

Article Type: Review

KRAS: reasons for optimism in lung cancer

C.R. Lindsay^{1,2,3}, M. Jamal-Hanjani^{3,4}, M. Forster^{3,4}, F. Blackhall^{1,2,3}

1. Division of Molecular and Clinical Cancer Sciences, University of Manchester, Manchester, UK.
2. Department of Medical Oncology, The Christie NHS Foundation Trust, Wilmslow Road, Manchester, M20 4BX, UK.
3. Cancer Research UK Lung Cancer Centre of Excellence London and Manchester.
4. Department of Oncology, University College of London Hospital and UCL Cancer Institute, London, UK.

Corresponding Author: Dr Colin Lindsay, Division of Molecular and Clinical Cancer Sciences, University of Manchester, Manchester, UK. Tel: +44 (0)161 446 3745.

colin.lindsay@manchester.ac.uk

Abstract

Despite being the most frequent gain-of-function genetic alteration in human cancer, *KRAS* mutation has to date offered only limited potential as a prognostic and predictive biomarker. Results from the phase III SELECT-1 trial in non-small cell lung cancer (NSCLC) recently added to a number of historical and more contemporary disappointments in targeting *KRAS* mutant disease, including farnesyl transferase inhibition and synthetic lethality partners such as STK33. This narrative review uses the context of these previous failures to demonstrate how the knowledge gained from these experiences can be used as a platform for exciting advances in NSCLC on the horizon. It now seems clear that mutational subtype (most commonly *G12C*) of individual mutations is of greater relevance than the categorical evaluation of *KRAS* mutation presence or otherwise. A number of direct small molecules targeted to these subtypes are in development and have shown promising biological activity, with some in the late stages of preclinical validation.

154 words

Key words: KRAS, NSCLC, RAS, non-small cell lung cancer, G12C

RAS is the most common oncogene in cancer, with its mutation occurring in approximately 30% of all cases [1]. This rate of mutation can vary substantially in different cancer types, with its most frequent rates of modification found to affect *KRAS* in pancreatic, colorectal and lung adenocarcinomas [2-4]. Its function and importance as a GTPase is evidenced by its central coordinating role in the cell, where it connects upstream signals from cell surface receptors such as FGFR, EGFR and ERBB2-4 to downstream cancer-associated pathways such as MAPK, PI3K and Ral [5]. Introduction of *RAS* mutations in mice using either chemical and/or environmental induction or genetic modification has been shown to induce cancers such as lung adenocarcinoma and melanoma, with perhaps the most well characterised lung cancer model demonstrating that lung-specific *KRAS G12D* expression can allow mediation of both tumor initiation and multiplicity [6-9]. More recent studies of tumor heterogeneity have identified *RAS* mutations as both 'truncal' and 'branch' drivers, depending on genetic subtype and the cancer cell context [10].

Predominant isoforms and genetic subtypes of mutant *RAS* vary with cancer histology. In terms of isoforms, *NRAS* is the most frequently mutated in melanoma, whereas *KRAS* mutation occurs most frequently in adenocarcinomas [11]. Mutant *HRAS* occurs in a small percentage of head and neck squamous cancers [12]. Why certain cancers are driven by specific *RAS* isoforms remains unknown, and it is also unclear whether this variation occurs as a consequence of cell lineage and/or other factors. In lung cancer, *KRAS* mutation occurs in 20-40% of adenocarcinomas, with codons 12 and 13 mutations being the most frequent, of which the most common subtypes are *G12C*, *G12V* and *G12D*. *G12C* and *G12V* have been epidemiologically associated with a smoking history, whereas the *G12D* subtype occurs more frequently in never smokers [13].

Following its description as a key cancer mutation in 1982, clinical investigation of *RAS* has been blighted by conflicting results regarding its prognostic relevance as well as unsuccessful clinical trial programmes, most recently evidenced by the large phase III SELECT-1 trial which showed no improvement in progression-free or overall survival with the addition of selumetinib, an oral small-

molecule MEK inhibitor, to 2nd line docetaxel chemotherapy in advanced *KRAS*-mutant NSCLC [14,15]. The context of this failed progress has been particularly disappointing: lung cancer is the most common cause of cancer-associated mortality in the UK and worldwide, with treatment for its most common histological category, NSCLC, offering only limited survival gains in both early and metastatic disease settings [16,17]. This translational review will argue that these past difficulties are by no means representative of *KRAS* irrelevance, but more a consequence of insufficient understanding of its biology coupled with sub-optimal study design. Here, strategies to improve delineation of the prognostic and predictive potential of *KRAS* as a biomarker will be presented, as well as reasons to be optimistic for the future success of *KRAS* targeting in lung cancer.

