

AT A GLANCE

Molecular biomarkers of Alzheimer's disease: progress and prospects

Tammarny Lashley¹, Jonathan M. Schott², Philip Weston², Christina E. Murray¹, Henny Wellington^{3,4}, Ashvini Keshavan², Sandrine C. Foti¹, Martha Foiani^{3,4}, Jamie Toombs^{3,4}, Jonathan D. Rohrer², Amanda Heslegrave^{3,4} and Henrik Zetterberg^{3,4,5,6,*}

ABSTRACT

The neurodegenerative disorder Alzheimer's disease is characterised by the formation of β -amyloid plaques and neurofibrillary tangles in the brain parenchyma, which cause synapse and neuronal loss. This leads

to clinical symptoms, such as progressive memory deficits. Clinically, these pathological changes can be detected in the cerebrospinal fluid and with brain imaging, although reliable blood tests for plaque and tangle pathologies remain to be developed. Plaques and tangles often co-exist with other brain pathologies, including aggregates of transactive response DNA-binding protein 43 and Lewy bodies, but the extent to which these contribute to the severity of Alzheimer's disease is currently unknown. In this 'At a glance' article and poster, we summarise the molecular biomarkers that are being developed to detect Alzheimer's disease and its related pathologies. We also highlight the biomarkers that are currently in clinical use and include a critical appraisal of the challenges associated with applying these biomarkers for diagnostic and prognostic purposes of Alzheimer's disease and related neurodegenerative disorders, also in their prodromal clinical phases.

¹Queen Square Brain Bank for Neurological Disorders, Department of Molecular Neuroscience, UCL Institute of Neurology, London WC1N 3BG, UK. ²Dementia Research Centre, UCL Institute of Neurology, London WC1N 3BG, UK. ³Department of Molecular Neuroscience, UCL Institute of Neurology, Queen Square, London WC1N 3BG, UK. ⁴UK Dementia Research Institute, London WC1N 3BG, UK. ⁵Institute of Neuroscience and Physiology, Department of Psychiatry and Neurochemistry, the Sahlgrenska Academy at the University of Gothenburg, Mölndal S-431 80, Sweden. ⁶Clinical Neurochemistry Laboratory, Sahlgrenska University Hospital, Mölndal S-431 80, Sweden.

*Author for correspondence (henrik.zetterberg@gu.se)

id H.Z., 0000-0002-4671-6763

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution and reproduction in any medium provided that the original work is properly attributed.

KEY WORDS: Alzheimer's disease, Biomarkers, Cerebrospinal fluid, Blood, Plasma, Serum, Tau, Amyloid, Neurofilament, Neurogranin

Molecular biomarkers of Alzheimer's disease

Tammarny Lashley, Jonathan M. Schott, Philip Weston, Christina E. Murray, Henny Wellington, Ashvini Keshavan, Sandrine C. Foti, Martha Foiani, Jamie Toombs, Jonathan D. Rohrer, Amanda Heslegrave, Henrik Zetterberg

Biomarkers

What is a biomarker?

A characteristic that can be objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention.

What can currently be used as a biomarker in AD?

- Brain scans to investigate what is happening to the brain whilst the patient is alive
- Blood tests to quantify any disease-related marker
- Cerebrospinal fluid to quantify any disease-related marker
- Pathology is used in research only; brain tissue is analysed to identify disease-related markers that can be quantified in blood and CSF

Brain scans

Molecular biomarkers

Neuropathology of Alzheimer's disease

Normal control

Alzheimer's disease

Brain slices show brain shrinkage in Alzheimer's disease. The lateral ventricle is enlarged (asterisk) and the hippocampus, the brain area controlling memory, has shrunk (arrow).

Primary pathologies

The primary pathological hallmarks of Alzheimer's disease are A β plaques that form around the cells. The A β deposits can also be found in the blood vessels in the brain (asterisk), causing cerebral amyloid angiopathy. Neurofibrillary tangles form when tau becomes hyperphosphorylated and forms filaments within the neurons in the brain. As well as these two proteins being deposited in the brain, there is also a neuroinflammatory reaction in which the microglia become activated in response to the pathology.

Secondary pathologies

Secondary pathologies can also occur and include TDP-43 aggregates and Lewy bodies composed of α -synuclein.

Brain imaging

PET scans of an Alzheimer's disease brain compared with a normal control brain. ¹⁸F-Flortetapir binds to amyloid producing the red colour on the scan.

AV1451 binds to tau, producing the red colour on the scan. These PET images demonstrate tau deposition in late- and early-onset Alzheimer's disease brains compared with a cognitively normal brain.

Images courtesy of Professor Oskar Hansson at Lund University and Associate Professor Michael Schenk at University of Gothenburg

Current biomarkers of Alzheimer's disease: bench, bedside and needed breakthroughs

Pathology	Biomarkers	Blood	Brain scan
Aβ pathology	<ul style="list-style-type: none"> CSF Aβ42 (ELISA, MS) Pancreatic polypeptide Y IgM Chemokine ligand 13 Interleukin 17 Vascular adhesion protein 1 α2-macroglobulin Apolipoprotein A1 	<ul style="list-style-type: none"> None 	<ul style="list-style-type: none"> Amyloid PET
Tau pathology	<ul style="list-style-type: none"> P-tau (ELISA, MS) T-tau (ELISA) Neurofilament light (ELISA) 	<ul style="list-style-type: none"> None 	<ul style="list-style-type: none"> ¹⁸F-AV1451
Axonal degeneration	<ul style="list-style-type: none"> Neurogranin SNAP25 RAB5A 	<ul style="list-style-type: none"> None 	<ul style="list-style-type: none"> MRI
Synaptic degeneration	<ul style="list-style-type: none"> Secreted ectodomain TREM2 Choline oxidase CD14 YKL-40 CCL2 	<ul style="list-style-type: none"> None 	<ul style="list-style-type: none"> SV2A (¹¹C-UCB-J) FDG
Gliat activation	<ul style="list-style-type: none"> None 	<ul style="list-style-type: none"> YKL-40 	<ul style="list-style-type: none"> TSPO ¹¹C-β-PK11195 ¹⁸F-DPA-714 ¹¹C-PBR28
TDP43 pathology	<ul style="list-style-type: none"> None 	<ul style="list-style-type: none"> None 	<ul style="list-style-type: none"> None
α-Synuclein pathology	<ul style="list-style-type: none"> None 	<ul style="list-style-type: none"> None 	<ul style="list-style-type: none"> ¹⁸F-BP-227

Microglia and **TDP-43 NCI** are highlighted in green, those with no highlight are current research findings, and those in red highlight areas in which more research and development is needed.

Pros, cons and future considerations

Biomarker source	Pros	Cons
CSF	<ul style="list-style-type: none"> Closer to the brain Lower protease activity 	<ul style="list-style-type: none"> Less accessible Invasive lumbar puncture
Blood	<ul style="list-style-type: none"> Less invasive to collect sample 	<ul style="list-style-type: none"> Further away from the affected brain
Brain scan	<ul style="list-style-type: none"> With the correct tracer gives accurate diagnostic accuracy 	<ul style="list-style-type: none"> Costly method Radioactive probe injected in the bloodstream Invasive

The future of biomarkers

- Needed to detect the underlying pathology with 100% accuracy
- Accurate biomarkers for all pathologies are needed (A β , tau, TDP-43 and α -synuclein)
- Once developed, these will be able to track disease progression
- Will enable the sub-classification of diseases based on pathological signatures
- Will enable disease tracking when disease modifying drugs become available

Abbreviations: A β , Alzheimer's disease; CCL2, C-C chemokine ligand 2; CSF, cerebrospinal fluid; ELISA, enzyme-linked immunosorbent assay; FDG, ¹⁸F-Fluoro-2-deoxy-D-glucose; IgM, immunoglobulin M; IHC, immunohistochemistry; MCI, mild cognitive impairment; MS, mass spectrometry; NCI, neuronal cytoplasmic vacuolation; PET, positron emission tomography; P-tau, phosphorylated tau; T-tau, total tau.

© 2018. Published by The Company of Biologists Ltd
doi: 10.1242/dmm.031781

Box 1. Glossary

Amyloid positron emission tomography: an imaging technique that visualises amyloid senile plaques in the living human brain.

Axonal degeneration: a progressive degeneration and loss of axons.

β -amyloid: the cleavage product of amyloid precursor protein and the main constituent of senile (amyloid) plaques in Alzheimer's disease.

Cerebral amyloid angiopathy: a form of blood vessel disorder in which amyloid deposits form in the walls of the blood vessels of the central nervous system, affecting blood flow.

Cerebral β -amyloidosis: β -amyloid plaque pathology in the brain.

Creutzfeldt-Jakob disease (CJD): a rapidly progressive neurodegenerative disease caused by self-propagating aggregation of a normal brain protein (prion protein).

Dementia with Lewy bodies (DLB): a progressive brain disorder in which Lewy bodies, deposits of the protein α -synuclein build up in areas of the brain that regulate behaviour, cognition and movement.

Diffuse leukoaraiosis: an abnormal change in appearance of white matter near the lateral ventricles of the brain.

Frontotemporal dementia (FTD): the clinical presentation of frontotemporal lobar degeneration, which is characterised by progressive neuronal loss predominantly involving the frontal or temporal lobes.

Lacunar infarction: the most common type of stroke that results from the occlusion of small penetrating arteries that provide blood to the brain's deep structures.

Logopenic variant primary progressive aphasia: a language disorder that involves changes in the ability to speak, read, write and understand what others are saying.

Parkinson's disease dementia: Parkinson's disease that later progresses into dementia.

Progressive supranuclear palsy: a tauopathy that often starts with Parkinson-like symptoms but then progresses to involve other brain regions.

Proteopathies: a class of diseases in which certain proteins become structurally abnormal, and thereby disrupt the function of cells, tissues and organs of the body.

