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3. Article title: **Physical Properties and Biocompatibility Effects of Doping SiO₂ and TiO₂ into Phosphate-Based Glass for Bone Tissue Engineering**
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5. Abstract

Abstract

Phosphate glass is gaining more attention to serve as a bone substitute. More specifically, the ternary (P_2O_5 -CaO- Na_2O) was investigated in terms of both physical properties and biocompatibility by doping different percentages of SiO_2 and TiO_2 . Disks from two groups of different compositions of the ternary (P_2O_5 -CaO- Na_2O) were prepared. The first group has different percentages of TiO_2 and SiO_2 , whereas the second group compositions has 5mol% TiO_2 and 5mol% SiO_2 added to compensate the network-forming oxide P_2O_5 and the network-modifying oxide CaO.

Density, degradation, DTA, XRD, and cation release experiments were performed to study the physicochemical properties of the compositions, while MG63 and hMSC cells were used within *in vitro* cell culture studies to study their biocompatibility. Results showed that a decrease in SiO_2 content directly correlated with an increase in glass density, decreased degradation, increased trend of T_g and T_m values, and Na^+ and Ca^{2+} release in group 1. However, there was no improvement in the MG63 viability or the ability of hMSC's to differentiate into osteoblasts where TiO_2 was replaced with SiO_2 . Furthermore, in group 2, 50 P_2O_5 -25CaO was less dense than 45 P_2O_5 -30CaO, degraded dramatically less, had lower T_g and T_m values and released less Na^+ and Ca^{2+} . The synergistic effect of doping 5mol% TiO_2 and 5mol% SiO_2 increased the MG63 viability in both compositions and was found 45 P_2O_5 -30CaO to have superior results in terms of the ability of hMSC's to differentiate into osteoblasts. To conclude, substituting TiO_2 in place of SiO_2 improved the physical properties and the

biocompatibility of (P₂O₅-CaO-Na₂O) glass system whereas doping 5mol% SiO₂ and 5mol% TiO₂ together in place of P₂O₅ and CaO had a synergistic effect in controlling their degradation rate and improving their biological responses.

Key words: Tissue Engineering, Bone Augmentation, Synthetic Bone, Phosphate-based Glass, Silicate-based Glasses.

1. Introduction.

Regenerative medicine is focused on developing materials that can help clinicians to manage hard tissue loss. This is vital to patients suffering from trauma, neoplasms and congenital abnormalities, which can be associated with significant scarring and deformation with potential psychological impairments. The concept of hard tissue augmentation went through different phases of techniques and materials until formulating the concept used today.

Autogenous graft is still considered to be the gold standard of bone grafting materials (1). However, different synthetic materials have been studied and used extensively in research and later in clinical environments such as silicate-based glasses, phosphate and borosilicate based glasses and hydroxyapatite. Silicate based glasses or the now famous 45S5® Bioglass were introduced by Larry Hench in the late 1960s as a revolution in the development of biomaterials and bone tissue engineering (1). Its bioactive properties made it a potential material to be applied clinically for bone defect repair (2). Its favourable properties granted it great attention in tissue engineering research as it is biodegradable, has the ability to drive the differentiation of hMSC's and their proliferation into osteoblasts (1, 3-5). Phosphate glasses, in turn, are also considered good candidate materials for both bone tissue engineering and also can be antimicrobial (6, 7). This can be attributed to two main factors; the controllability of its degradation and the similarity of its inorganic composition, such as P, Na and Ca, to the human bone (8, 9). Although it has a hygroscopic nature, it has been modified by doping with many metal oxides such as TiO₂, GeO₂ and B₂O₃ (6, 10). An example of such glass is the ternary (P₂O₅– CaO – Na₂O) which was developed and modified to control its degradation rate. Adding TiO₂ might have contributed to

increasing the covalent nature of the bonds within the glass structure as well as playing a major role in attracting osteoblasts and improving their attachment and proliferation (6). This attracted more attention to this ternary system as a potential material for bone engineering application.

The present paper focuses on doping SiO₂ and TiO₂ into phosphate-based glass (P₂O₅-CaO-Na₂O). The main goal is to control their degradation rate and enhance their biological response to be suitable synthetic materials that can be applied in bone augmentation. The concept behind the choice of SiO₂ to serve as an additive oxide to phosphate-based glass (P₂O₅-CaO-Na₂O-TiO₂) was due to the biocompatibility of silicate-based glasses as shown in many previous studies, and the good results obtained from doping Mg and Al in similar researches (11, 12).

As Si shares a position on the third row of the periodic table with Mg, Al and Na, it was suggested to investigate doping it into quaternary based-glass. Three different aspects are discussed in this paper, which include the production of different phosphate glass compositions, their physicochemical properties, and their biocompatibility reaction within an *in vitro* environment using human osteoblast-like osteosarcoma cells (MG63) and human mesenchymal stem cells (hMSC's).

2. Materials and Methods.

2.1. Materials Preparation

Eight different glass rods of 15mm diameter were prepared using eight different compositions of sodium dihydrogen phosphate NaH₂PO₄, calcium carbonate CaCO₃, phosphorus pentoxide P₂O₅, titanium dioxide TiO₂, and silicon dioxide SiO₂ (BDH, Poole, UK, all chemicals were >98% purity) as shown in table 1.

The first group has six compositions with similar core components (50% P₂O₅-30% CaO-15% Na₂O) mol% though different TiO₂ to SiO₂ percentage, starting from (5% TiO₂ / 0% SiO₂) and gradually replacing TiO₂ with SiO₂ to reach (0% TiO₂ / 5% SiO₂). The second group has two compositions both contain (5% TiO₂ / 5% SiO₂) with different percentages; (45% P₂O₅-30% CaO-15% Na₂O-5% TiO₂-5% SiO₂) and (50% P₂O₅-25% CaO-15% Na₂O-5% TiO₂-5% SiO₂). The precursors were mixed thoroughly (Stomacher 400 Circulator, Seward, Worthing, UK) and then placed in a 200 ml crucible Pt/10%Rh type 71040 (Johnson Matthey, Royston, UK). The conventional melt quenching technique was used to prepare the glass rods.

Each composition was prepared and mixed, and then heated in two stages. The first cycle was at 750°C for 30 minutes aiming to eliminate CO₂ and H₂O, whereas the second cycle was at 1350°C for four hours to ensure chemical homogeneity. The melt then was poured into a 15mm diameter mold and left to cool down gradually at 250°C for 30 minutes and the furnace was then switched off and the mould and glass allowed to cool down over the next 24 hours in order to remove residual stress. Then, each rod was sectioned into approximately 2mm thick discs using a Testbourne diamond saw with ethanol as a coolant. Discs from each composition were subjected to a series of grinding steps on two sides using a series of waterproof silicon carbide papers: P no. 120 for 30 seconds at 300RPM to flatten the surface, then P no. 500, 1000, and 2400, respectively, for 1min at 500RPM to smoothen the surfaces, and finally P no. 4000 for 2 min at 500RPM to get a smooth mirror-finish surface (Struers, UK).

Table 1 the amounts of precursors used in the eight different compositions.

2.2. X-ray Powder Diffraction (XRD)

This rapid analytical technique was applied to ensure the glasses were fully amorphous and was measured by X-ray diffractometer (D8 Advance Diffractometer, Bruker, Coventry, UK). The specimens were finely ground for 10 minutes and then positioned in flat plate geometry and (Ni-filtered Cu K α) radiation was applied. Data were collected using a Lynx Eye detector with a step size of 0.02 over an angular range of 10–100° 2 θ and a count time of 12 seconds.

2.3. Density

An analytical balance and density determination apparatus with respect to Archimedes' principle were applied (Mettler Toledo, UK). Triplicate samples of each composition were used to calculate the density using the equation below:

$$\rho_{glass} = \frac{mass\ in\ air\ (g)}{mass\ in\ air - mass\ in\ ethanol} \times \rho_{ethanol}$$

where $\rho_{ethanol}$ is the density of ethanol in g.cm⁻³ at the temperature of which the measurements of m_{air} in g and $m_{ethanol}$ in g were performed. In our experiment the temperature was 20°C and ethanol used was 99.8% (Sigma Aldrich, Gillingham, UK) hence $\rho_{ethanol} = 0.781596$ g.cm⁻³.

2.4. Differential Thermal Analysis

Setaram differential thermal analyser was used to perform differential thermal analysis (DTA) studies (Setaram, France). Approximately 60mg of glass powder (discs were grinded and the size of the particles varies between 30 μ m-150 μ m) were placed in a 100 μ l platinum crucible prior to placing it in the analyser. The sample was then heated from room temperature to 1000°C at a heating rate of 20°C per minute using air as the purge gas. An empty platinum crucible was used as a reference. The parameters measured by DTA were glass transition temperature (T_g), crystallisation temperature (T_c) and melting temperature (T_m).

