Dementia associated with disorders of the basal ganglia

Dementia associated with disorders of the basal ganglia

Vitanova, K. S.\*1, Stringer, K. M.\*1,2, Benitez, D. P.\*1, Brenton, J.\*1 & Cummings, D. M.1

<sup>1</sup>Dept Neuroscience, Physiology & Pharmacology, University College London, Gower

Street, London, United Kingdom, WC1E 6BT.

<sup>2</sup>Department of Psychiatry and Neurochemistry, Sahlgrenska Academy at the University

of Gothenburg, 43180 Mölndal, Sweden

\*Equal contribution

**Funding** 

KSV is funded by a Medical Research Council (MRC) UK studentship (MR/N013867/1);

KMS is funded by Alzheimer Fonden, Sweden studentship; DPB is funded by Consejo

Nacional de Ciencia y Tecnologia (CONACyT), Mexico studentship; JB is funded by a

Biotechnology and Biological Sciences Research Council (BBSRC) UK studentship; and

DMC is funded by the Dementia Research Institute (UCL).

Correspondence to <a href="mailto:d.cummings@ucl.ac.uk">d.cummings@ucl.ac.uk</a>

The authors declare no conflicts of interest.

Author contributions

Conceptualization: DMC. Writing - Original Draft: KSV, KMS, DPB and JB. Writing - Review &

Editing: KSV and DMC. Supervision: DMC

1

# Significance statement

Dementia now accounts for over 12% of deaths in the United Kingdom, exceeding those caused by cancer or heart disease. With the risk of developing dementia doubling every five years from the age of 65 and a globally aging population, finding a cure or an effective treatment to delay onset or progression is ever more imperative. These structures have historically been overlooked but it is becoming increasingly apparent that they demonstrate both dysfunction and subsequent neurodegeneration in various dementias. We summarize some of this currently sparse evidence as a means to highlight areas for further research.

# **Abstract**

Dementia is now the leading cause of death in the United Kingdom, accounting for over 12% of all deaths and is the fifth most common cause of death world-wide. As treatments for heart disease and cancers improve and the population ages, the number of sufferers will only increase, with the chance of developing dementia doubling every 5 years after the age of 65. Finding an effective treatment is ever more critical to avert this pandemic health (and economic) crisis. To date, most dementia-related research has focused on cortex and hippocampus, however, with dementia becoming more fully recognized as aspects of diseases historically categorized as motor disorders (e.g. Parkinson's and Huntington's diseases), the role of the basal ganglia in dementia is coming to the fore. Conversely, it is highly likely that neuronal pathways in these structures traditionally considered as spared in Alzheimer's disease are also affected, particularly in later stages of the disease. In this review we examine some of the limited evidence linking the basal ganglia to dementia.

### Introduction

The basal ganglia are a series of interconnected subcortical structures in the forebrain consisting of several components, including the striatum (comprised of the caudate nucleus and the putamen), the globus pallidus (internal (GPi) and external (GPe) segments), the subthalamic nucleus and the substantia nigra (pars compacta (SNc) and pars reticulata (SNr)) (Graybiel, 2000). The primary roles of the basal ganglia are motor control and motor learning, however emotional processing and goal-directed motivation have also been related to basal ganglia functions, indicating the multifactorial role of the basal ganglia in a number of different behaviors. Given their central role in motor control, impairments in the basal ganglia have been associated with a number of neurodegenerative disorders which display altered motor behavior, including Parkinson's disease (PD), dementia with Lewy bodies (DLB) and Huntington's (HD) and Alzheimer's diseases (AD). In particular, degeneration of striatal neurons has been associated with HD, whereas loss of neurons in the substantia nigra has been related to PD and similar disorders (Bernheimer et al., 1973). In this review we will first briefly describe the basal ganglia circuit, followed by discussion of how impairments in basal ganglia structures can contribute towards disease development.

### Basal ganglia circuit

Much research has focused on elucidating the mechanism through which the basal ganglia structures interact and has provided a much better understanding as to how their activity and organization may be affected under neurodegenerative conditions. The basal ganglia receive their main inputs through the striatum and subthalamic nucleus. These structures receive afferents from the cortex, thalamus and brainstem

regions, placing the basal ganglia in a central position for integrating motor information and mediating the functions of the forebrain (Graybiel, 2000).

#### Input nuclei

The striatum, the main input nucleus of the basal ganglia, is primarily made up of Yaminobutyric acid (GABA)-ergic medium-sized spiny neurons (MSNs), neurons with high abundance of dendritic spines comprising around 90% of all striatal neurons and a small proportion of cholinergic and GABAergic interneurons (Kawaguchi, 1993). These neurons relay the excitatory input they receive from cortical regions either monosynaptically to the output region of the basal ganglia, that is the SNr or GPi, or to the GPe which subsequently projects to the output structures. The classical view is that the striatal neurons directly innervating the output regions of the basal ganglia express dopamine receptor subtype 1 (D1) receptors and form the direct pathway. On the other hand, the latter neurons, which project to the globus pallidus, contain dopamine receptor subtype 2 (D2) receptors and make up the indirect pathway (Albin et al., 1995; DeLong, 1990). This expression pattern allows for the dopaminergic neurons in the SNc to modulate striatal neuron activity. Dopamine release increases the activity of the direct pathway through the activation of D1 receptors, whilst also decreasing the activity of the indirect pathway through D2 receptor activation through L-type calcium channels (Hernandez-Lopez et al., 1997; Kravitz et al., 2010). Ultimately, despite the seemingly opposing actions of these pathways, the net outcome of dopamine release results in the activation of cortical motor regions.

The subthalamic nucleus, composed mainly of glutamatergic neurons, mainly receives corticothalamic inputs, as well as projections from within the basal ganglia and

specifically, from the GPe. This region projects to the globus pallidus or SNr, a projection known as the hyperdirect pathway, which has been suggested to be involved in the regulation and cessation of motor behavior and cognition (Nelson & Kreitzer, 2014; Wessel & Aron, 2013; Wessel et al., 2013; Wessel et al., 2016). Interestingly, the subthalamic nucleus has been highlighted as a target for deep brain stimulation in PD patients due to its ability to directly relay motor and premotor cortical information to the globus pallidus through this hyperdirect pathway (Dayal et al., 2017; Hamel et al., 2003).

# Output nuclei

The two main output regions of the basal ganglia are the GPi and the SNr. They are primarily composed of GABAergic neurons and therefore form inhibitory projections with several thalamic and brainstem nuclei. The GPi and SNr are innervated by inhibitory projections coming from striatal neurons through the direct pathway, as well as excitatory projections from the subthalamic nucleus through the indirect pathway (Deschenes et al., 1996). Some of the projections of the output nuclei are directed at the intralaminar nuclei of the thalamus which comprises of both motor and limbic circuits, highlighting the potential involvement of the basal ganglia in cognitive functions as well as motor control (Fazl & Fleisher, 2018).

The tight organization of the basal ganglia demonstrates the high interconnectivity of their circuits and how impairment of one region may lead to abnormal functioning of another. In fact, exactly this interdependency of basal ganglia activity is what has been shown to be a driving factor for several neurodegenerative diseases, as discussed below.

### The basal ganglia in neurodegeneration and dementia

The basal ganglia consist of a range of subcortical cell groups that serve together to drive a variety of functions, including motor control and learning, working and procedural memory, executive functions, such as attentional filtering and emotions (Lanciego et al., 2012). As previously outlined, due to the diverse range of inputs and outputs to and from these deep embedded nuclei, damage to these circuits may affect any number of functions. Disorders of the basal ganglia have helped with the understanding of their role, as many of them share a common pathological profile and have an overlap of cognitive and motor symptoms. The basal ganglia are particularly susceptible to neurodegeneration, evidenced by the fact that many neurodegenerative disorders have to an extent some form of damage to basal ganglia circuitry. For example, PD is characterized by the progressive loss of dopaminergic neurons in the substantia nigra and basal ganglia network, causing loss of control of voluntary movement (Poewe et al., 2017), whereas degeneration of MSNs in the striatum is the predominant basal ganglia pathology in HD (Ehrlich, 2012). Corticobasal degeneration is a rare progressive disease involving neuron loss and atrophy of multiple areas of the brain, including the cerebral cortex and the basal ganglia. Patients with corticobasal degeneration have overlapping symptoms with multiple other dementias, most notably progressive supranuclear palsy but also PD, AD and DLB (Reich & Grill, 2009). Improving knowledge of any shared pathology underlying these dementias will no doubt aid the search for effective therapies.

A common pathway in the basal ganglia underlying psychiatric disorders and dementia

Dementia refers to a set of symptoms caused by brain disease or injury, resulting in persistent memory, personality, mood and judgement impairment. Many of the neurodegenerative diseases, including AD, PD, HD and DLB, are characterized by deterioration into some form of dementia sooner or later over the course of the disease. Cell death caused by abnormal protein accumulation and aggregation is a common feature of these diseases, which then leads to cognitive disturbances and decline into dementia.

The basal ganglia interact closely with the frontal cortex and have crucial roles in cognition and behaviors such as judgement and reasoning (Frank et al., 2001) and damage to the basal ganglia can lead to some of the same cognitive impairments as damage to the frontal cortex. This is evidenced in individuals with frontotemporal dementia, particularly frontotemporal lobar degeneration, where atrophy in the frontal and temporal lobes of the brain lead to a profound decline in behavior and language ability (Rabinovici & Miller, 2010). Both frontotemporal dementia and frontotemporal lobar degeneration appear to have a shared pathological profile with certain psychiatric disorders, particularly bipolar disorder and major depressive disorder (Nascimento et al., 2019). This indicates that there may be a common underlying pathway driving cognitive disturbances in dementia and psychiatric disorders; the basal ganglia are highly likely to be involved as they have shown to have abnormalities in both. For example, significant shape differences are observed in the striatum in bipolar disorder (Hwang et al., 2006) and frontotemporal dementia patients exhibit structural changes in the striatum and thalamus (Bede et al., 2018).

Another psychiatric disorder that dementia may have overlapping mechanisms with is depression. Individuals with disorders of the basal ganglia are particularly

susceptible to depression and patients with AD, PD, HD, DLB, frontotemporal lobar degeneration and vascular dementia, to name a few, commonly experience depressive symptoms at some point during the disease (Baquero & Martin, 2015; Lanctot et al., 2017; Ring & Serra-Mestres, 2002). Up to 50% of PD (Cummings, 1992) and up to 30% of HD patients (Slaughter et al., 2001) have depression, making it likely that basal ganglia damage in neurodegeneration may impact pathways underlying depression. Depression may also be a risk factor for dementia (Stern et al., 1993) and neurodegenerative abnormalities in the basal ganglia may contribute to depression: in patients with major depressive disorder, length of illness and number of depressive episodes is associated with decreased left putamen and left globus pallidus grey matter volume (Lacerda et al., 2003). This suggests that basal ganglia damage impacts higher level cognitive functions that are affected in both depression and dementia. Understanding this link will help elucidate if drugs used to treat depression can alleviate some of the symptoms of dementia and *vice versa*.