Senile plaques: extracellular deposits of β -amyloid.

Superficial central nervous system siderosis: a disease of the brain resulting from chronic iron deposition in neuronal tissues associated with cerebrospinal fluid.

Synaptic degeneration: a progressive degeneration and loss of synapses.

Synucleinopathies: diseases in which α -synuclein inclusions accumulate inside neurons and other cell types of the brain.

Tau: a structural protein in axons that may form neurofibrillary tangles if truncated and hyperphosphorylated.

Tauopathies: neurodegenerative diseases in which abnormal inclusions of tau accumulate inside neurons.

Temporo-parietal association cortices: the cerebral cortex outside the primary areas in the temporal and parietal lobes.

Vascular dementia (VaD): a form of dementia caused by cerebrovascular disease, resulting in neuronal injury and degeneration.

Introduction

Neurodegenerative dementias constitute a broad category of brain diseases that are characterised by a typically gradual decline in cognitive function, ultimately leading to increased mortality. The most common type of dementia is Alzheimer's disease (AD), which accounts for 50-70% of prevalent neurodegenerative dementia cases (Winblad et al., 2016). AD causes a progressive loss of cognitive abilities with short-term memory impairment being the most typical initial symptom. However, there are also atypical clinical presentations of AD, such as logopenic variant primary progressive aphasia (see Glossary, Box 1) or posterior cortical atrophy (Mattsson et al., 2016a). Alongside AD, there are many other dementia-causing neurodegenerative diseases that

might be important for differential diagnoses (Schott and Warren, 2012).

Traditionally, a dementia diagnosis is based on the history of the illness, the pattern of cognitive deficits, and on additional parameters assessed through clinical investigations, including blood tests and structural imaging of the brain, to rule out nondegenerative causes of the symptoms. Increasingly, and with the prospect of disease modification, there has been a shift towards the use of biomarkers (Dubois et al., 2014) to diagnose specific forms of dementia earlier, also in the pre-dementia stages of the disease, and with more specificity. Currently, a definite diagnosis of a neurodegenerative dementia requires histopathological confirmation at autopsy, as different degenerative dementia-causing brain disorders are characterised by more or less distinct pathologies (Hyman et al., 2012). A striking common feature of most neurodegenerative dementias is the presence of aggregates or inclusions of misfolded endogenous proteins in the brain extracellular matrix or within neurons and other brain cell types (Kovacs, 2016). This common feature classifies these dementias as proteopathies (Box 1) (Walker and Jucker, 2015).

Neuropathologically, AD is characterised by: (1) neuronal loss in specific brain regions – notably the medial temporal lobe structures and the temporo-parietal association cortices (Box 1); (2) intraneuronal neurofibrillary tangles composed of aggregated and often truncated and hyperphosphorylated tau protein (Box 1); and (3) extracellular neuritic plaques, which consist of deposits of β -amyloid (Box 1) peptides, mainly its 42-amino-acid isoform (Blennow et al., 2006) (see poster). There are other neurodegenerative diseases with symptoms that might overlap with AD, such as frontotemporal dementia (FTD; Box 1), in which inclusions can consist of several different proteins, most typically tau (MAPT) and/or transactive response DNA-binding protein 43 (TDP-43; TARDBP); Parkinson's disease dementia (PDD; Box 1) and dementia with Lewy bodies (DLB; Box 1), in which α -synuclein inclusions represent an important part of the pathology (Box 1). These neurodegenerative pathologies often present with a considerable degree of co-morbidity, with several different pathological changes co-occurring in the same brain tissue, indicating that pathologically deposited proteins might interact and be influenced by other factors to cause cognitive decline and other clinical symptoms (Lashley et al., 2008). For example, cerebral small vessel disease, which might be caused by several different pathological processes, including lacunar infarction, diffuse leukoaraiosis and cerebral amyloid angiopathy (Box 1), can both cause dementia and co-exist with degenerative dementias, thus influencing their severity and phenotype. Because clinical phenotypes of dementia can be caused by different pathological changes, sometimes interacting with each other, it is important to develop biomarkers that can diagnose these changes to improve the possibility to monitor and treat the underlying cause.

In this 'At a glance' poster, we show the different neuropathological changes that might underlie neurodegenerative dementias, especially AD, and discuss the currently available molecular fluid- and imaging-based biomarkers for each pathology. We also discuss why biomarkers are necessary in the clinic, the challenges encountered when using them clinically, which patient populations the different biomarkers could serve, and the issues relating to the implementation of standardised sampling and handling protocols. The biomarker concept with special focus on neurodegenerative dementias is detailed in the far-left panel of the poster. The discussion of structural or functional imaging biomarkers is beyond the scope of this article, but we refer the

interested reader to an excellent recent review on these biomarkers (Rathore et al., 2017).

Biomarkers for β -amyloid pathology

The 42-amino-acid isoform of β -amyloid (A β 42) is a major component of senile plaques (Box 1) and contributes to cerebral amyloid angiopathy in AD (Masters et al., 1985) (see poster). A β 42 is a cleavage product of type I transmembrane amyloid precursor protein (APP) with no known physiological function. It is released from neurons when APP is cleaved by β - and γ -secretases in synaptic vesicles. APP is metabolised by many cell types, but A β 42 secretion is by far the highest from neurons; its neuronal secretion also appears to depend on synaptic activity (Cirrito et al., 2005).

Cerebrospinal fluid biomarkers

A β 42 concentration can be measured in the cerebrospinal fluid (CSF) by antibody-based techniques, such as enzyme-linked immunosorbent assay (ELISA), as well as by antibody-independent techniques, such as mass spectrometry (Kuhlmann et al., 2016). AD patients have decreased concentrations of A β 42 in their CSF, a finding that has been well verified and replicated by many studies (Olsson et al., 2016). This decrease reflects the sequestration of A β 42 in senile plaques in the brain, as evidenced by autopsy and by *in vivo* amyloid positron emission tomography (PET) imaging (Box 1) studies in patients (Blennow et al., 2015). Reduced levels of A β 42 can be detected in the CSF of patients with mild cognitive impairment (MCI), as well as in the pre-clinical stages of AD (Bateman et al., 2012; Olsson et al., 2016). A plaque pathology-associated decrease in A β 42 concentration in the CSF is also frequently seen in DLB, another neurodegenerative dementia that is very commonly accompanied by cerebral A β aggregation (Abdelnour et al., 2016).

Blood biomarkers

It has been difficult to establish robust blood biomarkers for A β pathology. A β proteins can be measured in plasma, but their correlation with cerebral β -amyloidosis (Box 1) is absent or weak when the latter is assessed immunochemically (Olsson et al., 2016). Plasma A β concentrations are probably influenced by its secretion from platelets and from other extracerebral tissues (Zetterberg, 2015). Nevertheless, three recent studies have reported a clinically significant correlation between plasma A β concentrations and cerebral β -amyloidosis measured by mass spectrometry (Kaneko et al., 2014; Nakamura et al., 2018; Ovod et al., 2017). A similar result has been obtained using an ultrasensitive assay (Janelidze et al., 2016). The results of these papers are promising and warrant further validation. Other studies have reported that various plasma proteins (including pancreatic polypeptide Y, immunoglobulin M, chemokine ligand 13, interleukin 17, vascular cell adhesion protein 1, α 2-macroglobulin, apolipoprotein A1 and complement proteins) are associated with A β 42 burden in the brain, irrespective of the clinical stage of AD (Burnham et al., 2016; Westwood et al., 2016; Voyle et al., 2015). However, these data should be interpreted with caution, as they are derived from multimarker panels and have not been replicated or examined in relation to other neurodegenerative dementias. Further, we are currently lacking a mechanistic understanding of these associations.

Biomarkers for PET

The first chemical probe for amyloid PET was an ^{11}C -labelled modified derivative of the amyloid-binding histological dye thioflavin-T called Pittsburgh Compound-B (PiB, also known as ^{11}C -PiB). This probe is retained in cortical brain regions of AD patients compared with healthy controls, with retention in the

cerebellum used as a reference region (Klunk et al., 2004). The increased retention of PiB in cortical brain regions of AD patients has since been verified in many scientific reports (reviewed in Blennow et al., 2015). However, the short half-life of ^{11}C hinders the use of ^{11}C -PiB outside of expert research centres that have access to an on-site cyclotron to generate the probe, as well as to radiochemistry expertise. As a result, ^{18}F -labelled probes have been developed that have a half-life of ~ 110 min. This longer half-life enables the centralised production of this probe and its regional distribution to medical centres that have a PET scanner. Three ^{18}F -PET amyloid tracers are licensed for clinical use: ^{18}F -florbetapir (Amyvid), ^{18}F -flutemetamol (Vizamyl) and ^{18}F -florbetaben (Neuraceq), all of which have shown good correlation with amyloid plaque burden at autopsy (Morbelli and Bauckneht, 2018), although they generally show higher levels of nonspecific binding to white matter compared with ^{11}C -PiB.

Biomarkers for tau pathology

Abnormally phosphorylated and truncated tau proteins are the major component of neurofibrillary tangles in AD and other tauopathies (Box 1) (Grundke-Iqbal et al., 1986) (see poster). The normal function of tau is to bind to and stabilise microtubules in neuronal axons (Zetterberg, 2017), a process that is inhibited when tau becomes phosphorylated.

CSF biomarkers

Tangle-containing neurons release phosphorylated tau, which can be measured in the CSF by ELISA, using antibody combinations that specifically recognise mid-domain phospho-tau (P-tau) epitopes. AD patients have increased concentrations of P-tau in their CSF (Olsson et al., 2016). However, P-tau concentration in the CSF correlates weakly with neurofibrillary tangle pathology in the brain of AD patients (Buerger et al., 2006; Seppala et al., 2012). This finding was replicated in a recent tau PET imaging study of AD patients (Chhatwal et al., 2016), although the results are less clear than the association of CSF A β 42 with amyloid PET.