Biomarker challenges

Within phase I of the Cancer Research UK Stratified Medicine Programme, *KRAS* mutation was detected in 287 of 774 lung adenocarcinomas (37%), a figure which contrasts with a prior large American study reporting *KRAS* mutations in 21% of 482 adenocarcinomas [18]; these percentages can reasonably be assumed to be the upper and lower limits of normal for *KRAS* mutation incidence in NSCLC. *KRAS* mutations of codons 12 and/or 13 'not otherwise specified' were noted in 79 patients, leaving smoking-associated mutational subtypes to represent at least 56% of cases, most commonly *G12C* (73/208 samples, 35.1%) and *G12V* (44/208 samples, ~21.2%), while the *G12D* subtype (40/208 samples, ~19.2%) has been closely associated with never smokers [13]. The remaining cases can be constituted by a wide variety of mutational subtypes, involving usually codon 12 or codon 13 (Figure 1).

Clinical reports from the past 5 years have progressively demonstrated the importance of interrogating *KRAS* mutation beyond a simple categorisation of whether it is present or not [19,20]. Its historical study as a prognostic biomarker in lung cancer frequently concluded, in line with its important biological context, that its presence conferred diminished survival. Many of these reports were limited by small numbers of patients, retrospective data, and lack of a validation set and/or multivariate analysis, but subsequent meta-analyses offered similar conclusions [21,22]. However, an examination in 2013 by the LACE bio-collaborative group offered the most comprehensive single study assessment of this question with conflicting results. In greater than 1,500 *KRAS*-tested patients recruited from four key lung cancer adjuvant clinical trials, no clear prognostic or predictive relevance of *KRAS* mutation was observed. There was a suggestion that a small number of patients with codon 13 *KRAS* mutations did not benefit from adjuvant chemotherapy, however this could not be confirmed due to the lack of a validation set. Perhaps as important as its findings, this study offered the first analysis of *KRAS* prognostication according to mutational subtype – a genetic context that would shape prognostic and treatment progress in subsequent years [23].

So why would *KRAS* mutation succeed as a biomarker now when it has failed in the past? There are three particular reasons to be optimistic regarding the development of *KRAS* as a clinical biomarker and target in lung cancer:

1. *It is predictive.*

Cancer biomarkers with routine clinical utility are generally either predictive (such as *BRAF* mutation in melanoma or *ALK/ROS-1* gene fusion in lung cancer), or have been more historically used for tumour monitoring (for example, CA125 in high grade serous ovarian carcinoma) [24-26]. *KRAS* mutation in lung adenocarcinoma has recently been characterised as a predictor of negative benefit for patients with *EGFR* mutant disease undergoing treatment with *EGFR* tyrosine kinase inhibitors, a

finding that mirrors similar predictive benefits of *KRAS* mutation in colorectal cancer [27,28]. With the anticipated influx of direct *KRAS* mutation inhibitors to the clinic (discussed below) over the next few years, the opportunity to exploit the predictive potential of *KRAS* mutation will undoubtedly assume increasing relevance in a manner that may be analogous to the successes achieved with *BRAF* targeting across cancer [29].

2. Improved characterisation of KRAS subtypes, amplification and genomic context.

More recent prognostic investigation of *KRAS* mutation at the level of its genetic subtypes in lung cancer has so far offered more conclusive results. Ihle and colleagues demonstrated reduced survival in patients with the more common smoking-related *G12C* and *G12V* subtypes, also showing that their pathogenicity was conferred by upregulation of downstream Ral and PI3K signalling [19]. This report was consistent with another study that retrospectively analysed over 800 patients with the *G12V* subtype, also concluding that this was associated with reduced prognosis [20]. Identification of *KRAS* mutation copy number gain and amplification in lung cancer has been reported in cancers from mouse models harbouring the *G12D* subtype, a change that was associated with increased disease aggression [30]: although traditionally considered to be a cancer driven by somatic mutation as a consequence of smoking and environmental insults, undoubtedly more will follow regarding the characterisation of lung adenocarcinoma as a cancer further defined by copy number change, as has been the case more recently with oesophageal and ovarian carcinomas [31,32]. This level of genetic examination, whether through assessment of genetic subtypes and/or copy number aberration, was omitted in historical biomarker studies of lung cancer and will be paramount to our future exploitation of *KRAS* as a lung cancer biomarker. Moreover, genetic background and context of each *RAS*-driven tumour will also assume further relevance, a complexity which has been one of a number of reasons cited for the lack of success observed so far with synthetic lethality screens in *KRAS* mutant disease [33]. One key report has categorised *KRAS*-mutant adenocarcinoma into three major

subgroups (co-mutation with aberrations in *LKB1*, *TP53* or *CDKN2A/B*), assisting characterisation of their relative immune profiles and therapeutic vulnerabilities [34].

