31.3$  [maximum. 100]). Eight patients were able to walk independently (20 of 20 points in the WISCI score); 2 patients were dependent on walking aids (5 of 20 and 9 of 20 points in the WISCI score); and 7 patients were not able to walk (0 of 20 points). All data are summarized in table 1.

All patients had neurophysiologic impairment of the spinothalamic tract, and a majority had impaired function of the dorsal column below the level of lesion as assessed by CHEPs and SSEPs, respectively. The mean  $\pm$  SD perception/pain thresholds and the amplitudes and latencies of the recorded signals are shown in table 2.

### Pathophysiologic changes in the cervical cord above the level of injury

Compared to healthy controls, patients showed a decreased SCA of 20.2% ( $p < 0.001$ , healthy controls  $92.30 \pm 8.49 \text{ mm}^2$ , patients  $73.71 \pm 20.04 \text{ mm}^2$ ). In patients, WMA was decreased by 16.9% ( $p = 0.001$ , healthy controls  $75.34 \pm 8.06 \text{ mm}^2$ , patients  $62.64 \pm 18.22 \text{ mm}^2$ ), and GMA was decreased by 30.0% ( $p < 0.001$ , healthy controls  $16.96 \pm 1.25 \text{ mm}^2$ , patients  $11.93 \pm 2.73 \text{ mm}^2$ ). In the white matter, DCA was decreased by 21.4% ( $p < 0.001$ , healthy controls  $23.73 \pm 2.99 \text{ mm}^2$ , patients  $18.65 \pm 4.76 \text{ mm}^2$ ). Within the gray matter, the bilateral VHA showed a 34.4% decrease in patients compared to healthy controls (left:  $p < 0.001$ , healthy controls  $3.84 \pm 0.29 \text{ mm}^2$ , patients  $2.56 \pm 0.62 \text{ mm}^2$ ; right:  $p < 0.001$ , healthy controls  $3.95 \pm 0.40 \text{ mm}^2$ , patients  $2.61 \pm 0.58 \text{ mm}^2$ ). In patients, the DHA was decreased bilaterally by

**Table 1** Clinical and epidemiologic data for all patients included in the study

| Patient | Sex    | Age, y | Years since injury | Radiologic level of injury | AIS grade | Neurologic level of injury | Motor score | Light-touch score | Pinprick score | SCIM score | GRASSP score | WISCI score |
|---------|--------|--------|--------------------|----------------------------|-----------|----------------------------|-------------|-------------------|----------------|------------|--------------|-------------|
| 1       | Male   | 29     | 1.0                | C4-5                       | A         | C4                         | 14          | 16                | 13             | 22         | 21           | 0           |
| 2       | Female | 40     | 7.0                | C3-5                       | A         | C4                         | 7           | 71                | 12             | 19         | 43           | 0           |
| 3       | Female | 39     | 25.0               | C6-7                       | B         | C5                         | 30          | 32                | 34             | 28         | 125          | 0           |
| 4       | Male   | 50     | 25.1               | C6-7                       | B         | C7                         | 46          | 62                | 26             | 63         | 188          | 5           |
| 5       | Male   | 70     | 0.7                | C5                         | C         | C2                         | 46          | 48                | 34             | 19         | 71           | 0           |
| 6       | Female | 32     | 1.2                | C6-7                       | C         | C6                         | 44          | 33                | 24             | 23         | 92           | 20          |
| 7       | Male   | 51     | 4.3                | NS                         | D         | C1                         | 82          | 90                | 57             | 100        | 130          | 20          |
| 8       | Male   | 56     | 5.6                | C4                         | D         | C2                         | 88          | 56                | 26             | 40         | 151          | 0           |
| 9       | Male   | 43     | 13.1               | C7                         | D         | C2                         | 76          | 55                | 55             | 74         | 225          | 0           |
| 10      | Male   | 60     | 0.3                | C3                         | D         | C3                         | 78          | 60                | 60             | 67         | NA           | 9           |
| 11      | Male   | 50     | 7.6                | C4                         | D         | C3                         | 84          | 10                | 10             | 97         | 136          | 20          |
| 12      | Male   | 48     | 1.8                | NS                         | D         | C4                         | 100         | 112               | 107            | 100        | 232          | 20          |
| 13      | Female | 63     | 0.3                | C5                         | D         | C6                         | 85          | 109               | 99             | 70         | 172          | 20          |
| 14      | Male   | 69     | 0.2                | C6                         | D         | C6                         | 99          | 109               | 99             | 87         | 206          | 20          |
| 15      | Male   | 67     | 12.6               | C6-7                       | D         | C7                         | 91          | 110               | 107            | 99         | 183          | 20          |
| 16      | Male   | 27     | 4.7                | C6                         | D         | C7                         | 92          | 72                | 45             | 75         | 189          | 20          |
| 17      | Male   | 33     | 3.0                | C7                         | D         | C8                         | 96          | 82                | 88             | 89         | 232          | 20          |

Abbreviations: AIS = American Spinal Injury Association Impairment Scale; GRASSP = Graded Redefined Assessment of Strength, Sensibility and Prehension; NA = not available; NS = no signal alteration within myelon; SCIM = Spinal Cord Independence Measure; WISCI = Walking Index for Spinal Cord Injury.

33.4% (left:  $p < 0.001$ , healthy controls  $3.63 \pm 0.41 \text{ mm}^2$ , patients  $2.42 \pm 0.66 \text{ mm}^2$ ; right:  $p < 0.001$ , healthy controls  $3.60 \pm 0.51 \text{ mm}^2$ , patients  $2.37 \pm 0.66 \text{ mm}^2$ ) (figure 1C), and smaller DHA was associated with smaller DCA ( $p < 0.001$ ,  $R^2 = 0.74$ , 95% confidence interval [CI] 2.21–4.28).

Voxel-based analysis of the cervical cord revealed a 16.6% decrease in FA in the left dorsolateral funiculus (e.g., containing spinothalamic and lateral corticospinal tracts,  $p = 0.003$ ; localization [x, y, z] -4.2, -18.5, 26; z score 4.42; cluster extent 154), 14.9% decrease in the right dorsolateral funiculus ( $p = 0.025$ ; localization [x, y, z] 6, -18.5, 37; z score 4.34; cluster extent 85), and 17.0% decrease in the posterior funiculus (i.e., containing dorsal columns;  $p = 0.004$ ; localization [x, y, z] 0.7, -22.3, 37; z score 3.80; cluster extent 145) in patients compared to healthy controls. AD was also decreased in patients compared to healthy controls in the same regions, namely by 12.8% in the left dorsolateral funiculus ( $p = 0.014$ ; localization [x, y, z] -3.1, -19.2, 26; z score 3.72; cluster extent 58), 12.8% the right dorsolateral funiculus ( $p = 0.002$ ; localization [x, y, z] 4.1, -18.8, 26; z score 4.70; cluster extent 94), and 9.9% in the posterior funiculus ( $p = 0.020$ ; localization [x, y, z] 0.7, -19.2, 32; z score 3.69; cluster extent 52). RD increased by 31.8% in the dorsal column ( $p = 0.022$ ;

localization [x, y, z]: 0.3, -20.7, 37; z score 3.47; cluster extent 70) and by 34.0% in the left dorsolateral funiculus ( $p = 0.023$ ; localization [x, y, z] -5, -19.2, 32; z score 3.22; cluster extent 69) in patients compared to healthy controls. MD was not significantly different between patients and healthy controls (figure 2).