A major outstanding research question is why other tauopathies, including some forms of FTD and associated disorders like progressive supranuclear palsy (Box 1), do not show increased P-tau concentration in the CSF, at least not as robustly as in AD (Zetterberg, 2017). It is possible that disease-specific phosphorylation of tau occurs in these disorders, or that tau is processed or truncated in a way that is not recognised by the available assays. Another potential explanation for why increased CSF P-tau is specific to AD is that this particular pathological change is simply more extensive and severe in AD than it is in other tauopathies. CSF P-tau is currently considered to be the most specific biomarker for AD. Except for herpes encephalitis and superficial CNS siderosis (Box 1), no other condition features a systematic increase in this biomarker (Zetterberg, 2017).

Blood biomarkers

To date, no reliable blood biomarkers for neurofibrillary tangle pathology have been identified. However, recent studies have reported increased P-tau concentrations in blood-borne neuron-derived exosomes (Shi et al., 2016; Winston et al., 2016). In this assay, the exosomes are isolated from serum using antibodies directed against neuron-enriched proteins. The isolated exosomes are then washed and lysed and their tau content is measured using immunochemical assays. Although new, this technique represents a promising approach for P-tau measurements in blood.

Biomarkers for PET

PET tracers (also known as probes) have been developed to visualise tau inclusions *in vivo* in patients. One of these tracers, ^{18}F -AV1451, binds to tau aggregates in AD (Marquie et al., 2015), can differentiate AD patients from healthy controls (Schöll et al., 2016) and correlates with regional changes in brain metabolism in different clinical variants of AD (Ossenkoppele et al., 2016) (see poster). However, AV1451 does not reliably bind to all pathological isoforms of tau, nor does its binding reliably correlate with pathological tau load (Sander et al., 2016). Similar results have been published for other tau PET probes, although they might recognise different forms of tau deposits because of differences in their structure and binding properties (Kolb and Andres, 2017). Preliminary evidence indicates that tau measurements in the CSF and via PET correlate in the dementia stage of AD, but less clearly in the preclinical and mild cognitive impairment stages of the disease (Mattsson et al., 2017). However, this correlation is not as well established as that between CSF and PET biomarkers for A β pathology.

Biomarkers for axonal degeneration

Axonal degeneration (Box 1) is a key feature of AD, and is more closely linked to the onset of cognitive decline than A β pathology is (see poster, 'Biomarkers'). In fact, according to some models, the onset of neurodegeneration marks the beginning of the toxic phase of A β pathology in the pathogenesis of AD (Jack and Holtzman, 2013).

CSF biomarkers

Total tau (T-tau), measured using antibodies against mid-domain tau epitopes that are not phosphorylated, can be used as a general marker of axonal degeneration or injury in AD. AD patients have increased concentrations of T-tau in their CSF (Olsson et al., 2016), and the higher the increase, the more intense the neurodegenerative process (Wallin et al., 2010). However, increased levels of CSF T-tau are not specific to AD; the increase is also seen, for example, in Creutzfeldt-Jakob disease (CJD; Box 1) (Riemenschneider et al., 2003) and following stroke (Hesse et al., 2001). Assays have also been developed to quantify visinin-like protein 1 (VLP-1; VSNL1) and members of the fatty acid-binding protein (FABP) family in the CSF. VLP-1 and FABP proteins are enriched in neurons, but their association with AD is less strong than that of CSF T-tau (Olsson et al., 2016). Neuron-specific enolase (NSE; ENO2) has also been proposed as a candidate biomarker for neuronal loss in AD, but its association with AD is weak and clinically irrelevant (Olsson et al., 2016). In addition, the results of NSE tests are easily confounded by blood contamination, because NSE (despite its name) is highly expressed in erythrocytes (Ramont et al., 2005).

Another CSF biomarker for axonal degeneration is neurofilament light (NF-L; NEFL), which is a structural protein present in long axons (Zetterberg, 2016). The concentration of NF-L is increased in the CSF of AD patients, especially so in those with rapid disease progression (Zetterberg et al., 2016). However, increased NF-L in the CSF is not specific to AD, and is detected in other dementias, with the highest concentrations seen in FTD and in vascular dementia (VaD; Box 1) (de Jong et al., 2007; Landqvist Waldö et al., 2013; Sjogren et al., 2000). These results were recently confirmed in a large retrospective analysis of data from the Swedish Dementia Registry (Skillback et al., 2014), as well as in atypical parkinsonian disorders (Hall et al., 2012; Magdalinou et al., 2015). The highest CSF concentrations of NF-L are seen in CJD, as is the case for T-tau (Steinacker et al., 2016; van Eijk et al., 2010).

Blood biomarkers

CSF assays for T-tau and NF-L have recently been redeveloped into ultrasensitive blood tests using single molecule array (Simoa) technology (Andreasson et al., 2016). Serum and plasma NF-L concentrations correlate with their concentrations in the CSF (correlation coefficients of 0.75 to 0.97), and most measurements in the CSF (increased NF-L concentrations in AD, FTD, VaD and in atypical parkinsonian disorders) have been replicated in blood (Zetterberg, 2016). For T-tau, such correlation is less clear, but promising. First, for unknown reasons, tau concentrations are higher in plasma than in serum (H.Z., unpublished). Second, the correlation with the corresponding CSF concentration is either absent (Zetterberg et al., 2013) or weak (Mattsson et al., 2016b). In AD, plasma T-tau levels are increased, but less so than in the CSF, and there is no detectable increase in the MCI stage of the disease (Mattsson et al., 2016b; Zetterberg et al., 2013).

PET biomarkers

There are presently no PET probes for axonal degeneration. There will likely not be any in the near future, as there are no targetable molecular assemblies that are specific to axonal degeneration and which could function as anchors for PET probe binding.

Fluid biomarkers for synaptic degeneration

In its earliest clinical phase, AD characteristically and consistently causes memory impairment. Mounting evidence suggests that memory impairment begins with subtle alterations to synaptic efficacy in the hippocampus, prior to frank neuronal degeneration (see poster). A reduction in synapse number is associated with numerous brain disorders, and with AD in particular (Selkoe, 2002).

CSF biomarkers

Neurogranin (Ng; NRG1) is a dendritic protein enriched in neurons that is involved in long-term potentiation of synapses, particularly so in the hippocampus and the basal forebrain (Reprea et al., 1990). Recently, several independent studies have shown that the CSF concentration of Ng is increased in AD (Hellwig et al., 2015; Kester et al., 2015; Kvartsberg et al., 2015a,b; Thorsell et al., 2010), but not in other neurodegenerative disorders (Wellington et al., 2016). Moreover, studies showed a quantitative correlation between the magnitude of Ng increase and the severity of cognitive decline, brain atrophy and reduction in glucose metabolism in the prodromal stage of the disease (Portelius et al., 2015; Tarawneh et al., 2016). Currently, CSF Ng is the best-established CSF biomarker for synapse loss or dysfunction associated with AD, although other promising markers of this pathological change are being characterised in single-centre studies awaiting independent replication. These markers include synaptosomal-associated protein 25 (SNAP25) and Ras-related protein RAB3A (Bereczki et al., 2016; Brinkmalm et al., 2014).

Blood biomarkers

There are so far no reliable blood biomarkers for synaptic pathology. Plasma Ng has been explored as a candidate marker in this context, but its concentration remained unchanged in AD patients compared with cognitively healthy controls (De Vos et al., 2015). Most likely, the extracerebral expression of Ng constitutes the major source of Ng in plasma (De Vos et al., 2015), confounding potential differences between the healthy and AD groups.

PET biomarkers

In a recent report, the synaptic vesicle glycoprotein 2A (SV2A) radioligand ^{11}C -UCB-J combined with PET was used to quantify

synaptic density in the living human brain (Finnema et al., 2016). The probe had excellent imaging properties and was sensitive enough to detect synaptic loss in patients with temporal lobe epilepsy. Its utility in AD, however, remains to be established. Another technique to monitor synaptic loss in neurodegenerative dementias is ^{18}F -2-fluoro-2-deoxy-D-glucose (FDG)-PET. This method detects brain region-specific impairment of cerebral glucose metabolism in neurodegenerative diseases (Payoux and Salabert, 2017). A recent systematic review and meta-analysis revealed varying diagnostic accuracies of FDG-PET, with sensitivities between 46% and 95%, while the specificities were between 29% and 100%, and the conclusion was that the method in its current stage of development cannot be recommended for clinical use in AD, with more research recommended (Smailagic et al., 2015).

Biomarkers for glial activation

Glial cells in the brain are astrocytes, the star-shaped cells that provide neurons with nutrients, form part of the blood-brain barrier and take part in repair mechanisms following CNS injury, and microglia, the resident macrophages of the brain, constituting the main form of active immune defence in the CNS. Most often, both cell types are activated in parallel, and glial activation has been linked to deficits in neuronal function and to synaptic plasticity in AD. The recent discovery of a genetic link between AD and variants of the triggering receptor expressed on myeloid cells 2 (*TREM2*; *TREML2*) gene (Guerreiro et al., 2013; Jonsson et al., 2013), which is selectively expressed on microglia in the CNS (Lue et al., 2015; Takahashi et al., 2005), has reignited the interest in identifying biomarkers of glial activation (see poster).