#### Basal ganglia and cognitive function in dementia

Individuals with dementia have a wide range of cognitive impairments, including memory loss as one of the first signs. This may be linked to the basal ganglia in some way due to their role in higher level cognitive functions. Procedural memory, or the ability to learn new skills, is particularly affected in PD and HD patients. In early PD, spatial learning and working memory, typically associated with nigrostriatal and dopaminergic pathways, are selectively impaired, indicating damage to networks connecting the prefrontal cortex and basal ganglia (Postle et al., 1997). In a study of PD patients with or without dementia, dementia was linked to an increased atrophy of

regions of the basal ganglia including the caudate and the putamen (Melzer et al., 2012). Overall this suggests that damage to the basal ganglia may drive certain symptoms of dementia but how this occurs remains unclear.

In the healthy brain, the basal ganglia may regulate the ability to multitask and filter out distractions: individuals with better working memory have increased activation in the globus pallidus when faced with distracting stimuli, suggesting an improved efficiency at filtering out unwanted information (McNab & Klingberg, 2008). AD patients in particular have impaired working memory and attentional deficits (MacPherson et al., 2007). It is therefore conceivable that basal ganglia pathways controlling executive function and memory are affected in neurodegenerative disorders.

In a drive to develop novel, effective and targeted therapies for the dementias which currently have limited to no cure, it is crucial to understand how specific brain areas are impacted and how this may be shared between diseases. This review addresses shared basal ganglia dysfunction across neurodegenerative diseases and dementias and their implications.

#### A note on genetic risk-factors of dementias

It is becoming ever clearer that a whole host of genetic variants contribute to one's risk of developing dementia. A huge driver in the understanding of genetic risk-factors has been the now relative ease and low-cost of performing genome-wide association studies (GWAS). These studies are gaining more statistical power as metanalyses are performed, combining studies to create large data sets. It has become clear that there limited overlap in the risk genes for the various diseases (Brainstorm Consortium, 2018) and this may contribute to the regional-specificity of neurodegeneration associated with each disease. While we do not intend to offer an exhaustive review of

the genetics that have been associated with dementias related to basal-ganglia diseases, we discuss some of the key risk-factors for each disease and refer the reader to recent reviews for a more extensive discussion.

#### Parkinson's disease

Among the dementias in which the basal ganglia are affected, PD has been the most extensively studied. PD occurs in between 5 and 35 in 100,000 individuals worldwide, with incidence increasing after the age of 65 (Poewe et al., 2017). It is a neurodegenerative disease which is associated with the progressive loss of midbrain dopaminergic neurons, coupled to slowness of motor movements (bradykinesia), rigidity, tremor, as well as postural defects. In addition to these motor defects, PD patients also demonstrate a number of non-motor symptoms, including depression, anxiety and in most cases, dementia (Hely et al., 2008). The earliest symptoms of PD, both motor and non-motor, progress slowly and are therefore very difficult to detect. The accumulation of protein aggregates, known as Lewy bodies, within and around neurons is a common pathological hallmark (Spillantini et al., 1997). Examinations of post-mortem brains of patients have demonstrated that these accumulations contain misfolded and aggregated  $\alpha$ -synuclein as well as other proteins, including hyperphosphorylated tau and amyloidβ in cases where patients are diagnosed with DLB (Wills et al., 2010). Perhaps the most prominent and unifying morphological feature of PD is the extensive loss of dopaminergic neurons in the SNc and hence their dopaminergic projections to the striatum (Hughes et al., 2002). Given that these projections have been highlighted as important modulators of striatal and hence basal ganglia activity and output to the motor cortex, their impairment has been postulated to play a key role in the emergence of the motor defects associated with PD.

Genes associated with Parkinson's disease

PD itself is a multifactorial disease, arising from both genetic and environmental factors, which together contribute to disease progression through their effects on the accumulation and aggregation of misfolded proteins, clearance mechanisms, mitochondrial damage and neuroinflammation. However, although more than 90% of cases are sporadic, a proportion of patients have a family history of the disease. To date, a number of genes have been identified as causative for PD, including mutations in the genes encoding for PARKIN, PINK1, SNCA (α-synuclein), DJ-1 and LRRK2. These proteins have been shown to play an important role in molecular pathways, which, when perturbed, can lead to neurodegeneration (Schapira, 2008). For instance, PINK1, DI-1 and LRRK2 have been shown to encode proteins localized to mitochondria, suggesting their potential involvement in mitochondrial function and dynamics (Hao et al., 2010; Pickrell & Youle, 2015; Truban et al., 2017). We refer to the reader to the review by Karimi-Moghadam et al. (2018). Animal models of PD have contributed much to the current understanding of how mutations in these proteins can lead to mitochondrial impairment, oxidative stress and disturbance in calcium homeostasis, which together can bring about disease pathology (Klein & Westenberger, 2012; Lill, 2016; Polymeropoulos et al., 1997).

Loss of dopamine and physiological changes in the basal ganglia in Parkinson's disease

Loss of nigrostriatal dopaminergic neurons is a prominent feature of the disease.

Interestingly, the dopaminergic neurons projecting to the striatum are more largely affected than those projecting to limbic regions, explaining the disturbances seen

primarily in motor control (Galvan et al., 2015; Kish et al., 1988). This loss of dopaminergic neurons occurs in the early stages of the disease, with indications of this phenotype to precede the diagnosed onset of motor symptoms, suggesting their crucial role in modulating motor behavior (Iacono et al., 2015; Poewe et al., 2017). A common diagnostic tool for PD is the use of the radio-labelled ligand <sup>123</sup>I-ioflupane in single-photon emission tomography, which binds to dopamine transporters in the presynaptic space. The <sup>123</sup>I-ioflupane signal is reduced in the SNc and striatum of PD patients, which correlates well with the disease state, highlighting the importance of dopaminergic neuron death in driving pathology (Brooks, 2016; Kraemmer et al., 2014; Marshall et al., 2009).

The specific susceptibility of these neurons has been suggested to be due to a number of factors, particularly their high energetic demand, rendering them highly susceptible to mitochondrial dysfunction and oxidative stress, as well as their high dependence on L-type Ca<sup>2+</sup> channels for their activity, making them vulnerable to altered Ca<sup>2+</sup> influx (Bishop et al., 2010; Chan et al., 2007; Foehring et al., 2009; Liss et al., 2005; Sulzer, 2007). The importance of these neurons is highlighted by the efficiency of L-DOPA, a dopamine precursor, at slowing, albeit not halting, disease progression (Barbeau, 1969). Moreover, the finding that nigrostriatal dopaminergic neurons are also selectively vulnerable to MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine), a toxin which leads to the development of a movement disorder very similar to PD, further highlights the pivotal role these neurons play in PD pathogenesis (Varastet et al., 1994).

SNc dopaminergic neurons harbor extensive axonal arbors that contact a large number of synapses, making their axons highly susceptible to alterations in axonal

trafficking and abnormal mitochondrial activity (Arbuthnott & Wickens, 2007). Thus, due to the high metabolic demand of axonal terminals, somatodendritic regions of these neurons become less populated with mitochondria (Liang et al., 2007) which reduces their ATP generating capacity and may subsequently result in an energy crisis (Nicholls, 2008). In support of this idea, human imaging and *post-mortem* studies have demonstrated that some changes in the basal ganglia, such as the emergence of dysfunctional nigrostriatal axonal terminals (Kordower et al., 2013) and changes in dopamine turnover (Sossi et al., 2004), occur even before disease diagnosis. Recent positron emission tomography imaging studies have also demonstrated the loss of vesicular monoamine transporter 2 activity in the posterior putamen of patients with mild PD (Fazio et al., 2015; Hsiao et al., 2014). These findings suggest a reduced innervation of striatal neurons by SNc dopaminergic neurons and hence support the idea of an early loss of their axonal terminals. Furthermore,  $\alpha$ -synuclein, the main component of the pathological Lewy bodies, has also been suggested to play a role in the early abnormal axonal pathology observed in PD patients through a yet unknown mechanism. This idea has arisen from the protein's increased propensity for fibrillation when mutated (Conway et al., 1998; Narhi et al., 1999), its ability to regulate dopamine metabolism and transmission (Chen et al., 2008; Lam et al., 2011; Perez et al., 2002) and its toxic influence on mitochondrial function, as shown in animal models (Martin et al., 2006; Ordonez et al., 2018). Given that  $\alpha$ -synuclein is in fact a synaptic protein and is therefore able to exert its toxic effects at synaptic sites, it is reasonable to suggest its potential role in dopaminergic axonal dysfunction in the disease.

The dopaminergic neurons in the SNc are autonomously active, generating broad action potentials (2-10 Hz) in the absence of synaptic input (Chan et al., 2007; Grace & Bunney, 1984; Mamelak, 2018). In the basal ganglia this pacemaker activity is

considered important for the maintenance of basal dopamine levels and is highly reliant on the activity of voltage-dependent L-type Ca<sup>2+</sup> channels with the Cav1.3 subunit (Striessnig et al., 2006). This subunit allows for Ca<sup>2+</sup> channels to be open during a hyperpolarized state, resulting in continuous oscillations of intracellular Ca<sup>2+</sup> levels. However, this continuous Ca<sup>2+</sup> influx can overload the Ca<sup>2+</sup> buffering ability of SNc dopaminergic neurons and thus may be considered a key contributor to their susceptibility in PD (Surmeier et al., 2012). Ca<sup>2+</sup> influx requires an equivalent amount of ATP molecules for its exclusion, making dopaminergic neurons at a constant high energetic demand. In fact, nigrostriatal dopaminergic neurons have also been shown to have a low Ca<sup>2+</sup> buffering capacity resulting from their relatively low levels of Ca<sup>2+</sup> binding proteins such as calbindin. This allows excess intracellular Ca<sup>2+</sup> to bind to intracellular proteins which can trigger cell death pathways and further enter the mitochondria to then exert continuous mitochondrial oxidant stress (German et al., 1992; Surmeier et al., 2017). In comparison, the dopaminergic neurons in the ventral tegmental area, which are relatively resistant to pathology, express high levels of this protein, highlighting the importance of Ca<sup>2+</sup> buffering capacity in cell susceptibility in PD.

It is important, however, to highlight that all the above mechanisms happen simultaneously, making it particularly challenging to target just one mechanism to improve neuronal viability. The fact that current treatments have been ineffective stresses the importance to understand what makes the basal ganglia susceptible in this disease and how they can be targeted for symptom alleviation.

## **Huntington's disease**

HD is characterized by chorea, involuntary and continuous movements of the body, especially of the extremities and face. Clinically it consists of choreoathetosis, dementia and behavioral syndromes. It is an autosomal dominant progressive neurodegenerative disorder caused by impairments in the basal ganglia and cortex. In the basal ganglia, there is a selective degeneration of MSNs, resulting in the lack of inhibition of undesired movements and leading to cognitive, behavioral and motor decline (Reiner et al., 2011).