### Relationship between lesion severity and remote tissue-specific neurodegeneration

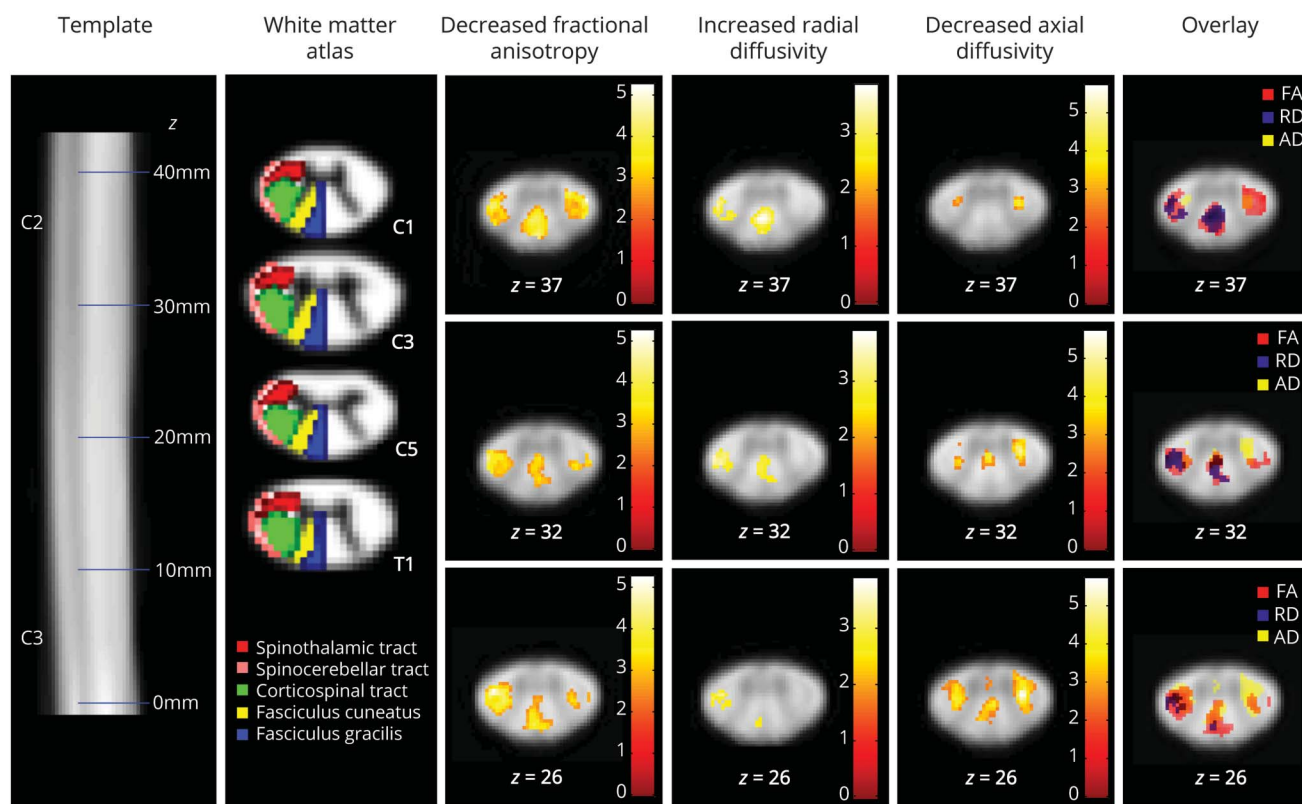
Greater lesion area and length were associated with greater SCA decrease above the level of injury (lesion area:  $p = 0.048$ ,  $R^2 = 0.25$ , 95% CI -3.64 to -0.23  $1/\text{mm}^3$ ; lesion length:  $p = 0.006$ ,  $R^2 = 0.42$ , 95% CI -0.55 to -0.11  $1/\text{mm}^2$ ) independently of age. The width of total midsagittal tissue bridges was associated with less SCA decrease ( $p = 0.007$ ,  $R^2 = 0.39$ , 95% CI 0.02–0.11  $1/\text{mm}^2$ ). Greater lesion length was associated with smaller GMA ( $p = 0.012$ ,  $R^2 = 0.40$ , 95% CI -3.74 to -0.57  $1/\text{mm}^2$ ), VHA ( $p = 0.039$ ,  $R^2 = 0.29$ , 95% CI -8.33 to -0.25  $1/\text{mm}^2$ ), and DHA ( $p = 0.004$ ,  $R^2 = 0.49$ , 95% CI -8.36 to -2.03  $1/\text{mm}^2$ ), while midsagittal tissue bridges were positively associated with GMA ( $p = 0.035$ ,  $R^2 = 0.28$ , 95% CI 0.33–0.781  $1/\text{mm}^2$ ) and DHA ( $p = 0.011$ ,  $R^2 = 0.38$ , 95% CI 0.26–1.74  $1/\text{mm}^2$ ) (figure 3A). Greater lesion length and preserved midsagittal tissue bridges were

**Table 2** Neurophysiologic data acquired in patients

|  | C4 Dermatome         |                       | C6 Dermatome         |                      | C8 Dermatome          |                     |
|--|----------------------|-----------------------|----------------------|----------------------|-----------------------|---------------------|
|  | Left                 | Right                 | Left                 | Right                | Left                  | Right               |
| <b>CHEPs</b>                             |                      |                       |                      |                      |                       |                     |
| <b>Heat perception threshold, °C</b>     | 43.97 ± 3.26 (15/15) | 44.94 ± 4.20 (13/14)  | 45.18 ± 5.69 (11/15) | 41.86 ± 4.36 (11/15) | 47.08 ± 3.82 (10/14)  | 45.27 ± 4.28 (8/14) |
| <b>Pain threshold, °C</b>                | 50.12 ± 2.58 (11/15) | 49.21 ± 3.53 (10/14)  | 49.56 ± 3.78 (6/15)  | 51.33 ± 2.75 (10/15) | 49.50 ± 3.60 (4/14)   | 51.51 ± 2.12 (7/14) |
| <b>Detectable signal</b>                 | 8/15                 | 8/14                  | 4/15                 | 7/15                 | 0/14                  | 5/14                |
| <b>Pathologic signal</b>                 | 3/8                  | 1/8                   | 1/4                  | 1/7                  | —                     | 0/5                 |
| <b>N2 latency, ms</b>                    | 409.12 ± 108.68      | 349.56 ± 78.45        | 357.38 ± 114.40      | 400.33 ± 63.15       | —                     | 352.88 ± 102.19     |
| <b>P2 latency, ms</b>                    | 506.63 ± 109.63      | 479.06 ± 90.75        | 461.38 ± 114.78      | 530.92 ± 45.90       | —                     | 466.13 ± 128.11     |
| <b>N2P2 amplitude, μV</b>                | 39.28 ± 34.59        | 40.57 ± 48.16         | 33.21 ± 29.96        | 25.72 ± 20.05        | —                     | 30.45 ± 26.18       |
| <b>SSEPs</b>                             |                      |                       |                      |                      |                       |                     |
| <b>Electric perception threshold, mA</b> | 4.26 ± 2.87 (11/12)  | 3.30 ± 1.94 (12/12)   | 6.49 ± 10.15 (13/14) | 8.18 ± 13.35 (12/14) | 7.28 ± 7.75 (12/14)   | 5.59 ± 4.52 (11/14) |
| <b>Pain threshold, mA</b>                | 24.1 ± 12.02 (11/12) | 26.24 ± 14.81 (12/12) | 17.72 ± 6.56 (12/14) | 17.63 ± 7.35 (11/14) | 19.06 ± 12.60 (11/14) | 15.7 ± 8.04 (10/14) |
| <b>Detectable signal</b>                 | 10/12                | 11/12                 | 11/14                | 9/14                 | 9/14                  | 9/14                |
| <b>Pathologic signal</b>                 | 1/10                 | 0/11                  | 1/11                 | 0/9                  | 2/9                   | 1/9                 |
| <b>N1 latency, ms</b>                    | 15.29 ± 3.07         | 15.78 ± 1.95          | 24.65 ± 2.23         | 23.92 ± 2.43         | 26.31 ± 3.20          | 26.08 ± 2.68        |
| <b>P1 latency, ms</b>                    | 21.57 ± 4.95         | 23.40 ± 3.10          | 29.71 ± 3.18         | 29.60 ± 2.69         | 31.07 ± 3.58          | 31.30 ± 2.87        |
| <b>N1P1 amplitude, μV</b>                | 1.04 ± 1.49          | 1.22 ± 1.64           | 1.06 ± 0.62          | 1.33 ± 0.75          | 0.81 ± 0.47           | 1.08 ± 0.57         |