Cerebrospinal fluid biomarkers

Recent reports suggest that the concentrations of the secreted ectodomain of *TREM2* are increased in the CSF of AD patients. This increase is disease specific and correlates with elevated CSF levels of T-tau and P-tau (Heslegrave et al., 2016; Piccio et al., 2016; Suárez-Calvet et al., 2016). These findings are supported by numerous studies that report the increased CSF concentrations of several other astrocyte-, microglia- and/or macrophage-derived proteins, including chitotriosidase (Mattsson et al., 2011; Watabe-Rudolph et al., 2012), CD14 (Yin et al., 2009) and YKL-40 (*CHI3LI*) (Craig-Schapiro et al., 2010; Olsson et al., 2013). Another glial marker, the C-C chemokine receptor 2, is expressed on monocytes, and one of its ligands, C-C chemokine ligand 2 (*CCL2*), which can be produced by microglia, is also present in increased concentrations in the CSF of AD patients (Corrêa et al., 2011; Galimberti et al., 2006a, b). Most studies suggest that these increases in glial proteins in AD are modest, with concentration ranges overlapping extensively between cases and controls, particularly when compared with the more prominent changes seen in 'traditional' neuroinflammatory conditions, such as multiple sclerosis (Öhrfelt et al., 2016) or human immunodeficiency virus (HIV)-associated neurocognitive dysfunction (Peluso et al., 2017). It should also be noted that all of the above-mentioned proteins, except *TREM2*, can also be released from activated astrocytes. Thus, microglial and astrocytic activation in AD are difficult to tease apart using only CSF-based biomarkers. This problem might not have any practical implications, as microglial and astrocytic activation are tightly linked and, as markers of the two processes, could potentially be used interchangeably to track AD.

Blood biomarkers

When biomarkers of microglial activation, such as those mentioned above, are measured in plasma or serum, their concentrations are

similar to those in the CSF and not 100-fold lower as would have been expected if they were CNS derived (Craig-Schapiro et al., 2010). This probably reflects their release from monocytes and macrophages in peripheral blood rather than reflecting CNS-related changes. However, a few studies indicate that YKL-40 concentration is slightly increased in the plasma of AD patients (Olsson et al., 2016); the overlap between AD patients and cognitively normal controls was, however, too large for the marker to be used clinically.

PET biomarkers

The mitochondrial translocator protein (TSPO) is known to be upregulated in activated microglia. Accurate visualisation and quantification of microglial density by PET imaging using the TSPO probe [^{11}C]-R-PK11195 has been challenging owing to the limitations of the probe, mainly its low brain permeability and the abundant expression of TSPO in extracerebral tissues. A number of new TSPO probes (e.g. [^{18}F]-DPA-714 and [^{11}C]-PBR28) have been evaluated in rodent and nonhuman primate models, but the literature on their clinical usefulness remains scant (Lagarde et al., 2017).

Biomarkers for TDP-43 pathology

Hyperphosphorylated TDP-43 proteinopathy occurs in ~50% of FTD patients and has recently been described both in studies of ageing and in association with the cognitive impairment of ageing patients, especially in the context of tau and A β pathology (James et al., 2016), providing some overlap with AD (see poster).

CSF biomarkers

Total TDP-43 can be measured in the CSF but, unfortunately, most of the protein appears to be blood derived, as it is ubiquitously expressed throughout the body. Thus, its concentration in the CSF does not reflect the presence of TDP-43 proteinopathy in the brain. Moreover, its levels in the CSF of FTD patients are also unaltered (Feneberg et al., 2014), further disqualifying this protein as a potential CSF biomarker.

Blood biomarkers

No reliable blood test for TDP-43 pathology in the CNS exists. Given the ubiquitous expression of TDP-43 throughout the body, brain inclusion-specific pathologic forms of TDP-43 would have to be targeted in the blood for this to become a feasible biomarker project.

Biomarkers for PET

There are presently no PET probes for TDP-43 inclusions. In 2016, the Amyotrophic Lateral Sclerosis Association announced a Grand Challenge grant to develop TDP-43 inclusion probes but, so far, there are no published data on this topic.

Biomarkers for α -synuclein pathology

The presynaptic neuronal protein α -synuclein can misfold and form seeds that can aggregate further into inclusions that are called Lewy bodies. These inclusions are characteristic of Parkinson's disease (PD) and of DLB (Mollenhauer et al., 2010), but also often feature in AD (Schneider et al., 2009) (see poster).

Cerebrospinal fluid biomarkers

In PD and in other synucleinopathies (Box 1), α -synuclein concentrations in the CSF are typically lower than in healthy controls (Hall et al., 2012; Mollenhauer et al., 2011), whilst in AD

and CJD, its concentrations are increased and correlate with T-tau, indicating that α -synuclein might also be a nonspecific marker of neurodegeneration (Mollenhauer et al., 2011; Öhrfelt et al., 2009; Slaets et al., 2014; Tateno et al., 2012; Wennström et al., 2013). Increased CSF levels of α -synuclein have also been reported in DLB, where a competition might exist between the aggregation of α -synuclein into Lewy bodies and its release from the degenerating synapses, which complicates data interpretation (Kapaki et al., 2013). In agreement with this hypothesis, a recently published multiple reaction monitoring mass spectrometry assay revealed significantly increased CSF concentrations of α -, β - and γ -synuclein in AD and CJD, but not in the 'classical' synucleinopathies like PD (Oeckl et al., 2016). Currently available assays for α -synuclein measure the total amounts of the protein and not the Lewy body-specific isoforms. The availability of sensitive and specific assays for these pathogenic isoforms would resolve the issue of having the biomarker results influenced or confounded by the release of native α -synuclein from degenerating synapses. However, there are some preliminary reports of increased concentrations of α -synuclein oligomers in the CSF of PD patients (Hansson et al., 2014; Tokuda et al., 2010). Recent studies described sensitive assays that detect the amplified biochemical signal of α -synuclein seeds that might be Lewy body derived in the CSF from PD patients but not in that from healthy controls (Fairfoul et al., 2016; Shah Nawaz et al., 2016), opening a promising avenue for using CSF α -synuclein as a biomarker.

Blood biomarkers

Because α -synuclein is highly expressed in red blood cells, blood contamination during CSF collection might limit its diagnostic value (Barbour et al., 2008; Hong et al., 2010). For the very same reason, blood tests for α -synuclein pathology in the brain might lack the specificity needed for it to be an informative clinical biomarker. Nevertheless, as peripheral Lewy body pathology, such as in the salivary gland and the gut, has been reported in PD (Uchihara and Giasson, 2016), blood or salivary tests for α -synuclein seeds might be worth exploring in the future as a biomarker of PD and other dementias associated with Lewy bodies, such as AD.

PET biomarkers

Efforts to develop PET probes for α -synuclein inclusions are ongoing but are still in their infancy. One of the compounds currently investigated for imaging α -synuclein inclusions is the ^{18}F -labelled compound BF-227 that was reported to bind to both synthetic α -synuclein aggregates as well as β -amyloid fibrils *in vitro* (Fodero-Tavoletti et al., 2009). A histopathological study demonstrated that BF-227 could stain α -synuclein-containing glial cytoplasmic inclusions in postmortem tissue (Fodero-Tavoletti et al., 2009). Moreover, a PET study with ^{11}C -labelled BF-227 showed its ability to detect α -synuclein deposits in the living brains of patients with multiple system atrophy (Kikuchi et al., 2010). However, the high affinity of this radiotracer for $\text{A}\beta$ plaques limits its usefulness for differential diagnosis (Kikuchi et al., 2010).

Pros and cons of the different biomarker modalities

A biomarker can be defined as 'a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes or pharmacologic responses to a therapeutic intervention' (Strimbu and Tavel, 2010). Biomarkers for AD should, thus, reflect the core pathogenic findings in the brain, i.e. plaque and tangle pathology, as well as the associated pathophysiological mechanisms, i.e. axonal and synaptic

degeneration (Box 1) and frequent co-pathologies, including TDP-43 and α -synuclein pathologies (see poster, 'Brain imaging'). We have discussed three broad categories of biomarker modalities in this 'At a glance' paper: CSF, blood and PET, and they differ in terms of accessibility and how closely they reflect the changes in the brain (see poster). In regards to the fluids, the CSF is closer to the brain and has a lower intrinsic protease activity than blood. However, the CSF is less accessible, as sampling requires a lumbar puncture. Regarding PET, this is a much more costly method. Further, it involves injecting a radioactive probe into the blood. This probe will cross the blood-brain barrier and remain bound to its target pathologies for an unknown time period. This technique can thus also be regarded as invasive. In regards to plaque pathology, the diagnostic accuracies of the CSF and PET tests are comparable (Blennow et al., 2015). However, for the other pathologies, prospective studies that directly compare the different biomarker modalities are needed to determine whether any one marker is better at reflecting the extent of the pathology than another. Regarding blood tests, one biomarker stands out as being particularly promising: NF-L, for which robust CSF and plasma/serum correlations have been established. For this particular protein, virtually the same information can be gathered from a CSF test and a blood test (Zetterberg, 2016). For the other blood tests, more research is needed before any of these could replace the corresponding CSF or PET test (see poster). For biomarkers, fluid- and imaging-based alike, to be implemented in clinical practice, the measurement techniques have to be well standardised and give stable results over time. For CSF $\text{A}\beta_{42}$ measurements, there are now certified reference methods and a reference material is under production (Kuhlmann et al., 2016). Similar work is ongoing for the CSF-based tau biomarkers. Pre-analytical standard operating procedures for CSF sampling and storage have been published (Blennow et al., 2010), and guidelines on how to interpret the results in different clinical stages of the disease are being developed (Herukka et al., 2017; Simonsen et al., 2017). Similarly, reliable quantitative analysis of amyloid PET scans acquired at multiple sites and over time requires rigorous standardisation of the acquisition protocols, subject management, tracer administration, image quality control, and image processing and analysis methods. Approaches to address these issues have been published (Schmidt et al., 2015). For most of the other candidate biomarkers discussed in this paper, such work is pending and will be required prior to the transition of these biomarkers into routine clinical practice.