2.5. Degradation

Three discs from each composition were weighed and then stored in 5 ml deionized water (pH 7.0 \pm 0.1) for 1, 4, 7 and 14 days. The ultrapure water was chosen parallel to the ion release experiment. This followed the protocol of previous studies. At each time point, the glass discs were taken out of the water, dried manually, weighed and then transferred into a fresh 5 ml of distilled water. In between the time points, samples were stored in a 37°C incubator. Mass change of each glass disc was calculated using the equation below:

$$\Delta m = \frac{m_0 - m_t}{A}$$

Where m_0 is the initial mass (mg) of the disc on day 0, m_t is the mass (g) of the disc at specific time point and A is the surface area (cm^2) of the glass disc. In our experiment $r(\text{disc}) = 0.75$ cm, hence $A = 2\pi rh + 2\pi r^2$ which is equal to about 4.45 cm^2 .

2.6. pH Changes

Three discs from each composition were stored in 1 ml Dulbecco Modified Eagles Medium (DMEM) (Gibco®, Life Technologies Ltd., Paisley, UK), pH 8.4 ± 0.1 , for 1, 4 and 7 days. At each time point, the pH of the specimen extract was measured using pH detector; Orion Star pH Meter (Thermo Scientific, UK). The culture media was changed every 3 days to mimic the environment of the cell culture study. Samples were stored in a 37°C/5% CO₂ incubator in between time points. The culture media without discs was used as control.

2.7. Ion Chromatography

The solutions collected above for the degradation measurements were collected for ion determination. The ultrapure water was chosen to eliminate the presence of any background readings might be created if any buffered solutions (e.g simulated body fluid) was used. This helped to focus on the ions of interest only. Due to the high levels of ions such as Ca²⁺, Na⁺ and P⁵⁺ in buffer solutions precluding the determination of the relatively low levels of ion release from the glass, the measurements were performed in ultrapure water. For cation determination, as the separator column possesses high sensitivity to phosphate species, all samples were filtered with OnGaurd IIA filters (Dionex, Thermo Scientific, Hemel Hempstead, UK). Prior to performing the experiment, the system was calibrated against a five-point calibration curve using a previously developed routine before carrying out the sample run for each time point. Na⁺ and Ca²⁺ ion release measurements were performed on a Dionex ICS-1000 ion chromatography system (Dionex, Surrey, UK) using 30 mM methanesulphonic acid (Fluka, Dorset, UK) incorporating a 4 x 250 mm Ion Pac® CS12A separator column and a sample loop of 25 µl.

Sodium chloride (Sigma–Aldrich, Dorset, UK) and calcium chloride (BDH, Poole, UK) were used as reagents to prepare a 100-ppm stock solution before serially diluting both to produce 50, 25, 10 and 5-ppm standard solutions

2.8. In *Vitro* Cytocompatibility Assessment

Samples for the biocompatibility test were cleaned with ethanol 98% and then sterilised on both sides using ultraviolet light for 10 minutes.

2.8.1. Human Osteoblast-Like Osteosarcoma Cells (MG-63)

Human osteoblast-like osteosarcoma cells MG63 were obtained from in-house stocks at the Eastman Dental Institute, University College London. Two different assays were applied; Alamar Blue and Live/Dead assay with confocal laser scanning microscopy. Cells were prepared and then seeded calculating a density of 3000 cells per cm^2 in 1 ml of medium. Then, three discs from each composition were placed in a conventional 24 well tissue culture test plate (Transwell® polyester inserts, Corning, USA) with one disc per well. Seeding of cells on the discs was carried out calculating 5300 cells in each well.

2.8.2. Human Mesenchymal Stem Cells (hMSc)

Human mesenchymal stem cells were obtained from in-house stocks at the Eastman Dental Institute, University College London. Two different assays were used which are alkaline phosphatase and cell counting Kit 8 (CCK). Cells were prepared and then seeded calculating a density of 3000 cells per cm^2 of 1 ml medium. Then, three discs from each composition were placed in a conventional 24 well tissue culture test plate (Transwell®

polyester inserts, Corning, USA) as in one disc per well. Seeding of cells on the discs was carried out calculating about 5000 cells in each well.

2.7.3 Statistical Analyses

All statistical analyses were performed using Microsoft Excel 2013 and IBM SPSS version 22 (SPSS Inc., USA). Initial normality tests showed that the data of the cell culture studies were not normally distributed; hence they were restructured using the square root to apply normal distribution tests. Consequently, Bonferroni correction method and Bunnnet test were applied. The assumed level of significance for the statistical tests was 0.05.

3. Results

3.1 X-ray powder Diffraction (XRD)

XRD patterns of the glass were measured. The broad peak confirmed the amorphous nature of all glass samples in both groups and indicated the absence of crystalline phases as shown in figure 1.

Figure 1 the pattern of the samples.

3.2 Density

The results presented in table 2 ranged from 2.54g/cm^3 to 2.59g/cm^3 in both groups. It was calculated that an increase in TiO_2 content correlated with an increase in glass density in group 1. In parallel, $45\text{P}_2\text{O}_5\text{-}30\text{CaO}$ was relatively denser than $50\text{P}_2\text{O}_5\text{-}25\text{CaO}$ in group 2 with densities ($\rho = 2.58 \mp 0.016$ and $\rho = 2.55 \mp 0.003 \text{ g/cm}^3$) respectively.

3.3 Differential Thermal Analysis (DTA)

DTA studies were performed on glass powder. Table 2 shows that the T_g of the glasses was in the range of 421–481°C while T_m was between 778–878°C with a decreased trend observed as SiO_2 the content was increased in both groups. T_c , in turn, was in the range of 681–777°C with no specific trend noted.

Table 2 density (g/cm^3) and different temperature values T_g , T_c and T_m of all samples.

3.4 Degradation (Mass changes)

Mass changes of each glass disc were calculated at four time-points as shown in figure 2. After 14 days, in group 1 Ti0Si5 had the highest mass loss ($m= 6.86 \pm 0.99$) $\text{mg}\cdot\text{cm}^{-2}$ followed by Ti1Si4, Ti2Si3, Ti3Si2, Ti4Si1 and Ti5Si0 respectively, whereas in group 2, 45P₂O₅-30CaO had higher mass loss ($m= 6.35 \pm 0.41$ $\text{mg}\cdot\text{cm}^{-2}$) than 50P₂O₅-25CaO. A comparison between the two groups showed that Ti0Si5 had the highest mass loss while Ti5Si0, Ti4Si1 and 50P₂O₅-25CaO were the least to be degraded with similar results ($m= 0.77 \pm 0.06$, $m= 0.98 \pm 0.13$, $m= 1.39 \pm 0.26$) mg/cm^2 . This indeed might have an impact on ion release and in *in vitro* culture studies.

Figure 2 the trend of mean averages with standard deviations of mass loss on day 1, 4, 7 and 14.

3.5 pH Changes

As the culture media was changed at every time point within *in vitro* atmosphere, a similar approach was applied to calculate the change in pH. The culture media was changed after each time point to mimic the environment of the cell culture studies.

All pH values recorded were alkaline as presented in table 3. While the highest pH recorded for control was on day 4 ($m=9.38 \mp 0.24$), the highest pH recorded for a composition was for 45 P₂O₅-30CaO on day 4 ($m= 9.74 \mp 0.17$). The lowest pH recorded for control was on day 1

($m=8.38 \mp 0.08$), whereas the lowest pH recorded for a composition was for Ti0Si5 on day 1 ($m=8.37 \mp 0.22$).

Table 3 the mean averages of pH on day 1, 4 and 7.

3.6 Ion Chromatography

All compositions released Na^+ and Ca^{2+} over the period of 7 days in a similar trend as shown in figure 3 and 4. In group 1, Ti0Si5 and Ti2Si3 released more Na^+ and Ca^{2+} than the rest, whereas 45P₂O₅-30CaO released more Na^+ and Ca^{2+} than 50P₂O₅-25CaO composition.

Figure 3 the mean average and standard deviations of Na^+ cations release on day 1, 4, 7.

Figure 4 the mean average and standard deviations of Ca^{2+} cations release on day 1, 4, 7.

3.7 In *Vitro* Cytocompatibility Assessment

3.7.1 Alamar Blue Assay

Results of alamar blue showed that for the same composition, cells in all samples underwent proliferation up to 7 days (Figure 5). While on day 7 the control group had ($n= 37127 \pm 3672$) cells, it was noticed that Ti5Si0 had the highest number of cells in group 1 $m \approx 33043.10$ cells. Other compositions were in between with Ti3Si2 and Ti4Si1 having higher cell numbers than Ti2Si3 and Ti1Si4. Whereas in group 2, both 50P₂O₅-25CaO and 45P₂O₅-30CaO had higher number of cells compared to the samples in group 1 ($m= 34855$, and 33547 cells respectively) and

were similar to Ti5Si0.