Abbreviations: CHEP = contact heat evoked potential; SSEP = somatosensory evoked potential. Mean ± SD values are shown. Numbers in parentheses refer to the number of patients with detectable threshold/signals over the total number of measured patients.

**Figure 2** Microstructural changes above the level of injury



Voxel-wise analysis of microstructural changes above the level of injury in patients compared to healthy controls. First row shows the spinal cord template; second row shows the white matter atlas. Note the spatial overlap of the different diffusion tensor imaging metrics showing that regions of decreased axonal diffusivity (AD; e.g., axonal degeneration) but unaltered radial diffusivity (RD; e.g., no demyelination) lie mostly adjacent to the gray matter, where unmyelinated propriospinal neurons are located. FA = fractional anisotropy. Reprinted from De Leener et al<sup>22</sup> with permission from Elsevier.

associated with WMA (lesion length:  $p = 0.014$ ,  $R^2 = 0.38$ , 95% CI  $-0.61$  to  $-0.08$   $1/\text{mm}^2$ ; tissue bridges:  $p = 0.011$ ,  $R^2 = 0.38$ , 95% CI  $0.02$ – $0.12$   $1/\text{mm}^2$ ) above the level of injury (figure 3B).

The width of total midsagittal tissue bridges was associated with DCA above the level of lesion ( $p = 0.019$ ,  $R^2 = 0.29$ , 95% CI  $0.25$ – $2.39$   $1/\text{mm}^2$ ). Neither lesion size nor midsagittal tissue bridges were associated with microstructural changes above the level of lesion.

### Relationship between remote neurodegeneration and neurophysiologic outcome

The size of the cross-sectional area of the dorsal columns identified those patients with bilateral recordable SSEPs of the dermatomes C6 and C8 (figure 4). This relationship was not evident for the WMA and CHEPs. Higher AD values within the dorsal column were associated with shorter SSEP N1P1 latency at the C4 dermatome ( $p = 0.0024$ ,  $R^2 = 0.83$ , 95% CI:  $-0.00007$  to  $-0.00001$   $10^{-3} \times \text{s}^2/\text{mm}^2$ ), corrected for age and lesion area. DTI metrics within the spinothalamic tracts were not associated with CHEPs recordings.

### Relationship between remote neurodegeneration and clinical outcome

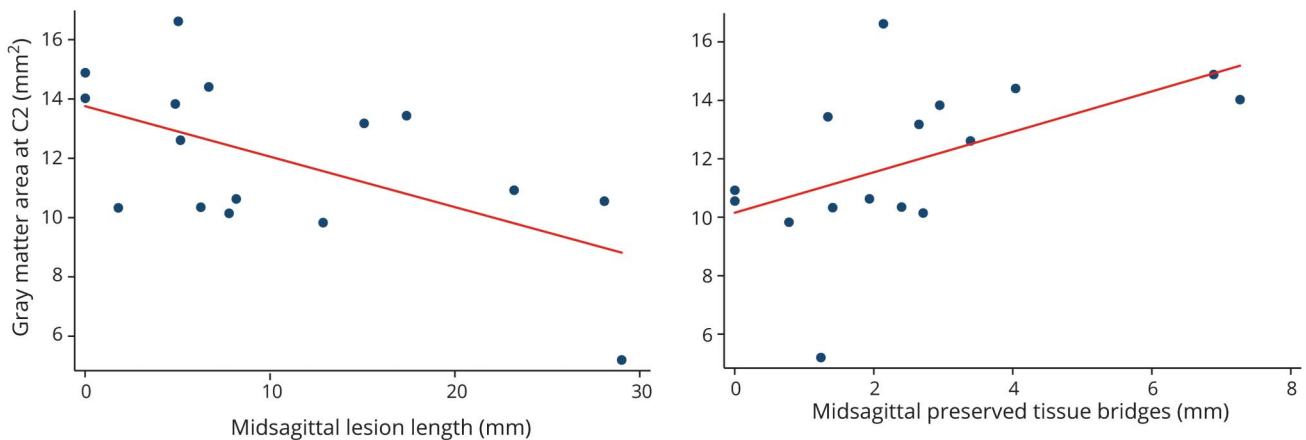
GMA was associated with motor score ( $p = 0.007$ ,  $R^2 = 0.72$ , 95% CI  $1.77$ – $9.26$ ) and pinprick score ( $p = 0.003$ ,  $R^2 = 0.58$ , 95% CI  $3.48$ – $13.90$ ); VHA area was associated with motor score ( $p = 0.001$ ,  $R^2 = 0.78$ , 95% CI  $6.74$ – $21.93$ ); and DHA was associated with pinprick score ( $p = 0.004$ ,  $R^2 = 0.57$ , 95% CI  $7.43$ – $31.52$ ) when corrected for lesion area and age (figure 5A).

To quantify tract-specific associations with appropriate clinical outcome, we used the extracted mean values of DTI indexes within the regions of interest (i.e., corticospinal tract, dorsal column, and spinothalamic tract). FA and RD within corticospinal tract and the dorsal columns were associated with SCIM score (corticospinal tract: FA:  $p = 0.002$ ,  $R^2 = 0.80$ , 95% CI  $105.82$ – $361.55$ ; RD:  $p = 0.001$ ,  $R^2 = 0.83$ , 95% CI  $-169829.70$  to  $-60547.48$ ; dorsal columns: FA:  $p = 0.002$ ,  $R^2 = 0.80$ , 95% CI  $106.07$ – $341.42$ ; RD:  $p = 0.003$ ,  $R^2 = 0.79$ , 95% CI  $-216944.1$  to  $-60854.36$ ) independently of lesion extent and age (figure 5B). AD within the dorsal columns was associated with GRASSP score independently of lesion extent and age ( $p = 0.30$ ,  $R^2 = 0.60$ , 95% CI  $40298.4$ – $646999.7$ ).