Conclusions

Disease biomarkers have been developed into clinically available methods to detect tangle and plaque pathology in the CSF and brains of AD patients, and there are also promising biomarkers to detect synaptic loss and dysfunction. Tau and $\text{A}\beta$ biomarkers can help to diagnose AD pathology in both the prodromal and the dementia stages of the disease. Moreover, a number of additional biomarkers have been identified that detect pathological changes common to AD and other neurodegenerative proteopathies, although reliable and accurate biomarkers for TDP-43 and Lewy body pathology remain to be identified. If identified in the future, such biomarkers could be employed in longitudinal studies to track the temporal development of different pathologies during neurodegenerative disease progression, and to assess how their interactions lead to clinical symptoms. As multimorbidity appears to be common not only in AD but also in other neurodegenerative dementias, one potential future scenario is that these biomarkers could be used to subclassify the clinical syndromes in individual

patients according to their pathological signature, allowing for personalised treatment.

Competing interests

The authors declare no competing or financial interests.

Funding

Work in the authors' laboratories is supported by Vetenskapsrådet (Swedish Research Council), the Medical Research Council (MRC) (MR/M008525/1), the European Research Council, Knut och Alice Wallenbergs Stiftelse (the Knut and Alice Wallenberg Foundation), Alzheimer's Research Trust, Alzheimer's Association, Swedish State Support for Clinical Research, the Wellcome Trust and the Leonard Wolfson Experimental Neurology Centre. J.D.R. is supported by an MRC Clinician Scientist Fellowship (MR/M008525/1) and has received funding from the National Institute for Health Research (NIHR) Rare Disease Translational Research Collaboration (BRC149/NS/MH).

At a glance

A high-resolution version of the poster is available for downloading in the online version of this article at <http://dmm.biologists.org/content/11/5/dmm031781/F1.poster.jpg>.

References

- Abdelnour, C., van Steenoven, I., Londos, E., Blanc, F., Auestad, B., Kramberger, M. G., Zetterberg, H., Mollenhauer, B., Boada, M. and Aarsland, D. (2016). Alzheimer's disease cerebrospinal fluid biomarkers predict cognitive decline in lewy body dementia. *Mov. Disord.* **31**, 1203-1208.
- Andreasson, U., Blennow, K. and Zetterberg, H. (2016). Update on ultrasensitive technologies to facilitate research on blood biomarkers for central nervous system disorders. *Alzheimers Dement.* **3**, 98-102.
- Barbour, R., Kling, K., Anderson, J. P., Banducci, K., Cole, T., Diep, L., Fox, M., Goldstein, J. M., Soriano, F., Seubert, P. et al. (2008). Red blood cells are the major source of alpha-synuclein in blood. *Neurodegener. Dis.* **5**, 55-59.
- Bateman, R. J., Xiong, C., Benzinger, T. L., Fagan, A. M., Goate, A., Fox, N. C., Marcus, D. S., Cairns, N. J., Xie, X., Blazey, T. M. et al. (2012). Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *N. Engl. J. Med.* **367**, 795-804.
- Bereczki, E., Francis, P. T., Howlett, D., Pereira, J. B., Höglund, K., Bogstedt, A., Cedazo-Minguez, A., Baek, J.-H., Hortobágyi, T., Attems, J. et al. (2016). Synaptic proteins predict cognitive decline in Alzheimer's disease and Lewy body dementia. *Alzheimers Dement.* **12**, 1149-1158.
- Blennow, K., de Leon, M. J. and Zetterberg, H. (2006). Alzheimer's disease. *Lancet* **368**, 387-403.
- Blennow, K., Hampel, H., Weiner, M. and Zetterberg, H. (2010). Cerebrospinal fluid and plasma biomarkers in Alzheimer disease. *Nat. Rev. Neurol.* **6**, 131-144.
- Blennow, K., Mattsson, N., Schöli, M., Hansson, O. and Zetterberg, H. (2015). Amyloid biomarkers in Alzheimer's disease. *Trends Pharmacol. Sci.* **36**, 297-309.
- Brinkmalm, A., Brinkmalm, G., Honer, W. G., Frölich, L., Hausner, L., Minthon, L., Hansson, O., Wallin, A., Zetterberg, H., Blennow, K. et al. (2014). SNAP-25 is a promising novel cerebrospinal fluid biomarker for synapse degeneration in Alzheimer's disease. *Mol. Neurodegener.* **9**, 53.
- Buerger, K., Ewers, M., Pirttilä, T., Zinkowski, R., Alafuzoff, I., Teipel, S. J., DeBernardis, J., Kerkman, D., McCulloch, C., Soininen, H. et al. (2006). CSF phosphorylated tau protein correlates with neocortical neurofibrillary pathology in Alzheimer's disease. *Brain* **129**, 3035-3041.
- Burnham, S. C., Rowe, C. C., Baker, D., Bush, A. I., Doecker, J. D., Faux, N. G., Laws, S. M., Martins, R. N., Maruff, P., Macaulay, S. L. et al. (2016). Predicting Alzheimer disease from a blood-based biomarker profile: a 54-month follow-up. *Neurology* **87**, 1093-1101.
- Chhatwal, J. P., Schultz, A. P., Marshall, G. A., Boot, B., Gomez-Isla, T., Dumurgier, J., LaPoint, M., Scherzer, C., Roe, A. D., Hyman, B. T. et al. (2016). Temporal T807 binding correlates with CSF tau and phospho-tau in normal elderly. *Neurology* **87**, 920-926.
- Cirrito, J. R., Yamada, K. A., Finn, M. B., Sloviter, R. S., Bales, K. R., May, P. C., Schoepp, D. D., Paul, S. M., Mennerick, S. and Holtzman, D. M. (2005). Synaptic activity regulates interstitial fluid amyloid-beta levels in vivo. *Neuron* **48**, 913-922.
- Corrêa, J. D., Starling, D., Teixeira, A. L., Caramelli, P. and Silva, T. A. (2011). Chemokines in CSF of Alzheimer's disease patients. *Arq. Neuropsiquiatr.* **69**, 455-459.
- Craig-Schapiro, R., Perrin, R. J., Roe, C. M., Xiong, C., Carter, D., Cairns, N. J., Mintun, M. A., Peskind, E. R., Li, G., Galasko, D. R. et al. (2010). YKL-40: a novel prognostic fluid biomarker for preclinical Alzheimer's disease. *Biol. Psychiatry* **68**, 903-912.