Figure 5 the mean average and standard deviations of alamar blue assay on day 1, 4, 7.

* The mean difference is significant at the 0.05 level, 95% Confidence Interval.

3.7.2 Live/Dead Assay

Confocal Laser Scanning Microscopy (CLSM) images of MG63 cells attached to all samples were performed using live/dead stain on day 1, 4 and 7 post-culture. This is subjective and the main reason of choosing such an assay was to determine the viability of MG63 cells based on plasma membrane integrity and esterase activity (green for live cells/ red for dead cells). All compositions showed relatively good results at all three time-points as shown in figure 6.

Figure 6 CLSM images of MG63 cells attached to different composition 1,4 &7 days post culture under 20X-wet lens.

3.7.3 Alkaline Phosphatase Assay

Results of alkaline phosphatase presented in figure 7 showed that the control group recorded $m = 0.343 \pm 0.107$ after 7 days. In group 1, Ti0Si5 and Ti1Si4 had significantly lower ALP values than the rest of the compositions ($m = 0.163 \pm 0.038$, 0.220 ± 0.053) respectively; while in group 2 both 50P₂O₅-25CaO and 45P₂O₅-30CaO had similar ALP values to the control group. On day 14, the situation had changed as Ti0Si5 and Ti1Si4 had similar ALP values to the control group however Ti5Si0, Ti4Si1, Ti3Si2 and Ti2Si3 had superior ALP values in comparison with both the control and the other compositions. In group 2, both 50P₂O₅-25CaO and 45P₂O₅-

30CaO had similar ALP values to the control group on day 7, however, on day 14 50P₂O₅-25CaO had significantly higher ALP values than both control and 45P₂O₅-30CaO.

Figure 7 the mean average and standard deviations of Alkaline Phosphatase on day 7 & 14.

*The mean difference is significant at the 0.05 level, 95% Confidence Interval.

3.7.4 CCK Assay

Results of the CCK presented in figure 8 showed that all samples had induced cell proliferation. While on day 1, Ti5Si0 and Ti3Si2 had a similar number of cells to the control group and higher compared to the rest of the compositions in group 1. This however had changed on day 4 and 7 as all samples had similar results and were lower than the control group. In group 2, both compositions had lower numbers of cells in comparison with the control group with 50P₂O₅-25CaO having lower number of cells than 45P₂O₅-30CaO though the difference was insignificant to the rest of the compositions.

Figure 8 the mean average and standard deviations of CCK assay over 1, 4 and 7 days. *The mean difference is significant at the 0.05 level, 95% Confidence Interval.

4. Discussion

4.1 Composition Selection

This paper focused on studying the effect of doping TiO₂ and SiO₂ into the ternary (P₂O₅-CaO -Na₂O). While many studies (13-15) focused on adding different metallic oxides such as

TiO₂, Fe₂O₃ and SrO, into (P₂O₅ -CaO -Na₂O) glass system, this paper tried to combine doping two different oxides; metallic and nonmetallic. While compositions in group 1 were prepared to investigate the effect of adding different percentages of SiO₂ and TiO₂ to the system (50%P₂O₅-30%CaO -15%Na₂O), group 2 compositions were also studied to give a better understanding of this effect knowing the fact that silicate-based glass is more stable than the phosphate based version. Therefore, 5%SiO₂ and 5%TiO₂ were added into (50%P₂O₅-30%CaO-15%Na₂O) compensating the network-forming oxide in P₂O₅ (45%P₂O₅-30%CaO-15%Na₂O-5%TiO₂-5%SiO₂) and the network-modifying oxide in Ca (50%P₂O₅-25%CaO-15%Na₂O-5%TiO₂-5%SiO₂). The main aim was to create a more complex system, increasing the glass stability and improving the cytocompatibility.

4.2 X-ray Powder Diffraction (XRD)

X-ray diffraction was used to provide information about the structure of all compositions. Our results showed that all patterns had broad peaks, which confirmed the amorphous nature of all glass samples and indicated that they lacked a crystalline phase.

4.3 Density

In terms of group 1 compositions, their core components were the same (50%P₂O₅-30%CaO -15%Na₂O) with different SiO₂ to TiO₂ ratios. Results showed that an increase in TiO₂ content correlated with an increase in glass density. This was due to the fact that the relative atomic weight of titanium (47.867) is greater than silicon (28.085), hence Ti has a larger sample mass, whereas the atomic radius of silicon (117.6pm) is smaller than titanium (176pm); hence Si

has a reduced inter-atomic space. Both effects jointly led to an increase in glass density with the increase in TiO₂ content.

In terms of group 2 compositions, their core components were also the same (15%Na₂O-5%TiO₂-5%SiO₂) with different P₂O₅ to CaO ratios. Results showed that 45P₂O₅-30CaO ($\rho = 2.58 \text{ g.cm}^{-3} \pm 0.016$) was slightly denser than 50P₂O₅-25CaO ($\rho = 2.55 \text{ g.cm}^{-3} \pm 0.003$), which did not follow the trend of the relative atomic weight and the atomic radius described previously. For example, the relative atomic weight of calcium (40.078) is larger than phosphorus (30.973), whereas the atomic radius of phosphorus (98pm) is slightly smaller than calcium (118pm) and hence 50P₂O₅-25CaO should be denser than 45P₂O₅-30CaO; however, results were the opposite. The justification of such results may be attributed to the fact that the density may be controlled not only by the density of the element but also the bond lengths (16).

4.4 Differential Thermal Analysis (DTA)

DTA trends were obtained followed by identification of T_g, T_c and T_m values. Group 1 compositions had two changing variables; TiO₂ and SiO₂. Knowing the fact that the bond enthalpy of Ti–O (672 kcal/mol) is greater than Si–O (452 kcal/mol) may explain the decreased trend observed as the TiO₂ content was decreased and SiO₂ content was increased. On the other hand, group 2 compositions had also two changing variables; P₂O₅ and CaO. As Ca–O has a stronger ionic bond compared to the Van Der Waals bond between P=O....P=O, P₂O₅ needed higher energy to break these bonds.

Furthermore, the trend of T_m values was similar to those of T_g . This was due to the high melting point of titanium (1668°C) in comparison with other components. Consequently, the less TiO_2 , the lower the melting point. Similarly, as 45 P_2O_5 -30CaO contained more calcium than 50 P_2O_5 -25CaO, its T_m was higher. This was also due to the stronger ionic bond in Ca–O in comparison with the Van Der Waals bond between P atoms. On the other hand, T_c readings did not follow any specific trend. This might be related to the differences in particle size (17).

4.5 Degradation

One of the main aims of doping SiO_2 and TiO_2 into phosphate glasses was to control their degradation. As glass degradation behaviour is closely linked to glass chemistry, alteration of the glass composition may allow a better control of the degradation rate over time (6). Results showed that degradation increased with decreasing titanium oxide and increasing silicon oxide in group 1 compositions. Therefore, TiO_5Si_5 degraded the most in comparison to other compositions. Ti_1Si_4 , however, degraded slowly on day 1 before catching up with the rest of the compositions. These results concurred with previous studies (18).

Furthermore, results of group 2 compositions showed that 50 P_2O_5 -25CaO degraded dramatically less than 45 P_2O_5 -30CaO. Such results may be related to the fact that the latter has a lower percentage of network-forming oxide, but there is a possible more complex interaction at play. Although many studies indicated that solubility decreases with decreasing P_2O_5 and increasing CaO (19), this research has doped two different oxides in favour of the network-forming and the network modifying oxides. This might have alter the result suggesting new concept to be investigated in the future.

4.6 pH Changes

Although previous papers have used deionized water to detect pH changes, this paper used 1 ml of Dulbecco Modified Eagles Medium (DMEM), pH 8.4 ± 0.1 , as an immersion liquid. The main idea of this approach was to copy the conditions arranged for the cell culture study. After each time-point, the culture media for all glass discs was changed as well as triplicate of DMEM to act as a control. Samples were stored in a $37^{\circ}\text{C}/5\%$ CO_2 incubator in-between time-points. Our results concluded that the solutions obtained from all samples were found to be alkaline. In compare to control group, all samples had an increased pH values at all time points with the highest scores recorded on day 4 for all of them. The trend obtained from the sample values was similar to the one obtained from the control values.