3. *There is a paradigm for routine KRAS testing in the clinic.*

The backdrop for introduction of any new biomarker to the hospital setting has become significantly more accommodating in recent years - to the extent that cancer and lung cancer-specific organisations such as ASCO, ESMO, and the IASLC have issued position statements and guidelines on clinical molecular testing to be performed routinely on lung cancer patients. Targeted sequencing involving a panel of core cancer genes such as *RAS* is now possible at substantially reduced time and cost, with many molecular pathology departments across publicly-funded hospitals in the Western world incorporating this technology into their short to medium-term plans for implementation. Such a setup will foster optimisation of other biomarker test qualities stipulated in REMARK guidelines: an increase in reproducibility allied to a reduction of complexity [35]. A number of academically led lung cancer stratified-medicine programmes rolled out in the US, UK and Europe will have a key role in informing this process, and the example of lung cancer molecular testing implemented across France has set a standard for others to follow [36].

Why has there been failure to date to effectively target KRAS in NSCLC?

The structural and biochemical challenges of targeting *RAS* have held back a research environment which has been characterised for a number of years by its lack of a breakthrough. *KRAS* in particular has been coined as an 'undruggable' target when in reality structural biochemists and drug developers have considered it to be challenging but possible. The recent success of targeting *BCL-2* (another 'undruggable' target) has highlighted how progress can be made with difficult targets by

applying an emerging understanding of structural biochemistry characterised through an increased success of NMR and X-Ray crystallography [37].

Several lessons have been learned in recent years that represent challenges for effective KRAS targeting. First, it binds to GTP with nanomolar affinity in its active state, an interaction that requires more potent inhibition than that conferred by traditional tyrosine kinase inhibitors. Second, it is structurally a relatively smooth protein, with few clefts for potential lead compounds to 'hook' on to in order to exert their effects [1]. As a consequence, KRAS mutation targeting has been focused for many years on downstream inhibitors and, more recently, targeting of identified synthetic lethality partners. For different reasons, neither approach has yet offered significant successes:

1. Downstream inhibition.

Farnesyl transferase inhibitors (FTIs) offer the most recited example of treatment failure in targeting *RAS* mutant cancer. Preclinical research had suggested their potential efficacy through inhibition of membrane association by mutant Ras and its subsequent activation (mediated by farnesyl transferase), yet clinical trials interrogating their role in *KRAS* mutant pancreas and lung cancer were subsequently considered a significant failure [38-41]. Further analysis following the clinical abandonment of FTIs offered more hope, emphasising the importance of considering *RAS* mutant isoforms when designing and delivering clinical trials: farnesylation was concluded to be of particular relevance to *HRAS*-driven cancer (with alternative mechanisms of membrane association/activation employed in the context of *NRAS* and *KRAS* mutations), and application of FTIs to *HRAS* mutant cell lines has subsequently proven effective [42].

More recently, the combination of docetaxel and the MEK inhibitor selumetinib in the phase II setting appeared to offer an incremental survival advantage in pretreated metastatic *KRAS* mutant

NSCLC, but the subsequent phase III SELECT-1 trial proved negative [15,43]. A phase II trial examining trametinib, another MEK inhibitor, was also shown to offer no discernible survival advantage compared to second line docetaxel in *KRAS* mutant disease [44].

One common feature of these studies which could help explain their lack of success was that contemporaneous biopsies were not mandated for *KRAS* testing – thus, prescient information on the background genetic context of individual tumours was unobtainable in the vast majority of patients. A new generation of NMR-developed small molecules which interact with the RAS-binding domains (RBDs) of proteins immediately downstream to *KRAS* now offer a potential route to success in this area, with one compound rigosertib undergoing clinical trial evaluation in haematological malignancies having demonstrated activity against a number of Ras effectors [45].

2. *Synthetic lethality partners.*

A number of synthetic lethality screens have been performed using *KRAS* mutant cancer cell lines, offering potential novel targets that include BCL-XL, TANK binding kinase-1 and CDK4 [46-48]. Many of these targets have associated direct small molecule inhibitors under preclinical assessment, and inhibitors of CDK4 are currently being tested in the clinic for treatment of 2nd to 3rd line NSCLC patients with *KRAS* mutant disease [49]. However the example of a previously identified synthetic lethality partner, *STK33*, suggested there are reasons to be cautious with this approach: following its identification using an RNA interference-based screen in *KRAS* mutant cell lines, a kinase inhibitor of *STK33* proved ineffective [50]. A number of legitimate reasons have been offered to explain why there has been no breakthrough with synthetic lethality screens of *RAS* mutant cancer that match the success of PARP inhibition in *BRCA* mutant disease, including inconsistency of methodology, technology and cell line choice [33].

