

1 **Type of manuscript:** Contributed Paper

2 **Manuscript title:** Sex-biased disease dynamics increase extinction risk by impairing
3 population recovery

4 **Authors:** Gonçalo M. Rosa^{1,2,3*}, Jaime Bosch⁴, An Martel⁵, Frank Pasmans⁵, Rui Rebelo³,
5 Richard A. Griffiths¹, Trenton W.J. Garner²

6 ¹Durrell Institute of Conservation and Ecology, School of Anthropology and
7 Conservation, University of Kent, Canterbury, Kent, CT2 7NR, UK

8 ². Institute of Zoology, Zoological Society of London, Regent's Park, NW1 4RY, London,
9 UK

10 ³. Centre for Ecology, Evolution and Environmental Changes (CE3C), Faculdade de
11 Ciências da Universidade de Lisboa, Campo Grande, 1749-016 Lisboa, Portugal

12 ⁴. Museo Nacional de Ciencias Naturales, CSIC, José Gutiérrez Abascal 2. 28006 Madrid,
13 Spain

14 ⁵. Department of Pathology, Bacteriology and Avian Diseases, Faculty of Veterinary
15 Medicine, Ghent University, Salisburylaan 133, 9820 Merelbeke, Belgium

16 **Author for correspondence:**

17 Gonçalo M. Rosa

18 e-mail: goncalo.m.rosa@gmail.com

19 ORCID id: <https://orcid.org/0000-0002-8658-8436>

20

21 Running head: Sex-biased disease dynamics and extinction risk

22

23 **Sex-biased disease dynamics increase extinction risk by impairing**
24 **population recovery**

25 **Abstract**

26 The periodicity of life-cycle events (phenology) modulates host availability to pathogens
27 in a repeatable pattern. The effects of sexual differences in host phenology have been
28 little explored in wildlife epidemiological studies. A recent series of ranavirosis
29 outbreaks led to serious declines of Boscas' newt populations at Serra da Estrela
30 (Portugal). The peculiar phenology of this species, in which a large number of females
31 remain in the aquatic habitat after the breeding season, turns it into a suitable model to
32 test how sex-biased mortality can affect host population persistence in the context of
33 infectious diseases. We investigated how the phenology of Bosca's newt (i.e. biased
34 number of females) mediated the impact of *Ranavirus*. We then evaluated the risk of
35 extinction of the population under different scenarios of sex-biased mortality using a
36 population viability analysis. Two newt populations (one subject to yearly outbreaks and
37 a comparative site where outbreaks have not been recorded) were tracked for trends
38 over time following emergence of ranaviral disease, allowing us to assess the differential
39 impact of the disease on both sexes. In addition to a significant decline in abundance of
40 adult newts, our data suggest that phenology can affect disease dynamics indirectly,
41 leading to reduction in females and a reversal of the sex ratio of the breeding
42 population. Our models suggest that female-biased mortality does not exacerbate
43 *Ranavirus*-driven population declines in the short-term, but is likely to have a
44 deleterious impact during the recovery process once the lethal effect of disease is
45 removed from the system.

46

47

48 **Keywords.** Bosca's newt; emerging infectious diseases; host-pathogen dynamics;

49 phenology; *Lissotriton boscai*; population viability analysis; *Ranavirus*

50

Introduction

51 Variation in virulence amongst host species or populations (e.g., Daszak *et al.*,
52 2000; Filotas & Hajek, 2004) can be mediated by environmental factors (Dowell, 2001;
53 Roberts & Wiedmann, 2003; Schmeller *et al.*, 2014; Raffel *et al.*, 2015), pathogen type or
54 genotype (Farrer *et al.*, 2011; Price *et al.*, 2014), or host community structure (Begon,
55 2008). However, within-population host heterogeneity may also produce differences in
56 susceptibility to a pathogen as well as recovery, altering the population-level impact of
57 disease, and driving ecological phenomena such as pathogen amplification or dilution
58 (Schmidt & Ostfeld, 2001; Keesing *et al.*, 2006; Searle *et al.*, 2011). There are
59 evolutionary (immunogenetic) explanations for variability of host susceptibility, but
60 ecological factors are more commonly invoked as drivers behind, for example, seasonal
61 patterns of disease prevalence (Altizer *et al.*, 2006; Grassly & Fraser, 2006; Brunner *et al.*,
62 2015). Among these, differences in life-history strategies have been associated with
63 changing rates of pathogen transmission or growth, with phenology potentially playing
64 an important role in host-pathogen dynamics (Visser *et al.*, 2010; Searle *et al.*, 2011).
65 Severe consequences may arise for the host population when pathogen phenology is
66 synchronized with a particularly vulnerable phase of the host life cycle. Nevertheless,
67 there are few empirical studies that explore the relationship between phenology and
68 infectious disease dynamics.

69 Ranaviruses are emerging pathogens of fish, reptiles and amphibians that have
70 caused mass host mortality in North and South America, Australia, Europe and Asia
71 (Schock *et al.*, 2008; Chinchar *et al.*, 2009; Teacher *et al.*, 2010; Chinchar *et al.*, 2011;
72 Chinchar & Waltzek, 2014; Price *et al.*, 2014, 2017; Rosa *et al.*, 2017). Although
73 ranaviruses has been reported for decades, lethal forms of the disease appear to be
74 increasing in incidence and affecting new host populations (Gray *et al.*, 2009; Price *et al.*,

75 2014, 2016), but impacts on host populations are inconsistent. For example, annual
76 ranaviruses epizootics in North American amphibian populations do not appear to be
77 causing population declines (Brunner *et al.*, 2004; Greer *et al.*, 2005; Sutton *et al.*, 2015),
78 contrasting with the pattern in Europe, where catastrophic host population or
79 amphibian community declines have been recorded (Teacher *et al.*, 2010; Kik *et al.*,
80 2011; Price *et al.*, 2014; Miaud *et al.*, 2016; Rosa *et al.*, 2017). The range of European
81 hosts seems to be, at least in part, determined by pathogen genotype (Price *et al.*, 2014;
82 Price, 2015), but single host species population-level (and presumably community-level)
83 responses may be influenced by other ecological factors (Teacher *et al.*, 2010; North *et al.*,
84 2015; Rosa *et al.*, 2017). European amphibian populations experiencing extensive,
85 persistent and recurring mass mortality may, or may not, undergo population declines,
86 despite ongoing episodes of high mortality being expected to decrease the estimated
87 time to population extinction (Lafferty & Gerber, 2002; Teacher *et al.*, 2010; Price *et al.*,
88 2016).

89 In both North America and Europe, increased prevalence of infection and
90 virulence are often associated to specific life history stages, whereas mass mortality
91 events are linked to breeding aggregations (Cunningham *et al.*, 1996; Brunner *et al.*,
92 2004; Whittington *et al.*, 2010; Hoverman *et al.*, 2011; Price *et al.*, 2014). Amphibians
93 are notable for seasonal activity patterns dictated by the relationships between an
94 ectothermic physiology and environmental factors such temperature, rainfall, humidity
95 and sunlight (Duellman & Trueb, 1994). In temperate regions, the majority of amphibian
96 species migrate to breeding sites, forming high density and typically transient adult
97 aggregations. Because in most of these species' reproduction involves the production of
98 single, or at best few egg clutches, and females migrate to breeding sites more or less
99 synchronously, aggregations may form and dissipate in a matter of days or a few weeks.