4.6 Ion Chromatography

Results showed that all sample released Na^+ and Ca^{2+} cations over the period of 7 days. Complying with the results of degradation, TiO_2Si_5 and $45\text{P}_2\text{O}_5\text{-}30\text{CaO}$ released more Na^+ than the rest as they degraded the most, whereas Ti_5Si_0 and Ti_4Si_1 released the least as they were the less degraded samples. Similarly, TiO_2Si_5 and $45\text{P}_2\text{O}_5\text{-}30\text{CaO}$ released the most Ca^{2+} , while Ti_5Si_0 and Ti_4Si_1 released the least. Degradation and ion release proportionality was discussed thoroughly in the literature with all in agreement that the more degraded the sample is, the higher its ability to release ions.

4.7 In *Vitro* Biocompatibility Assessment

4.7.1 Alamar Blue Assay

This assay was used to investigate the interactions between cells and glass discs of all samples and to identify compositions that would elicit the most favourable cellular responses. Consequently, using different known numbers of cells with their alamar blue readings, an equation was derived to count the number of cells indirectly using the alamar blue readings obtained.

In compare to control, results of group 1 compositions showed that the alamar blue readings increased with decreasing SiO₂ and increasing TiO₂ after 7 days. Therefore, on day 7 Ti₅Si₀ had the highest values within this group (n=33043, P value = 0.205) while Ti₀Si₅ had the lowest (n=9626, P value=0.211). It is worth mentioning that Ti₃Si₂ had a high alamar blue reading on day 1 (n= 8492, P value= 0.377) in comparison to the control, however, it did not continue at the same rate until day 7 (n=26761, P value= 0.245). On the other hand, group 2 results showed that both compositions had superior alamar blue readings over group 1 compositions at all three time-points. On day 7, both 50P₂O₅-25CaO (n= 34855, P value = 0.829) and 45P₂O₅-30CaO (n= 33547, P value =0.333) had the closest results to control.

Relying on the aforementioned, doping TiO₂ into the glass composition showed to have potential effects on the cellular biocompatibility as previously seen (17). This could be related to the ability of TiO₂ to consolidate the glass structure and eventually to yield in decreasing the rate of degradation and consequently to enhance the cellular metabolic response as was founded in previous studies (20). SiO₂ did not improve the cellular responses when compensated with TiO₂ in group 1 compositions, and hence doping with SiO₂ instead of TiO₂ did not have a positive

effect. However, doping both 5%SiO₂ and 5%TiO₂ into group 2 compositions (45P₂O₅-30CaO and 50P₂O₅-25CaO) had a positive effect in increasing cell number.

4.7.2 Live/Dead Assay

The principle of the Live/Dead assay is that membrane-permeable calcein-AM is cleaved by esterases in live cells to yield cytoplasmic green fluorescence, and membrane-impermeable ethidium homodimer-1 labels nucleic acids of membrane-compromised cells with red fluorescence. Consequently, live and dead cells display green and red fluorescence respectively. All samples showed promising results in terms of keeping the MG63 cells alive after 7 days.

4.7.3 Alkaline Phosphatase Assay

The aim of using this assay was to investigate the ALP values for all compositions. Using different dilutions of the alkaline phosphatase assay kit (Sensolyte[®] pNPP), an equation was derived to measure the ALP concentration for each composition based on the fluorescence signal obtained.

While on day 7 Ti5Si0, Ti1Si4, Ti3Si2 and Ti4Si1 had significant lower levels of proliferation compared to the other compositions (P value < 0.05); the situation was different on day 14. Ti5Si0, Ti4Si1, Ti3Si2, Ti2Si3 and 45P₂O₅-30CaO had higher ALP concentration (P value < 0.05) than control group as well as the rest compositions. This significant difference in alkaline phosphatase concentrations between day 7 and day 14 may be due to the fact that the expression of this enzyme is usually highest between day 5 to day 14 during the matrix maturation stage of stem cell differentiation process (21). Although Ti1Si4, Ti0Si5 and 50P₂O₅-25CaO had

lower values compared to the other compositions in the whole study, their results can also be considered promising as they were similar to control.

Although substituting TiO_2 with SiO_2 did not improve the ability of hMSC's to differentiate into osteoblasts in group 1, it did not cause cytotoxicity. However, doping both 5% SiO_2 and 5% TiO_2 in group 2 compositions had a significantly better effect in 45 P_2O_5 -30CaO although 50 P_2O_5 -25CaO composition recorded similar ALP results to the control.

4.7.4 CCK Assay

The aim of using CCK was to count the number of viable cells for all compositions at different time points. Therefore, different known numbers of cells with their CCK readings were used to derive an equation that would help count the number of cells indirectly using the CCK readings obtained.

While on day 1 both Ti_5Si_0 , and Ti_3Si_2 had higher readings than the rest compositions and similar to the control sample (P value = 0.956 and 0.927 respectively); all samples had similar results on day 4 and 7. On day 7, for example, the control sample had a value of (56973 ± 2726) and with respect to it; 50 P_2O_5 -25CaO had the lowest reading (42189.74 ± 1432.71) while Ti_5Si_0 had the highest (48447.42 ± 2259.43) . As the mean differences between the control sample and the highest and the lowest compositions were similar, no negative effect was found of doping SiO_2 .

Consequently, compensating TiO_2 with SiO_2 in group 1 was found to give good results. No specific trend was found although compositions with less SiO_2 had insignificant better

readings. Doping both 5%SiO₂ and 5%TiO₂ in group 2 compositions had a better result in 45P₂O₅-30CaO; 50P₂O₅-25CaO having a lower cell numbers.

5. Conclusion

XRD studies confirmed the success of the glass production process, the amorphous nature of all glass samples and indicated that they lacked a crystalline phase. Other results showed that an increase in TiO₂ content directly correlated with an increase in glass density, decrease in degradation, increased trend of T_g and T_m values and Na⁺ and Ca²⁺ release in group 1 compositions. Whereas in group 2 compositions, 50P₂O₅-25CaO was less dense than 45P₂O₅-30CaO, it degraded dramatically less, had less T_g and T_m values and released less Na⁺ and Ca²⁺.

Regarding cell culture studies, live/dead assay showed that all compositions had promising results in terms of keeping the MG63 cells alive after 7 days. However, the rest of the studies showed differences between compositions in terms of cell viability and proliferation.

Within the first group, replacing TiO₂ with SiO₂ did not increase the number of MG63 cells, or the ability of hMSC's cells to differentiate into osteoblasts though it did not cause cytotoxicity. Whereas, within the second group, doping both 5%SiO₂ and 5%TiO₂ into group 2 compositions had a positive effect in increasing the number of MG63 cells in both compositions with 45P₂O₅-30CaO to have better results in terms of cell numbers and improving the ability of hMSC's cells to differentiate into osteoblasts.

Declaration.

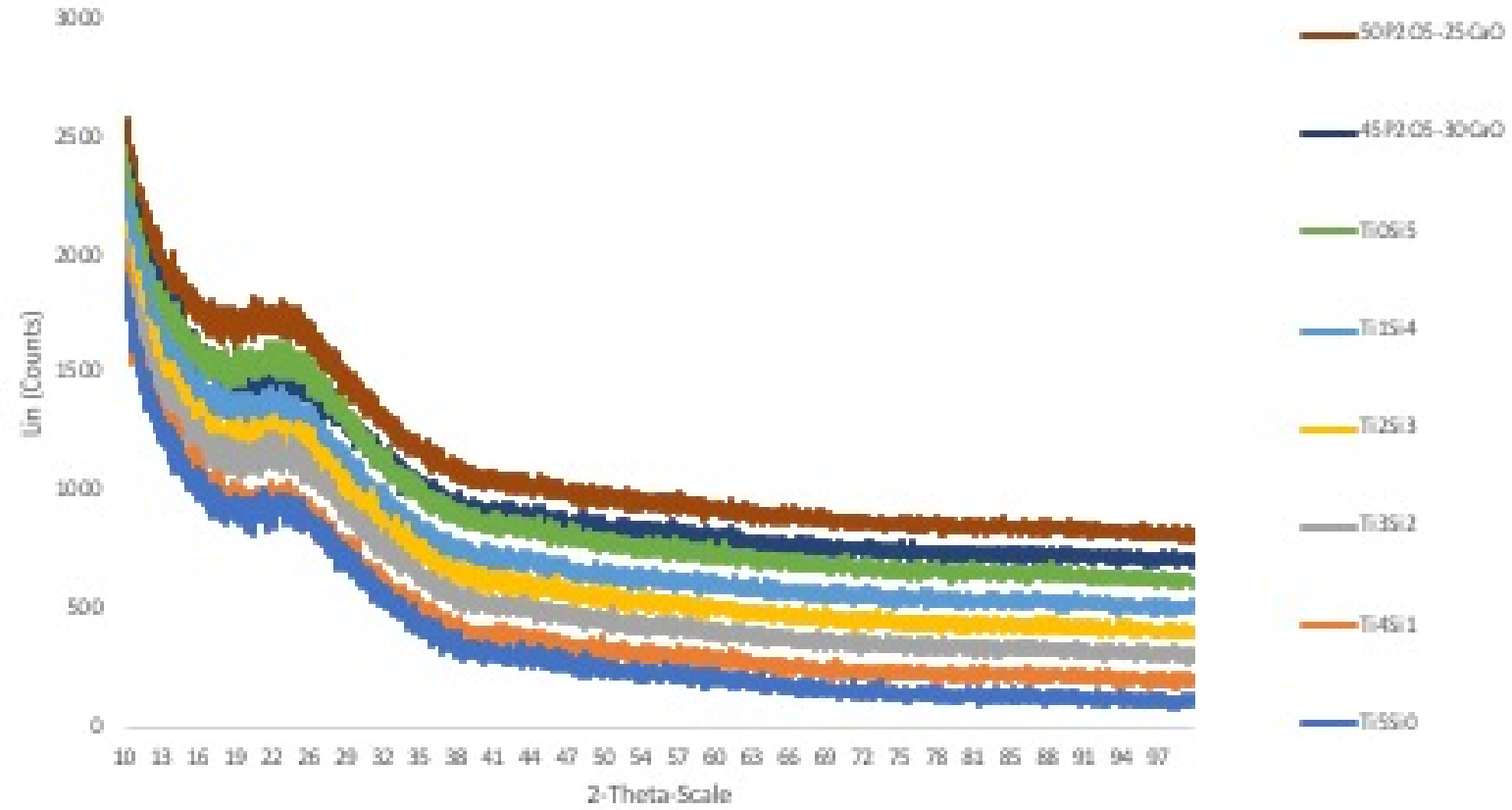
The researcher confirms the absence of any conflict of interest.