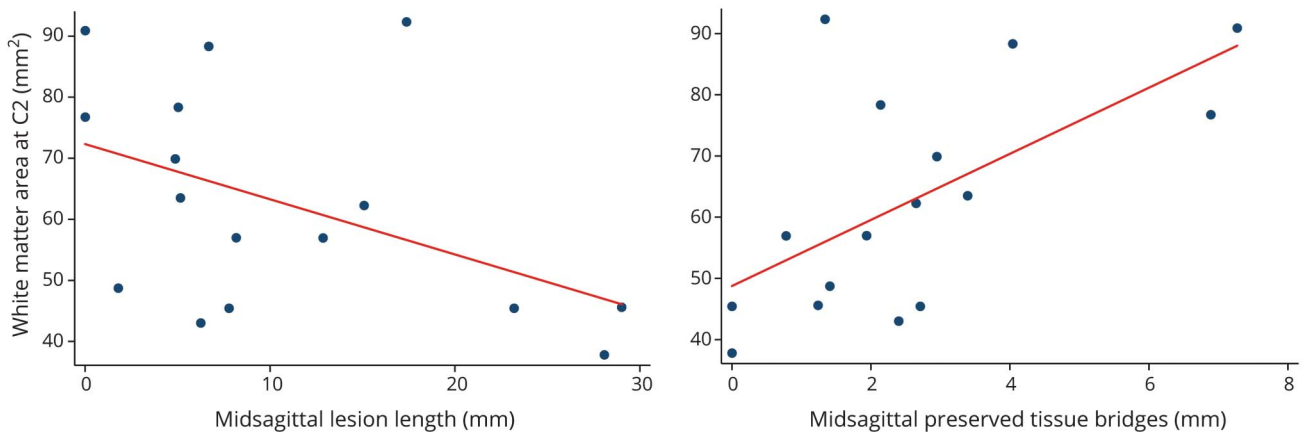


**Figure 3** Relationship between lesion severity and neurodegeneration above the level of injury

**A.** Relationship between lesion parameters and remote grey matter changes



**B.** Relationship between lesion parameters and remote white matter changes



The magnitude of tissue damage at the lesion site is associated with the amount of neurodegeneration above the level of injury. Lesion length and midsagittal tissue bridges are associated with (A) remote gray matter and (B) white matter atrophy.

## Discussion

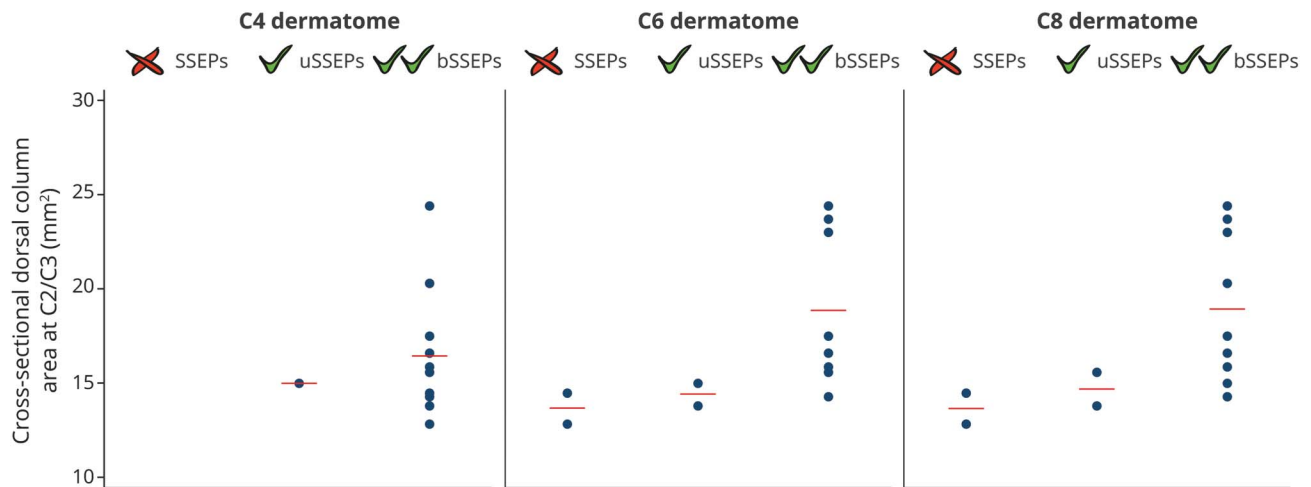
This study shows the *in vivo* structure-function relationship between the extent of tissue-specific cord pathology and neurophysiologic and clinical impairment after traumatic SCI. Crucially, we show that the magnitude of tissue damage at the lesion epicenter is associated with the extent of neurodegeneration above the level of lesion, which, in turn, is associated with clinically relevant impairment and neurophysiologic abnormalities. These findings allow us to investigate the extent of tissue-specific neurodegeneration above the level of injury, its relationship to neuronal tissue loss at the site of the lesion, and its effect on neurophysiologic and clinical outcome.

Tissue damage at the epicenter of a traumatic SCI results both from the direct effect of the traumatic insult and from damage to the vascular architecture and the ensuing ischemic effects on the neuronal and glial cell populations within the acute

phase of injury.<sup>23</sup> Remote from the epicenter of the lesion, secondary neurodegeneration within white<sup>24,25</sup> and gray matter<sup>26</sup> follows with a time lag and is driven by a multiphasic response to cellular inflammation.<sup>27</sup> While the extent of secondary remote atrophy has been quantified *in vivo* after injury,<sup>2,28-30</sup> we provide evidence that changes within both gray and white matter contribute to cord atrophy above the level of injury. This is in agreement with spinal gray matter degeneration distant to the initial site of damage in patients with multiple sclerosis<sup>31</sup> and experimental SCI.<sup>32</sup> Although the relative decrease is larger within ventral and dorsal horns (i.e., gray matter), the absolute magnitude of change is larger within white matter, contributing more to the overall loss of SCA by 20.2%.

We uncovered an *in vivo* relationship between neuronal tissue loss (i.e., lesion severity) and remote tissue-specific cord pathology above the level of injury. Moreover, we show an interdependence of remote white and gray matter atrophy

**Figure 4** Relationship between neurodegeneration above the level of injury and electrophysiologic outcome



Patients were grouped into 3 cohorts: without or with unilateral (u) or bilateral (b) recordable dermatomal somatosensory-evoked potentials (SSEPs) at the C4, C6, and C8 level. Patients with recordable dermatomal SSEPs showed a tendency toward larger dorsal column area above the level of injury.

(i.e., DHA and DCA). Neurodegenerative changes within gray matter above the level of injury are not likely to be specific for any single pathologic process but rather are likely to represent a combination of different pathologic mechanisms taking place after SCI. Possible mechanisms involve transsynaptic/transneuronal degeneration affecting the propriospinal systems<sup>33,34</sup> and motor neurons located in the proximity of the spinal injury.<sup>35</sup> Next to direct effects of neurodegenerative processes, a reduction in muscle activity of the upper extremity could lead to a reduction of neuronal activity above the level of injury, which may translate into shrinkage of the neuron soma size.