Why is there new optimism for *KRAS* mutation targeting?

Other than the encouraging progress seen with RBD inhibition described above, a number of direct inhibitors of mutant *KRAS* are now in early to late stages of preclinical development, with some expected to reach preclinical and clinical testing shortly (Table 1). Many of these inhibitors were designed with an appreciation that the crystal/NMR structures for each individual *KRAS* mutant subtype would be vital for progress.

The most advanced lead compound reported so far targets the most common *G12C* smoking-related subtype based on its crystal structure, using allosteric inhibition of a cysteine residue to prevent nucleoside exchange and regulation of *G12C*, thus holding it in the inactive GDP-bound state [51]. Second and third-generation iterations of this compound have demonstrated increasing biological activity measured by their ability to inhibit *KRAS* signalling [52,53]. Given its key role in cell homeostasis, avoiding off-target effects on *wild-type KRAS* will be a particular concern going forward, with not all compounds specifically targeted to its mutant subtypes and the Ras superfamily in general consisting of a network of 150 related proteins. However, the example of synthetic lethality screens, where no putative co-target could offer as potent an effect in *KRAS*-mutant cell lines as targeting *KRAS* itself with RNAi, suggests there is good reason to pursue this approach vigorously [33].

Other promising advances include pan-Ras compounds (which bind at least two sites on the Ras protein and have so far reported activity in the low micromolar range) and the identification of intracellular antibodies which can target oncogenic *KRAS* (Table 1) [54,55]. Although perhaps at an earlier stage of development compared to some other agents, the latter of these two prospects offers particular cause for excitement given it could broadly represent a new class of antibody which homes in on a tumour and internalises through receptor-mediated endocytosis.

Applying the same principals to what was observed 5-10 years ago with BRAF inhibitors in *BRAF* mutant cancer, it is expected that the ultimate success of these drugs will be contingent on their stratification to *KRAS* mutational subtypes such as *G12C*. In general, the increasing pursuit of direct *KRAS* inhibitors by both biotechs and academic institutions offer reasons to be hopeful, and the National Cancer Institute RAS initiative is another development which can support this programme of research in a number of different contexts.

Conclusions

Whether through an increased understanding of its genetic detail and molecular structure, or an improved practicality of identifying genetic subtypes with consistency in a large number of patients, the emergence of *KRAS* as a biomarker and target for biological agents should offer one of the next major breakthroughs in genetically-targeted treatment of cancer. However this will only represent the beginning of the process given a number of further complexities to delineate in the years ahead, including the potentiation of *KRAS* mutant-mediated malignant transformation by smoke-induced epigenetic changes, the role of *KRAS* in modulating the host immune response, and developing stories of resistance involving *KRAS* alteration or targeting [56-58]. Given its central role in mediating lung cancer, the world's biggest cancer killer, as well as a number of other difficult-to-treat cancers, such a development could have an impact beyond anything seen before in molecular therapeutics.

2,663 words

CONFLICT OF INTEREST STATEMENT The authors have declared no conflicts of interest.

REFERENCES

1. Cox AD, Fesik SW, Kimmelman AC, Luo J, Der CJ. Drugging the undruggable RAS: Mission possible? *Nat Rev Drug Discov* 2014; 13(11): 828-51.
2. Lau ST, Leung PS. Role of the RAS in pancreatic cancer. *Curr Cancer Drug Targets* 2011; 11(4): 412-20.
3. Fakih MG. Metastatic colorectal cancer: current state and future directions. *J Clin Oncol* 2015; 33(16): 1809-24.
4. Kempf E, Rousseau B, Besse B, Paz-Ares L. KRAS oncogene in lung cancer: focus on molecularly driven clinical trials. *Eur Respir Rev* 2016; 25:71-76
5. Downward J. Targeting RAS signalling pathways in cancer therapy. *Nat Rev Cancer* 2003; 3(1): 11-22.
6. Jackson EL, Willis N, Mercer K, Bronson RT, Crowley D, Montoya R et al. Analysis of lung tumor initiation and progression using conditional expression of oncogenic K-ras. *Genes Dev* 2001; 15(24): 3243-8.
7. Tuveson DA, Shaw AT, Willis NA, Silver DP, Jackson EL, Chang S et al. Endogenous oncogenic K-ras(G12D) stimulates proliferation and widespread neoplastic and developmental defects. *Cancer Cell* 2004; 5(4): 375-87.
8. Lindsay CR, Lawn S, Campbell AD, Faller WJ, Rambow F, Mort RL et al. P-Rex1 is required for efficient melanoblast migration and melanoma metastasis. *Nat Commun* 2011; 2: 555.
9. Westcott PM, Halliwill KD, To MD, Rashid M, Rust AG, Keane TM et al. The mutational landscapes of genetic and chemical models of Kras-driven lung cancer. *Nature* 2015; 517(7535): 489-92.
10. McGranahan N, Swanton C. Biological and therapeutic impact of intratumor heterogeneity in cancer evolution. *Cancer Cell*. 2015 Jan 12;27(1):15-26.