123 north-central Portugal (Daveau, 1971; Mora *et al.*, 2001), and comprising the largest
124 protected area in Portugal: Serra da Estrela Natural Park (PNSE). Disease outbreaks
125 causing mass mortality emerged in the area of Folgoso in the early autumn of 2011
126 (Rosa *et al.*, 2017) in a 255 m² artificial pond of spring water with constant flow
127 (40°29'37.09"N, 7°31'47.61"W, 1079 m a.s.l.). We monitored another spring water
128 artificial pond at a similar elevation where *Ranavirus* outbreaks have never been
129 recorded but where infection occurs: the 50 m² artificial pond in the Sazes area
130 (40°20'39.70"N, 7°42'52.63"W; 985 m a.s.l.). The two ponds are about 23 km apart, both
131 approximately 1.2-1.7 m deep, located in mountain slopes with the same orientation
132 (facing west), and have the same amphibian assemblage composition breeding regularly
133 (Laurentino *et al.*, 2016; Rosa *et al.*, 2017).

134 *Batrachochytrium dendrobatidis*, a fungal pathogen commonly associated with
135 amphibian die-offs in Iberia and elsewhere, has been present in both sites at least since
136 2010, but no signs of mass mortality associated with chytridiomycosis were recorded in
137 Bosca's newt before (Rosa *et al.*, 2013) or during the study (Rosa *et al.*, 2017).

138 **Host phenology**

139 Bosca's newt, *Lissotriton boscai* (Lataste, 1879) is endemic to the western half of
140 the Iberian Peninsula and relatively common in suitable habitats (Pleguezuelos *et al.*,
141 2002; Loureiro *et al.*, 2008). This species is subject to increasing anthropogenic threats,
142 including loss of breeding pools through drainage for agriculture and urbanization,
143 introduction of non-native predators, and also local outbreaks of emerging pathogens
144 (Soares *et al.*, 2003; Arntzen *et al.*, 2009; Rosa *et al.*, 2017).

145 The breeding migration of *L. boscai* does not involve mass movements (Caetano
146 & Leclair, 1999), and at around 1000 m elevation locations in Serra da Estrela males start

147 migrating to the ponds in November, although most wait until February. Mating takes
148 place in April-June and larvae are present between May-December, although a few
149 might overwinter. The adults start leaving the ponds in June, but some individuals,
150 mostly females, stay in the water year-round (Fig. 1). This sex-specific pattern of
151 breeding site occupation is similar to what has been described in other populations (e.g.,
152 Caetano & Leclair, 1999). In Serra de Sintra, Malkmus (1980-81) also recorded aquatic
153 activity by this species throughout the year, with male abundance as low as 10%
154 between August and December. After metamorphosis, juveniles are predominantly
155 terrestrial until sexual maturity and rarely encountered in ponds before this (Lizana *et*
156 *al.*, 1989).

157 **Survey and disease screening**

158 Newts were sampled in the two sites four times (seasons) per year for 5 years,
159 from 2011 to 2015, with the exception of summer 2014 through to winter 2015. We
160 sampled for 2 to 3 days during each sampling event (capture effort: 2 persons/ 2 hour/
161 site) and newts were captured using dip nets. The largest count per site within the same
162 season was considered the peak abundance for that season and used for analyses. To
163 assess the disease status of each living individual, a small piece of tail tissue or toe was
164 clipped, and from dead specimens a piece of liver was collected. All tissue samples were
165 stored in 70% ethanol for *Ranavirus* assay (St-Amour & Lesbarrères, 2007). Before
166 release, we applied the antiseptic/analgesic Bactine (Bayer, USA) to the clipped tail/ toe
167 (Martin & Hong, 1991). The clipping mark prevented the animals from being re-sampled
168 within a season.

169 DNA was extracted from tissue samples using the DNEasy Tissue Kit (Qiagen,
170 Hilden, Germany). *Ranavirus* was detected by PCR using the MCP4 and 5 primers

171 targeting the viral MCP gene (CMTV ORF 16L; major capsid protein; AFA44920) as
172 described by Mao *et al.* (1996). All PCR assays were run with a negative control (HPLC
173 water) and positive control (DNA extracted from an infected and confirmed ranavirus-
174 positive tadpole) served as controls for the PCR runs. We have previously reported the
175 results of sequence analysis for products generated from Bosca's newt as PNTRV (Stöhr
176 *et al.*, 2015; Rosa *et al.*, 2017).

177 To prevent cross-contamination and the spread of pathogens across sites, we
178 sterilized tools between samples and wore disposable vinyl gloves to handle animals,
179 changing between specimens. Other field equipment used during the surveys, including
180 hiking boots, was periodically immersed in a 1% solution of Virkon® (Antec International
181 Ltd., Sudbury, Suffolk, UK) according to the protocol suggested by Phillott *et al.* (2010).

182 **Population Viability Analyses**

183 Population viability analyses (PVA) were used to evaluate the risk of extinction
184 and probability of population recovery under different scenarios of *Ranavirus* outbreaks.
185 Models were built using VORTEX v. 10.0, which is primarily used to model the probability
186 of extinction of small populations and the relative effects of differing actions and/or
187 perturbations, allowing for individual age- and sex-structured demographic models
188 (<http://www.vortex10.org>; Lacy *et al.*, 2005). A detailed software description and all
189 demographic parameters used in our models are described in the Supporting
190 Information.

191 To simulate the effects of ranavirosis-induced mortality on population stability
192 we tested different "harvest" rates (see below) for larvae and adults (the life stages
193 predominantly making use of the aquatic environment), while holding subadult stage
194 abundance constant (0). Models were constructed considering different scenarios of

195 sex-biased mortality. Stochastic simulation of demographic (and epidemiological)
196 processes was carried out with 10000 iterations and extinction risk was measured as the
197 proportion of populations going extinct within 20 years. We considered extinction to
198 have occurred when only one sex remained. Seven scenarios were considered, testing
199 specific hypotheses generated from field observations:

200 *Scenario 1: no Ranavirus outbreaks;*

201 *Scenario 2: annual Ranavirus outbreaks within the first 5 years of simulation with a*
202 *biased effect on females; at-risk stages were larvae and >3-year-olds; disease*
203 *outbreak caused constant annual offtake of 40% of larvae and 55% of all mature*
204 *individuals (90% females and 10% males; estimation of annual offtake based on*
205 *Rosa et al. (2017) and this study);*

206 *Scenario 3: same as Scenario 2, but with Ranavirus outbreaks affecting both males and*
207 *females equally: constant annual offtake of 40% of larvae, 55% of all mature*
208 *individuals (equal numbers of males and females);*

209 *Scenario 4: annual Ranavirus outbreaks within the first 10 years of simulation with a*
210 *biased effect on females; at-risk stages were larvae and >3-year-olds; disease*
211 *outbreak caused constant annual offtake of 40% of larvae and 55% of all mature*
212 *individuals (90% females and 10% males);*

213 *Scenario 5: same as Scenario 4, but with Ranavirus outbreaks affecting both males and*
214 *females equally: constant annual offtake of 40% of larvae, 55% of all mature*
215 *individuals (equal numbers of males and females).*

216 *Scenario 6: annual Ranavirus outbreaks over the total time of simulation (20 years) with*
217 *a biased effect on females; at-risk stages were larvae and >3-year-olds; disease*

218 outbreak was a constant annual offtake of 40% of larvae and 55% of all mature
219 individuals (90% females and 10% males);

220 *Scenario 7*: same as for *Scenario 6*, but with *Ranavirus* outbreaks affecting both males
221 and females equally: constant annual offtake of 40% of larvae, 55% of all mature
222 individuals (equal numbers of males and females).