Furthermore, demyelination of corticospinal projections to the dorsal horns,<sup>36</sup> the expression of neurotrophic factors from nonneuronal cells around neighboring degenerating axons,<sup>36</sup> growth factor dysregulation,<sup>37</sup> and vascular remodeling<sup>38</sup> could contribute to gray matter pathology. As white matter damage is known to induce microglial activation altering glutamate signaling, this process is thought to be responsible for the dying back of axons and their parternal neurons,<sup>39</sup> which might be a shared underlying disease mechanism. Thus, next to anterograde and retrograde degeneration of white matter tracts<sup>30,40,41</sup> the neuronal circuits within the spinal cord far above the level of injury undergo a temporary structured neurodegeneration.<sup>25</sup>

Within the microstructure of the atrophied white matter, we found indications of both axonal degeneration and demyelination,<sup>42</sup> represented by decreased FA and AD and increased RD in the dorsolateral funiculus (e.g., containing the lateral corticospinal and spinothalamic tracts) and posterior funiculus (e.g., containing the dorsal columns). Within the corticospinal tract and the dorsal column, leg function is represented most laterally, whereas arm function is located

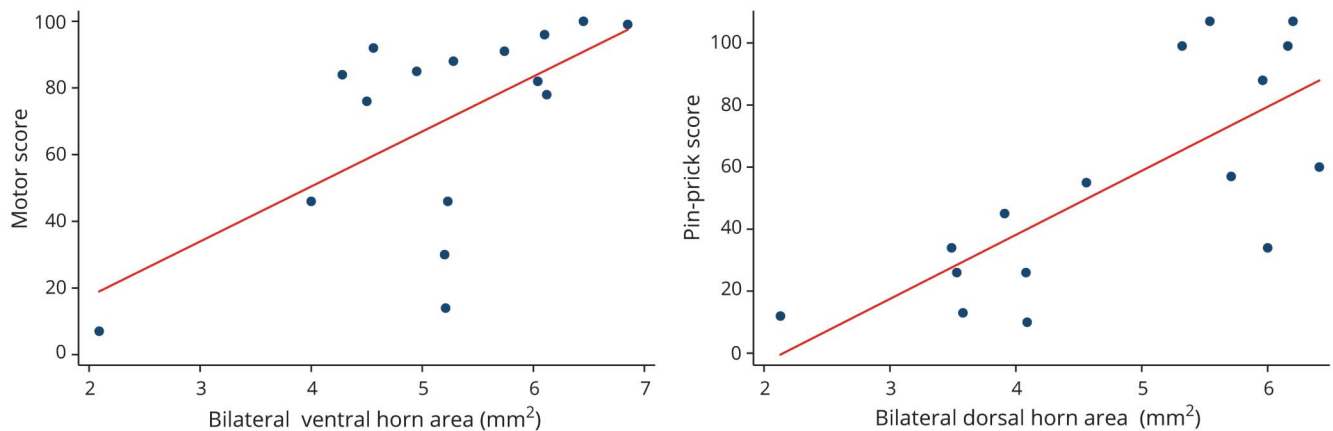
either medially (i.e., corticospinal tract) or centrally (i.e., dorsal column). Our observed changes cover the entire lateral corticospinal tract and the dorsal columns, indicating neurodegenerative processes affecting axons that convey information relating to leg and arm function. Spatially overlaying the different DTI metrics changes revealed that regions showing decreased AD (e.g., axonal degeneration) but unaltered RD (e.g., no demyelination) lie mostly adjacent to the gray matter border. This region contains the fasciculi proprii and contains mostly short, mainly unmyelinated propriospinal neurons.<sup>43</sup> This underlies our hypothesis that SCI might lead to degeneration affecting interneurons within the spinal cord.

Our findings complement previous studies in patients with SCI<sup>20,44–46</sup> in that they now locate these changes to gray and white matter rather than being nonspecific in terms of location and tissue. Thus, our in vivo cord MRI measurements demonstrate a combination of structural and functional processes occurring over several segments above the level of injury affecting both gray and white matter that is driven by lesion severity.

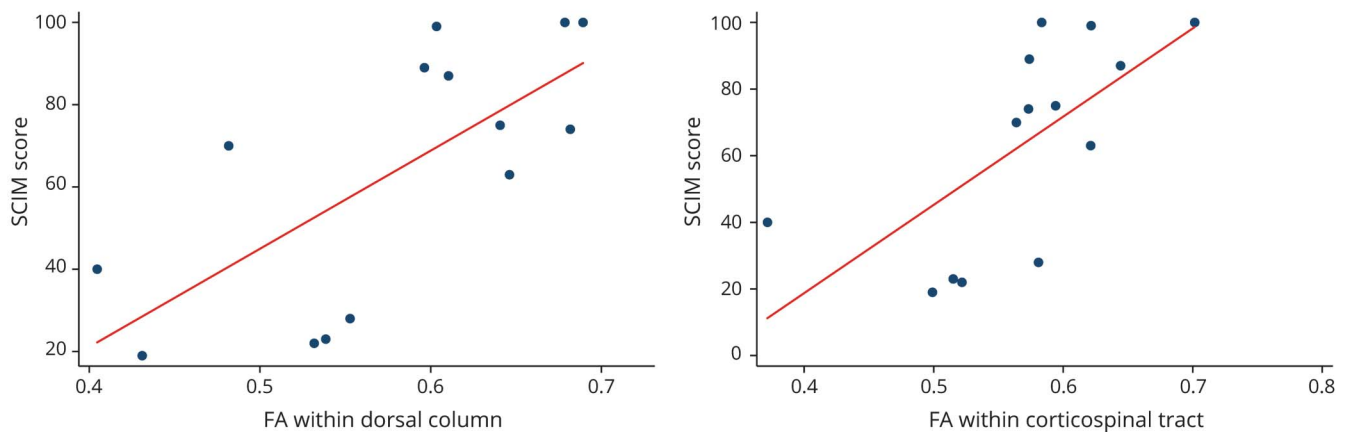
We show that tract-specific microstructural and macrostructural changes are associated with prolonged conduction of appropriate electrophysiologic recordings. This association suggests a structure-function relationship because the amount of neurodegeneration was directly associated with impairment of neurophysiologic information flow. Because the DTI signal is sensitive to altered diffusion properties occurring as a response to CNS damage (i.e., demyelination/degeneration) and because neuronal excitability is affected by morphometry of the axon and its myelin, it seems plausible that neurophysiologic changes might be reflected in remote macrostructural and

**Figure 5** Relationship between neurodegeneration above the level of injury and clinical outcome

**A.** Relationship between remote macrostructural neurodegeneration and clinical impairment



**B.** Relationship between remote microstructural neurodegeneration and clinical impairment



Associations between (A) remote tract-specific macrostructural MRI parameters above the level of injury and clinical impairment and (B) remote tract-specific microstructural MRI indexes above the level of injury and clinical impairment. FA = fractional anisotropy; SCIM = Spinal Cord Independence Measure.

microstructural changes above the level of injury. Our findings complement previous findings showing that the topography and the excitability of corticomotor projections were associated with cervical cord atrophy.<sup>2,47</sup>