HD onset is around the age of 40 and early symptoms include depression, small uncontrolled movements, poor coordination, learning difficulties and behavioral changes. As the disease progress the undesired movements become more pronounced and daily activities like walking, speaking or swallowing are affected. Additionally, HD can also be found in the younger population (<20 years) with similar symptoms but with some variations, for instance hypokinetic signs, rigidity, drooling or seizures (Roos, 2010; Tereshchenko et al., 2019).

The genetic cause for this disease is clear: mutations in the *HTT* gene located on chromosome 4, which is responsible for the production of the huntingtin protein, involving the DNA segment CAG trinucleotide repeat in exon 1. Affected individuals possess a sequence of more than 36 CAG repeats and longer expansions result in earlier disease onset. The CAG encodes glutamine and thus results in longer versions of huntingtin. In turn, this abnormal protein is cleaved into toxic fragments that disrupt normal neuronal functions and eventual death of projection neurons of the basal ganglia occurs (Reiner et al., 2011). More recently, it has become apparent through GWAS that disease presentation is moderated through variants of genes other than *HTT*. A review is provided in Holmans et al. (2017).

Structural alterations of basal ganglia in Huntington's disease

Brain abnormalities in HD develop before evident symptoms or clinical diagnosis. There is 25% brain weight loss at advanced stages of the disease. The neuropathology consists of atrophy, neuronal loss and astrogliosis of the basal ganglia and cortical structures. Striatal pathology starts caudally and then spreads rostrally into a dorsomedial to ventromedial directions (Fazio et al., 2015). Additionally, other regions of the brain are affected while the disease progresses, such as hippocampus, amygdala and cerebellum (Rosas et al., 2003).

The loss of striatal and pallidal neurons in HD is selective; early in the disease there is loss of putaminal GABAergic cells projecting to the lateral globus pallidus which affects the indirect pathway of the basal ganglia. The balance between indirect and direct pathways is consequently in favor of the direct pathway: there is unsuccessful inhibition and thus excitation of thalamic neurons is enhanced, resulting in random and inappropriate neuronal activity. In contrast, when the disease is advanced, cholinergic signals are lost and the caudate nuclei is atrophied. Detection of this state of the disease is visible through MRI or computerized tomography in the brain, where enlarged lateral ventricles of the caudate frontal horns is observed (Reiner et al., 2011).

In HD there is also a significant shrinkage of the amygdala. This region plays a crucial role in the limbic circuit of the basal ganglia to trigger specific cognitive responses linked to motor activities. Indeed, declining emotion recognition in patients correlating with amygdala volume loss has been reported (Kipps et al., 2007). On the other hand, the thalamus, subthalamus and substantia nigra undergo cell loss, shrinkage and astrogliosis as well but at lower levels. About 40% of GABAergic and dopaminergic

neurons of the substantia nigra are lost, which could contribute to the akinesia symptoms seen in both HD and PD (Vonsattel, 2008).

#### Alzheimer's disease

It is estimated that there are currently over 50 million people in the world living with dementia, with the number expected to triple to 152 million by 2050 (Patterson, 2018). AD is the most common cause of dementia, accounting for over 60% of all dementia cases. The global and societal impact of dementia is huge and there are still no effective treatments in halting or improving symptoms of AD, so understanding the pathology of this disorder represents a crucial step towards finding a cure.

Over 97% of AD cases are sporadic and occur later than 60 years of age. No common causative gene has been identified for the late-onset disease; however, the APOE gene influences risk of developing the disease, with the  $\epsilon 4$  allele increasing and the  $\epsilon 2$  allele decreasing risk with respect to the  $\epsilon 3$  allele (Verghese et al., 2011). However, sporadic AD is thought to arise from a complex interplay between genetic and environmental factors. There are three genes in which mutations have been identified that cause rare familial forms of AD: amyloid precursor protein (APP), presenilin 1 and presenilin 2 (PSEN1/PSEN2). More recently, GWAS has highlighted a number of risk-factor variants of genes. Many of the encoded proteins are associated with the immune response but also a number of other cellular functions (reviewed in Verheijen and Sleegers, 2018).

The defining hallmark pathologies of AD are amyloid- $\beta$  plaques and neurofibrillary tangles alongside substantial neurodegeneration. Amyloid- $\beta$  plaques accumulate first and result from cleavage of APP into (amongst many other metabolites) amyloid $\beta$ -40 and amyloid $\beta$ -42, which abnormally fold and aggregate in the

brain. Neurofibrillary tangles arise from hyperphosphorylation of tau protein, leading to neuronal damage and neurodegeneration (Selkoe & Hardy, 2016).

Neuroinflammation is also a classic feature of the disease, involving reactive microglia and astrocytes. Indeed, microglia have been found to colocalize with plaques in *post-mortem* human brain (Calsolaro & Edison, 2016) and in mouse models of AD (e.g. Medawar et al., 2019).

Clinical features of AD include initial problems with episodic memory and difficulties multi-tasking, followed by more profound cognitive impairment as the disease progresses, including behavioral and mood changes, and eventual decline into full dementia (Jost & Grossberg, 1995).

Structural abnormalities of the basal ganglia in Alzheimer's disease

The prevalence of AD undoubtedly makes it a crucial disease to find a cure for; however, much of the research to date has focused primarily on the hippocampus and the neocortex. The basal ganglia have received little attention in this context, despite evidence of degeneration of structures in the deep grey matter which may contribute to AD symptoms (de Jong et al., 2008). Early evidence from Braak and Braak (1991b) indicated that, in cases of severe AD, there are large numbers of extracellular amyloid- $\beta$  deposits in all thalamic nuclei and severe changes in the limbic nuclei due to the presence of neurofibrillary tangles, indicating an impairment of the limbic circuits (Braak & Braak, 1991a).

Continuing research has consistently demonstrated that AD patients have shape changes in the basal ganglia, aided by recent advances in brain imaging techniques.

Patients exhibit more rapid shape changes of the thalamus and basal ganglia compared with age-matched controls (Cho et al., 2014); the medial heads on the caudate nucleus

and ventral lateral putamen are selectively degenerated (de Jong et al., 2011); and MRI indicates that volumes of caudate nucleus (Almeida et al., 2003), putamen and thalamus (de Jong et al., 2011) are reduced in AD patients. Moreover, in all cases, the structural changes were correlated with cognitive decline. Thus, it is clear that, contrary to the standard declaration that the basal ganglia and associated subcortical structures are spared in AD, these regions are susceptible to structural changes during disease progression and may well underlie aspects of cognitive impairment.

The striatum, comprising of the nucleus accumbens, caudate nucleus and putamen, appears to be particularly vulnerable in AD. Indeed, both diffuse plaques and neurofibrillary tangles are observed in striatal areas and amyloid- $\beta$  deposits are found in the striatum in young, asymptomatic members of families with *PSEN1* mutations (Klunk et al., 2007). Additionally, amyloid- $\beta$  levels may increase in the basal ganglia over 10 years before expected symptom onset (Bateman et al., 2012), suggesting that it may be an early-affected brain region in the disease and perhaps an underexplored therapeutic target.

As the basal ganglia are believed to drive higher level cognitive processes such as attention and memory, it is perhaps unsurprising that degeneration and shape changes would affect cognitive function in AD. Little, however, is known about this, as robust studies correlating alterations in subcortical volumes in AD with cognitive function are lacking.

Iron-associated basal ganglia dysfunction in Alzheimer's disease

Metabolic processing is known to dysregulate with age and is particularly affected in neurodegeneration. The basal ganglia (in particular the putamen, caudate nucleus and globus pallidus) have a high metabolic rate and contain the highest levels of iron in the

brain (Ramos et al., 2014), which may render them particularly sensitive to ageassociated metabolic dysregulation. Even in the healthy ageing brain, iron accumulates
in cells leading to a region-specific increase in total iron levels (Connor et al., 1990). As
ageing represents the most significant risk-factor for neurodegenerative diseases, iron
may be involved as either a cause or a consequence. This is evidenced by studies
showing that iron accumulation in the basal ganglia is a common feature of many
neurodegenerative diseases and results in both cognitive and motor dysfunction. For
example, iron has been found to colocalize with the insoluble protein aggregates that
characterize neurodegenerative diseases such as AD, PD, HD and DLB (Agrawal et al.,
2018; Joppe et al., 2019; van Bergen et al., 2016), suggesting a dysfunction in iron
trafficking or metabolism as a cause or consequence of pathology. The heavy iron load
in the basal ganglia may explain its increased susceptibility to neurodegeneration, so
this represents an important brain region to further examine in this context.

Certain disorders are characterized by iron dysregulation (Dusek et al., 2012) but the impact of this in AD is not fully understood. An early study of the basal ganglia in AD revealed that patients who have basal ganglia mineralization have a higher rate of psychotic and motor disturbances, such as delusions, depression, parkinsonism and myoclonus, compared with age- and gender-matched AD patients without mineralization in the basal ganglia (Forstl et al., 1992). Autopsy studies have shown that the highest concentrations of iron in the brain exist in the basal ganglia including the globus pallidus, putamen and substantia nigra (Conway et al., 1998). Amyloid- $\beta$  deposition and aggregation has been shown to be accelerated by metal ions such as iron and zinc (Lovell et al., 1998) and iron has been demonstrated *in vitro* to increase the aggregation and cytotoxicity of amyloid- $\beta$  peptide (Galante et al., 2018). Iron also appears to be involved with plaques and tangles, the two hallmark pathologies of AD.

Amyloid- $\beta$  plaques colocalize with iron (van Bergen et al., 2016), suggesting that iron may either exacerbate amyloid- $\beta$  aggregation or that the plaques may trap iron in some way. In mouse models, iron has been demonstrated to correlate with amyloid- $\beta$  plaque morphology, with evidence for the formation of an iron-amyloid complex (Telling et al., 2017). Iron has also been found to bind to tau protein, inducing phosphorylation and aggregation (Liu et al., 2018), thereby promoting the accumulation of hyperphosphorylated tau which leads to neurofibrillary tangles. Neuropathological studies appear to support this idea as iron levels in eight regions of the AD brain, including the globus pallidus and caudate nucleus, are raised compared to healthy controls, implicating the basal ganglia as part of the pathological profile of AD (Tao et al., 2014). It appears iron metabolism is impaired in some way in AD and other neurodegenerative diseases but how this relates to the basal ganglia or precisely to disease progression requires further studies. An improved understanding of this would aid the development of treatment strategies, such as developing iron chelators to sequester excess iron in the brain.

#### **Dementia with Lewy bodies**

DLB is the second most common form of dementia and occurs in approximately 15 out of 100 patients with dementia, when diagnosed by autopsy (McKeith et al., 2005) and accounts for 4-8% clinically (Vann Jones & O'Brien, 2014). Clinically, the disease is characterized by increasing deficits in executive, visual–spatial and memory function (Lippa et al., 2007). Diagnostically, the core features of the disease are fluctuation in cognition and attention (especially between periods of lucidity and confusion), recurrent visual hallucinations and rapid eye movement sleep behavior disorder that precede or occur within a year of parkinsonian motor symptoms (Walker et al., 2015).