223 Scenarios #2 and #3 were set to simulate what we observed in our system but also in
224 other Iberian CMTV-like *Ranavirus* systems, where the pathogen has been present in the
225 community for at least 5 years (Price *et al.*, 2014). Scenarios #4 to #7 reflected mid- to
226 long-term pathogen persistence in the population as long as there are suitable hosts
227 and/ or environmental conditions, as suggested by Teacher *et al.* (2010), and where
228 *Ranavirus* decreases in virulence in the host species, which we modelled as the end of
229 the epidemic after 5 or 10 years. Additionally, the two cut-off points assume the
230 possibility of eventual conservation intervention/mitigation actions that prevent
231 ongoing mortality. We compared female-biased mortality as this reflected what we
232 observed in nature (see Results).

233 **Statistical analysis**

234 Density was calculated by dividing the highest number of individuals captured in
235 a single day per life stage per sampling season by the area of the aquatic habitat
236 ($n/\text{sampled area}$). We used a binary logistic regression to assess the effect of season
237 (season*year) and sex (alone and over time) on the response variable (prevalence of
238 infection). Sex ratio was expressed as the proportion males/(males + females).
239 Comparisons of sex ratios were performed using a Generalized Linear Model (site*time
240 as fixed effects) assuming a binomial error distribution with a logit link function. Post-
241 hoc pairwise comparisons were performed using Bonferroni correction. We used the

242 nonparametric Kruskal-Wallis test to ascertain the differences of extinction probabilities
243 between PVA scenarios. Post-hoc testing was then performed through Dunn-Bonferroni
244 tests to ascertain which pairs of groups differed significantly. Statistical analysis was
245 carried out with software IBM SPSS 20.0 (IBM corp. Chicago, USA).

246 **Results**

247 **Ranavirus and mortality**

248 *Ranavirus* with disease and associated mass mortality was first observed in
249 November (autumn) 2011, where 92.3% (48/52) of Bosca's newts found at Folgosinho
250 were dead. The same scenario occurred annually at about the same time of the year
251 (late summer/early autumn) throughout our field surveys (Figs. 1, 2). Prevalence of
252 infection in live animals broadly mapped with disease dynamics. Prevalence over the
253 two seasons immediately preceding outbreaks or during outbreaks (summer/autumn)
254 consistently averaged out to >75%, while winter/spring averages were significantly
255 lower and, in 2013, close to zero (Fig. 2 and Table S1: Wald $\chi^2 = 7.446$; $df = 1$; $p = 0.006$).
256 Sex-specific prevalences averaged across the entire study (males 34.2%, and females
257 31.2%), with sex not having a significant effect on infection prevalence over time (Wald
258 $\chi^2 = 0.008$; $df = 1$; $p = 0.928$). The majority of dead and dying adult and larval Bosca's
259 newts tested positive for *Ranavirus* (96%). Sick and dead/ moribund animals exhibited
260 all gross signs typical of lethal ranavirosis (see Rosa *et al.*, 2017). Mortality was not
261 recorded during springtime but some positives were detected (e.g., 5%, $n = 20$ in 2013).
262 However, when occasional visits were made to the pond early in the year, dead
263 individuals were observed in the water (> 50 in January and > 10 in March 2012). In
264 contrast, no outbreaks of ranavirosis or mass mortality events were ever recorded at

265 Sazes. Despite virus presence being recorded at the site since 2012 (in salamanders)
266 with detection in Bosca's in 2014 (2/12, 16.7% prevalence), no animals have shown signs
267 of disease (Rosa *et al.*, 2017). At Folgoso, the adult newt population suffered a
268 decline of 45.5% between 2011 and 2012 and of 68.8% between 2011 and 2013. In
269 spring 2014 the decline of the Folgoso population was of 95.5% when compared to
270 the 2011 numbers, before the *Ranavirus* outbreak.

271 **Sex ratio**

272 Populations of newts from both sites showed no differences in the sex ratio in
273 springtime 2011, before the first outbreak of ranavirosis (Fig. 3, Table 1 and S2). At
274 Sazes, where no outbreaks of disease were detected, there was no significant change in
275 newt sex ratios (spring: Wald $\chi^2 = 3.342$; $df = 4$; $p = 0.502$; autumn: Wald $\chi^2 = 2.706$; $df =$
276 2 ; $p = 0.258$; Fig. 3, Table 1 and S2) over the period of this study.

277 During the first outbreak in Folgoso, the majority of the adults (91.7%) found
278 dead were females. Females also comprised the majority of dead animals (64.4% across
279 all life stages) throughout the course of the study (Fig. S1). In the following years we
280 noted a reversal in the sex ratio, which shifted from 25% males in late spring 2011 to
281 over 60% in the subsequent years (Fig. 3). There was a significant effect of time after the
282 first outbreak on the sex ratio in Folgoso in springtime (Wald $\chi^2 = 38.075$; $df = 3$; $p <$
283 0.001), with a marked difference between the first year and subsequent years (Table
284 1 and S2). The effect was detectable in the first autumn (2011), such that the proportion
285 of males found in the autumn across years did not vary significantly (Wald $\chi^2 = 0.638$; df
286 $= 2$; $p = 0.727$; see Table 1). Larvae comprised a smaller proportion of dead individuals
287 over the entire study (28%) than adults (70%; Fig. S1).

288 **Risk assessment and population viability**

289 In all scenarios including disease outbreaks, there was a rapid decline in
290 abundance to approximately 15% of the pre-outbreak population size, paralleling values
291 observed in the field (95.5% decline until 2014 and 70.6% by 2015 compared to 2011;
292 Fig. 2). Increasing persistence of annual mortality events caused by ranaviruses and
293 altering the sex bias in probability of mortality had no strong effect on patterns of
294 population declines over the first five years of the simulations. The effect of a 5-year
295 period of female-biased mortality was not significantly different from the effect of mid-
296 and long-term persistence of unbiased mortality (Fig. 4; Fig. S2; Table 3). However, both
297 the persistence of declines and post-decline recovery rates were significantly affected by
298 sex-biased mortality, as well as by sex-unbiased mortality (K-W: $\chi^2 = 55.163$; $df = 6$; $p <$
299 0.001 ; Table 3). Specifically, and perhaps unsurprisingly, population recoveries began as
300 soon as annual mortality events ceased. However, the rate of recovery was significantly
301 slower when ranaviruses events lasted ten years when compared to populations where
302 disease ceased after five years (Fig. 4; Fig. S2). Rates were further depressed when
303 mortality was sex-biased: for example, five years after the last outbreak, population size
304 estimated in scenario #3 was 1.8 times greater than for projection #2 (Fig. 4; Fig. S2). In
305 scenarios where disease persisted, a high likelihood of population extirpation was
306 predicted, greater so when mortality was sex-biased (98%, versus 88% when risk of
307 mortality was equal across the sexes). This difference in risk reflected a difference in
308 rate of decline after the 5-year period: median time to extinction was 11 years when
309 mortality was female biased versus 16 years (Table 2).

310 Discussion

311 Seasonality of mortality events is not uncommon in ranavirus epidemics and is
312 often invoked as a covariate that affects viral growth dynamics (Pfennig *et al.*, 1991;

313 Dowell, 2001; Rojas *et al.*, 2005; Gray *et al.*, 2007, 2009; Price *et al.*, 2018). Our data
314 suggest seasonality can also affect disease dynamics indirectly through the breeding
315 phenology of amphibians, e.g., by eliciting sex-specific mortality schedules. Sex-biased
316 mortality is not uncommon across different vertebrate groups (e.g., Müller *et al.*, 2005;
317 Sperry & Weatherhead, 2009), and when skewed towards females is expected to lead to
318 accelerated population declines. For example, Gruebler *et al.* (2008) showed that
319 female-biased mortality of whinchats could lead to a 1.7 times faster local population
320 decline. In support of this, we recorded a significant decline in abundance of adult newts
321 at Folgoso (Portugal), but also a reversal of the sex ratio of the breeding population
322 and concurrent disproportionate reduction in female abundance (Fig. 2, 3). Although we
323 cannot determine whether population decline was a simple direct effect of adult (and
324 larval) mortality or if it was also affected by a reduction in recruitment due to a
325 decreasing availability of breeding females, our models show that either of the two
326 mechanisms is enough to drive the rate of decline we observed at Folgoso.