6. References

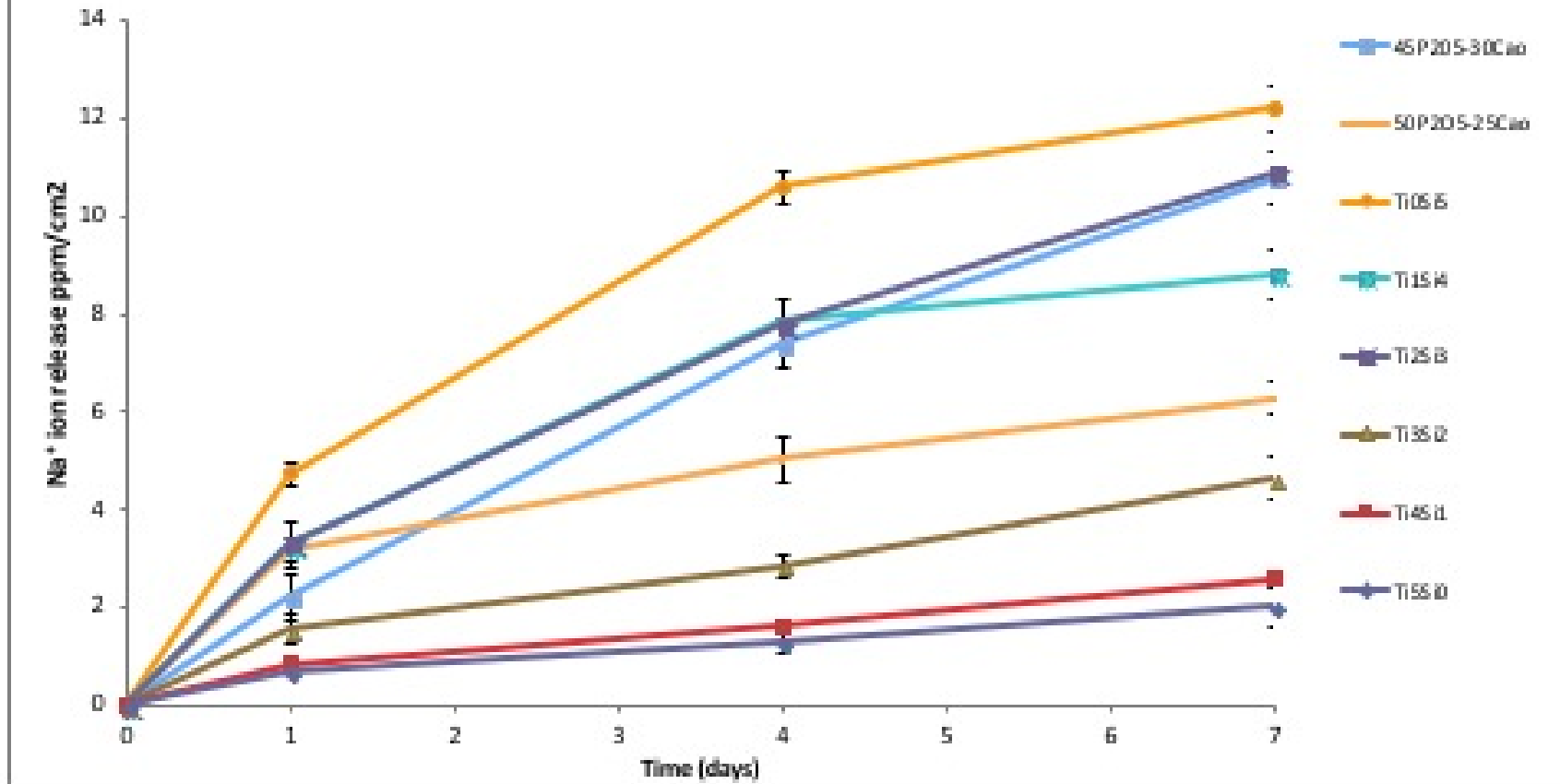
1. Rezwan K, Chen QZ, Blaker JJ, Boccaccini AR. Biodegradable and bioactive porous polymer/inorganic composite scaffolds for bone tissue engineering. *Biomaterials*. 2006;27(18):3413-31.
2. Kaur G, Pandey OP, Singh K, Homa D, Scott B, Pickrell G. A review of bioactive glasses: Their structure, properties, fabrication and apatite formation. *J Biomed Mater Res A*. 2014;102(1):254-74.
3. Lin K, Chang J, Liu Z, Zeng Y, Shen R. Fabrication and characterization of 45S5 bioglass reinforced macroporous calcium silicate bioceramics. *Journal of the European Ceramic Society*. 2009;29(14):2937-43.
4. Demirkiran H, Mohandas A, Dohi M, Fuentes A, Nguyen K, Aswath P. Bioactivity and mineralization of hydroxyapatite with bioglass as sintering aid and bioceramics with Na₃Ca₆(PO₄)₅ and Ca₅(PO₄)₂SiO₄ in a silicate matrix. *Materials Science and Engineering: C*. 2010;30(2):263-72.
5. Thakur S, Garg S, Kaur G, Pandey OP. Effect of strontium substitution on the cytocompatibility and 3-D scaffold structure for the xSrO-(10-x) MgO-60SiO₂-20CaO-10 P₂O₅ (2 ≤ x ≤ 8) sol-gel glasses. *Journal of Materials Science: Materials in Medicine*. 2017;28(6):89.
6. Knowles JC. Phosphate based glasses for biomedical applications. *Journal of Materials Chemistry*. 2003;13(10):2395-401.
7. Abou Neel EA, Chrzanowski W, Valappil SP, O'Dell LA, Pickup DM, Smith ME, et al. Doping of a high calcium oxide metaphosphate glass with titanium dioxide. *Journal of Non-Crystalline Solids*. 2009;355(16-17):991-1000.
8. Anselme K. Osteoblast adhesion on biomaterials. *Biomaterials*. 2000;21(7):667-81.
9. Brow RK. Review: the structure of simple phosphate glasses. *NOC* *Journal of Non-Crystalline Solids*. 2000;263:1-28.
10. Kasuga T, Hattori T, Niinomi M. PHOSPHATE GLASSES AND GLASS-CERAMICS FOR BIOMEDICAL APPLICATIONS. *Phosphorus Research Bulletin Phosphorus Research Bulletin*. 2012;26(0):8-15.
11. Franks K, Salih V, Knowles JC, Olsen I. The effect of MgO on the solubility behavior and cell proliferation in a quaternary soluble phosphate based glass system. *J Mater Sci Mater Med*. 2002;13(6):549-56.
12. Shah R, Sinanan AC, Knowles JC, Hunt NP, Lewis MP. Craniofacial muscle engineering using a 3-dimensional phosphate glass fibre construct. *Biomaterials*. 2005;26(13):1497-505.
13. Reis ST, Faria DLA, Martinelli JR, Pontuschka WM, Day DE, Partiti CSM. Structural features of lead iron phosphate glasses. *Journal of Non-Crystalline Solids*. 2002;304(1-3):188-94.