This is highly similar to a condition called Parkinson's disease dementia, however, dementia in Parkinson's disease dementia is diagnosed only when it occurs in the presence of long-established motor symptoms (Jellinger & Korczyn, 2018). Indeed, pathological features of both disorders are also highly similar (Jellinger, 2018). This disorder is frequently misdiagnosed due to its similarity of symptoms with AD; however, the presence of visual hallucinations is a distinguishing factor (Thomas et al., 2018).

Like PD, both DLB and Parkinson's disease dementia are classified as Lewy body disorders. Lewy body disorders are characterized by the presence of protein aggregates predominantly in the soma, Lewy bodies, or processes, Lewy neurites, of neurons (Braak et al., 2004; Forno, 1996). Lewy bodies measure 8-30  $\mu$ m in diameter with a spherical shape, with or without a dense granular core and a periphery of radiating filaments, which appear pink with a typical haematoxylin and eosin staining (Olanow et al., 2004). In the brainstem, Lewy bodies possess a densely stained core and a lightly stained peripheral halo, while in the cortex Lewy bodies lack the halo (Forno, 1996; Kosaka et al., 1976). The inner core stains more intensely for  $\alpha$ -synuclein, while the periphery stains more strongly for ubiquitin, with the  $\alpha$ -synuclein staining being more specific (Spillantini et al., 1997). Approximately two thirds of Lewy bodies in the substantia nigra stain for  $\alpha$ -synuclein and approximately one third in the cortex (Sakamoto et al., 2002). More than 76 other proteins have been found in Lewy bodies and neurites (Wakabayashi et al., 2007).

Mutations in the  $\alpha$ -synuclein gene (*SNCA*) are linked to familial cases in PD and DLB (Polymeropoulos et al., 1997; Zarranz et al., 2004). Genes linked to both AD and PD are also linked to DLB, for example *GBA* and *APOE* genes (Guerreiro et al., 2018; Nalls et

al., 2013; Tsuang et al., 2012). Mutations in other genes linked to PD. such as *LRRK2* and *SCARB2*, may also play a role in disease risk (Alcalay et al., 2016; Bras et al., 2014; Hyun et al., 2013; Orme et al., 2018). A recent review of the genetics of DLB can be found in Guerreiro et al., 2019).

α-synuclein is highly expressed in the brain, located in pre- and postsynaptic terminals, and may make up as much as 1% of the cytosol (Kim et al., 2014). It is a 140 amino acid-long typically unfolded protein, with an amino-terminal lipid binding αhelix, a non-amyloidogenic core domain and an unstructured carboxy-terminal. Although its exact physiological role is unclear, α-synuclein likely has a role in neurotransmitter release, potentially by regulating the exocytosis of vesicles by interactions with the SNARE complex (Murphy et al., 2000; Orme et al., 2018). When  $\alpha$ synuclein becomes misfolded, the random coil of the non-amyloidogenic core domain forms β-sheets, which are important for protofibril and fibril formation (Serpell et al., 2000; Uversky & Eliezer, 2009). The spread of Lewy bodies in a rostro-caudal direction in PD, suggests a prion-like transmission (Braak et al., 2004; George et al., 2013; Seidel et al., 2015). Strong evidence for this spread comes from studies in which embryonic neurons grafted into PD patients to replace lost nigral cells revealed α-synuclein/Lewy body pathology upon post-mortem examination (Kordower et al., 2008; Kurowska et al., 2011; Li et al., 2008). Furthermore, propagation of Lewy bodies has been shown in animal and cell culture models (Karpowicz et al., 2019; Kordower et al., 2011; Ngolab et al., 2017).

### **Pathology**

Lewy bodies have been linked to cell death and the extent of Lewy body pathology correlates with clinical pathology in both DLB (Tiraboschi et al., 2015) and Parkinson's disease with dementia (Hurtig et al., 2000). Lewy body pathology is found widespread in *post-mortem* brains from both groups of patients, especially affecting the frontal cortex, midbrain and brainstem nuclei, dorsal efferent nucleus of the vagus, basal forebrain nuclei and limbic regions (Jellinger & Korczyn, 2018; Neef & Walling, 2006). This widespread pathology makes the spreading of the disorder more difficult to define than in PD, as it does not appear to spread in a rostro-caudal manner (Walker et al., 2015) although systems to stage the disorder have been proposed (McKeith et al., 2005).

Grey matter atrophy occurs in frontotemporal, occipital and parietal cortical areas and subcortical atrophy of amygdala, caudate, putamen and pallidum (Almeida et al., 2003; Beyer et al., 2009; Gazzina et al., 2016; Sanchez-Castaneda et al., 2009; Watson et al., 2012). Like PD, dopaminergic and cholinergic neurons appear especially vulnerable in the disease and in some cases have been found to be more degenerated than in PD (Colloby et al., 2012; Muenter et al., 1998; Outeiro et al., 2019; Thomas et al., 2017). Dopamine synthesis, transporter binding and storage are all decreased in patients with DLB (Gomperts et al., 2016; Marquie et al., 2014; Nikolaus et al., 2009). Cholinergic deficits in the cortex are observed due to the loss of neurons in the basal forebrain, leading to cholinergic denervation and signaling deficits (Grothe et al., 2014; Mazere et al., 2017).

Cell death due to an impairment of axonal transport, autophagy-lysosomal, ubiquitin-proteasome and mitochondrial pathways have all been strongly implicated in the disease (Manecka et al., 2017; Power et al., 2017; Sato et al., 2018; Volpicelli-Daley,

2017). Importantly, whether this is directly due to Lewy bodies is unclear, as they may play a role in protecting the cell through the up-take of misfolded and non-functional proteins (Beyer et al., 2009), as there are cases of individuals with significant Lewy body pathology at autopsy that do not demonstrate symptoms of dementia (Parkkinen et al., 2008). Oligomers of  $\alpha$ -synuclein may also mediate the cellular toxicity in the disease (Outeiro et al., 2019). An additional factor that may play a role in cell death is the co-occurrence of AD pathology in patients with DLB, which is suggested to be an increasingly important part of the disorder rather than a comorbidity (Deramecourt et al., 2006; Jellinger & Korczyn, 2018). This may lead to synergistic effects and increased cell death. Indeed, AD and DLB co-pathologies have been shown to lead to a greater temporal lobe atrophy (van der Zande et al., 2018) and greater baseline amyloid- $\beta$  imaging predicts more severe neurodegeneration in DLB patients (Sarro et al., 2016).

## **Concluding remarks**

The evidence that the basal ganglia are implicated in dementia is limited but, particularly in a subset of neurodegenerative diseases, it is becoming ever more apparent that their role is not restricted to motor control. As the physiological roles of the basal ganglia and associated structures in cognition and other higher functions become better appreciated, the dysfunction associated with their pathology is broadening. Whether targeting the basal ganglia for treatments of late stages of the less prominent disorders of the basal ganglia, such as AD, is a worthwhile approach is yet to be ascertained but understanding how they are affected in the disease may indeed reveal disease modifying targets.

#### References

- Agrawal, S., Fox, J., Thyagarajan, B., & Fox, J. H. (2018). Brain mitochondrial iron accumulates in Huntington's disease, mediates mitochondrial dysfunction, and can be removed pharmacologically. *Free Radic Biol Med, 120,* 317-329. doi:10.1016/j.freeradbiomed.2018.04.002
- Albin, R. L., Young, A. B., & Penney, J. B. (1995). The functional anatomy of disorders of the basal ganglia. *Trends Neurosci*, 18(2), 63-64. doi:10.1016/0166-2236(95)80020-3
- Alcalay, R. N., Levy, O. A., Wolf, P., Oliva, P., Zhang, X. K., Waters, C. H., . . . Gan-Or, Z. (2016). SCARB2 variants and glucocerebrosidase activity in Parkinson's disease. *NPJ Parkinsons Dis*, 2. doi:10.1038/npjparkd.2016.4
- Almeida, O. P., Burton, E. J., McKeith, I., Gholkar, A., Burn, D., & O'Brien, J. T. (2003). MRI study of caudate nucleus volume in Parkinson's disease with and without dementia with Lewy bodies and Alzheimer's disease. *Dement Geriatr Cogn Disord, 16*(2), 57-63. doi:10.1159/000070676
- Arbuthnott, G. W., & Wickens, J. (2007). Space, time and dopamine. *Trends Neurosci*, *30*(2), 62-69. doi:10.1016/j.tins.2006.12.003
- Baquero, M., & Martin, N. (2015). Depressive symptoms in neurodegenerative diseases. *World J Clin Cases*, *3*(8), 682-693. doi:10.12998/wjcc.v3.i8.682
- Barbeau, A. (1969). L-dopa therapy in Parkinson's disease: a critical review of nine years' experience. *Can Med Assoc J.*, 101(13), 59-68.
- Bateman, R. J., Xiong, C., Benzinger, T. L., Fagan, A. M., Goate, A., Fox, N. C., . . . Dominantly Inherited Alzheimer, N. (2012). Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *The New England journal of medicine, 367*(9), 795-804. doi:10.1056/NEJMoa1202753
- Bede, P., Finegan, E., Chipika, R., Omer, T., Pender, N., Hutchinson, S., & Hardiman, O. (2018). Characterisation of basal ganglia pathology in frontotemporal dementia: a connectivity-based multimodal neuroimaging study. *Neurology, 90*.
- Bernheimer, H., Birkmayer, W., Hornykiewicz, O., Jellinger, K., & Seitelberger, F. (1973). Brain dopamine and the syndromes of Parkinson and Huntington. Clinical, morphological and neurochemical correlations. *J Neurol Sci, 20*(4), 415-455. doi:10.1016/0022-510X(73)90175-5
- Beyer, K., Domingo-Sabat, M., & Ariza, A. (2009). Molecular pathology of Lewy body diseases. *Int J Mol Sci, 10*(3), 724-745. doi:10.3390/ijms10030724
- Bishop, M. W., Chakraborty, S., Matthews, G. A., Dougalis, A., Wood, N. W., Festenstein, R., & Ungless, M. A. (2010). Hyperexcitable substantia nigra dopamine neurons in PINK1- and HtrA2/Omi-deficient mice. *J Neurophysiol*, *104*(6), 3009-3020. doi:10.1152/jn.00466.2010
- Braak, H., & Braak, E. (1991a). Alzheimer's disease affects limbic nuclei of the thalamus. *Acta Neuropathol*, 81(3), 261-268.
- Braak, H., & Braak, E. (1991b). Neuropathological stageing of Alzheimer-related changes. *Acta Neuropathol*, 82(4), 239-259.
- Braak, H., Ghebremedhin, E., Rub, U., Bratzke, H., & Del Tredici, K. (2004). Stages in the development of Parkinson's disease-related pathology. *Cell Tissue Res, 318*(1), 121-134. doi:10.1007/s00441-004-0956-9
- Brainstorm Consortium, The (2018). Analysis of shared heritability in common disorders of the brain. *Science*, 360(6395), eaap8757. doi: 10.1126/science.aap8757