327 Although recovery after declines due to ranavirus can occur (Greer *et al.*,
328 2008), ranaviruses do meet the conditions required to cause host extinction (Miller *et al.*,
329 2011). A study by Earl & Gray (2014) modelled the likelihood of extinction of a closed
330 population of wood frogs (*Lithobates sylvaticus*) and showed that extinction could
331 theoretically occur as quickly as 5 years if ranavirus persisted over that time span. Our
332 analyses and models suggest that even if the recurring ranavirus outbreaks do not
333 drive the host to complete extinction, they can reduce population size to a point where
334 stochastic events can eliminate host populations (de Castro & Bolker, 2005). Even if
335 extirpation is avoided, recovery may not occur (Schock & Bollinger, 2005; Teacher *et al.*,
336 2010). This is also a key finding of our modelling, where we obtained a >88% probability

337 that extinction will occur within a median of 11 to 16 years if outbreaks occur yearly for
338 10 years, even if they are totally controlled after that.

339 More importantly, we found that female-biased mortality did not exacerbate
340 *Ranavirus*-driven population decline in the short-term, but impaired population recovery
341 once the lethal effect of disease was removed from the system. With that in mind, and
342 assuming the possibility of eventual conservation intervention/mitigation actions based
343 in our projections, this finding suggests that pathogen mitigation does not imply
344 population recovery. As a result of reduced recruitment rates, our models predict a
345 slower population growth, which could be offset if the female population is augmented.

346 What is not considered in our study, though, is the effect of other possible
347 reservoir hosts on both declines and recovery. Our study species exists in a multi-host
348 community system (Rosa *et al.*, 2017), as are other CMTV-affected sites in Western
349 Europe (see Kik *et al.*, 2011; Price *et al.*, 2014; Miaud *et al.*, 2016). Given the broad host
350 range and the potential for ranaviruses to persist in the environment, we are likely
351 underestimating the risk of extinction for Bosca's newts.

352 Previous reports have shown that other populations of *L. boscai* have been
353 affected by ranavirosis outbreaks throughout the northern half of the species
354 distribution range (Soares *et al.*, 2003; Price *et al.*, 2014; Rosa *et al.*, 2017). Moreover,
355 increasing threats to ponds by wildfires, desiccation and other human activities, even
356 within protected areas like Serra da Estrela (ICNB, 2008; Vicente *et al.*, 2013), raise
357 additional concerns from the population management point of view. Vulnerability of
358 these freshwater habitats raises challenges for the conservation of newts and other
359 species affected by *Ranavirus*. However, declines due to ranavirosis are not
360 deterministic, as newts at Sazes have experienced low-level infections over a similar
361 time span and also exhibit similar sex-specific breeding phenologies, but prevalence has

362 yet to reach saturation and newt mass mortality does not occur. Locations like Sazes
363 offer the opportunity to characterize the factors allowing host-*Ranavirus* coexistence.
364 This is important, as unlike the situation with *Batrachochytrium dendrobatidis* (see
365 Bosch *et al.*, 2015; Garner *et al.*, 2016), no successful treatment or mitigation measure
366 for ranavirosis has been published.

367

368 **Supporting Information**

369 A detailed PVA software description and all demographic parameters used in our models
370 are described in the Supporting Information, and can be found online at <https://XXXXXX>

371

372 **Acknowledgements**

373 We thank José Conde (CISE), Marco Saraiva (ICNF), and Ricardo Brandão (CERVAS) for all
374 the support and logistics. Ana Ferreira, Ana Marques, Andreia Penado, Diogo Veríssimo,
375 Isabela Berbert, Joana Sabino Pinto, Maria Alho, Marta Palmeirim, Marta Sampaio,
376 Miguel Pais, Ninda Baptista, Pedro Patrício, and Telma G. Laurentino for all the help in
377 the field; Madalena Madeira for the unconditional help in the field but also with the
378 schematic graph design. We also thank Bob Lacy and Phil Miller for advice about the
379 software VORTEX. Research permits were provided by the Instituto da Conservação da
380 Natureza e das Florestas. G. M. Rosa held a doctoral scholarship (SFRH/BD/69194/2010)
381 from Fundação para a Ciência e a Tecnologia (FCT). J. Bosch was supported by Spanish
382 Ministry of Economy and Competitiveness grant CGL2015-70070-R.

383 **References**

- 384 Altizer, S., Dobson, A., Hosseini, P., Hudson, P., Pascual, M. & Rohani, P. (2006).
385 Seasonality and the dynamics of infectious diseases. *Ecol. Lett.* **9**, 467–484.
- 386 Arntzen, J.W., Beja, P., Jehle, R., Bosch, J., Tejedo, M., Lizana, M., Martínez-Solano, I.,
387 Salvador, A., García-París, M., Recuero Gil, E., Sá-Sousa, P. & Marquez, R. (2009).
388 *Lissotriton boscai*. The IUCN Red List of Threatened Species 2009,
389 e.T59473A11947331.
390 <http://dx.doi.org/10.2305/IUCN.UK.2009.RLTS.T59473A11947331.en>.
391 <www.iucnredlist.org>. Downloaded on April 2018.
- 392 Begon, M. (2008). Effects of host diversity on disease dynamics. in *Effects of ecosystems*
393 *on disease and of disease on ecosystems*: 12–29. Ostfeld, R.S., Keesing, F. &
394 Eviner, V.T. (Eds.). Princeton: Princeton University Press.
- 395 Bosch, J., Sanchez-Tomé, E., Fernández-Loras, A., Oliver, J.A., Fisher, M.C. & Garner,
396 T.W.J. (2015). Successful elimination of a lethal wildlife infectious disease in
397 nature. *Biol. Lett.* **11**, 20150874.
- 398 Brockes, J. & Kumar, A. (2005). Newts. *Curr. Biol.* **15**, R42–R44.
- 399 Brunner, J.L., Schock, D.M., Davidson, E.W. & Collins, J.P. (2004). Intraspecific reservoirs:
400 complex life history and the persistence of a lethal *Ranavirus*. *Ecology* **85**, 560–
401 566.
- 402 Brunner, J.L., Storfer, A., Gray, M.J. & Hoverman, J.T. (2015). *Ranavirus* ecology and
403 evolution: From epidemiology to extinction. in *Ranaviruses: Lethal pathogens of*
404 *ectothermic vertebrates*: 71–104. Gray, M.J. & Chinchar, V.G. (Eds.). Secaucus:
405 Springer.

406 Caetano, M.H. & Leclaire, R. (1999). Comparative phenology and demography of *Triturus*
407 *boscai* from Portugal. *J. Herpetol.* **33**, 192–202.

408 Chinchar, V.G., Hyatt, A., Miyazaki, T. & Williams, T. (2009). Family Iridoviridae: poor
409 viral relations no longer. *Curr. Top. Microbiol. Immunol.* **328**, 123–170.

410 Chinchar, V.G., Robert, J. & Storfer, A.T. (2011). Ecology of viruses infecting ectothermic
411 vertebrates: The impact of *Ranavirus* infections on amphibians. in *Studies in*
412 *Viral Ecology: Animal Host Systems*: **2**, 231–259. Hurst, C.J. (Ed.). Wiley-
413 Blackwell.