14. Abou Neel EA, Mizoguchi T, Ito M, Bitar M, Salih V, Knowles JC. In vitro bioactivity and gene expression by cells cultured on titanium dioxide doped phosphate-based glasses. *Biomaterials*. 2007;28(19):2967-77.
15. Al Qaysi M, Walters NJ, Foroutan F, Owens GJ, Kim H-W, Shah R, et al. Strontium- and calcium-containing, titanium-stabilised phosphate-based glasses with prolonged degradation for orthopaedic tissue engineering. *Journal of Biomaterials Applications*. 2015;30(3):300-10.
16. Kukhareenko SA, Shilo AE, Itsenko PP, Kutsai AN. The effect of titanium dioxide on the structure of silicate multicomponent glasses. *Journal of Superhard Materials*. 2010;32(6):396-405.
17. Abou Neel EA, Chrzanowski W, Knowles JC. Effect of increasing titanium dioxide content on bulk and surface properties of phosphate-based glasses. *Acta Biomaterialia*. 2008;4(3):523-34.
18. Abou Neel EA, Chrzanowski W, Georgiou G, Dalby MJ, Knowles JC. In Vitro Biocompatibility and Mechanical Performance of Titanium Doped High Calcium Oxide Metaphosphate-Based Glasses. *Journal of Tissue Engineering*. 2010;2010:390127.
19. Knowles JC, Franks K, Abrahams I. Investigation of the solubility and ion release in the glass system K₂O-Na₂O-CaO-P₂O₅. *Biomaterials*. 2001;22(23):3091-6.
20. Lakhkar NJ, Park JH, Mordan NJ, Salih V, Wall IB, Kim HW, et al. Titanium phosphate glass microspheres for bone tissue engineering. *Acta Biomater*. 2012;8(11):4181-90.
21. Aubin JE. Regulation of osteoblast formation and function. *Reviews in endocrine & metabolic disorders*. 2001;2(1):81-94.