Current assessments in patients with SCI lack sensitivity to minimal changes in motor and sensory function<sup>48</sup> in that they cannot detect subtle changes due to remyelination and axonal regeneration. Neuroimaging biomarkers have the potential to track these subtle abnormalities because they are sensitive to microstructural changes.<sup>1</sup> In this study, the magnitude of both remote macrostructural and microstructural changes within gray and white matter was significantly associated with clinical impairment, independently of lesion extent. In particular, the extent of remote ventral horn atrophy was associated with motor impairment, whereas dorsal horn atrophy was associated with sensory disturbance. Microstructural tract-specific changes above

the level of injury were related to measures of functional independence (i.e., SCIM) and strength, sensibility, and prehension of the upper limbs (i.e., GRASSP). This suggests that high-resolution MRI sequences applied above the level of injury provide superior information on the patient's clinical status compared to standard clinical sequences at the lesion site. In addition, the latter findings are striking in that they suggest that remote neurodegeneration within gray matter above the level of injury contributes, in addition to white matter pathology, to motor and sensory impairment. This multilevel interaction supports the view that SCI leads to a cascade of neurodegenerative changes affecting the entire spinal cord and brain.<sup>49</sup> Characterizing these secondary neurodegenerative events has the potential to provide insights into new therapeutic interventions, in addition to providing opportunities for monitoring treatment effects in trials conducted in patients with acute and chronic SCI.

This study had several limitations. Although our cohorts did not show a significant age difference, the mean age was on average higher in the patient group, which could potentially affect the analysis. We therefore adjusted for age as a potential confounder of no interest in all analyses. Furthermore, unbiased voxel-based morphometry of DTI indexes in the spinal cord has just started emerging,<sup>3</sup> and the automated post-processing methods for spatial normalization of the spinal cord into common space are in their infancy. To increase the reliability of our analysis, we therefore manually corrected the spatial normalization to the template.

This study shows that the magnitude of dorsal and ventral horn and white matter structural changes above the level of injury is associated with appropriate clinical and neurophysiologic impairment and is driven by lesion severity. These findings suggest a combination of different pathologic processes affecting both gray and white matter several segments above the level of injury that are clinically eloquent. Therefore, these neuroimaging biomarkers might serve as promising surrogate markers for future clinical trials supplementing (or complementing) clinical outcome measures.

### Author contributions

Eveline Huber: study concept and design; acquisition, analysis and interpretation of data, statistical analysis, writing the manuscript. Gergely David: analysis of data, critical revision of manuscript for intellectual content. Alan Thompson: design of study, critical revision of manuscript for intellectual content. Nikolas Weiskopf and Siawoosh Mohammadi: study concept and design, critical revision of manuscript for intellectual content. Patrick Freund: study concept and design, interpretation of data, writing the manuscript, study supervision.

### Acknowledgment

The authors thank all the participants for taking part in this study, as well as the staff of the Department of Radiology and Neurology at the University Hospital Balgrist. They also thank Dr. Patrick Grabher for his help with recruiting the healthy controls.

### Study funding

Funded by Wings for Life, Austria (WFL-CH-007/14), the International Foundation for Research in Paraplegia (IRP-P158), European Union's project (Horizon 2020 "NISCI" grant agreement 681094), Clinical Research Priority Program Neurorehab UZH, and the European Research Council (ERC grant agreement n616905). The Wellcome Trust Centre for Neuroimaging is supported by core funding from the Wellcome Trust 0915/Z/10/Z. S.M. received funding from the European Union's Horizon 2020 research and innovation program under the Marie Skłodowska-Curie grant agreement 658589 and was supported by the Deutsche Forschungsgemeinschaft, grant MO 2397/4-1. N.W., P.F., and S.M. received funding from the BMBF (01 EW1711A & B) in the framework of ERA-NET NEURON. A.J.T. acknowledges support from the University College London/University

College London Hospitals National Institute of Health Biomedical Research Centre.

### Disclosure

E. Huber and G. David report no disclosures relevant to the manuscript. A. Thompson has received honoraria and support for travel for consultancy from Biogen Idec, MedDay, Eisai, and Novartis and for teaching from Teva, Novartis, and EXCEMED. He receives an honorarium as editor-in-chief of *Multiple Sclerosis Journal*. N. Weiskopf reports that the Wellcome Trust Centre for Neuroimaging has an institutional research agreement with and receives support from Siemens Healthcare. S. Mohammadi and P. Freund report no disclosures relevant to the manuscript. Go to [Neurology.org/N](http://Neurology.org/N) for full disclosures.

Received November 20, 2017. Accepted in final form January 24, 2018.

### References

1. Huber E, Curt A, Freund P. Tracking trauma-induced structural and functional changes above the level of spinal cord injury. *Curr Opin Neurol* 2015;28:365–372.
2. Freund P, Weiskopf N, Ward NS, et al. Disability, atrophy and cortical reorganization following spinal cord injury. *Brain* 2011;134:1610–1622.
3. Martin AR, Aleksanderek I, Cohen-Adad J, et al. Translating state-of-the-art spinal cord MRI techniques to clinical use: a systematic review of clinical studies utilizing DTI, MT, MWF, MRS, and fMRI. *NeuroImage Clin* 2016;10:192–238.
4. Mohammadi S, Hutton C, Nagy Z, Josephs O, Weiskopf N. Retrospective correction of physiological noise in DTI using an extended tensor model and peripheral measurements. *Magn Reson Med* 2013;70:358–369.
5. David G, Freund P, Mohammadi S. The efficiency of retrospective artifact correction methods in improving the statistical power of between-group differences in spinal cord DTI. *Neuroimage* 2017;158:296–307.
6. Grabher P, Mohammadi S, David G, Freund P. Neurodegeneration in the spinal ventral horn prior to motor impairment in cervical spondylotic myelopathy. *J Neurotrauma* 2017;34:2329–2334.
7. Huber E, Lachappelle P, Sutter R, Curt A, Freund P. Are midsagittal tissue bridges predictive of outcome after cervical spinal cord injury? *Ann Neurol* 2017;81:740–748.
8. Grabher P, Mohammadi S, Trachsler A, et al. Voxel-based analysis of grey and white matter degeneration in cervical spondylotic myelopathy. *Sci Rep* 2016;6:24636.
9. Kirshblum SC, Burns SP, Biering-Sorensen F, et al. International Standards for Neurological Classification of Spinal Cord Injury (revised 2011). *J Spinal Cord Med* 2011;34:535–546.
10. Itzkovich M, Gelernter I, Biering-Sorensen F, et al. The Spinal Cord Independence Measure (SCIM) version III: reliability and validity in a multi-center international study. *Disabil Rehabil* 2007;29:1926–1933.
11. Kalsi-Ryan S, Beaton D, Curt A, et al. The Graded Redefined Assessment of Strength, Sensibility and Prehension: reliability and validity. *J Neurotrauma* 2012;29:905–914.
12. Dittuno PL, Ditunno JF Jr. Walking Index for Spinal Cord Injury (WISCI II): scale revision. *Spinal Cord* 2001;39:654–656.
13. Kramer JL, Haefeli J, Jutzeler CR, Steeves JD, Curt A. Improving the acquisition of nociceptive evoked potentials without causing more pain. *Pain* 2013;154:235–241.
14. Kramer JL, Moss AJ, Taylor P, Curt A. Assessment of posterior spinal cord function with electrical perception threshold in spinal cord injury. *J Neurotrauma* 2008;25:1019–1026.
15. Schmid MR, Pfirrmann CW, Koch P, Zanetti M, Kuehn B, Hodler J. Imaging of patellar cartilage with a 2D multiple-echo data image combination sequence. *AJR Am J Roentgenol* 2005;184:1744–1748.
16. Morelli JN, Runge VM, Feiweier T, Kirsch JE, Williams KW, Attenberger UI. Evaluation of a modified Stejskal-Tanner diffusion encoding scheme, permitting a marked reduction in TE, in diffusion-weighted imaging of stroke patients at 3 T. *Invest Radiol* 2010;45:29–35.
17. Ashburner J, Ridgway GR. Symmetric diffeomorphic modeling of longitudinal structural MRI. *Front Neurosci* 2012;6:197.
18. Horsfield MA, Sala S, Neema M, et al. Rapid semi-automatic segmentation of the spinal cord from magnetic resonance images: application in multiple sclerosis. *Neuroimage* 2010;50:446–455.
19. Mohammadi S, Moller HE, Kugel H, Muller DK, Deppe M. Correcting eddy current and motion effects by affine whole-brain registrations: evaluation of three-dimensional distortions and comparison with slice-wise correction. *Magn Reson Med* 2010;64:1047–1056.
20. Kamble RB, Venkataramana NK, Naik AL, Rao SV. Diffusion tensor imaging in spinal cord injury. *Indian J Radiol Imaging* 2011;21:221–224.
21. Fonov VS, Le Troter A, Taso M, et al. Framework for integrated MRI average of the spinal cord white and gray matter: the MNI-Poly-AMU template. *Neuroimage* 2014;102:817–827.