11. Hobbs GA, Der CJ, Rossman KL. RAS isoforms and mutations in cancer at a glance. *J Cell Sci* 2016; 129(7): 1287-92.
12. Hah JH, Zhao M, Pickering CR, Frederick MJ, Andrews GA, Jasser SA et al. HRAS mutations and resistance to the epidermal growth factor receptor tyrosine kinase inhibitor erlotinib in head and neck squamous cell carcinoma cells. *Head Neck* 2014; 36(11): 1547-54.
13. Dogan S, Shen R, Ang DC, Johnson ML, D'Angelo SP, Paik PK et al. Molecular epidemiology of EGFR and KRAS mutations in 3,026 lung adenocarcinomas: higher susceptibility of women to smoking-related KRAS-mutant cancers. *Clin Cancer Res* 2012; 18(22): 6169-77.
14. Aviel-Ronen S, Blackhall FH, Shepherd FA, Tsao MS. K-ras mutations in non-small-cell lung carcinoma: a review. *Clin Lung Cancer* 2006; 8(1): 30-8.
15. Jänne PA, van den Heuvel MM, Barlesi F, Cobo M, Mazieres J, Crinò L et al. Selumetinib Plus Docetaxel Compared With Docetaxel Alone and Progression-Free Survival in Patients With KRAS-Mutant Advanced Non-Small Cell Lung Cancer: The SELECT-1 Randomized Clinical Trial. *JAMA* 2017; 317(18): 1844-1853.
16. <http://www.cancerresearchuk.org/health-professional/cancer-statistics/statistics-by-cancer-type/lung-cancer/survival>
17. Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. *CA Cancer J Clin* 2011; 61(2): 69-90.
18. Riely GJ, Kris MG, Rosenbaum D et al. Frequency and distinctive spectrum of KRAS mutations in never smokers with lung adenocarcinoma. *Clin Cancer Res* 2008; 14(18): 5731-4.
19. Ihle NT, Byers LA, Kim ES, Marks J, Li A, Chitale DA et al. Effect of KRAS oncogene substitutions on protein behavior: implications for signaling and clinical outcome. *J Natl Cancer Inst* 2012; 104(3):228-39.
20. Renaud S, Falcoz PE, Schaëffer M, Guenot D, Romain B, Olland A et al. Prognostic value of the KRAS G12V mutation in 841 surgically resected Caucasian lung adenocarcinoma cases. *Br J Cancer* 2015; 113(8): 1206-15.

21. Huncharek M, Muscat J, Geschwind JF. K-ras oncogene mutation as a prognostic marker in non-small cell lung cancer: a combined analysis of 881 cases. *Carcinogenesis* 1999; 20(8): 1507-10.
22. Mascaux C, Iannino N, Martin B, Paesmans M, Berghmans T, Dusart M et al. The role of RAS oncogene in survival of patients with lung cancer: a systematic review of the literature with meta-analysis. *Br J Cancer* 2005; 92(1): 131-9.
23. Shepherd FA, Domerg C, Hainaut P, Jänne PA, Pignon JP, Graziano S et al. Pooled analysis of the prognostic and predictive effects of KRAS mutation status and KRAS mutation subtype in early-stage resected non-small-cell lung cancer in four trials of adjuvant chemotherapy. *J Clin Oncol* 2013; 31(17): 2173-81.
24. De Bono JS, Ashworth A. Translating cancer research into targeted therapeutics. *Nature* 2010; 467(7315): 543-9.
25. Kwak EL, Bang YJ, Camidge DR, Shaw AT, Solomon B, Maki RG et al. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med* 2010; 363(18): 1693-703.
26. Chapman PB, Hauschild A, Robert C, Haanen JB, Ascierto P, Larkin J et al. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. *N Engl J Med* 2011; 364(26): 2507-16.
27. Van Cutsem E, Köhne CH, Láng I, Folprecht G, Nowacki MP, Cascinu S et al. Cetuximab plus irinotecan, fluorouracil, and leucovorin as first-line treatment for metastatic colorectal cancer: updated analysis of overall survival according to tumor KRAS and BRAF mutation status. *J Clin Oncol* 2011; 29(15): 2011-9.
28. Fiala O, Pesek M, Finek J, Benesova L, Belsanova B, Minarik M. The dominant role of G12C over other KRAS mutation types in the negative prediction of efficacy of epidermal growth factor receptor tyrosine kinase inhibitors in non-small cell lung cancer. *Cancer Genet* 2013; 206(1-2): 26-31.