414 Chinchar, V.G. & Waltzek, T.B. (2014). Ranaviruses: Not just for frogs. *PLoS Pathog.* **10**,
415 e1003850.

416 Cunningham, A.A., Langton, T.E.S., Bennett, P.M., Lewin, J.F., Drury, S.E.N., Gough, R.E.
417 & Macgregor, S.K. (1996). Pathological and microbiological findings from
418 incidents of unusual mortality of the common frog (*Rana temporaria*). *Phil.*
419 *Trans. R. Soc. Lond. B Biol. Sci.* **351**, 1539–1557.

420 Daveau, S. (1971). La glaciation de la Serra da Estrela. *Finisterra* **6**, 5–40.

421 Daszak, P., Cunningham, A.A. & Hyatt, A.D. (2000). Emerging infectious diseases of
422 wildlife—threats to biodiversity and human health. *Science* **287**, 443–449.

423 de Castro, F. & Bolker, B. (2005). Mechanisms of disease-induced extinction. *Ecol. Lett.* **8**,
424 117–126.

425 Díaz-Paniagua, C. (1998). Reproductive dynamics of a population of small marbled newts
426 (*Triturus marmoratus pygmaeus*) in south-western Spain. *Herpetol. J.* **8**, 93–98.

427 Díaz-Paniagua, C. & Mateo, J.A. (1999). Geographic variation in body size and life history
428 traits in *Triturus boscai*. *Herpetol. J.* **9**, 21–27.

429 Dowell, S.F. (2001). Seasonal variation in host susceptibility and cycles of certain
430 infectious diseases. *Emerg. Infect. Dis.* **7**, 369–374.

431 Duellman, W.E. & Trueb, L. (1994). *Biology of the amphibians*. Baltimore & London:
432 Johns Hopkins University Press.

433 Earl, J.E. & Gray, M.J. (2014). Introduction of Ranavirus to isolated wood frog
434 populations could cause local extinction. *EcoHealth* **11**, 581–592.

435 Farrer, R.A., Weinert, L.A., Bielby, J., Garner, T.W.J., Balloux, F., Clare, F., Bosch, J.,
436 Cunningham, A.A., Weldon, C., du Preez, L.H., Anderson, L., Pond S.L.K., Shahar-
437 Golan, R., Henk, D.A. & Fisher, M.C. (2011). Multiple emergences of amphibian
438 chytridiomycosis include a globalised hypervirulent recombinant lineage. *Proc.*
439 *Natl. Acad. Sci.* **108**, 18732–18736

440 Filotas, M.J. & Hajek, A.E. (2004). Influence of temperature and moisture on infection of
441 forest tent caterpillars (Lepidoptera: Lasiocampidae) exposed to resting spores
442 of the entomopathogenic fungus *Furia gastropachae* (Zygomycetes:
443 Entomophthorales). *Environ. Entomol.* **33**, 1127–1136.

444 Garner, T.W.J., Schmidt, B.R., Martel, A., Pasmans, F., Muths, E., Cunningham, A.A.,
445 Weldon, C., Fisher, M.C. & Bosch, J. (2016). Mitigating amphibian
446 chytridiomycoses in nature. *Philos. Trans. R. Soc. Lond B Biol. Sci.* **371**, 20160207.

447 Grassly, N.C. & Fraser, C. (2006). Seasonal infectious disease epidemiology. *Proc. R. Soc.*
448 *Lond. B Biol. Sci.* **273**, 2541–2550.

449 Gray, M.J., Miller, D.L. & Hoverman, J.T. (2009). Ecology and pathology of amphibian
450 ranaviruses. *Dis. Aquat. Organ.* **87**, 243–266.

451 Gray, M.J., Miller, D.L., Schmutzer, A.C. & Baldwin, C.A. (2007). Frog virus 3 prevalence in
452 tadpole populations inhabiting cattle-access and non-access wetlands in
453 Tennessee, USA. *Dis. Aquat. Organ.* **77**, 97–103.

454 Greer, A.L., Berrill, M. & Wilson, P.J. (2005). Five amphibian mortality events associated
455 with *Ranavirus* infection in south central Ontario, Canada. *Dis. Aquat. Organ.* **67**,
456 9–14.

457 Greer, A.L., Briggs, C.J. & Collins, J.P. (2008). Testing a key assumption of host-pathogen
458 theory: Density and disease transmission. *Oikos* **117**, 1667–1673.

459 Griffiths, R.A. (1997). *The newts and salamanders of Europe*. London & San Diego: T. &
460 A.D. Poyser/Academic Press.

461 Gruebler, M.U., Schuler, H., Müller, M., Spaar, R., Horch, P. & Naef-Daenzer, B. (2008).
462 Female biased mortality caused by anthropogenic nest loss contributes to
463 population decline and adult sex ratio of a meadow bird. *Biol. Conserv.* **141**,
464 3040–3049.

465 Hoverman, J.T., Gray, M.J., Haislip, N.A. & Miller, D.L. (2011). Phylogeny, life history, and
466 ecology contribute to differences in amphibian susceptibility to ranaviruses.
467 *EcoHealth* **8**, 301–319.

468 ICNB (2008). Relatório Ambiental: Plano de Ordenamento do Parque Natural da Serra da
469 Estrela. Instituto de Conservação da Natureza e Biodiversidade, 47pp (available
470 from:
471 [http://www.icnf.pt/portal/pn/biodiversidade/ordgest/poap/popnse/resource/o](http://www.icnf.pt/portal/pn/biodiversidade/ordgest/poap/popnse/resource/ordenam/relat-ambiental)
472 [rdenam/relat-ambiental](http://www.icnf.pt/portal/pn/biodiversidade/ordgest/poap/popnse/resource/ordenam/relat-ambiental))

473 Keesing, F., Holt, R.D. & Ostfeld, R.S. (2006). Effects of species diversity on disease risk.
474 *Ecol. Lett.* **9**, 485–498.

- 475 Kik, M., Martel, A., Spitzen-van der Sluijs, A., Pasmans, F., Wohlsein, P., Gröne, A. & Rijks,
476 J.M. (2011). Ranavirus-associated mass mortality in wild amphibians, The
477 Netherlands, 2010: A first report. *Vet. J.* **190**, 284–286.
- 478 Lacy, R.C., Borbat, M. & Pollack, J.P. (2005). *VORTEX: A Stochastic Simulation of the*
479 *Extinction Process*. Brookfield, IL: Chicago Zoological Society.
- 480 Lafferty, K.D. & Gerber, L.R. (2002). Good medicine for conservation biology: the
481 intersection of epidemiology and conservation theory. *Conserv. Biol.* **16**, 593–
482 604.
- 483 Laurentino, T.G., Pais, M.P. & Rosa, G.M. (2016). From a local observation to a
484 European-wide phenomenon: amphibian deformities at Serra da Estrela Natural
485 Park, Portugal. *Basic Appl. Herpetol.* **30**, 7–23.
- 486 Lizana, M., Ciudad, M.J. & Pérez-Mellado, V. (1989). Actividad, reproducción y uso del
487 espacio en una comunidad de anfibios. *Treb. Soc. Cat. Ictio. Herp.* **2**, 92–127.
- 488 Loureiro, A., Ferrand de Almeida, N., Carretero, M.A., Paulo, O.S. (Eds.). (2008). *Atlas dos*
489 *Anfibios e Répteis de Portugal*. Lisboa: Instituto da Conservação da Natureza e da
490 Biodiversidade.
- 491 Malkmus, R. (1980–81). Bemerkungen zu einer *Triturus boscai* population in einem
492 brunnenbecken der Serra de Sintra. *Bol. Soc. Port. Ci. Nat.* **20**, 25–40.
- 493 Mao, J., Tham, T.N., Gentry, G.A., Aubertin, A. & Chinchar, V.G. (1996). Cloning,
494 sequence analysis, and expression of the major capsid protein of the iridovirus
495 frog virus 3. *Virology* **216**, 431–436.
- 496 Martin, D. & Hong, H. (1991). The use of Bactine in the treatment of open wounds and
497 other lesions in captive anurans. *Herpetol. Rev.* **22**, 21–21.