22. De Leener B, Levy S, Dupont SM, et al. SCT: Spinal Cord Toolbox, an open-source software for processing spinal cord MRI data. *Neuroimage* 2017;145:24–43.
23. Schwab ME, Bartholdi D. Degeneration and regeneration of axons in the lesioned spinal cord. *Physiol Rev* 1996;76:319–370.
24. Crowe MJ, Bresnahan JC, Shuman SL, Masters JN, Beattie MS. Apoptosis and delayed degeneration after spinal cord injury in rats and monkeys. *Nat Med* 1997;3:73–76.
25. Ward RE, Huang W, Kostusiak M, Pallier PN, Michael-Titus AT, Priestley JV. A characterization of white matter pathology following spinal cord compression injury in the rat. *Neuroscience* 2014;260:227–239.
26. Huang WL, George KJ, Ibba V, et al. The characteristics of neuronal injury in a static compression model of spinal cord injury in adult rats. *Eur J Neurosci* 2007;25:362–372.
27. Beck KD, Nguyen HX, Galvan MD, Salazar DL, Woodruff TM, Anderson AJ. Quantitative analysis of cellular inflammation after traumatic spinal cord injury: evidence for a multiphasic inflammatory response in the acute to chronic environment. *Brain* 2010;133:433–447.
28. Lundell H, Barthelemy D, Skimminge A, Dyrby TB, Biering-Sorensen F, Nielsen JB. Independent spinal cord atrophy measures correlate to motor and sensory deficits in individuals with spinal cord injury. *Spinal Cord* 2011;49:70–75.
29. Freund P, Schneider T, Nagy Z, et al. Degeneration of the injured cervical cord is associated with remote changes in corticospinal tract integrity and upper limb impairment. *PLoS One* 2012;7:e51729.
30. Freund P, Wheeler-Kingshott CA, Nagy Z, et al. Axonal integrity predicts cortical reorganisation following cervical injury. *J Neurol Neurosurg Psychiatry* 2012;83:629–637.
31. Calabrese M, Magliozzi R, Ciccarelli O, Geurts JJ, Reynolds R, Martin R. Exploring the origins of grey matter damage in multiple sclerosis. *Nat Rev Neurosci* 2015;16:147–158.
32. Pearse DD, Lo TP Jr, Cho KS, et al. Histopathological and behavioral characterization of a novel cervical spinal cord displacement contusion injury in the rat. *J Neurotrauma* 2005;22:680–702.
33. Yu H, Li L, Liu R, et al. Autophagy in long propriospinal neurons is activated after spinal cord injury in adult rats. *Neurosci Lett* 2016;634:138–145.
34. Grumbles RM, Thomas CK. Motoneuron death after human spinal cord injury. *J Neurotrauma* 2017;34:581–590.
35. Filli L, Schwab ME. Structural and functional reorganization of propriospinal connections promotes functional recovery after spinal cord injury. *Neural Regen Res* 2015;10:509–513.
36. Lemon RN, Griffiths J. Comparing the function of the corticospinal system in different species: organizational differences for motor specialization? *Muscle Nerve* 2005;32:261–279.
37. Bareyre FM, Schwab ME. Inflammation, degeneration and regeneration in the injured spinal cord: insights from DNA microarrays. *Trends Neurosci* 2003;26:S55–S63.
38. Cao Y, Zhou Y, Ni S, et al. Three dimensional quantification of microarchitecture and vessel regeneration by synchrotron radiation microcomputed tomography in a rat model of spinal cord injury. *J Neurotrauma* 2017;34:1187–1199.
39. Matute C, Alberdi E, Domercq M, et al. Excitotoxic damage to white matter. *J Anat* 2007;210:693–702.
40. Buss A, Pech K, Merkler D, et al. Sequential loss of myelin proteins during Wallerian degeneration in the human spinal cord. *Brain* 2005;128:356–364.
41. Kerschensteiner M, Schwab ME, Lichtman JW, Misgeld T. In vivo imaging of axonal degeneration and regeneration in the injured spinal cord. *Nat Med* 2005;11:572–577.
42. Brennan FH, Cowin GJ, Kurniawan ND, Ruitenber MJ. Longitudinal assessment of white matter pathology in the injured mouse spinal cord through ultra-high field (16.4 T) in vivo diffusion tensor imaging. *Neuroimage* 2013;82:574–585.
43. Flynn JR, Graham BA, Galea MP, Callister RJ. The role of propriospinal interneurons in recovery from spinal cord injury. *Neuropharmacology* 2011;60:809–822.
44. Cohen-Adad J, El Mendili MM, Lehericy S, et al. Demyelination and degeneration in the injured human spinal cord detected with diffusion and magnetization transfer MRI. *Neuroimage* 2011;55:1024–1033.
45. Petersen JA, Wilm BJ, von Meyenburg J, et al. Chronic cervical spinal cord injury: DTI correlates with clinical and electrophysiological measures. *J Neurotrauma* 2012;29:1556–1566.
46. Koskinen E, Brander A, Hakulinen U, et al. Assessing the state of chronic spinal cord injury using diffusion tensor imaging. *J Neurotrauma* 2013;30:1587–1595.
47. Freund P, Rothwell J, Craggs M, Thompson AJ, Bestmann S. Corticomotor representation to a human forearm muscle changes following cervical spinal cord injury. *Eur J Neurosci* 2011;34:1839–1846.
48. Ellaway PH, Kuppaswamy A, Balasubramaniam AV, et al. Development of quantitative and sensitive assessments of physiological and functional outcome during recovery from spinal cord injury: a clinical initiative. *Brain Res Bull* 2011;84:343–345.
49. Freund P, Friston K, Thompson AJ, et al. Embodied neurology: an integrative framework for neurological disorders. *Brain* 2016;139:1855–1861.