- Bras, J., Guerreiro, R., Darwent, L., Parkkinen, L., Ansorge, O., Escott-Price, V., . . . Hardy, J. (2014). Genetic analysis implicates APOE, SNCA and suggests lysosomal dysfunction in the etiology of dementia with Lewy bodies. *Hum Mol Genet, 23*(23), 6139-6146. doi:10.1093/hmg/ddu334
- Brooks, D. J. (2016). Molecular imaging of dopamine transporters. *Ageing Res Rev, 30*, 114-121. doi:10.1016/j.arr.2015.12.009
- Calsolaro, V., & Edison, P. (2016). Neuroinflammation in Alzheimer's disease: Current evidence and future directions. *Alzheimers Dement, 12*(6), 719-732. doi:10.1016/j.jalz.2016.02.010
- Chan, C. S., Guzman, J. N., Ilijic, E., Mercer, J. N., Rick, C., Tkatch, T., . . . Surmeier, D. J. (2007). 'Rejuvenation' protects neurons in mouse models of Parkinson's disease. *Nature*, *447*(7148), 1081-1086. doi:10.1038/nature05865
- Chen, M. K., Kuwabara, H., Zhou, Y., Adams, R. J., Brasic, J. R., McGlothan, J. L., . . . Guilarte, T. R. (2008). VMAT2 and dopamine neuron loss in a primate model of Parkinson's disease. *J Neurochem*, 105(1), 78-90. doi:10.1111/j.1471-4159.2007.05108.x
- Cho, H., Kim, J. H., Kim, C., Ye, B. S., Kim, H. J., Yoon, C. W., . . . Seo, S. W. (2014). Shape changes of the basal ganglia and thalamus in Alzheimer's disease: a three-year longitudinal study. *J Alzheimers Dis*, 40(2), 285-295. doi:10.3233/JAD-132072
- Colloby, S. J., McParland, S., O'Brien, J. T., & Attems, J. (2012). Neuropathological correlates of dopaminergic imaging in Alzheimer's disease and Lewy body dementias. *Brain*, 135(Pt 9), 2798-2808. doi:10.1093/brain/aws211
- Connor, J. R., Menzies, S. L., St Martin, S. M., & Mufson, E. J. (1990). Cellular distribution of transferrin, ferritin, and iron in normal and aged human brains. *J Neurosci Res, 27*(4), 595-611. doi:10.1002/jnr.490270421
- Conway, K. A., Harper, J. D., & Lansbury, P. T. (1998). Accelerated in vitro fibril formation by a mutant alpha-synuclein linked to early-onset Parkinson disease. *Nat Med, 4*(11), 1318-1320. doi:10.1038/3311
- Cummings, J. L. (1992). Depression and Parkinson's disease: a review. *Am J Psychiatry*, 149(4), 443-454. doi:10.1176/ajp.149.4.443
- Dayal, V., Limousin, P., & Foltynie, T. (2017). Subthalamic Nucleus Deep Brain Stimulation in Parkinson's Disease: The Effect of Varying Stimulation Parameters. *J Parkinsons Dis*, 7(2), 235-245. doi:10.3233/JPD-171077
- de Jong, L. W., Ferrarini, L., van der Grond, J., Milles, J. R., Reiber, J. H., Westendorp, R. G., . . . van Buchem, M. A. (2011). Shape abnormalities of the striatum in Alzheimer's disease. *J Alzheimers Dis*, 23(1), 49-59. doi:10.3233/JAD-2010-101026
- de Jong, L. W., van der Hiele, K., Veer, I. M., Houwing, J. J., Westendorp, R. G., Bollen, E. L., . . . van der Grond, J. (2008). Strongly reduced volumes of putamen and thalamus in Alzheimer's disease: an MRI study. *Brain, 131*(Pt 12), 3277-3285. doi:10.1093/brain/awn278
- DeLong, M. R. (1990). Primate models of movement disorders of basal ganglia origin. *Trends Neurosci*, 13(7), 281-285. doi:10.1016/0166-2236(90)90110-V
- Deramecourt, V., Bombois, S., Maurage, C. A., Ghestem, A., Drobecq, H., Vanmechelen, E., . . Delacourte, A. (2006). Biochemical staging of synucleinopathy and amyloid deposition in dementia with Lewy bodies. *J Neuropathol Exp Neurol, 65*(3), 278-288. doi:10.1097/01.jnen.0000205145.54457.ea
- Deschenes, M., Bourassa, J., Doan, V. D., & Parent, A. (1996). A single-cell study of the axonal projections arising from the posterior intralaminar thalamic nuclei in the rat.

- *The European journal of neuroscience, 8*(2), 329-343. doi:10.1111/j.1460-9568.1996.tb01217.x
- Dusek, P., Jankovic, J., & Le, W. (2012). Iron dysregulation in movement disorders. *Neurobiol Dis*, 46(1), 1-18. doi:10.1016/j.nbd.2011.12.054
- Ehrlich, M. E. (2012). Huntington's disease and the striatal medium spiny neuron: cell-autonomous and non-cell-autonomous mechanisms of disease. *Neurotherapeutics*, 9(2), 270-284. doi:10.1007/s13311-012-0112-2
- Fazio, P., Svenningsson, P., Forsberg, A., Jonsson, E. G., Amini, N., Nakao, R., . . . Varrone, A. (2015). Quantitative Analysis of (1)(8)F-(E)-N-(3-Iodoprop-2-Enyl)-2beta-Carbofluoroethoxy-3beta-(4'-Methyl-Phenyl) Nortropane Binding to the Dopamine Transporter in Parkinson Disease. *J Nucl Med*, *56*(5), 714-720. doi:10.2967/jnumed.114.152421
- Fazl, A., & Fleisher, J. (2018). Anatomy, Physiology, and Clinical Syndromes of the Basal Ganglia: A Brief Review. *Semin Pediatr Neurol*, *25*, 2-9. doi:10.1016/j.spen.2017.12.005
- Foehring, R. C., Zhang, X. F., Lee, J. C., & Callaway, J. C. (2009). Endogenous calcium buffering capacity of substantia nigral dopamine neurons. *J Neurophysiol*, *102*(4), 2326-2333. doi:10.1152/jn.00038.2009
- Forno, L. S. (1996). Neuropathology of Parkinson's disease. *J Neuropathol Exp Neurol, 55*(3), 259-272. doi:10.1097/00005072-199603000-00001
- Forstl, H., Burns, A., Cairns, N., Luthert, P., & Levy, R. (1992). Basal ganglia mineralization in Alzheimer's disease: a comparative study of clinical, neuroradiological and neuropathological findings. *Behav Neurol*, *5*(1), 53-57. doi:10.3233/BEN-1992-5110
- Frank, M. J., Loughry, B., & O'Reilly, R. C. (2001). Interactions between frontal cortex and basal ganglia in working memory: a computational model. *Cogn Affect Behav Neurosci*, 1(2), 137-160.
- Galante, D., Cavallo, E., Perico, A., & D'Arrigo, C. (2018). Effect of ferric citrate on amyloid-beta peptides behavior. *Biopolymers*, 109(6), e23224. doi:10.1002/bip.23224
- Galvan, A., Devergnas, A., & Wichmann, T. (2015). Alterations in neuronal activity in basal ganglia-thalamocortical circuits in the parkinsonian state. *Frontiers in Neuroanatomy*, *9*, 5. doi:10.3389/fnana.2015.00005
- Gazzina, S., Premi, E., Turrone, R., Acosta-Cabronero, J., Rizzetti, M. C., Cotelli, M. S., . . . Borroni, B. (2016). Subcortical matter in the alpha-synucleinopathies spectrum: an MRI pilot study. *J Neurol*, *263*(8), 1575-1582. doi:10.1007/s00415-016-8173-5
- George, S., Rey, N. L., Reichenbach, N., Steiner, J. A., & Brundin, P. (2013). alpha-Synuclein: the long distance runner. *Brain Pathol*, 23(3), 350-357. doi:10.1111/bpa.12046
- German, D. C., Manaye, K. F., Sonsalla, P. K., & Brooks, B. A. (1992). Midbrain dopaminergic cell loss in Parkinson's disease and MPTP-induced parkinsonism: sparing of calbindin-D28k-containing cells. *Ann N Y Acad Sci, 648*, 42-62. doi:10.1111/j.1749-6632.1992.tb24523.x
- Gomperts, S. N., Marquie, M., Locascio, J. J., Bayer, S., Johnson, K. A., & Growdon, J. H. (2016). PET Radioligands Reveal the Basis of Dementia in Parkinson's Disease and Dementia with Lewy Bodies. *Neurodegener Dis, 16*(1-2), 118-124. doi:10.1159/000441421
- Grace, A. A., & Bunney, B. S. (1984). The control of firing pattern in nigral dopamine neurons: burst firing. *J Neurosci, 4*(11), 2877-2890. doi:10.1523/JNEUROSCI.04-11-02877.1984