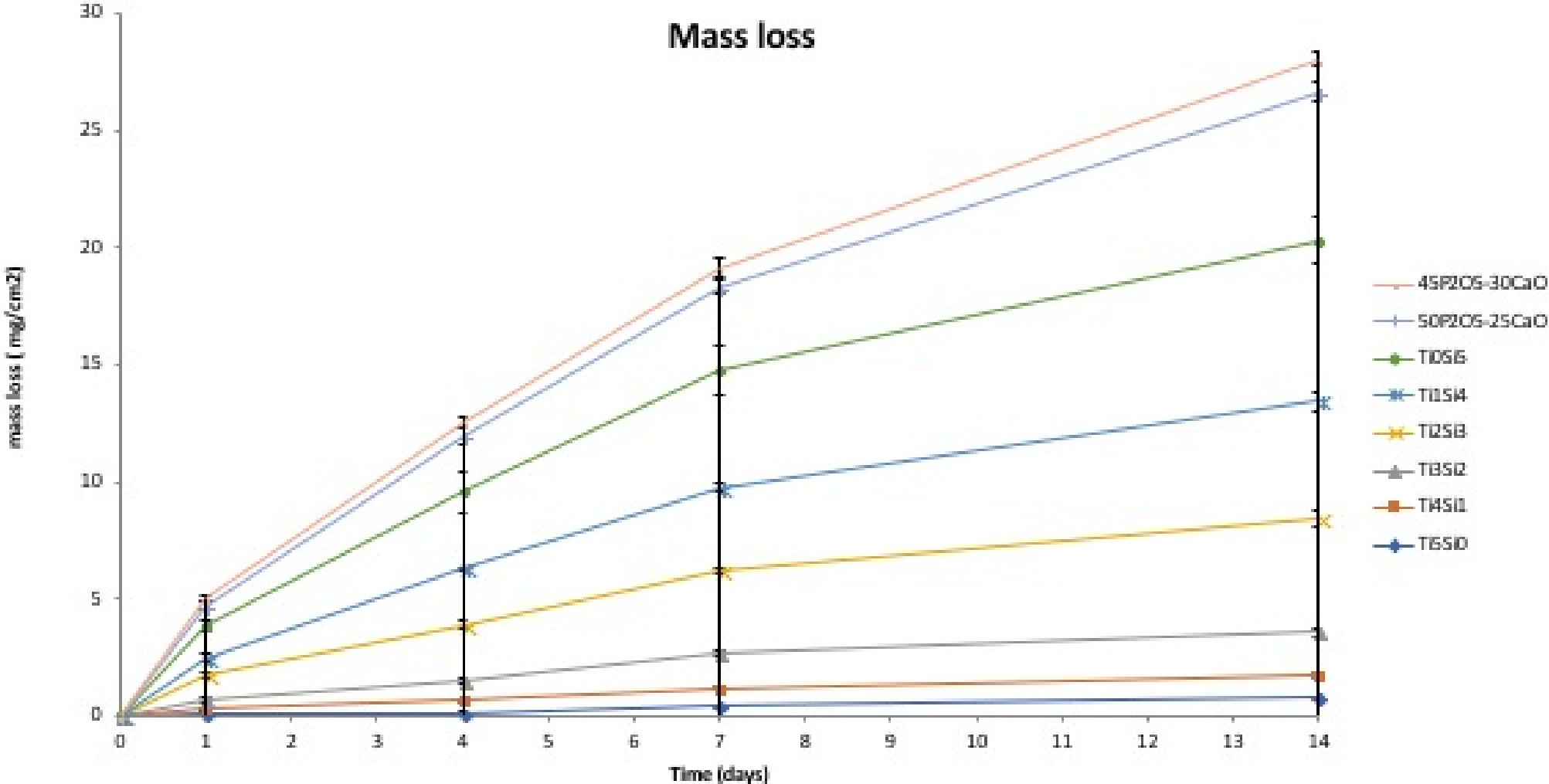
XRD Patterns

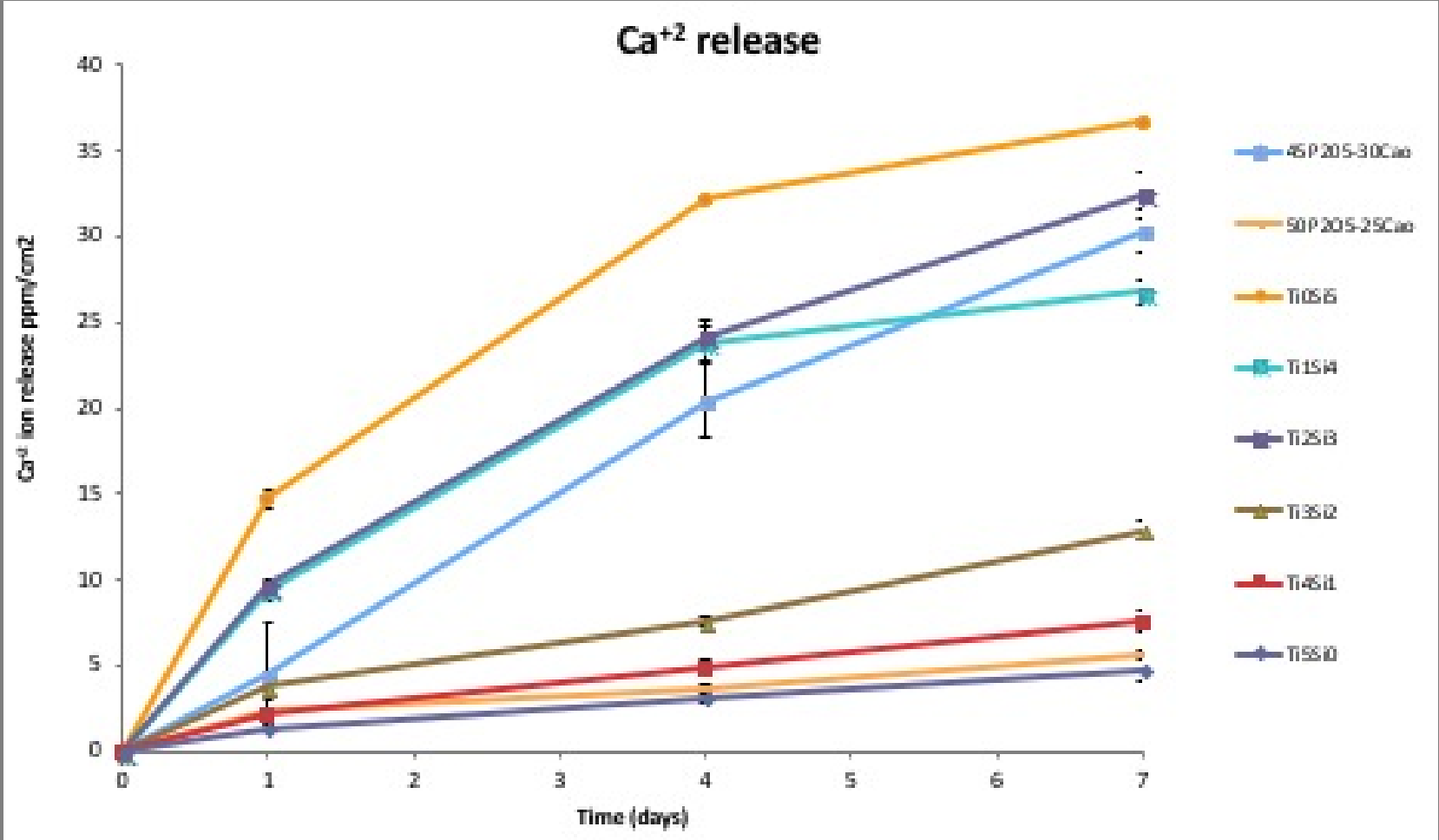


Na⁺ release

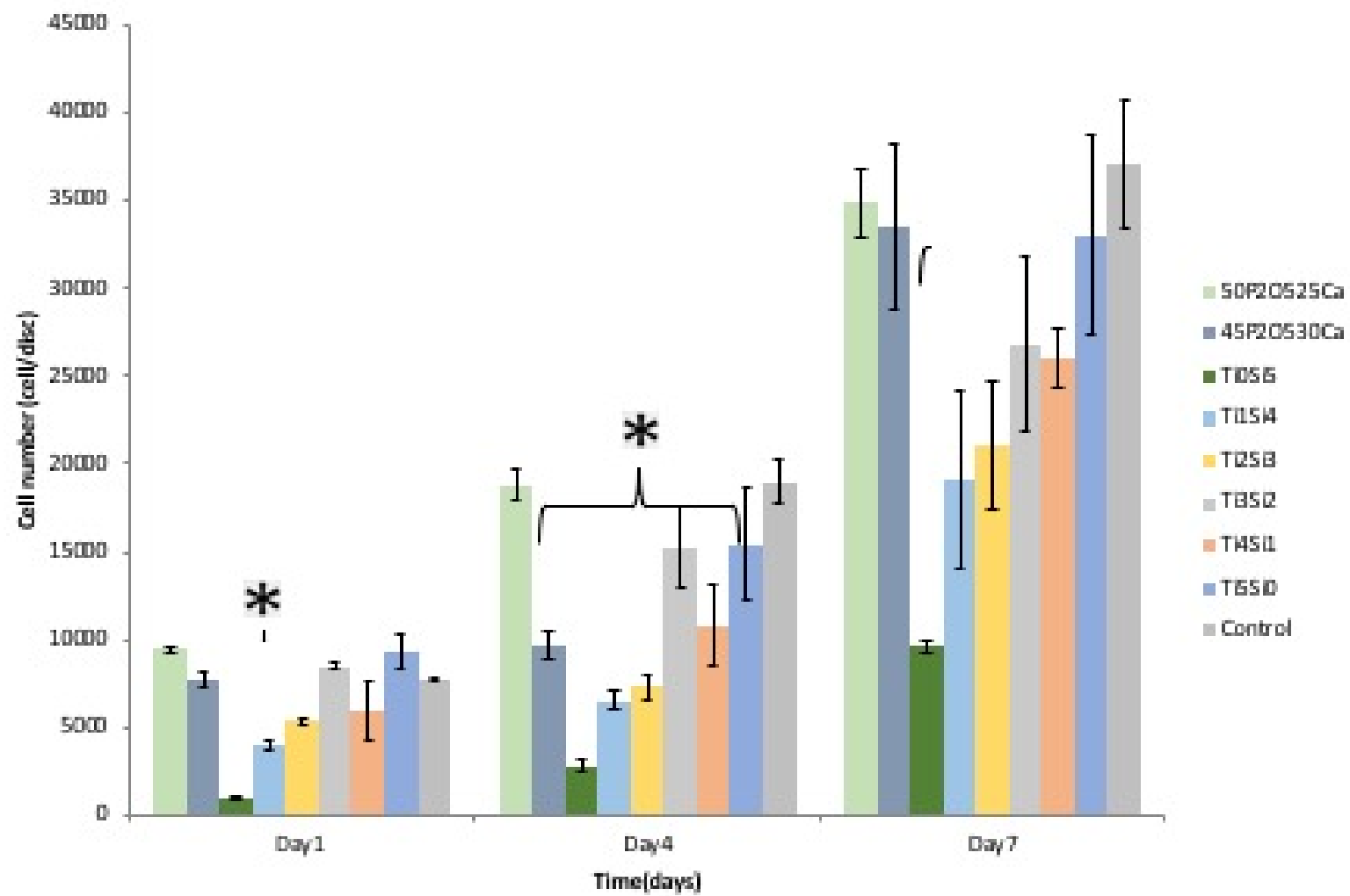


Mass loss

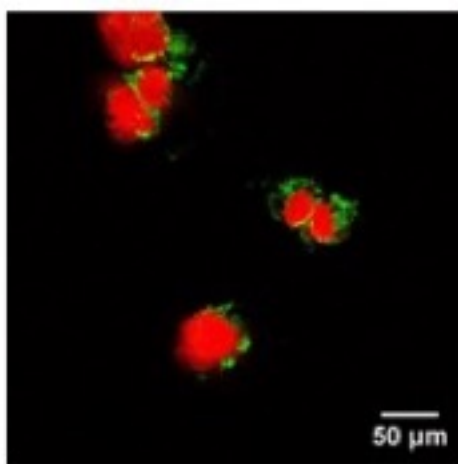




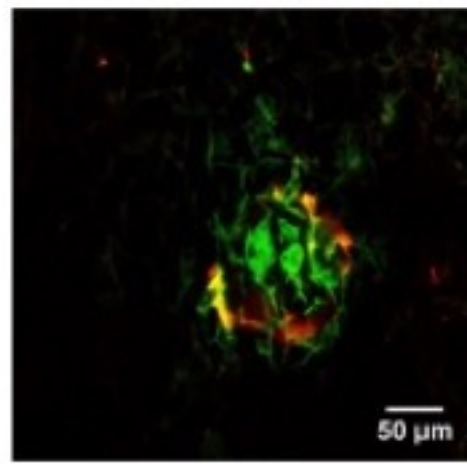
Alamar Blue Assay



Day 4

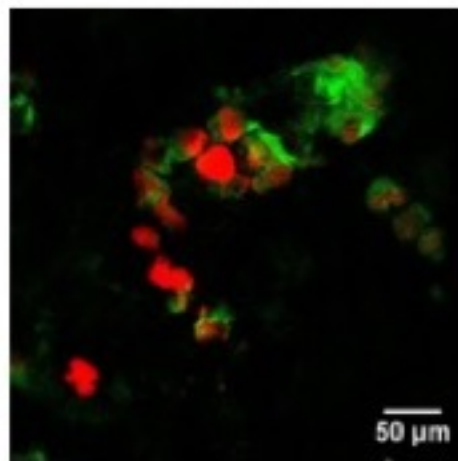


Ti0Si5

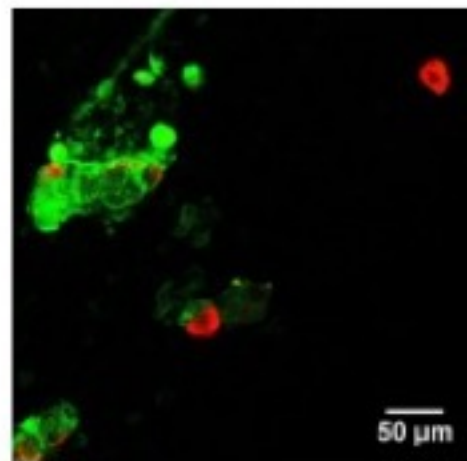


Ti5Si0

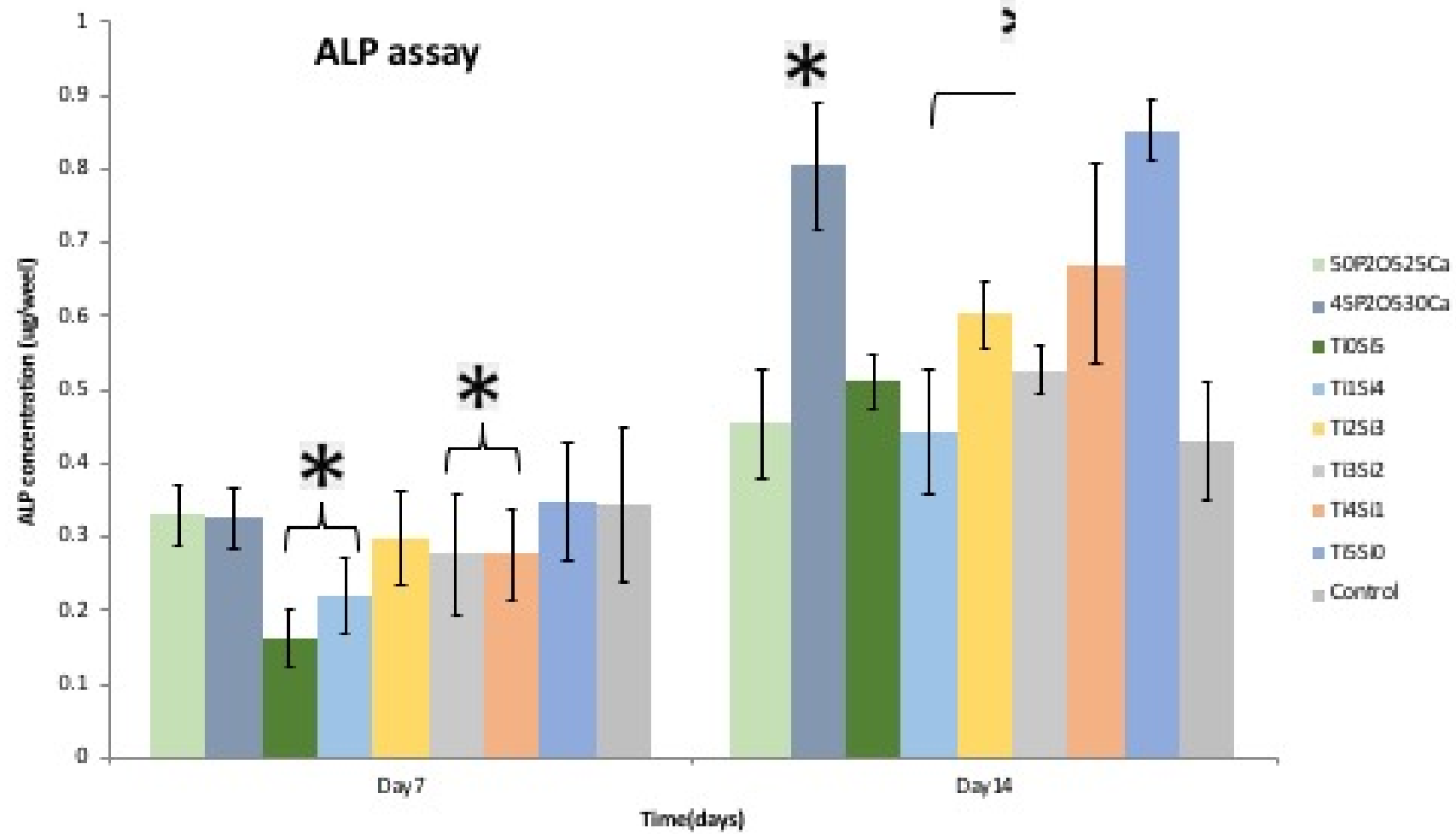
Day 7



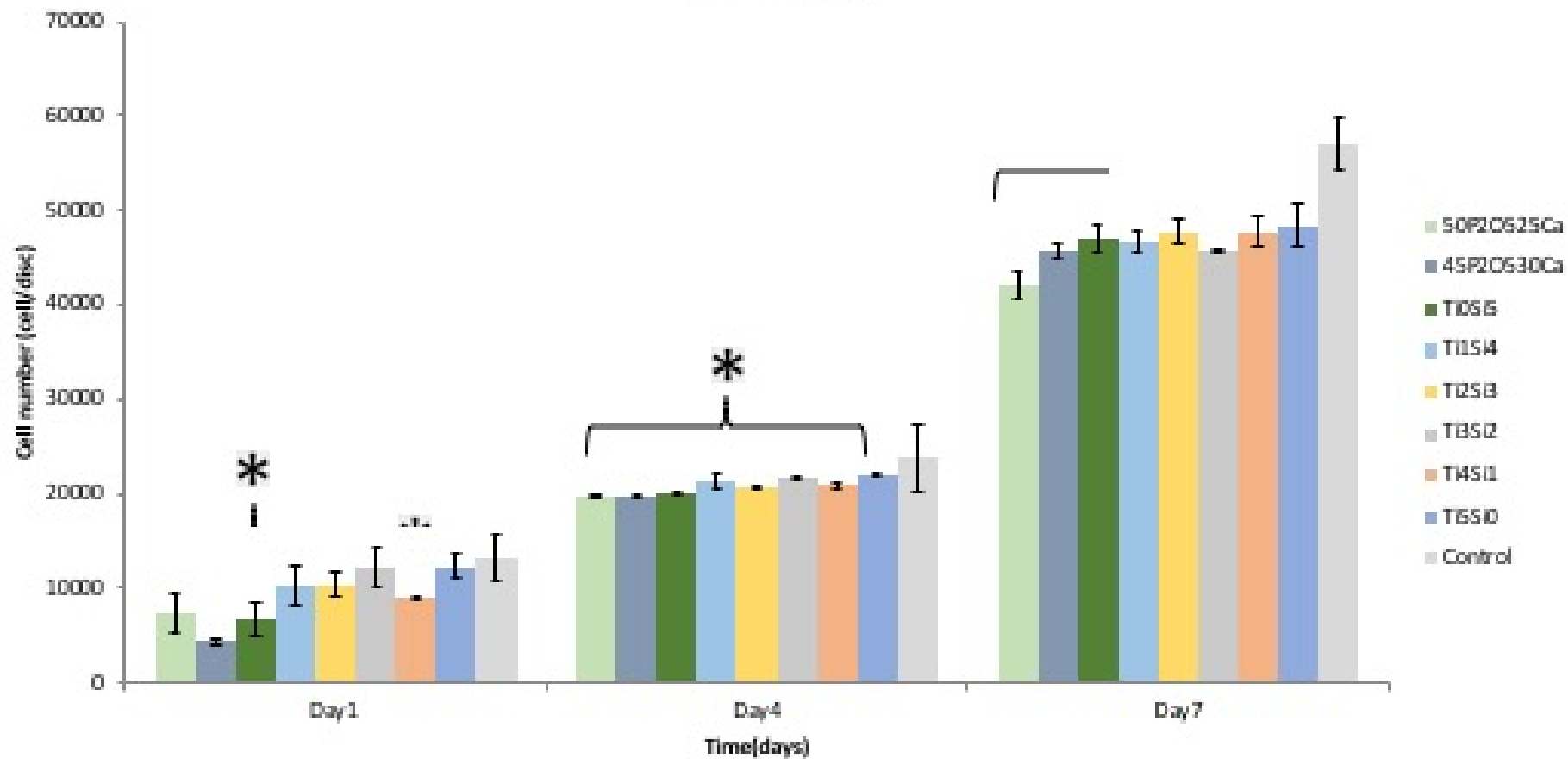
Ti0Si5



Ti5Si0



CCK Assay



Compositions	Code	Glass Compositions (mol%)				
		P ₂ O ₅	CaO	Na ₂ O	TiO ₂	SiO ₂
Group 1						
Ti 0% Si 5%	Ti0Si5	50	30	15	0	5
Ti 1% Si 4%	Ti1Si4	50	30	15	1	4
Ti 2% Si 3%	Ti2Si3	50	30	15	2	3
Ti 3% Si 2%	Ti3Si2	50	30	15	3	2
Ti 4% Si 1%	Ti4Si1	50	30	15	4	1