- de Jong, D., Jansen, R. W. M. M., Pijnenburg, Y. A. L., van Geel, W. J. A., Borm, G. F., Kremer, H. P. H. and Verbeek, M. M. (2007). CSF neurofilament proteins in the differential diagnosis of dementia. *J. Neurol. Neurosurg. Psychiatry* **78**, 936-938.
- De Vos, A., Jacobs, D., Struyfs, H., Franssen, E., Andersson, K., Portelius, E., Andreasson, U., De Surgelese, D., Hernalsteen, D., Sleegers, K. et al. (2015). C-terminal neurogranin is increased in cerebrospinal fluid but unchanged in plasma in Alzheimer's disease. *Alzheimers Dement.* **11**, 1461-1469.
- Dubois, B., Feldman, H. H., Jacova, C., Hampel, H., Molinuevo, J. L., Blennow, K., DeKosky, S. T., Gauthier, S., Selkoe, D., Bateman, R. et al. (2014). Advancing research diagnostic criteria for Alzheimer's disease: the IWG-2 criteria. *Lancet Neurol.* **13**, 614-629.
- Fairfoul, G., McGuire, L. I., Pal, S., Ironside, J. W., Neumann, J., Christie, S., Joachim, C., Esiri, M., Evetts, S. G., Rolinski, M. et al. (2016). Alpha-synuclein RT-QuIC in the CSF of patients with alpha-synucleinopathies. *Ann. Clin. Transl. Neurol.* **3**, 812-818.
- Feneberg, E., Steinacker, P., Lehnert, S., Schneider, A., Walther, P., Thal, D. R., Linsenmeier, M., Ludolph, A. C. and Otto, M. (2014). Limited role of free TDP-43 as a diagnostic tool in neurodegenerative diseases. *Amyotroph. Lateral Scler. Frontotemporal. Degener.* **15**, 351-356.
- Finnema, S. J., Nabulsi, N. B., Eid, T., Detyniecki, K., Lin, S. F., Chen, M. K., Dhaer, R., Matuskey, D., Baum, E., Holden, D. et al. (2016). Imaging synaptic density in the living human brain. *Sci. Transl. Med.* **8**, 348ra96.
- Fodero-Tavoletti, M. T., Mulligan, R. S., Okamura, N., Furumoto, S., Rowe, C. C., Kudo, Y., Masters, C. L., Cappai, R., Yanai, K. and Villemagne, V. L. (2009). In vitro characterisation of BF227 binding to alpha-synuclein/Lewy bodies. *Eur. J. Pharmacol.* **617**, 54-58.
- Galimberti, D., Schoonenboom, N., Scheltens, P., Fenoglio, C., Bouwman, F., Venturelli, E., Guidi, I., Blankenstein, M. A., Bresolin, N. and Scarpini, E. (2006a). Intrathecal chemokine synthesis in mild cognitive impairment and Alzheimer disease. *Arch. Neurol.* **63**, 538-543.
- Galimberti, D., Schoonenboom, N., Scheltens, P., Fenoglio, C., Venturelli, E., Pijnenburg, Y. A. L., Bresolin, N. and Scarpini, E. (2006b). Intrathecal chemokine levels in Alzheimer disease and frontotemporal lobar degeneration. *Neurology* **66**, 146-147.
- Grundke-Iqbal, I., Iqbal, K., Tung, Y. C., Quinlan, M., Wisniewski, H. M. and Binder, L. I. (1986). Abnormal phosphorylation of the microtubule-associated protein tau (tau) in Alzheimer cytoskeletal pathology. *Proc. Natl. Acad. Sci. USA* **83**, 4913-4917.
- Guerreiro, R., Wojtas, A., Bras, J., Carrasquillo, M., Rogaeva, E., Majounie, E., Cruchaga, C., Sassi, C., Kauwe, J. S. K., Younkin, S. et al. (2013). TREM2 variants in Alzheimer's disease. *N. Engl. J. Med.* **368**, 117-127.
- Hall, S., Öhrfelt, A., Constantinescu, R., Andreasson, U., Surova, Y., Bostrom, F., Nilsson, C., Hakan, W., Decraemer, H., Nägga, K. et al. (2012). Accuracy of a panel of 5 cerebrospinal fluid biomarkers in the differential diagnosis of patients with dementia and/or parkinsonian disorders. *Arch. Neurol.* **69**, 1445-1452.
- Hansson, O., Hall, S., Öhrfelt, A., Zetterberg, H., Blennow, K., Minthon, L., Nägga, K., Londos, E., Varghese, S., Majbour, N. K. et al. (2014). Levels of cerebrospinal fluid alpha-synuclein oligomers are increased in Parkinson's disease with dementia and dementia with Lewy bodies compared to Alzheimer's disease. *Alzheimers Res. Ther.* **6**, 25.
- Hellwig, K., Kvarnberg, H., Portelius, E., Andreasson, U., Oberstein, T. J., Lewczuk, P., Blennow, K., Kornhuber, J., Maler, J. M., Zetterberg, H. et al. (2015). Neurogranin and YKL-40: independent markers of synaptic degeneration and neuroinflammation in Alzheimer's disease. *Alzheimers Res. Ther.* **7**, 74.
- Herukka, S.-K., Simonsen, A. H., Andreasen, N., Baldeiras, I., Bjerke, M., Blennow, K., Engelborghs, S., Frisoni, G. B., Gabrylewicz, T., Galluzzi, S. et al. (2017). Recommendations for cerebrospinal fluid Alzheimer's disease biomarkers in the diagnostic evaluation of mild cognitive impairment. *Alzheimers Dement.* **13**, 285-295.
- Heslegrave, A., Heywood, W., Paterson, R., Magdalinou, N., Svensson, J., Johansson, P., Öhrfelt, A., Blennow, K., Hardy, J., Schott, J. et al. (2016). Increased cerebrospinal fluid soluble TREM2 concentration in Alzheimer's disease. *Mol. Neurodegener.* **11**, 3.
- Hesse, C., Rosengren, L., Andreasen, N., Davidsson, P., Vanderstichele, H., Vanmechelen, E. and Blennow, K. (2001). Transient increase in total tau but not phospho-tau in human cerebrospinal fluid after acute stroke. *Neurosci. Lett.* **297**, 187-190.
- Hong, Z., Shi, M., Chung, K. A., Quinn, J. F., Peskind, E. R., Galasko, D., Jankovic, J., Zabetian, C. P., Leverenz, J. B., Baird, G. et al. (2010). DJ-1 and alpha-synuclein in human cerebrospinal fluid as biomarkers of Parkinson's disease. *Brain* **133**, 713-726.
- Hyman, B. T., Phelps, C. H., Beach, T. G., Bigio, E. H., Cairns, N. J., Carrillo, M. C., Dickson, D. W., Duyckaerts, C., Frosch, M. P., Masliah, E. et al. (2012). National Institute on Aging-Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease. *Alzheimers Dement.* **8**, 1-13.
- Jack, C. R., Jr. and Holtzman, D. M. (2013). Biomarker modeling of Alzheimer's disease. *Neuron* **80**, 1347-1358.
- James, B. D., Wilson, R. S., Boyle, P. A., Trojanowski, J. Q., Bennett, D. A. and Schneider, J. A. (2016). TDP-43 stage, mixed pathologies, and clinical Alzheimer's-type dementia. *Brain* **139**, 2983-2993.