498 Miller, D., Gray, M. & Storfer, A. (2011). Ecopathology of ranaviruses infecting
499 amphibians. *Viruses* **3**: 2351–2373.

500 Minchella, D. & Scott, M. (1991). Parasitism: a cryptic determinant of animal community
501 structure. *Trends Ecol. Evol.* **6**, 250–254.

502 Mora, C., Vieira, G. & Alcoforado, M.J. (2001). Daily minimum air temperatures in the
503 Serra da Estrela, Portugal. *Finisterra* **36**, 49–59.

504 Müller, W., Grootuis, T.G.G., Eising, C.M., Dijkstra, C. (2005). An experimental study on
505 the causes of sex-biased mortality in the black-headed gull – the possible role of
506 testosterone. *J. Anim. Ecol.* **74**, 735–741.

507 North, A.C., Hodgson, D.J., Price, S.J. & Griffiths, A.G.F. (2015). Anthropogenic and
508 ecological drivers of amphibian disease (ranavirosis). *PLoS ONE* **10**, e0127037.

509 Paterson, S., Vogwill, T., Buckling, A., Benmayor, R., Spiers, A.J., Thomson, N.R., Quail,
510 M., Smith, F., Walker, D., Libberton, B., Fenton, A., Hall, N. & Brockhurst, M.A.
511 (2010). Antagonistic coevolution accelerates molecular evolution. *Nature* **464**,
512 275–278.

513 Pfennig, D.W., Loeb, M.L.G. & Collins, J.P. (1991). Pathogens as a factor limiting the
514 spread of cannibalism in tiger salamanders. *Oecologia* **88**, 161–166.

515 Phillott, A.D., Speare, R., Hines, H.B., Skerratt, L.F., Meyer, E., McDonald, K.R., Cashins,
516 S.D., Mendez, D. & Berger, L. (2010). Minimising exposure of amphibians to
517 pathogens during field studies. *Dis. Aquat. Organ.* **92**, 175–185.

518 Pleguezuelos, J.M., Márquez, R. & Lizana, M. (Eds.). (2002). *Atlas y libro rojo de los*
519 *anfíbios y reptiles de España*. Madrid: Dirección General de Conservación de la
520 Naturaleza / Asociación Herpetológica Española (2ª impresión).

521 Price, S.J. (2015). Comparative genomics of amphibian-like ranaviruses,
522 nucleocytoplasmic large DNA viruses of poikilotherms. *Evol. Bioinform.* (Suppl. 2)
523 **11**, 71–82.

524 Price, S.J., Ariel, E., Maclain, A., Rosa, G.M., Gray, M.J., Brunner, J.L. & Garner, T.W.J.
525 (2017). From fish to frogs and beyond: Impact and host range of emergent
526 ranaviruses. *Virology* **511**, 272–279.

527 Price, S.J., Garner, T.W.J., Nichols, R.A., Balloux, F., Ayres, C., Mora-Cabello de Alba, A. &
528 Bosch, J. (2014). Collapse of amphibian communities due to an introduced
529 *Ranavirus*. *Curr. Biol.* **24**, 2586–2591.

530 Price, S.J., Garner, T.W.J., Cunningham, A.A., Langton, T.E.S., Nichols, R.A. (2016).
531 Reconstructing the emergence of an infectious disease of wildlife supports a key
532 role for spread through translocations by humans. *Proc. R. Soc. Lond. B Biol. Sci.*
533 **283**, 20160952.

534 Price, S.J., Leung, W.T.M., Owen, C., Sergeant, C., Cunningham, A.A., Balloux, F., Garner,
535 T.W.J. & Nichols, R.A. (2018). Temperature is a key driver of a wildlife epidemic
536 and future warming will increase impacts. *BioRxiv*
537 <https://doi.org/10.1101/272369>

538 Raffel, T.R., Halstead, N.T., McMahon, T.A., Davis, A.K. & Rohr, J.R. (2015). Temperature
539 variability and moisture synergistically interact to exacerbate an epizootic
540 disease. *Proc. R. Soc. Lond. B Biol. Sci.* **282**, 20142039.

541 Roberts, A.J. & Wiedmann, M. (2003). Pathogen, host and environmental factors
542 contributing to the pathogenesis of listeriosis. *Cell. Mol. Life Sci.* **60**, 904–918.

543 Rojas, S., Richards, K., Jancovich, J.K. & Davidson, E.W. (2005). Influence of temperature
544 on *Ranavirus* infection in larval salamanders *Ambystoma tigrinum*. *Dis. Aquat.*
545 *Organ.* **63**, 95–100.

546 Rosa, G.M., Anza, I., Moreira, P.L., Conde, J., Martins, F., Fisher, M.C. & Bosch, J. (2013).
547 Evidence of chytrid-mediated population declines in common midwife toad in
548 Serra da Estrela, Portugal. *Anim. Conserv.* **16**, 306–315.

549 Rosa, G.M., Sabino-Pinto, J., Laurentino, T.G., Martel, A., Pasmans, F., Rebelo, R.,
550 Griffiths, R.A., Stöhr, A.C., Marschang, R.E., Price, S.J., Garner, T.W.J. & Bosch, J.
551 (2017). Impact of asynchronous emergence of two lethal pathogens on
552 amphibian assemblages. *Sci. Rep.* **7**, 43260.

553 Schmeller, D.S., Blooi, M., Martel, A., Garner, T.W.J., Fisher, M.C., Azemar, F., Clare, F.C.,
554 Leclerc, C., Jäger, L., Guevara-Nieto, M., Loyau, A. & Pasmans, F. (2014).
555 Microscopic aquatic predators strongly affect infection dynamics of a globally
556 emerged pathogen. *Curr. Biol.* **24**, 176–180.

557 Schmidt, K.A. & Ostfeld, S. (2001). Biodiversity and the dilution effect in disease ecology.
558 *Ecology* **82**, 609–619.

559 Schock, D.M. & Bollinger, T.K. (2005). An apparent decline of northern leopard frogs
560 (*Rana pipiens*) on the Rafferty Dam mitigation lands near Estevan,
561 Saskatchewan. *Blue Jay* **63**: 144–154.

562 Schock, D.M., Bollinger, T.K., Chinchar, V.G., Jancovich, J.K. & Collins, J.P. (2008).
563 Experimental evidence that amphibian ranaviruses are multi-host pathogens.
564 *Copeia* **2008**, 133–143.

565 Searle, C.L., Gervasi, S.S., Hua, J., Hammond, J.I., Relyea, R.A., Olson, D.H. & Blaustein,
566 A.R. (2011). Differential host susceptibility to *Batrachochytrium dendrobatidis*,
567 an emerging amphibian pathogen. *Conserv. Biol.* **25**, 965–974.

568 Soares, C., Alves de Matos, A., Arntzen, J.W., Carretero, M. & Loureiro, A. (2003).
569 Amphibian mortality in a national park in the north of Portugal. *FrogLog* **56**, 1–1.