# Dorsal and ventral horn atrophy is associated with clinical outcome after spinal cord injury

Eveline Huber, MSc, Gergely David, MSc, Alan J. Thompson, MD, Nikolaus Weiskopf, PhD, Siawoosh Mohammadi, PhD, and Patrick Freund, MD, PhD

## Correspondence

Dr. Freund  
patrick.freund@balgrist.ch

Cite as: *Neurology*® 2018;90:e1510-e1522. doi:10.1212/WNL.0000000000005361

## Study question

Does grey matter pathology above the level of injury contribute to functional/sensorimotor impairments in patients with spinal cord injury (SCI)?

## Summary answer

The magnitude of tissue damage to both grey and white matter at the lesion epicenter is associated with the extent of neurodegeneration above the level of the lesion, which is in turn associated with clinical impairments and neuropsychologic abnormalities, in SCI.

## What is known and what this paper adds

White matter pathology within the spinal cord contributes to sensorimotor impairments. This study provides evidence that changes within grey matter—driven by the severity of the lesion—also contribute to atrophy above the level of injury and relate to clinical impairment, suggesting that neuroimaging biomarkers can be used to supplement clinical outcome measures in future clinical trials.

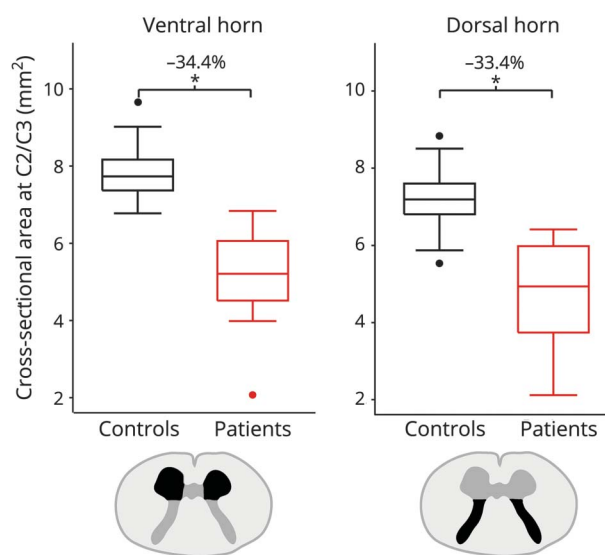
## Participants and setting

The study included 17 patients with SCI (mean age: 48.7 ± 14.1 years) who had been admitted to University Hospital Balgrist (Zurich, Switzerland) and 21 healthy controls (mean age: 41.7 ± 11.3 years) recruited from among the local population. Inclusion criteria for patients with SCI were as follows: traumatic cervical SCI, absence of other neurologic or psychologic disorders, age 18–70 years, absence of contraindications for MRI, and absence of pregnancy.

## Design, size, and duration

All patients underwent clinical assessments for functional/neurologic impairment (e.g., Spinal Cord Independence Measure [SCIM], the International Standards for Neurological Classification of Spinal Cord Injury [ISNCSCI], Graded Redefined Assessment of Strength, Sensibility, and Prehension [GRASSP], as well as neurophysiologic assessments such as Contact heat-evoked potentials [CHEPs] and somatosensory-evoked potentials [SSEPs]). Contact heat-evoked potentials (CHEPs), somatosensory-evoked potentials (SSEPs), and imaging findings were examined an average of 6.7 ± 7.8 years after injury.

A draft of the short-form article was written by D. Drobish, a writer with Editage, a division of Cactus Communications. The authors of the full-length article and the journal editors edited and approved the final version.



## Main results and the role of chance

Greater lesion area and length were associated with greater decreases in spinal cord area above the level of injury ( $p = 0.048$  and  $p = 0.006$ , respectively). Tissue-specific neurodegeneration was associated with electrophysiologic abnormalities. Dorsal and ventral horn atrophy were associated with sensory and motor outcomes, respectively. White matter integrity was associated with functional independence.

## Bias, confounding, and other reasons for caution

Mean age was higher in the patient group, and manual correction was performed for spatial normalization.

## Generalizability to other populations

The results may be generalizable to nontraumatic SCI forms.

## Study funding/potential competing interests

This study was supported by Wings for Life, (Austria), the International Foundation for Research in Paraplegia, the EU Project, the Clinical Research Priority Program, the European Research Council, and the Wellcome Trust Centre for Neuroimaging, the Horizon 2020 Research and Innovation Programme, the BMBF, and the UCL/UCLH National Institute of Health Biomedical Research Centre. Go to [Neurology.org/N](http://Neurology.org/N) for full disclosures.

# Neurology®

## Dorsal and ventral horn atrophy is associated with clinical outcome after spinal cord injury

Eveline Huber, Gergely David, Alan J. Thompson, et al.

*Neurology* 2018;90:e1510-e1522 Published Online before print March 28, 2018

DOI 10.1212/WNL.0000000000005361

**This information is current as of March 28, 2018**

|   |   |
|---|---|
| <b>Updated Information &amp; Services</b> | including high resolution figures, can be found at:<br><a href="http://n.neurology.org/content/90/17/e1510.full">http://n.neurology.org/content/90/17/e1510.full</a>  |
| <b>References</b>                         | This article cites 49 articles, 1 of which you can access for free at:<br><a href="http://n.neurology.org/content/90/17/e1510.full#ref-list-1">http://n.neurology.org/content/90/17/e1510.full#ref-list-1</a>   |
| <b>Subspecialty Collections</b>           | This article, along with others on similar topics, appears in the following collection(s):<br><b>DWI</b><br><a href="http://n.neurology.org/cgi/collection/dwi">http://n.neurology.org/cgi/collection/dwi</a><br><b>Evoked Potentials/Somatosensory</b><br><a href="http://n.neurology.org/cgi/collection/evoked_potentials-somatosensory">http://n.neurology.org/cgi/collection/evoked_potentials-somatosensory</a><br><b>Plasticity</b><br><a href="http://n.neurology.org/cgi/collection/plasticity">http://n.neurology.org/cgi/collection/plasticity</a><br><b>Spinal cord trauma; see Trauma/spinal cord trauma</b><br><a href="http://n.neurology.org/cgi/collection/spinal_cord_trauma-see_trauma-s">http://n.neurology.org/cgi/collection/spinal_cord_trauma-see_trauma-s</a><br><a href="http://n.neurology.org/cgi/collection/spinal_cord_trauma">pinal_cord_trauma</a><br><b>Volumetric MRI</b><br><a href="http://n.neurology.org/cgi/collection/volumetric_mri">http://n.neurology.org/cgi/collection/volumetric_mri</a> |
| <b>Permissions &amp; Licensing</b>        | Information about reproducing this article in parts (figures, tables) or in its entirety can be found online at:<br><a href="http://www.neurology.org/about/about_the_journal#permissions">http://www.neurology.org/about/about_the_journal#permissions</a>   |
| <b>Reprints</b>                           | Information about ordering reprints can be found online:<br><a href="http://n.neurology.org/subscribers/advertise">http://n.neurology.org/subscribers/advertise</a>   |

*Neurology*® is the official journal of the American Academy of Neurology. Published continuously since 1951, it is now a weekly with 48 issues per year. Copyright © 2018 The Author(s). Published by Wolters Kluwer Health, Inc. on behalf of the American Academy of Neurology. All rights reserved. Print ISSN: 0028-3878. Online ISSN: 1526-632X.