29. Girotti MR, Saturno G, Lorigan P, Marais R. No longer an untreatable disease: how targeted and immunotherapies have changed the management of melanoma patients. *Mol Oncol* 2014; 8(6): 1140-58.
30. Kerr EM, Gaude E, Turrell FK, Frezza C, Martins CP. Mutant Kras copy number defines metabolic reprogramming and therapeutic susceptibilities. *Nature* 2016; 531(7592): 110-3.
31. Cooke SL, Ng CK, Melnyk N, Garcia MJ, Hardcastle T, Temple J et al. Genomic analysis of genetic heterogeneity and evolution in high-grade serous ovarian carcinoma. *Oncogene* 2010; 29(35): 4905-13.
32. Ross-Innes CS, Becq J, Warren A, Cheetham RK, Northen H, O'Donovan M et al. Whole-genome sequencing provides new insights into the clonal architecture of Barrett's esophagus and esophageal adenocarcinoma. *Nat Genet* 2015; 47(9): 1038-46.
33. Downward J. RAS synthetic lethal screen revisited: still seeking the elusive prize? *Clin Cancer Res* 2015; 21(8): 1802-9.
34. Skoulidis F, Byers LA, Diao L, Papadimitrakopoulou VA, Tong P, Izzo J et al. Co-occurring genomic alterations define major subsets of KRAS-mutant lung adenocarcinoma with distinct biology, immune profiles, and therapeutic vulnerabilities. *Cancer Discov* 2015; 5(8): 860-77.
35. Altman DG, McShane LM, Sauerbrei W, Taube SE. Reporting Recommendations for Tumor Marker Prognostic Studies (REMARK): explanation and elaboration. *PLoS Med* 2012; 9(5): e1001216.
36. Barlesi F, Mazieres J, Merlio JP, Debieuvre D, Mosser J, Lena H et al. Routine molecular profiling of patients with advanced non-small-cell lung cancer: results of a 1-year nationwide programme of the French Cooperative Thoracic Intergroup (IFCT). *Lancet* 2016; 387(10026): 1415-1426.
37. Merino D, Lok SW, Visvader JE, Lindeman GJ. Targeting BCL-2 to enhance vulnerability to therapy in estrogen receptor-positive breast cancer. *Oncogene* 2016; 35(15): 1877-87.

38. Kohl NE, Omer CA, Conner MW, Anthony NJ, Davide JP, deSolms SJ et al. Inhibition of farnesyltransferase induces regression of mammary and salivary carcinomas in ras transgenic mice. *Nat Med* 1995; 1(8): 792-7.
39. Adjei AA, Mauer A, Bruzek L, Marks RS, Hillman S, Geyer S et al. Phase II study of the farnesyl transferase inhibitor R115777 in patients with advanced non-small-cell lung cancer. *J Clin Oncol* 2003; 21(9): 1760-6.
40. Heymach JV, Johnson DH, Khuri FR, Safran H, Schlabach LL, Yunus F et al. Phase II study of the farnesyl transferase inhibitor R115777 in patients with sensitive relapse small-cell lung cancer. *Ann Oncol* 2004; 15(8): 1187-93.
41. Van Cutsem E, van de Velde H, Karasek P, Oettle H, Vervenne WL, Szawlowski A et al. Phase III trial of gemcitabine plus tipifarnib compared with gemcitabine plus placebo in advanced pancreatic cancer. *J Clin Oncol* 2004; 22(8): 1430-8.
42. Whyte DB, Kirschmeier P, Hockenberry TN, Nunez-Oliva I, James L, Catino JJ et al. K- and N-Ras are geranylgeranylated in cells treated with farnesyl protein transferase inhibitors. *J Biol Chem* 1997; 272(22): 14459-64.
43. Jänne PA, Shaw AT, Pereira JR, Jeannin G, Vansteenkiste J, Barrios C et al. Selumetinib plus docetaxel for KRAS-mutant advanced non-small-cell lung cancer: a randomised, multicentre, placebo-controlled, phase 2 study. *Lancet Oncol* 2013; 14(1): 38-47.
44. Blumenschein GR Jr, Smit EF, Planchard D, Kim DW, Cadranet J, De Pas T et al. A randomized phase II study of the MEK1/MEK2 inhibitor trametinib (GSK1120212) compared with docetaxel in KRAS-mutant advanced non-small-cell lung cancer (NSCLC). *Ann Oncol* 2015; 26(5): 894-901.
45. Athuluri-Divakar SK, Vasquez-Del Carpio R, Dutta K, Baker SJ, Cosenza SC, Basu I et al. A Small Molecule RAS-Mimetic Disrupts RAS Association with Effector Proteins to Block Signaling. *Cell* 2016; 165(3): 643-55.