570 Sperry, J.H. & Weatherhead, P.J. (2009). Sex differences in behavior associated with sex-
571 biased mortality in an oviparous snake species. *Oikos* **118**, 627–633.

572 St-Amour, V. & Lesbarrères, D. (2007). Genetic evidence of *Ranavirus* in toe clips: An
573 alternative to lethal sampling methods. *Conserv. Genet.* **8**, 1247–1250.

574 Stöhr, A.C., López-Bueno, A., Blahak, S., Caeiro, M.F., Rosa, G.M., Alves de Matos, A.P.,
575 Martel, A., Alejo, A. & Marschang, R. (2015). Phylogeny and differentiation of
576 reptilian and amphibian ranaviruses detected in Europe. *PLoS ONE* **10**,
577 e0118633.

578 Sutton, W.B., Gray, M.J., Hoverman, J.T., Secrist, R.G., Super, P., Hardman, R.H., Tucker,
579 J.L. & Miller, D.L. (2015). Trends in *Ranavirus* prevalence among plethodontid
580 salamanders in the Great Smoky Mountains National Park. *EcoHealth* **12**, 320–
581 329.

582 Teacher, A.G.F., Cunningham, A.A. & Garner, T.W.J. (2010). Assessing the long-term
583 impact of *Ranavirus* infection in wild common frog populations. *Anim. Conserv.*
584 **13**, 514–522.

585 Vicente, F., Cesari, M., Serrano, A. & Bertolani, R. (2013). The impact of fire on terrestrial
586 tardigrade biodiversity: A first case-study from Portugal. *J. Limnol.* **72**. 152–159.

- 587 Visser, M.E., Caro, S.P., van Oers, K., Schaper, S.V. & Helm, B. (2010). Phenology,
588 seasonal timing and circannual rhythms: towards a unified framework. *Philos.*
589 *Trans. R. Soc. Lond. B Biol. Sci.* **365**, 3113–3127.
- 590 Whittington, R.J., Becker, J.A. & Dennis, M.M. (2010). Iridovirus infections in finfish –
591 critical review with emphasis on ranaviruses. *J. Fish Dis.* **33**: 95–122.

CAPTIONS FOR TABLES

593 **Table 1** Pairwise comparisons of springtime sex ratios between two populations of
 594 *Lissotriton boscai* over time at Serra da Estrela (Portugal) using a Generalized Linear
 595 Model (site*year). Folgoso: yearly outbreaks of *Ranavirus* record (2011 represent the
 596 sex ratio before the first outbreak); Sazes: no *Ranavirus* outbreak recorded. A *p* value <
 597 0.05 was considered significant and highlighted in bold after Bonferroni correction.

Folgoso	2011	2012	2013	Sazes	2011	2012	2013	2014
2012	< 0.01			2012	1.00			
2013	< 0.01	1.00		2013	1.00	1.00		
2014	††	††	††	2014	1.00	1.00	1.00	
2015	< 0.01	1.00	1.00	2015	1.00	1.00	1.00	1.00

Sites	year	Bonferroni adj. <i>p</i> value
	2011	1.00
Folgoso*Sazes	2012	< 0.01
	2013	0.02
	2015	< 0.01

598 †† No data were considered for Folgoso site in spring 2014.

599 **Table 2** PVA outputs from simulation of disease outbreaks in a population of *Lissotriton*
600 *boscai* under seven different disease mortality offtakes. Adult (55%) and larvae mortality
601 (40%) is constant across scenarios #2–#7 (varying in time and sex offtake) but absent in
602 #1. PE: probability of population extinction after 20 yrs; GR: stochastic growth rate; N:
603 mean population size after 20 yrs; TE: time to first population extinction; SD: standard
604 deviation. Grey shading highlights scenarios with sex-biased mortality.

Scenario	PE	GR ± SD	N ± SD	Median TE
#1. no <i>Ranavirus</i> outbreaks	0.001	0.206 ±0.606	4104.95 ±1551.87	0
#2. female biased mortality within 5 yrs	0.132	-0.026 ±0.661	2325.63 ±2112.10	0
#3. equal sex mortality within 5 yrs	0.009	0.046 ±0.677	3390.51 ±1916.13	0
#4. female biased mortality within 10 yrs	0.580	-0.293 ±0.634	237.41 ±714.17	12
#5. equal sex mortality within 10 yrs	0.164	-0.149 ±0.687	769.65 ±1246.74	0
#6. female biased mortality within 20 yrs	0.984	-0.516 ±0.393	0.12 ±0.84	11
#7. equal sex mortality within 20 yrs	0.880	-0.452 ±0.472	0.67 ±2.44	16

605

606 **Table 3** Dunn-Bonferroni test between pairs of different scenarios generated from PVA,
 607 to look at significant differences in probability of extinction. Adjusted p value < 0.05 was
 608 considered significant and highlighted in bold.

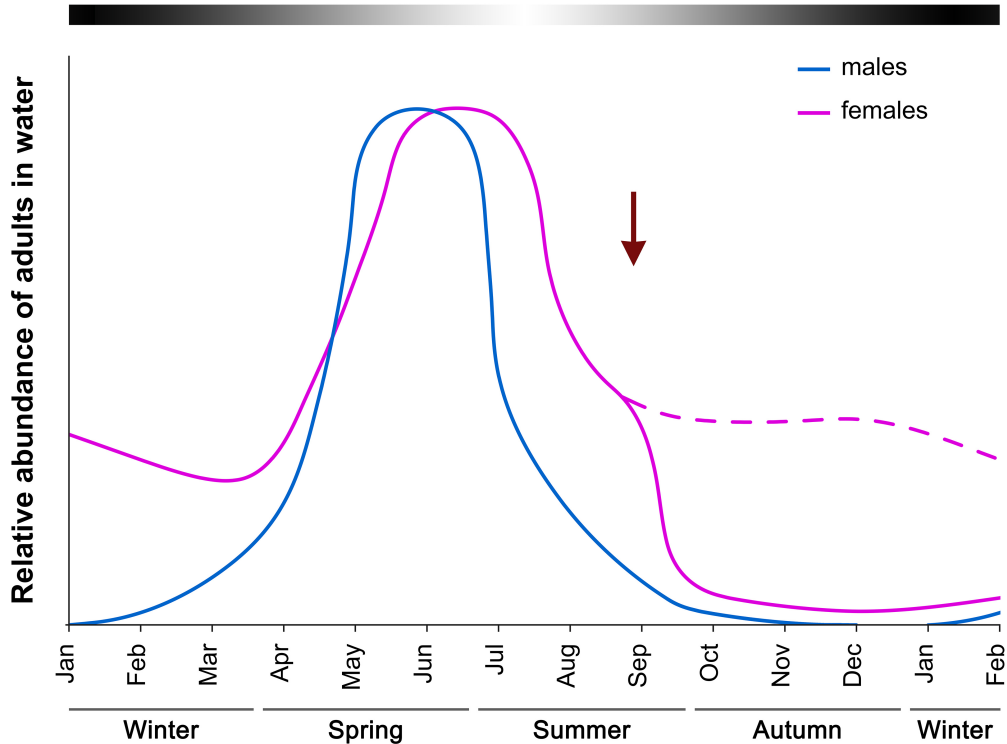
Scenario	1	2	3	4	5	6
2	< 0.01					
3	1.00	0.26				
4	< 0.01	1.00	< 0.01			
5	0.03	1.00	1.00	0.31		
6	< 0.01	1.00	< 0.01	1.00	0.10	
7	< 0.01	1.00	0.22	1.00	1.00	1.00