- Graybiel, A. M. (2000). The basal ganglia. *Curr Biol, 10*(14), R509-511. doi:10.1016/S0960-9822(00)00593-5
- Grothe, M. J., Schuster, C., Bauer, F., Heinsen, H., Prudlo, J., & Teipel, S. J. (2014). Atrophy of the cholinergic basal forebrain in dementia with Lewy bodies and Alzheimer's disease dementia. *J Neurol*, 261(10), 1939-1948. doi:10.1007/s00415-014-7439-z
- Guerreiro, R., Ross, O. A., Kun-Rodrigues, C., Hernandez, D. G., Orme, T., Eicher, J. D., . . . Bras, J. (2018). Investigating the genetic architecture of dementia with Lewy bodies: a two-stage genome-wide association study. *Lancet Neurol*, *17*(1), 64-74. doi:10.1016/S1474-4422(17)30400-3
- Guerreiro, R., Escott-Price, V., Hernandez, D. G., Kun-Rodrigues, C., Ross, O. A., Orme, T., . . . Bras, J. (2019). Heritability and genetic variance of dementia with Lewy bodies. *Neurobiol Dis*, 127:492-501. doi: 10.1016/j.nbd.2019.04.004
- Hamel, W., Fietzek, U., Morsnowski, A., Schrader, B., Herzog, J., Weinert, D., . . . Mehdorn, H. M. (2003). Deep brain stimulation of the subthalamic nucleus in Parkinson's disease: evaluation of active electrode contacts. *J Neurol Neurosurg Psychiatry*, 74(8), 1036-1046. doi:10.1136/jnnp.74.8.1036
- Hao, L. Y., Giasson, B. I., & Bonini, N. M. (2010). DJ-1 is critical for mitochondrial function and rescues PINK1 loss of function. *Proc Natl Acad Sci U S A, 107*(21), 9747-9752. doi:10.1073/pnas.0911175107
- Hely, M. A., Reid, W. G., Adena, M. A., Halliday, G. M., & Morris, J. G. (2008). The Sydney multicenter study of Parkinson's disease: the inevitability of dementia at 20 years. *Mov Disord*, *23*(6), 837-844. doi:10.1002/mds.21956
- Hernandez-Lopez, S., Bargas, J., Surmeier, D. J., Reyes, A., & Galarraga, E. (1997). D1 receptor activation enhances evoked discharge in neostriatal medium spiny neurons by modulating an L-type Ca2+ conductance. *J Neurosci, 17*(9), 3334-3342. doi:10.1523/JNEUROSCI.17-09-03334.1997
- Holmans, P. A., Massey, T. H., Jones, L. (2017). Genetic modifiers of Mendelian disease: Huntington's disease and the trinucleotide repeat disorders. *Hum Mol Gen*, 26(R2):R83-R90. doi: 10.1093/hmg/ddx261
- Hsiao, I. T., Weng, Y. H., Hsieh, C. J., Lin, W. Y., Wey, S. P., Kung, M. P., . . . Lin, K. J. (2014). Correlation of Parkinson disease severity and 18F-DTBZ positron emission tomography. *Jama Neurology*, 71(6), 758-766. doi:10.1001/jamaneurol.2014.290
- Hughes, A. J., Daniel, S. E., Ben-Shlomo, Y., & Lees, A. J. (2002). The accuracy of diagnosis of parkinsonian syndromes in a specialist movement disorder service. *Brain, 125*(Pt 4), 861-870. doi:10.1093/brain/awf080
- Hurtig, H. I., Trojanowski, J. Q., Galvin, J., Ewbank, D., Schmidt, M. L., Lee, V. M., . . . Arnold, S. E. (2000). Alpha-synuclein cortical Lewy bodies correlate with dementia in Parkinson's disease. *Neurology*, *54*(10), 1916-1921. doi:10.1212/wnl.54.10.1916
- Hwang, J., Lyoo, I. K., Dager, S. R., Friedman, S. D., Oh, J. S., Lee, J. Y., . . . Renshaw, P. F. (2006). Basal ganglia shape alterations in bipolar disorder. *Am J Psychiatry*, *163*(2), 276-285. doi:10.1176/appi.ajp.163.2.276
- Hyun, C. H., Yoon, C. Y., Lee, H. J., & Lee, S. J. (2013). LRRK2 as a Potential Genetic Modifier of Synucleinopathies: Interlacing the Two Major Genetic Factors of Parkinson's Disease. *Exp Neurobiol*, 22(4), 249-257. doi:10.5607/en.2013.22.4.249
- Iacono, D., Geraci-Erck, M., Rabin, M. L., Adler, C. H., Serrano, G., Beach, T. G., & Kurlan, R. (2015). Parkinson disease and incidental Lewy body disease: Just a question of time? *Neurology*, 85(19), 1670-1679. doi:10.1212/WNL.000000000002102

- Jellinger, K. A. (2018). Dementia with Lewy bodies and Parkinson's disease-dementia: current concepts and controversies. *J Neural Transm (Vienna), 125*(4), 615-650. doi:10.1007/s00702-017-1821-9
- Jellinger, K. A., & Korczyn, A. D. (2018). Are dementia with Lewy bodies and Parkinson's disease dementia the same disease? *BMC Med*, *16*(1), 34. doi:10.1186/s12916-018-1016-8
- Joppe, K., Roser, A. E., Maass, F., & Lingor, P. (2019). The Contribution of Iron to Protein Aggregation Disorders in the Central Nervous System. *Front Neurosci*, *13*, 15. doi:10.3389/fnins.2019.00015
- Jost, B. C., & Grossberg, G. T. (1995). The natural history of Alzheimer's disease: a brain bank study. *J Am Geriatr Soc, 43*(11), 1248-1255. doi:10.1111/j.1532-5415.1995.tb07401.x
- Karimi-Moghadam, A., Charsouei, S., & Jabalameli, M. R. (2018). Parkinson Disease from Mendelian Forms to Genetic Susceptibility: New Molecular Insights into the Neurodegeneration Process. *Cell Mol Neurobiol*, 38(6):1153-1178. doi: 10.1007/s10571-018-0587-4
- Karpowicz, R. J., Jr., Trojanowski, J. Q., & Lee, V. M. (2019). Transmission of alpha-synuclein seeds in neurodegenerative disease: recent developments. *Lab Invest*. doi:10.1038/s41374-019-0195-z
- Kawaguchi, Y. (1993). Physiological, morphological, and histochemical characterization of three classes of interneurons in rat neostriatum. *J Neurosci*, *13*(11), 4908-4923. doi:10.1523/JNEUROSCI.13-11-04908.1993
- Kim, W. S., Kagedal, K., & Halliday, G. M. (2014). Alpha-synuclein biology in Lewy body diseases. *Alzheimers Res Ther*, 6(5), 73. doi:10.1186/s13195-014-0073-2
- Kipps, C. M., Duggins, A. J., McCusker, E. A., & Calder, A. J. (2007). Disgust and happiness recognition correlate with anteroventral insula and amygdala volume respectively in preclinical Huntington's disease. *J Cogn Neurosci*, 19(7), 1206-1217. doi:10.1162/jocn.2007.19.7.1206
- Kish, S. J., Shannak, K., & Hornykiewicz, O. (1988). Uneven pattern of dopamine loss in the striatum of patients with idiopathic Parkinson's disease. Pathophysiologic and clinical implications. *The New England journal of medicine*, *318*(14), 876-880. doi:10.1056/NEJM198804073181402
- Klein, C., & Westenberger, A. (2012). Genetics of Parkinson's disease. *Cold Spring Harb Perspect Med*, *2*(1), a008888. doi:10.1101/cshperspect.a008888
- Klunk, W. E., Price, J. C., Mathis, C. A., Tsopelas, N. D., Lopresti, B. J., Ziolko, S. K., . . . DeKosky, S. T. (2007). Amyloid deposition begins in the striatum of presenilin-1 mutation carriers from two unrelated pedigrees. *J Neurosci, 27*(23), 6174-6184. doi:10.1523/JNEUROSCI.0730-07.2007
- Kordower, J. H., Chu, Y., Hauser, R. A., Freeman, T. B., & Olanow, C. W. (2008). Lewy body-like pathology in long-term embryonic nigral transplants in Parkinson's disease. *Nat Med*, *14*(5), 504-506. doi:10.1038/nm1747
- Kordower, J. H., Dodiya, H. B., Kordower, A. M., Terpstra, B., Paumier, K., Madhavan, L., . . . Collier, T. J. (2011). Transfer of host-derived alpha synuclein to grafted dopaminergic neurons in rat. *Neurobiol Dis*, *43*(3), 552-557. doi:10.1016/j.nbd.2011.05.001
- Kordower, J. H., Olanow, C. W., Dodiya, H. B., Chu, Y., Beach, T. G., Adler, C. H., . . . Bartus, R. T. (2013). Disease duration and the integrity of the nigrostriatal system in Parkinson's disease. *Brain*, *136*(Pt 8), 2419-2431. doi:10.1093/brain/awt192

- Kosaka, K., Oyanagi, S., Matsushita, M., & Hori, A. (1976). Presenile dementia with Alzheimer-, Pick- and Lewy-body changes. *Acta Neuropathol*, *36*(3), 221-233.
- Kraemmer, J., Kovacs, G. G., Perju-Dumbrava, L., Pirker, S., Traub-Weidinger, T., & Pirker, W. (2014). Correlation of striatal dopamine transporter imaging with post mortem substantia nigra cell counts. *Mov Disord*, *29*(14), 1767-1773. doi:10.1002/mds.25975
- Kravitz, A. V., Freeze, B. S., Parker, P. R., Kay, K., Thwin, M. T., Deisseroth, K., & Kreitzer, A. C. (2010). Regulation of parkinsonian motor behaviours by optogenetic control of basal ganglia circuitry. *Nature*, *466*(7306), 622-626. doi:10.1038/nature09159
- Kurowska, Z., Englund, E., Widner, H., Lindvall, O., Li, J. Y., & Brundin, P. (2011). Signs of degeneration in 12-22-year old grafts of mesencephalic dopamine neurons in patients with Parkinson's disease. *J Parkinsons Dis, 1*(1), 83-92. doi:10.3233/JPD-2011-11004
- Lacerda, A. L., Nicoletti, M. A., Brambilla, P., Sassi, R. B., Mallinger, A. G., Frank, E., . . . Soares, J. C. (2003). Anatomical MRI study of basal ganglia in major depressive disorder. *Psychiatry Res*, *124*(3), 129-140. doi:10.1016/S0925-4927(03)00123-9
- Lam, H. A., Wu, N., Cely, I., Kelly, R. L., Hean, S., Richter, F., . . . Maidment, N. T. (2011). Elevated tonic extracellular dopamine concentration and altered dopamine modulation of synaptic activity precede dopamine loss in the striatum of mice overexpressing human alpha-synuclein. *J Neurosci Res, 89*(7), 1091-1102. doi:10.1002/jnr.22611
- Lanciego, J. L., Luquin, N., & Obeso, J. A. (2012). Functional neuroanatomy of the basal ganglia. *Cold Spring Harb Perspect Med, 2*(12), a009621. doi:10.1101/cshperspect.a009621
- Lanctot, K. L., Amatniek, J., Ancoli-Israel, S., Arnold, S. E., Ballard, C., Cohen-Mansfield, J., . . . Boot, B. (2017). Neuropsychiatric signs and symptoms of Alzheimer's disease: New treatment paradigms. *Alzheimers Dement (N Y), 3*(3), 440-449. doi:10.1016/j.trci.2017.07.001
- Li, J. Y., Englund, E., Holton, J. L., Soulet, D., Hagell, P., Lees, A. J., . . . Brundin, P. (2008). Lewy bodies in grafted neurons in subjects with Parkinson's disease suggest host-to-graft disease propagation. *Nat Med*, *14*(5), 501-503. doi:10.1038/nm1746
- Liang, C. L., Wang, T. T., Luby-Phelps, K., & German, D. C. (2007). Mitochondria mass is low in mouse substantia nigra dopamine neurons: implications for Parkinson's disease. *Exp Neurol*, 203(2), 370-380. doi:10.1016/j.expneurol.2006.08.015
- Lill, C. M. (2016). Genetics of Parkinson's disease. *Mol Cell Probes, 30*(6), 386-396. doi:10.1016/j.mcp.2016.11.001
- Lippa, C. F., Duda, J. E., Grossman, M., Hurtig, H. I., Aarsland, D., Boeve, B. F., . . . Group, D. P. W. (2007). DLB and PDD boundary issues: diagnosis, treatment, molecular pathology, and biomarkers. *Neurology*, *68*(11), 812-819. doi:10.1212/01.wnl.0000256715.13907.d3
- Liss, B., Haeckel, O., Wildmann, J., Miki, T., Seino, S., & Roeper, J. (2005). K-ATP channels promote the differential degeneration of dopaminergic midbrain neurons. *Nat Neurosci*, 8(12), 1742-1751. doi:10.1038/nn1570
- Liu, J. L., Fan, Y. G., Yang, Z. S., Wang, Z. Y., & Guo, C. (2018). Iron and Alzheimer's Disease: From Pathogenesis to Therapeutic Implications. *Front Neurosci*, *12*, 632. doi:10.3389/fnins.2018.00632