Ti 5% Si 0%	Ti5Si0	50	30	15	5	0
Group 2						
P ₂ O ₅ 45%	45P ₂ O ₅ -30CaO	45	30	15	5	5
Ca 25%	50P ₂ O ₅ -25CaO	50	25	15	5	5

	ρ glass g/cm ³	T _g °C	T _c °C	T _m °C
Group 1				
TiO ₅ Si	2.55 ± 0.005	421	681	782
Ti ₁ Si ₄	2.55 ± 0.003	459	756	823
Ti ₂ Si ₃	2.56 ± 0.002	466	719	839
Ti ₃ Si ₂	2.56 ± 0.0003	466	693	844
Ti ₄ Si ₁	2.59 ± 0.0009	481	733	878
Ti ₅ Si ₀	2.59 ± 0.002	477	715	867
Group 2				
45P ₂ O ₅ -30CaO	2.58 ± 0.016	481	777	846
50P ₂ O ₅ -25CaO	2.55 ± 0.003	445	714	778

	Group 1						Group 2		Control
	Ti5Si0	Ti4Si1	Ti3Si2	Ti2Si3	Ti1Si4	Ti0Si5	50P2O5- 25CaO	45P2O5-30CaO	Control
Day1	9.30	9.17	8.50	9.47	8.83	8.37	9.46	9.38	8.38
Day4	9.44	9.54	8.77	9.36	9.54	9.65	9.25	9.74	9.38
Day7	9.21	9.47	8.63	9.18	9.28	9.01	9.44	9.49	8.94