46. Barbie DA, Tamayo P, Boehm JS, Kim SY, Moody SE, Dunn IF et al. Systematic RNA interference reveals that oncogenic KRAS-driven cancers require TBK1. *Nature* 2009; 462(7269): 108-12.
47. Puyol M, Martín A, Dubus P, Mulero F, Pizcueta P, Khan G et al. A synthetic lethal interaction between K-Ras oncogenes and Cdk4 unveils a therapeutic strategy for non-small cell lung carcinoma. *Cancer Cell* 2010; 18(1): 63-73.
48. Corcoran RB, Cheng KA, Hata AN, Faber AC, Ebi H, Coffee EM et al. Synthetic lethal interaction of combined BCL-XL and MEK inhibition promotes tumor regressions in KRAS mutant cancer models. *Cancer Cell* 2013; 23(1): 121-8.
49. Middleton G, Crack LR, Popat S, Swanton C, Hollingsworth SJ, Buller R et al. The National Lung Matrix Trial: translating the biology of stratification in advanced non-small-cell lung cancer. *Ann Oncol* 2015; 26(12): 2464-9.
50. Luo T, Masson K, Jaffe JD, Silkworth W, Ross NT, Scherer CA et al. STK33 kinase inhibitor BRD-8899 has no effect on KRAS-dependent cancer cell viability. *Proc Natl Acad Sci U S A* 2012; 109(8): 2860-5.
51. Ostrem JM, Peters U, Sos ML, Wells JA, Shokat KM. K-Ras(G12C) inhibitors allosterically control GTP affinity and effector interactions. *Nature* 2013; 503(7477): 548-51.
52. Patricelli MP, Janes MR, Li LS, Hansen R, Peters U, Kessler LV et al. Selective Inhibition of Oncogenic KRAS Output with Small Molecules Targeting the Inactive State. *Cancer Discov* 2016; 6(3): 316-29.
53. Lito P, Solomon M, Li LS, Hansen R, Rosen N. Allele-specific inhibitors inactivate mutant KRAS G12C by a trapping mechanism. *Science* 2016; 351(6273): 604-8.
54. Shin SM, Choi DK, Jung K, Bae J, Kim JS, Park SW et al. Antibody targeting intracellular oncogenic Ras mutants exerts anti-tumour effects after systemic administration. *Nat Commun* 2017; 8: 15090.