- Lovell, M. A., Robertson, J. D., Teesdale, W. J., Campbell, J. L., & Markesbery, W. R. (1998). Copper, iron and zinc in Alzheimer's disease senile plaques. *J Neurol Sci, 158*(1), 47-52. doi:10.1016/S0022-510X(98)00092-6
- MacPherson, S. E., Della Sala, S., Logie, R. H., & Wilcock, G. K. (2007). Specific AD impairment in concurrent performance of two memory tasks. *Cortex, 43*(7), 858-865. doi:10.1016/S0010-9452(08)70685-3
- Mamelak, M. (2018). Parkinson's Disease, the Dopaminergic Neuron and Gammahydroxybutyrate. *Neurol Ther, 7*(1), 5-11. doi:10.1007/s40120-018-0091-2
- Manecka, D. L., Vanderperre, B., Fon, E. A., & Durcan, T. M. (2017). The Neuroprotective Role of Protein Quality Control in Halting the Development of Alpha-Synuclein Pathology. *Frontiers in Molecular Neuroscience, 10,* 311. doi:10.3389/fnmol.2017.00311
- Marquie, M., Locascio, J. J., Rentz, D. M., Becker, J. A., Hedden, T., Johnson, K. A., . . . Gomperts, S. N. (2014). Striatal and extrastriatal dopamine transporter levels relate to cognition in Lewy body diseases: an (11)C altropane positron emission tomography study. *Alzheimers Res Ther*, 6(5-8), 52. doi:10.1186/s13195-014-0052-7
- Marshall, V. L., Reininger, C. B., Marquardt, M., Patterson, J., Hadley, D. M., Oertel, W. H., . . . Grosset, D. G. (2009). Parkinson's disease is overdiagnosed clinically at baseline in diagnostically uncertain cases: a 3-year European multicenter study with repeat [1231]FP-CIT SPECT. *Mov Disord*, *24*(4), 500-508. doi:10.1002/mds.22108
- Martin, L. J., Pan, Y., Price, A. C., Sterling, W., Copeland, N. G., Jenkins, N. A., . . . Lee, M. K. (2006). Parkinson's disease alpha-synuclein transgenic mice develop neuronal mitochondrial degeneration and cell death. *J Neurosci, 26*(1), 41-50. doi:10.1523/JNEUROSCI.4308-05.2006
- Mazere, J., Lamare, F., Allard, M., Fernandez, P., & Mayo, W. (2017). 123Ilodobenzovesamicol SPECT Imaging of Cholinergic Systems in Dementia with Lewy Bodies. *J Nucl Med*, *58*(1), 123-128. doi:10.2967/jnumed.116.176180
- McKeith, I. G., Dickson, D. W., Lowe, J., Emre, M., O'Brien, J. T., Feldman, H., . . . Consortium on, D. L. B. (2005). Diagnosis and management of dementia with Lewy bodies: third report of the DLB Consortium. *Neurology*, *65*(12), 1863-1872. doi:10.1212/01.wnl.0000187889.17253.b1
- McNab, F., & Klingberg, T. (2008). Prefrontal cortex and basal ganglia control access to working memory. *Nat Neurosci, 11*(1), 103-107. doi:10.1038/nn2024
- Medawar, E., Benway, T. A., Liu, W., Hanan, T. A., Haslehurst, P., James, O. T., . . . Edwards, F. A. (2019). Effects of rising amyloidbeta levels on hippocampal synaptic transmission, microglial response and cognition in APPSwe/PSEN1M146V transgenic mice. *EBioMedicine*, *39*, 422-435. doi:10.1016/j.ebiom.2018.12.006
- Melzer, T. R., Watts, R., MacAskill, M. R., Pitcher, T. L., Livingston, L., Keenan, R. J., . . . Anderson, T. J. (2012). Grey matter atrophy in cognitively impaired Parkinson's disease. *J Neurol Neurosurg Psychiatry*, 83(2), 188-194. doi:10.1136/jnnp-2011-300828
- Muenter, M. D., Forno, L. S., Hornykiewicz, O., Kish, S. J., Maraganore, D. M., Caselli, R. J., . . . Calne, D. B. (1998). Hereditary form of parkinsonism--dementia. *Ann Neurol, 43*(6), 768-781. doi:10.1002/ana.410430612
- Murphy, D. D., Rueter, S. M., Trojanowski, J. Q., & Lee, V. M. (2000). Synucleins are developmentally expressed, and alpha-synuclein regulates the size of the

- presynaptic vesicular pool in primary hippocampal neurons. *J Neurosci, 20*(9), 3214-3220. doi:10.1523/JNEUROSCI.20-09-03214.2000
- Nalls, M. A., Duran, R., Lopez, G., Kurzawa-Akanbi, M., McKeith, I. G., Chinnery, P. F., . . . Sidransky, E. (2013). A multicenter study of glucocerebrosidase mutations in dementia with Lewy bodies. *Jama Neurology*, *70*(6), 727-735. doi:10.1001/jamaneurol.2013.1925
- Narhi, L., Wood, S. J., Steavenson, S., Jiang, Y., Wu, G. M., Anafi, D., . . . Citron, M. (1999).

  Both familial Parkinson's disease mutations accelerate alpha-synuclein aggregation. *J Biol Chem, 274*(14), 9843-9846. doi:10.1074/jbc.274.14.9843
- Nascimento, C., Villela Nunes, P., Diehl Rodriguez, R., Takada, L., Suemoto, C. K., Tenenholz Grinberg, L., . . . Lafer, B. (2019). A review on shared clinical and molecular mechanisms between bipolar disorder and frontotemporal dementia. *Prog Neuropsychopharmacol Biol Psychiatry*, 93, 269-283. doi:10.1016/j.pnpbp.2019.04.008
- Neef, D., & Walling, A. D. (2006). Dementia with Lewy bodies: an emerging disease. *Am Fam Physician*, 73(7), 1223-1229.
- Nelson, A. B., & Kreitzer, A. C. (2014). Reassessing models of basal ganglia function and dysfunction. *Annu Rev Neurosci, 37*, 117-135. doi:10.1146/annurev-neuro-071013-013916
- Ngolab, J., Trinh, I., Rockenstein, E., Mante, M., Florio, J., Trejo, M., . . . Rissman, R. A. (2017). Brain-derived exosomes from dementia with Lewy bodies propagate alphasynuclein pathology. *Acta Neuropathol Commun*, *5*(1), 46. doi:10.1186/s40478-017-0445-5
- Nicholls, D. G. (2008). Oxidative stress and energy crises in neuronal dysfunction. *Ann N Y Acad Sci*, 1147, 53-60. doi:10.1196/annals.1427.002
- Nikolaus, S., Antke, C., & Muller, H. W. (2009). In vivo imaging of synaptic function in the central nervous system: I. Movement disorders and dementia. *Behav Brain Res*, 204(1), 1-31. doi:10.1016/j.bbr.2009.06.008
- Olanow, C. W., Perl, D. P., DeMartino, G. N., & McNaught, K. S. (2004). Lewy-body formation is an aggresome-related process: a hypothesis. *Lancet Neurol*, *3*(8), 496-503. doi:10.1016/S1474-4422(04)00827-0
- Ordonez, D. G., Lee, M. K., & Feany, M. B. (2018). alpha-synuclein Induces Mitochondrial Dysfunction through Spectrin and the Actin Cytoskeleton. *Neuron*, *97*(1), 108-124 e106. doi:10.1016/j.neuron.2017.11.036
- Orme, T., Guerreiro, R., & Bras, J. (2018). The Genetics of Dementia with Lewy Bodies: Current Understanding and Future Directions. *Curr Neurol Neurosci Rep, 18*(10), 67. doi:10.1007/s11910-018-0874-y
- Outeiro, T. F., Koss, D. J., Erskine, D., Walker, L., Kurzawa-Akanbi, M., Burn, D., . . . McKeith, I. (2019). Dementia with Lewy bodies: an update and outlook. *Mol Neurodegener,* 14(1), 5. doi:10.1186/s13024-019-0306-8
- Parkkinen, L., Pirttila, T., & Alafuzoff, I. (2008). Applicability of current staging/categorization of alpha-synuclein pathology and their clinical relevance. *Acta Neuropathol, 115*(4), 399-407. doi:10.1007/s00401-008-0346-6
- Patterson, C. (2018). The state of the art of dementia research: New frontiers. *Alzheimer's Disease International World Alzheimer Report 2018*.

- Perez, R. G., Waymire, J. C., Lin, E., Liu, J. J., Guo, F., & Zigmond, M. J. (2002). A role for alpha-synuclein in the regulation of dopamine biosynthesis. *J Neurosci, 22*(8), 3090-3099. doi:10.1523/JNEUROSCI.22-08-03090.2002
- Pickrell, A. M., & Youle, R. J. (2015). The roles of PINK1, parkin, and mitochondrial fidelity in Parkinson's disease. *Neuron*, *85*(2), 257-273. doi:10.1016/j.neuron.2014.12.007
- Poewe, W., Seppi, K., Tanner, C. M., Halliday, G. M., Brundin, P., Volkmann, J., . . . Lang, A. E. (2017). Parkinson disease. *Nat Rev Dis Primers*, *3*, 17013. doi:10.1038/nrdp.2017.13
- Polymeropoulos, M. H., Lavedan, C., Leroy, E., Ide, S. E., Dehejia, A., Dutra, A., . . . Nussbaum, R. L. (1997). Mutation in the alpha-synuclein gene identified in families with Parkinson's disease. *Science*, *276*(5321), 2045-2047. doi:10.1126/science.276.5321.2045
- Postle, B. R., Locascio, J. J., Corkin, S., & Growdon, J. H. (1997). The time course of spatial and object learning in Parkinson's disease. *Neuropsychologia*, 35(10), 1413-1422. doi:10.1016/S0028-3932(97)00054-7
- Power, J. H., Barnes, O. L., & Chegini, F. (2017). Lewy Bodies and the Mechanisms of Neuronal Cell Death in Parkinson's Disease and Dementia with Lewy Bodies. *Brain Pathol*, 27(1), 3-12. doi:10.1111/bpa.12344
- Ramos, P., Santos, A., Pinto, N. R., Mendes, R., Magalhaes, T., & Almeida, A. (2014). Iron levels in the human brain: a post-mortem study of anatomical region differences and age-related changes. *J Trace Elem Med Biol, 28*(1), 13-17. doi:10.1016/j.jtemb.2013.08.001
- Reich, S. G., & Grill, S. E. (2009). Corticobasal degeneration. *Curr Treat Options Neurol, 11*(3), 179-185.
- Reiner, A., Dragatsis, I., & Dietrich, P. (2011). Genetics and neuropathology of Huntington's disease. *Int Rev Neurobiol*, *98*, 325-372. doi:10.1016/B978-0-12-381328-2.00014-6
- Ring, H. A., & Serra-Mestres, J. (2002). Neuropsychiatry of the basal ganglia. *J Neurol Neurosurg Psychiatry*, 72(1), 12-21. doi:10.1136/jnnp.72.1.12
- Roos, R. A. (2010). Huntington's disease: a clinical review. *Orphanet journal of rare diseases,* 5, 40. doi:10.1186/1750-1172-5-40
- Rosas, H. D., Koroshetz, W. J., Chen, Y. I., Skeuse, C., Vangel, M., Cudkowicz, M. E., . . . Goldstein, J. M. (2003). Evidence for more widespread cerebral pathology in early HD: an MRI-based morphometric analysis. *Neurology*, *60*(10), 1615-1620. doi:10.1212/01.wnl.0000065888.88988.6e
- Sakamoto, M., Uchihara, T., Hayashi, M., Nakamura, A., Kikuchi, E., Mizutani, T., . . . Hirai, S. (2002). Heterogeneity of nigral and cortical Lewy bodies differentiated by amplified triple-labeling for alpha-synuclein, ubiquitin, and thiazin red. *Exp Neurol*, *177*(1), 88-94. doi:10.1006/exnr.2002.7961
- Sanchez-Castaneda, C., Rene, R., Ramirez-Ruiz, B., Campdelacreu, J., Gascon, J., Falcon, C., . . . Junque, C. (2009). Correlations between gray matter reductions and cognitive deficits in dementia with Lewy Bodies and Parkinson's disease with dementia. *Mov Disord*, 24(12), 1740-1746. doi:10.1002/mds.22488
- Sarro, L., Senjem, M. L., Lundt, E. S., Przybelski, S. A., Lesnick, T. G., Graff-Radford, J., . . . Kantarci, K. (2016). Amyloid-beta deposition and regional grey matter atrophy rates