- Janelidze, S., Stomrud, E., Palmqvist, S., Zetterberg, H., van Westen, D., Jeromin, A., Song, L., Hanlon, D., Tan Hehir, C. A., Baker, D. et al. (2016). Plasma beta-amyloid in Alzheimer's disease and vascular disease. *Sci. Rep.* **6**, 26801.
- Jonsson, T., Stefansson, H., Steinberg, S., Jonsdottir, I., Jonsson, P. V., Snaedal, J., Bjornsson, S., Huttenlocher, J., Levey, A. I., Lah, J. J. et al. (2013). Variant of TREM2 associated with the risk of Alzheimer's disease. *N. Engl. J. Med.* **368**, 107-116.
- Kaneko, N., Nakamura, A., Washimi, Y., Kato, T., Sakurai, T., Arahata, Y., Bundo, M., Takeda, A., Niida, S., Ito, K. et al. (2014). Novel plasma biomarker surrogating cerebral amyloid deposition. *Proc. Jpn. Acad. Ser. B Phys. Biol. Sci.* **90**, 353-364.
- Kapaki, E., Paraskevas, G. P., Emmanouilidou, E. and Vekrellis, K. (2013). The diagnostic value of CSF alpha-synuclein in the differential diagnosis of dementia with Lewy bodies vs. normal subjects and patients with Alzheimer's disease. *PLoS ONE* **8**, e81654.
- Kester, M. I., Teunissen, C. E., Crimmins, D. L., Herries, E. M., Ladenson, J. H., Scheltens, P., van der Flier, W. M., Morris, J. C., Holtzman, D. M. and Fagan, A. M. (2015). Neurogranin as a cerebrospinal fluid biomarker for synaptic loss in symptomatic Alzheimer disease. *JAMA Neurol.* **72**, 1275-1280.
- Kikuchi, A., Takeda, A., Okamura, N., Tashiro, M., Hasegawa, T., Furumoto, S., Kobayashi, M., Sugeno, N., Baba, T., Miki, Y. et al. (2010). In vivo visualization of alpha-synuclein deposition by carbon-11-labelled 2-[2-(2-dimethylaminothiazol-5-yl)ethenyl]-6-[2-(fluoro)ethoxy]benzoxazole positron emission tomography in multiple system atrophy. *Brain* **133**, 1772-1778.
- Klunk, W. E., Engler, H., Nordberg, A., Wang, Y., Blomqvist, G., Holt, D. P., Bergström, M., Savitcheva, I., Huang, G.-F., Estrada, S. et al. (2004). Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B. *Ann. Neurol.* **55**, 306-319.
- Kolb, H. C. and Andres, J. I. (2017). Tau positron emission tomography imaging. *Cold Spring Harb. Perspect. Biol.* **9**, a023721.
- Kovacs, G. G. (2016). Molecular pathological classification of neurodegenerative diseases: turning towards precision medicine. *Int. J. Mol. Sci.* **17**, 189.
- Kuhlmann, J., Andreasson, U., Pannee, J., Bjerke, M., Portelius, E., Leinenbach, A., Bittner, T., Korecka, M., Jenkins, R. G., Vanderstichele, H. et al. (2016). CSF Abeta1-42 - an excellent but complicated Alzheimer's biomarker - a route to standardisation. *Clin. Chim. Acta* **467**, 27-33.
- Kvartsberg, H., Duits, F. H., Ingelsson, M., Andreasen, N., Öhrfelt, A., Andersson, K., Brinkmalm, G., Lannfelt, L., Minthon, L., Hansson, O. et al. (2015a). Cerebrospinal fluid levels of the synaptic protein neurogranin correlates with cognitive decline in prodromal Alzheimer's disease. *Alzheimers Dement.* **11**, 1180-1190.
- Kvartsberg, H., Portelius, E., Andreasson, U., Brinkmalm, G., Hellwig, K., Leleental, N., Kornhuber, J., Hansson, O., Minthon, L., Spitzer, P. et al. (2015b). Characterization of the postsynaptic protein neurogranin in paired cerebrospinal fluid and plasma samples from Alzheimer's disease patients and healthy controls. *Alzheimers Res. Ther.* **7**, 40.
- Lagarde, J., Sarazin, M. and Bottlaender, M. (2017). In vivo PET imaging of neuroinflammation in Alzheimer's disease. *J. Neural. Transm. (Vienna)* **125**, 847-867.
- Landqvist Waldö, M., Frizell Santillo, A., Passant, U., Zetterberg, H., Rosengren, L., Nilsson, C. and Englund, E. (2013). Cerebrospinal fluid neurofilament light chain protein levels in subtypes of frontotemporal dementia. *BMC Neurol.* **13**, 54.
- Lashley, T., Holton, J. L., Gray, E., Kirkham, K., O'Sullivan, S. S., Hilbig, A., Wood, N. W., Lees, A. J. and Revesz, T. (2008). Cortical alpha-synuclein load is associated with amyloid-beta plaque burden in a subset of Parkinson's disease patients. *Acta Neuropathol.* **115**, 417-425.
- Lue, L.-F., Schmitz, C. T., Serrano, G., Sue, L. I., Beach, T. G. and Walker, D. G. (2015). TREM2 protein expression changes correlate with Alzheimer's disease neurodegenerative pathologies in post-mortem temporal cortices. *Brain Pathol.* **25**, 469-480.
- Magdalinou, N. K., Paterson, R. W., Schott, J. M., Fox, N. C., Mummery, C., Blennow, K., Bhatia, K., Morris, H. R., Giunti, P., Warner, T. T. et al. (2015). A panel of nine cerebrospinal fluid biomarkers may identify patients with atypical parkinsonian syndromes. *J. Neurol. Neurosurg. Psychiatry* **86**, 1240-1247.
- Marquie, M., Normandin, M. D., Vanderburg, C. R., Costantino, I. M., Bien, E. A., Rycyna, L. G., Klunk, W. E., Mathis, C. A., Ikonomic, M. D., Debnath, M. L. et al. (2015). Validating novel tau positron emission tomography tracer [F-18]-AV-1451 (T807) on postmortem brain tissue. *Ann. Neurol.* **78**, 787-800.
- Masters, C. L., Simms, G., Weinman, N. A., Multhaup, G., McDonald, B. L. and Beyreuther, K. (1985). Amyloid plaque core protein in Alzheimer disease and Down syndrome. *Proc. Natl. Acad. Sci. USA* **82**, 4245-4249.
- Mattsson, N., Tabatabaei, S., Johansson, P., Hansson, O., Andreasson, U., Månsson, J.-E., Johansson, J.-O., Olsson, B., Wallin, A., Svensson, J. et al. (2011). Cerebrospinal fluid microglial markers in Alzheimer's disease: elevated chitotriosidase activity but lack of diagnostic utility. *Neuromolecular Med.* **13**, 151-159.
- Mattsson, N., Schott, J. M., Hardy, J., Turner, M. R. and Zetterberg, H. (2016a). Selective vulnerability in neurodegeneration: insights from clinical variants of Alzheimer's disease. *J. Neurol. Neurosurg. Psychiatry* **87**, 1000-1004.
- Mattsson, N., Zetterberg, H., Janelidze, S., Insel, P. S., Andreasson, U., Stomrud, E., Palmqvist, S., Baker, D., Tan Hehir, C. A., Jeromin, A. et al. (2016b). Plasma tau in Alzheimer disease. *Neurology* **87**, 1827-1835.
- Mattsson, N., Schöll, M., Strandberg, O., Smith, R., Palmqvist, S., Insel, P. S., Hägerström, D., Ohlsson, T., Zetterberg, H., Jögi, J. et al. (2017). (18)F-AV-1451 and CSF T-tau and P-tau as biomarkers in Alzheimer's disease. *EMBO Mol. Med.* **9**, 1212-1223.
- Mollenhauer, B., El-Agnaf, O. M. A., Marcus, K., Trenkwalder, C. and Schlossmacher, M. G. (2010). Quantification of alpha-synuclein in cerebrospinal fluid as a biomarker candidate: review of the literature and considerations for future studies. *Biomark. Med.* **4**, 683-699.
- Mollenhauer, B., Locascio, J. J., Schulz-Schaeffer, W., Sixel-Döring, F., Trenkwalder, C. and Schlossmacher, M. G. (2011). alpha-Synuclein and tau concentrations in cerebrospinal fluid of patients presenting with parkinsonism: a cohort study. *Lancet Neurol.* **10**, 230-240.
- Morbelli, S. and Bauckneht, M. (2018). Amyloid PET imaging: standardization and integration with other Alzheimer's disease biomarkers. *Methods Mol. Biol.* **1750**, 203-212.
- Nakamura, A., Kaneko, N., Villemagne, V. L., Kato, T., Doecke, J., Doré, V., Fowler, C., Li, Q.-X., Martins, R., Rowe, C. et al. (2018). High performance plasma amyloid-beta biomarkers for Alzheimer's disease. *Nature* **554**, 249-254.
- Oeckl, P., Metzger, F., Nagl, M., von Arnim, C. A. F., Halbgebauer, S., Steinacker, P., Ludolph, A. C. and Otto, M. (2016). Alpha-, Beta-, and gamma-synuclein quantification in cerebrospinal fluid by multiple reaction monitoring reveals increased concentrations in Alzheimer's and Creutzfeldt-Jakob disease but no alteration in synucleinopathies. *Mol. Cell. Proteomics* **15**, 3126-3138.
- Öhrfelt, A., Grognet, P., Andreasen, N., Wallin, A., Vanmechelen, E., Blennow, K. and Zetterberg, H. (2009). Cerebrospinal fluid alpha-synuclein in neurodegenerative disorders—a marker of synapse loss? *Neurosci. Lett.* **450**, 332-335.
- Öhrfelt, A., Axelsson, M., Malmeström, C., Novakova, L., Heslegrave, A., Blennow, K., Lycke, J. and Zetterberg, H. (2016). Soluble TREM-2 in cerebrospinal fluid from patients with multiple sclerosis treated with natalizumab or mitoxantrone. *Mult. Scler.* **22**, 1587-1595.
- Olsson, B., Hertz, J., Lautner, R., Zetterberg, H., Nagga, K., Högglund, K., Basun, H., Annas, P., Lannfelt, L., Andreasen, N. et al. (2013). Microglial markers are elevated in the prodromal phase of Alzheimer's disease and vascular dementia. *J. Alzheimers Dis.* **33**, 45-53.
- Olsson, B., Lautner, R., Andreasson, U., Öhrfelt, A., Portelius, E., Bjerke, M., Hölttä, M., Rosén, C., Olsson, C., Strobel, G. et al. (2016). CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. *Lancet Neurol.* **15**, 673-684.
- Ossenkoppele, R., Schonhaut, D. R., Schöll, M., Lockhart, S. N., Ayakta, N., Baker, S. L., O'Neil, J. P., Janabi, M., Lazaris, A., Cantwell, A. et al. (2016). Tau PET patterns mirror clinical and neuroanatomical variability in Alzheimer's disease. *Brain* **139**, 1551-1567.
- Ovod, V., Ramsey, K. N., Mawuenyega, K. G., Bollinger, J. G., Hicks, T., Schneider, T., Sullivan, M., Paumier, K., Holtzman, D. M., Morris, J. C. et al. (2017). Amyloid beta concentrations and stable isotope labeling kinetics of human plasma specific to central nervous system amyloidosis. *Alzheimers Dement.* **13**, 841-849.
- Payoux, P. and Salabert, A. S. (2017). New PET markers for the diagnosis of dementia. *Curr. Opin. Neurol.* **30**, 606-616.
- Peluso, M. J., Valcour, V., Phanuphak, N., Ananworanich, J., Fletcher, J. L. K., Chalermchai, T., Krebs, S. J., Robb, M. L., Hellmuth, J., Gisslén, M. et al. (2017). Immediate initiation of cART is associated with lower levels of cerebrospinal fluid YKL-40, a marker of microglial activation, in HIV-1 infection. *AIDS* **31**, 247-252.
- Piccio, L., Deming, Y., Del-Aguila, J. L., Ghezzi, L., Holtzman, D. M., Fagan, A. M., Fenoglio, C., Galimberti, D., Borroni, B. and Cruchaga, C. (2016). Cerebrospinal fluid soluble TREM2 is higher in Alzheimer disease and associated with mutation status. *Acta Neuropathol.* **131**, 925-933.
- Portelius, E., Zetterberg, H., Skillbäck, T., Törnqvist, U., Andreasson, U., Trojanowski, J. Q., Weiner, M. W., Shaw, L. M., Mattsson, N. and Blennow, K. (2015). Cerebrospinal fluid neurogranin: relation to cognition and neurodegeneration in Alzheimer's disease. *Brain* **138**, 3373-3385.
- Ramont, L., Thoannes, H., Volondat, A., Chastang, F., Millet, M.-C. and Maquart, F.-X. (2005). Effects of hemolysis and storage condition on neuron-specific enolase (NSE) in cerebrospinal fluid and serum: implications in clinical practice. *Clin. Chem. Lab. Med.* **43**, 1215-1217.
- Rathore, S., Habes, M., Iftikhar, M. A., Shacklett, A. and Davatzikos, C. (2017). A review on neuroimaging-based classification studies and associated feature extraction methods for Alzheimer's disease and its prodromal stages. *Neuroimage* **155**, 530-548.