609

CAPTIONS FOR FIGURES

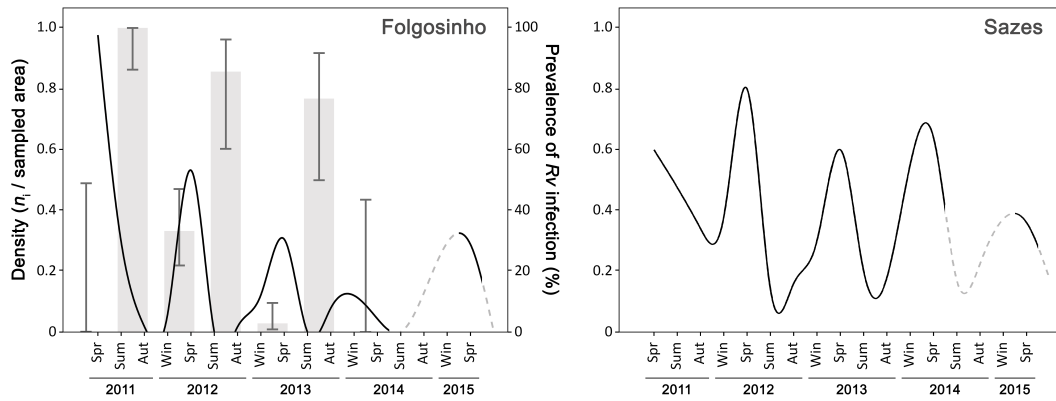
610

611



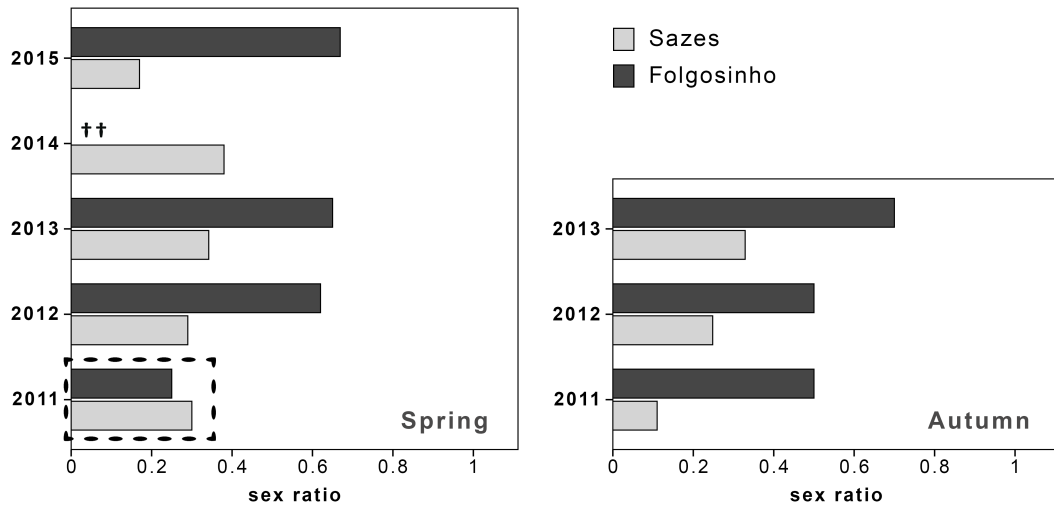
612

613 **Figure 1** Schematic phenology of the host species, Bosca's newt (*Lissotriton boscai*) in
614 Serra da Estrela (Portugal) in ponds located at about 800–1100 m of elevation showing a
615 representation of expected relative proportion of males and females in the water
616 throughout the year. Arrow indicates time of the first outbreak of ranavirosis (2011) and
617 dashed line illustrates the expected relative abundance of females in the absence of
618 outbreak (based on relative data from previous years and other similar ponds, collected
619 as part of ongoing amphibian monitoring studies; Rosa *et al.*, 2013, 2017; Laurentino *et*
620 *al.*, 2016). Top bar represents the temperature throughout the year from cool (dark) to
621 warm (light).



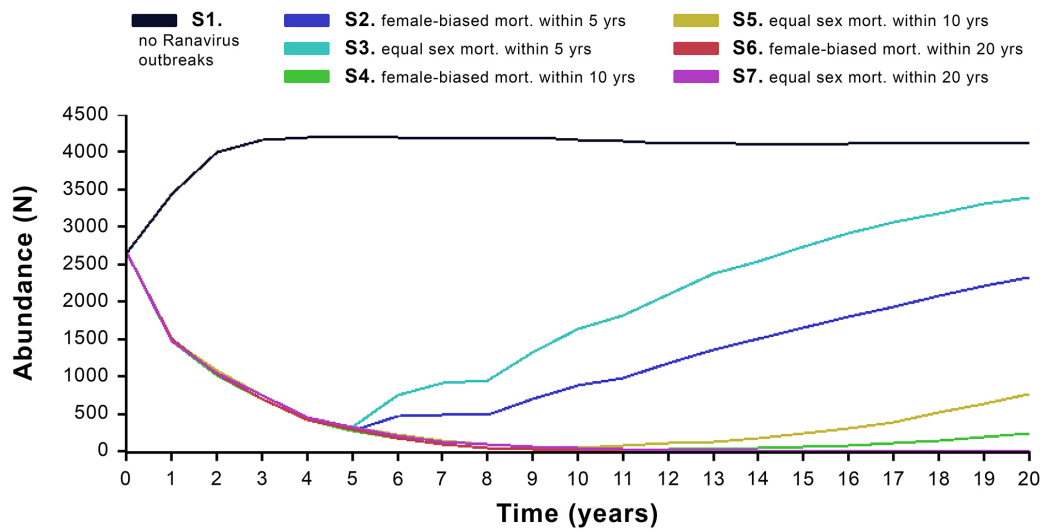
622

623 **Figure 2** Density of adult Bosca's newt (*Lissotriton boscai*) in two sites in Serra da Estrela
 624 (Portugal) over five years (spline interpolation with data missing between summer 2014
 625 and winter 2015): Folgosinho shows the density of newts with yearly outbreaks of
 626 ranaviruses, while Sazes shows a natural population fluctuation in an area where
 627 outbreaks have not been recorded. Prevalence of *Ranavirus* infection in Bosca's newts is
 628 also provided for Folgosinho pond (grey bars) as the total proportion of infected
 629 individuals by two grouped seasons (summer+autumn and winter+spring), where error
 630 bars indicate the 95% confidence intervals.



631

632 **Figure 3** Comparative sex ratio of adult Bosca's newts (*Lissotriton boscai*) in two sites in
 633 Serra da Estrela (Portugal) over five years. Folgosinho shows the sex ratio of a
 634 population facing yearly outbreaks of ranavirosis after spring 2011, while Sazes shows
 635 the expected sex ratio of a population where outbreaks have not been recorded. Sex
 636 ratio is expressed as the proportion of males/(males + females) in two different seasons:
 637 spring and autumn. Dashed rectangle highlights the sex ratio in both populations before
 638 the first outbreak of ranavirosis, when there was no difference between them (*L. boscai*
 639 $p > 0.05$). †† No data for Folgosinho site in spring 2014.



640

641 **Figure 4** 20-year population projection for Bosca's newt (*Lissotriton boscai*) in VORTEX
 642 under seven different disease scenarios with no management: S1. No *Ranavirus*
 643 outbreaks; S2. Female-biased mortality during 5 years; S3. Equal sex mortality during 5
 644 years; S4. Female-biased mortality during 10 years; S5. Equal sex mortality during 10
 645 years; S6. Female-biased mortality during 20 years; S7. Equal sex mortality during 10
 646 years.