55. Welsch ME, Kaplan A, Chambers JM, Stokes ME, Bos PH, Zask A et al. Multivalent Small-Molecule Pan-RAS Inhibitors. *Cell* 2017; 168(5): 878-889.
56. Bettgowda C, Sausen M, Leary RJ, Kinde I, Wang Y, Agrawal N et al. Detection of circulating tumor DNA in early- and late-stage human malignancies. *Sci Transl Med* 2014; 6(224): 224ra24.
57. Ortiz-Cuaran S, Scheffler M, Plenker D, Dahmen L, Scheel AH, Fernandez-Cuesta L et al. Heterogeneous Mechanisms of Primary and Acquired Resistance to Third-Generation EGFR Inhibitors. *Clin Cancer Res* 2016; 22(19): 4837-4847.
58. Vaz M, Hwang SY, Kagiampakis I, Phallen J, Patil A, O'Hagan HM et al. Chronic Cigarette Smoke-Induced Epigenomic Changes Precede Sensitization of Bronchial Epithelial Cells to Single-Step Transformation by KRAS Mutations. *Cancer Cell* 2017; 32(3): 360-376.
59. Guillard S, Kolasinska-Zwierz P, Debreczeni J, Breed J, Zhang J, Bery N et al. Structural and functional characterization of a DARPin which inhibits Ras nucleotide exchange. *Nat Commun* 2017; 8:16111.
60. Leshchiner ES, Parkhitko A, Bird GH, Luccarelli J, Bellairs JA, Escudero S et al. Direct inhibition of oncogenic KRAS by hydrocarbon-stapled SOS1 helices. *Proc Natl Acad Sci U S A* 2015; 112(6): 1761-6.
61. Lim SM, Westover KD, Ficarro SB, Harrison RA, Choi HG, Pacold ME et al. Therapeutic targeting of oncogenic K-Ras by a covalent catalytic site inhibitor. *Angew Chem Int Ed Engl* 2014; 53(1): 199-204.
62. Shima F, Yoshikawa Y, Ye M, Araki M, Matsumoto S, Liao J et al. In silico discovery of small-molecule Ras inhibitors that display antitumor activity by blocking the Ras-effector interaction. *Proc Natl Acad Sci U S A* 2013; 110(20): 8182-7.
63. Hocker HJ, Cho KJ, Chen CY, Rambahal N, Sagineedu SR, Shaari K et al. Andrographolide derivatives inhibit guanine nucleotide exchange and abrogate oncogenic Ras function. *Proc Natl Acad Sci U S A* 2013; 110(25): 10201-6.

64. Maurer T, Garrenton LS, Oh A, Pitts K, Anderson DJ, Skelton NJ et al. Small-molecule ligands bind to a distinct pocket in Ras and inhibit SOS-mediated nucleotide exchange activity. *Proc Natl Acad Sci U S A* 2012; 109(14): 5299-304.
65. Sun Q, Burke JP, Phan J, Burns MC, Olejniczak ET, Waterson AG et al. Discovery of small molecules that bind to K-Ras and inhibit Sos-mediated activation. *Angew Chem Int Ed Engl* 2012; 51(25): 6140-3.
66. Patgiri A, Yadav KK, Arora PS, Bar-Sagi D. An orthosteric inhibitor of the Ras-Sos interaction. *Nat Chem Biol* 2011; 7(9): 585-7.
67. Taveras AG, Remiszewski SW, Doll RJ, Cesarz D, Huang EC, Kirschmeier P et al. Ras oncoprotein inhibitors: the discovery of potent, ras nucleotide exchange inhibitors and the structural determination of a drug-protein complex. *Bioorg Med Chem* 1997; 5(1): 125-33.

TABLES

Name	Mechanism	Reference
ARS-853	Irreversible binding of allosteric site on G12C	Ostrem et al, Nature 2013 [51] Patricelli et al, Cancer Discov 2016 [52] Lito et al, Science 2016 [53]
3144	Pan-Ras inhibition	Welsch et al, Cell 2017 [55]
RT11	Intracellular antibody	Shin et al, Nature Comm 2016 [54]
<i>DARPin</i>	Antibody mimetic inhibiting wild-type Ras nucleotide exchange	Guillard et al, Nat Commun 2017 [59]
SAH-SOS1 _A , SAH-SOS1 _B	Peptides to disrupt wild-type KRAS/SOS1 interaction	Leshchiner et al, PNAS 2015 [60]
SML-8-73-1, SML-10-70-1	GDP analogue that inhibits G12C	Lim et al, Angew Chem Int Ed Engl. 2014 [61]
Kobe 0065 and Kobe 2602	Small molecule inhibition of HRas-GTP-c-Raf-1 binding	Shima et al, PNAS 2013 [62]
<i>Andrographis paniculata</i> (AGP) and its derivatives	Inhibition of GTP-GDP exchange in KRAS wild-type and G12V	Hocker et al, PNAS 2013 [63]
Ras-DCAI	Small molecule to interfere with wild-type Ras-SOS interaction	Maurer et al, PNAS 2012 [64]
Not given	Small molecule to interfere with G12D Ras-SOS interaction	Sun et al, Angew Chem Int Ed Engl. 2012 [65]
F929, N944	Mimics protein-protein interaction to interfere with wild-type Ras-SOS interaction	Patgiri et al, Nature Chem Biol 2011 [66]
SCH 54292	Nucleotide exchange inhibitor of wild-type protein	Taveras et al, Bioorg Med Chem 1997 [67]

Table 1. Reported compounds in development that have directly targeted RAS.

FIGURE LEGENDS

Figure 1. *KRAS* mutational subtypes of lung adenocarcinomas analysed in phase I of the Cancer Research UK stratified medicine programme. NOS = mutation of *KRAS* codon 12 and/or 13 not otherwise specified.