- Represa, A., Deloulme, J. C., Sensenbrenner, M., Ben-Ari, Y. and Baudier, J. (1990). Neurogranin: immunocytochemical localization of a brain-specific protein kinase C substrate. *J. Neurosci.* **10**, 3782-3792.
- Riemschneider, M., Wagenpfeil, S., Vanderstichele, H., Otto, M., Wiltfang, J., Kretzschmar, H., Vanmechelen, E., Förstl, H. and Kurz, A. (2003). Phospho-tau/total tau ratio in cerebrospinal fluid discriminates Creutzfeldt-Jakob disease from other dementias. *Mol. Psychiatry* **8**, 343-347.
- Sander, K., Lashley, T., Gami, P., Gendron, T., Lythgoe, M. F., Rohrer, J. D., Schott, J. M., Revesz, T., Fox, N. C. and Årstad, E. (2016). Characterization of tau positron emission tomography tracer [18F]AV-1451 binding to postmortem tissue in Alzheimer's disease, primary tauopathies, and other dementias. *Alzheimers Dement.* **12**, 1116-1124.
- Schmidt, M. E., Chiao, P., Klein, G., Matthews, D., Thurfjell, L., Cole, P. E., Margolin, R., Landau, S., Foster, N. L., Mason, N. S. et al. (2015). The influence of biological and technical factors on quantitative analysis of amyloid PET: points to consider and recommendations for controlling variability in longitudinal data. *Alzheimers Dement.* **11**, 1050-1068.
- Schneider, J. A., Arvanitakis, Z., Leurgans, S. E. and Bennett, D. A. (2009). The neuropathology of probable Alzheimer disease and mild cognitive impairment. *Ann. Neurol.* **66**, 200-208.
- Schöll, M., Lockhart, S. N., Schonhaut, D. R., O'Neil, J. P., Janabi, M., Ossenkoppele, R., Baker, S. L., Vogel, J. W., Faria, J., Schwimmer, H. D. et al. (2016). PET imaging of Tau deposition in the aging human brain. *Neuron* **89**, 971-982.
- Schott, J. M. and Warren, J. D. (2012). Alzheimer's disease: mimics and chameleons. *Pract. Neurol.* **12**, 358-366.
- Selkoe, D. J. (2002). Alzheimer's disease is a synaptic failure. *Science* **298**, 789-791.
- Seppala, T. T., Nerg, O., Koivisto, A. M., Rummukainen, J., Puli, L., Zetterberg, H., Pyykko, O. T., Helisalmi, S., Alafuzoff, I., Hiltunen, M. et al. (2012). CSF biomarkers for Alzheimer disease correlate with cortical brain biopsy findings. *Neurology* **78**, 1568-1575.
- Shahnawaz, M., Tokuda, T., Waragai, M., Mendez, N., Ishii, R., Trenkwalder, C., Mollenhauer, B. and Soto, C. (2016). Development of a biochemical diagnosis of Parkinson disease by detection of alpha-Synuclein misfolded aggregates in cerebrospinal fluid. *JAMA Neurol.* **74**, 163-172.
- Shi, M., Kovac, A., Korff, A., Cook, T. J., Ginghina, C., Bullock, K. M., Yang, L., Stewart, T., Zheng, D., Aro, P. et al. (2016). CNS tau efflux via exosomes is likely increased in Parkinson's disease but not in Alzheimer's disease. *Alzheimers Dement.* **12**, 1125-1131.
- Simonsen, A. H., Herukka, S.-K., Andreasen, N., Baldeiras, I., Bjerke, M., Blennow, K., Engelborghs, S., Frisoni, G. B., Gabryelewicz, T., Galluzzi, S. et al. (2017). Recommendations for CSF AD biomarkers in the diagnostic evaluation of dementia. *Alzheimers Dement.* **13**, 274-284.
- Sjogren, M., Rosengren, L., Minthon, L., Davidsson, P., Blennow, K. and Wallin, A. (2000). Cytoskeleton proteins in CSF distinguish frontotemporal dementia from AD. *Neurology* **54**, 1960-1964.
- Skillback, T., Farahmand, B., Bartlett, J. W., Rosen, C., Mattsson, N., Nagga, K., Kilander, L., Religa, D., Wimo, A., Winblad, B. et al. (2014). CSF neurofilament light differs in neurodegenerative diseases and predicts severity and survival. *Neurology* **83**, 1945-1953.
- Slaets, S., Vanmechelen, E., Le Bastard, N., Decraemer, H., Vandijck, M., Martin, J.-J., De Deyn, P. P. and Engelborghs, S. (2014). Increased CSF alpha-synuclein levels in Alzheimer's disease: correlation with tau levels. *Alzheimers Dement.* **10**, S290-S298.
- Smailagic, N., Vacante, M., Hyde, C., Martin, S., Ukoumunne, O. and Sachpekidis, C. (2015). (1)(8)F-FDG PET for the early diagnosis of Alzheimer's disease dementia and other dementias in people with mild cognitive impairment (MCI). *Cochrane Database Syst. Rev.* **1**, CD010632.
- Steinacker, P., Blennow, K., Halbgebauer, S., Shi, S., Ruf, V., Oeckl, P., Giese, A., Kuhle, J., Slivarichova, D., Zetterberg, H. et al. (2016). Neurofilaments in blood and CSF for diagnosis and prediction of onset in Creutzfeldt-Jakob disease. *Sci. Rep.* **6**, 38737.
- Strimbu, K. and Tavel, J. A. (2010). What are biomarkers? *Curr. Opin. HIV/AIDS* **5**, 463-466.
- Suárez-Calvet, M., Kleinberger, G., Araque Caballero, M. Á., Brendel, M., Rominger, A., Alcolea, D., Fortea, J., Lleó, A., Blesa, R., Gisbert, J. D. et al. (2016). sTREM2 cerebrospinal fluid levels are a potential biomarker for microglia activity in early-stage Alzheimer's disease and associate with neuronal injury markers. *EMBO Mol. Med.* **8**, 466-476.
- Takahashi, K., Rochford, C. D. P. and Neumann, H. (2005). Clearance of apoptotic neurons without inflammation by microglial triggering receptor expressed on myeloid cells-2. *J. Exp. Med.* **201**, 647-657.
- Tarawneh, R., D'Angelo, G., Crimmins, D., Herries, E., Griest, T., Fagan, A. M., Zipfel, G. J., Ladenson, J. H., Morris, J. C. and Holtzman, D. M. (2016). Diagnostic and prognostic utility of the synaptic marker neurogranin in Alzheimer disease. *JAMA Neurol.* **73**, 561-571.
- Tateno, F., Sakakibara, R., Kawai, T., Kishi, M. and Murano, T. (2012). Alpha-synuclein in the cerebrospinal fluid differentiates synucleinopathies (Parkinson Disease, dementia with Lewy bodies, multiple system atrophy) from Alzheimer disease. *Alzheimer Dis. Assoc. Disord.* **26**, 213-216.
- Thorsell, A., Bjerke, M., Gobom, J., Brunhage, E., Vanmechelen, E., Andreasen, N., Hansson, O., Minthon, L., Zetterberg, H. and Blennow, K. (2010). Neurogranin in cerebrospinal fluid as a marker of synaptic degeneration in Alzheimer's disease. *Brain Res.* **1362**, 13-22.
- Tokuda, T., Qureshi, M. M., Ardah, M. T., Varghese, S., Shehab, S. A. S., Kasai, T., Ishigami, N., Tamaoka, A., Nakagawa, M. and El-Agnaf, O. M. A. (2010). Detection of elevated levels of alpha-synuclein oligomers in CSF from patients with Parkinson disease. *Neurology* **75**, 1766-1772.
- Uchihara, T. and Giasson, B. I. (2016). Propagation of alpha-synuclein pathology: hypotheses, discoveries, and yet unresolved questions from experimental and human brain studies. *Acta Neuropathol.* **131**, 49-73.
- van Eijk, J. J. J., van Everbroeck, B., Abdo, W. F., Kremer, B. P. H. and Verbeek, M. M. (2010). CSF neurofilament proteins levels are elevated in sporadic Creutzfeldt-Jakob disease. *J. Alzheimers Dis.* **21**, 569-576.
- Voyle, N., Baker, D., Burnham, S. C., Covin, A., Zhang, Z., Sangurdekar, D. P., Tan Hehir, C. A., Bazenet, C., Lovestone, S., Kiddle, S. et al. (2015). Blood protein markers of neocortical amyloid-beta burden: a candidate study using SOMAscan technology. *J. Alzheimers Dis.* **46**, 947-961.
- Walker, L. C. and Jucker, M. (2015). Neurodegenerative diseases: expanding the prion concept. *Annu. Rev. Neurosci.* **38**, 87-103.
- Wallin, A. K., Blennow, K., Zetterberg, H., Londos, E., Minthon, L. and Hansson, O. (2010). CSF biomarkers predict a more malignant outcome in Alzheimer disease. *Neurology* **74**, 1531-1537.
- Watabe-Rudolph, M., Song, Z., Lausser, L., Schnack, C., Begus-Nahrman, Y., Scheithauer, M.-O., Rettinger, G., Otto, M., Tumani, H., Thal, D. R. et al. (2012). Chitinase enzyme activity in CSF is a powerful biomarker of Alzheimer disease. *Neurology* **78**, 569-577.
- Wellington, H., Paterson, R. W., Portelius, E., Törnqvist, U., Magdalinou, N., Fox, N. C., Blennow, K., Schott, J. M. and Zetterberg, H. (2016). Increased CSF neurogranin concentration is specific to Alzheimer disease. *Neurology* **86**, 829-835.
- Wennström, M., Surova, Y., Hall, S., Nilsson, C., Minthon, L., Boström, F., Hansson, O. and Nielsen, H. M. (2013). Low CSF levels of both alpha-synuclein and the alpha-synuclein cleaving enzyme neurosin in patients with synucleinopathy. *PLoS ONE* **8**, e53250.
- Westwood, S., Leoni, E., Hye, A., Lynham, S., Khondoker, M. R., Ashton, N. J., Kiddle, S. J., Baird, A. L., Sainz-Fuertes, R., Leung, R. et al. (2016). Blood-based biomarker candidates of cerebral amyloid using PIB PET in non-demented elderly. *J. Alzheimers Dis.* **52**, 561-572.
- Winblad, B., Amouyel, P., Andrieu, S., Ballard, C., Brayne, C., Brodaty, H., Cedazo-Minguez, A., Dubois, B., Edvardsson, D., Feldman, H. et al. (2016). Defeating Alzheimer's disease and other dementias: a priority for European science and society. *Lancet Neurol.* **15**, 455-532.
- Winston, C. N., Goetzl, E. J., Akers, J. C., Carter, B. S., Rockenstein, E. M., Galasko, D., Masliah, E. and Rissman, R. A. (2016). Prediction of conversion from mild cognitive impairment to dementia with neuronally derived blood exosome protein profile. *Alzheimers Dement.* **3**, 63-72.
- Yin, G. N., Jeon, H., Lee, S., Lee, H. W., Cho, J.-Y. and Suk, K. (2009). Role of soluble CD14 in cerebrospinal fluid as a regulator of glial functions. *J. Neurosci. Res.* **87**, 2578-2590.
- Zetterberg, H. (2015). Plasma amyloid beta-quo vadis? *Neurobiol. Aging* **36**, 2671-2673.
- Zetterberg, H. (2016). Neurofilament light: a dynamic cross-disease fluid biomarker for neurodegeneration. *Neuron* **91**, 1-3.
- Zetterberg, H. (2017). Review: Tau in biofluids - relation to pathology, imaging and clinical features. *Neuropathol. Appl. Neurobiol.* **43**, 194-199.
- Zetterberg, H., Wilson, D., Andreasson, U., Minthon, L., Blennow, K., Randall, J. and Hansson, O. (2013). Plasma tau levels in Alzheimer's disease. *Alzheimers Res. Ther.* **5**, 9.
- Zetterberg, H., Skillbäck, T., Mattsson, N., Trojanowski, J. Q., Portelius, E., Shaw, L. M., Weiner, M. W. and Blennow, K. (2016). Association of cerebrospinal fluid neurofilament light concentration with Alzheimer disease progression. *JAMA Neurol.* **73**, 60-67.