- in dementia with Lewy bodies. *Brain, 139*(Pt 10), 2740-2750. doi:10.1093/brain/aww193
- Sato, S., Uchihara, T., Fukuda, T., Noda, S., Kondo, H., Saiki, S., . . . Hattori, N. (2018). Loss of autophagy in dopaminergic neurons causes Lewy pathology and motor dysfunction in aged mice. *Sci Rep*, *8*(1), 2813. doi:10.1038/s41598-018-21325-w
- Schapira, A. H. (2008). Mitochondria in the aetiology and pathogenesis of Parkinson's disease. *Lancet Neurol*, 7(1), 97-109. doi:10.1016/S1474-4422(07)70327-7
- Seidel, K., Mahlke, J., Siswanto, S., Kruger, R., Heinsen, H., Auburger, G., . . . Rub, U. (2015). The brainstem pathologies of Parkinson's disease and dementia with Lewy bodies. *Brain Pathol*, 25(2), 121-135. doi:10.1111/bpa.12168
- Selkoe, D. J., & Hardy, J. (2016). The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol Med*, 8(6), 595-608. doi:10.15252/emmm.201606210
- Serpell, L. C., Berriman, J., Jakes, R., Goedert, M., & Crowther, R. A. (2000). Fiber diffraction of synthetic alpha-synuclein filaments shows amyloid-like cross-beta conformation. *Proc Natl Acad Sci U S A, 97*(9), 4897-4902. doi:10.1073/pnas.97.9.4897
- Slaughter, J. R., Martens, M. P., & Slaughter, K. A. (2001). Depression and Huntington's disease: prevalence, clinical manifestations, etiology, and treatment. *CNS Spectr*, 6(4), 306-326.
- Sossi, V., de la Fuente-Fernandez, R., Holden, J. E., Schulzer, M., Ruth, T. J., & Stoessl, J. (2004). Changes of dopamine turnover in the progression of Parkinson's disease as measured by positron emission tomography: their relation to disease-compensatory mechanisms. *J Cereb Blood Flow Metab*, *24*(8), 869-876. doi:10.1097/01.WCB.0000126563.85360.75
- Spillantini, M. G., Schmidt, M. L., Lee, V. M., Trojanowski, J. Q., Jakes, R., & Goedert, M. (1997). Alpha-synuclein in Lewy bodies. *Nature*, *388*(6645), 839-840. doi:10.1038/42166
- Stern, Y., Marder, K., Tang, M. X., & Mayeux, R. (1993). Antecedent clinical features associated with dementia in Parkinson's disease. *Neurology*, *43*(9), 1690-1692. doi:10.1212/wnl.43.9.1690
- Striessnig, J., Koschak, A., Sinnegger-Brauns, M. J., Hetzenauer, A., Nguyen, N. K., Busquet, P., . . . Singewald, N. (2006). Role of voltage-gated L-type Ca2+ channel isoforms for brain function. *Biochem Soc Trans*, *34*(Pt 5), 903-909. doi:10.1042/BST0340903
- Sulzer, D. (2007). Multiple hit hypotheses for dopamine neuron loss in Parkinson's disease. *Trends Neurosci*, *30*(5), 244-250. doi:10.1016/j.tins.2007.03.009
- Surmeier, D. J., Guzman, J. N., Sanchez, J., & Schumacker, P. T. (2012). Physiological phenotype and vulnerability in Parkinson's disease. *Cold Spring Harb Perspect Med,* 2(7), a009290. doi:10.1101/cshperspect.a009290
- Surmeier, D. J., Schumacker, P. T., Guzman, J. D., Ilijic, E., Yang, B., & Zampese, E. (2017). Calcium and Parkinson's disease. *Biochem Biophys Res Commun, 483*(4), 1013-1019. doi:10.1016/j.bbrc.2016.08.168
- Tao, Y., Wang, Y., Rogers, J. T., & Wang, F. (2014). Perturbed iron distribution in Alzheimer's disease serum, cerebrospinal fluid, and selected brain regions: a systematic review and meta-analysis. *J Alzheimers Dis*, 42(2), 679-690. doi:10.3233/JAD-140396
- Telling, N. D., Everett, J., Collingwood, J. F., Dobson, J., van der Laan, G., Gallagher, J. J., . . . Hitchcock, A. P. (2017). Iron Biochemistry is Correlated with Amyloid Plaque Morphology in an Established Mouse Model of Alzheimer's Disease. *Cell Chem Biol*, 24(10), 1205-1215 e1203. doi:10.1016/j.chembiol.2017.07.014

- Tereshchenko, A., Magnotta, V., Epping, E., Mathews, K., Espe-Pfeifer, P., Martin, E., . . . Nopoulos, P. (2019). Brain structure in juvenile-onset Huntington disease. *Neurology*, 92(17), e1939-e1947. doi:10.1212/WNL.000000000007355
- Thomas, A. J., Attems, J., Colloby, S. J., O'Brien, J. T., McKeith, I., Walker, R., . . . Walker, Z. (2017). Autopsy validation of 123I-FP-CIT dopaminergic neuroimaging for the diagnosis of DLB. *Neurology*, 88(3), 276-283. doi:10.1212/WNL.000000000003512
- Thomas, A. J., Mahin-Babaei, F., Saidi, M., Lett, D., Taylor, J. P., Walker, L., & Attems, J. (2018). Improving the identification of dementia with Lewy bodies in the context of an Alzheimer's-type dementia. *Alzheimers Res Ther*, 10(1), 27. doi:10.1186/s13195-018-0356-0
- Tiraboschi, P., Attems, J., Thomas, A., Brown, A., Jaros, E., Lett, D. J., . . . McKeith, I. G. (2015). Clinicians' ability to diagnose dementia with Lewy bodies is not affected by beta-amyloid load. *Neurology*, *84*(5), 496-499. doi:10.1212/WNL.00000000001204
- Truban, D., Hou, X., Caulfield, T. R., Fiesel, F. C., & Springer, W. (2017). PINK1, Parkin, and Mitochondrial Quality Control: What can we Learn about Parkinson's Disease Pathobiology? *J Parkinsons Dis, 7*(1), 13-29. doi:10.3233/JPD-160989
- Tsuang, D., Leverenz, J. B., Lopez, O. L., Hamilton, R. L., Bennett, D. A., Schneider, J. A., . . . Zabetian, C. P. (2012). GBA mutations increase risk for Lewy body disease with and without Alzheimer disease pathology. *Neurology*, *79*(19), 1944-1950. doi:10.1212/WNL.0b013e3182735e9a
- Uversky, V. N., & Eliezer, D. (2009). Biophysics of Parkinson's disease: structure and aggregation of alpha-synuclein. *Curr Protein Pept Sci, 10*(5), 483-499. doi:10.2174/138920309789351921
- van Bergen, J. M., Li, X., Hua, J., Schreiner, S. J., Steininger, S. C., Quevenco, F. C., . . . Unschuld, P. G. (2016). Colocalization of cerebral iron with Amyloid beta in Mild Cognitive Impairment. *Sci Rep, 6*, 35514. doi:10.1038/srep35514
- van der Zande, J. J., Steenwijk, M. D., Ten Kate, M., Wattjes, M. P., Scheltens, P., & Lemstra, A. W. (2018). Gray matter atrophy in dementia with Lewy bodies with and without concomitant Alzheimer's disease pathology. *Neurobiol Aging*, 71, 171-178. doi:10.1016/j.neurobiolaging.2018.07.005
- Vann Jones, S. A., & O'Brien, J. T. (2014). The prevalence and incidence of dementia with Lewy bodies: a systematic review of population and clinical studies. *Psychol Med,* 44(4), 673-683. doi:10.1017/S0033291713000494
- Varastet, M., Riche, D., Maziere, M., & Hantraye, P. (1994). Chronic MPTP treatment reproduces in baboons the differential vulnerability of mesencephalic dopaminergic neurons observed in Parkinson's disease. *Neuroscience*, *63*(1), 47-56. doi:10.1016/0306-4522(94)90006-X
- Verghese, P. B., Castellano, J. M., & Holtzman, D. M. (2011). Apolipoprotein E in Alzheimer's disease and other neurological disorders. *Lancet Neurol*, *10*(3), 241-252. doi:10.1016/S1474-4422(10)70325-2
- Verheijen, J., and Sleegers, K. (2018) Understanding Alzheimer Disease at the Interface between Genetics and Transcriptomics. *Trends in Gen*, 34(6), 434-447. doi:10.1016/j.tig.2018.02.007
- Volpicelli-Daley, L. A. (2017). Effects of alpha-synuclein on axonal transport. *Neurobiol Dis,* 105, 321-327. doi:10.1016/j.nbd.2016.12.008
- Vonsattel, J. P. (2008). Huntington disease models and human neuropathology: similarities and differences. *Acta Neuropathol, 115*(1), 55-69. doi:10.1007/s00401-007-0306-6

- Wakabayashi, K., Tanji, K., Mori, F., & Takahashi, H. (2007). The Lewy body in Parkinson's disease: molecules implicated in the formation and degradation of alpha-synuclein aggregates. *Neuropathology*, *27*(5), 494-506. doi:10.1111/j.1440-1789.2007.00803.x
- Walker, Z., Possin, K. L., Boeve, B. F., & Aarsland, D. (2015). Lewy body dementias. *Lancet,* 386(10004), 1683-1697. doi:10.1016/S0140-6736(15)00462-6
- Watson, R., O'Brien, J. T., Barber, R., & Blamire, A. M. (2012). Patterns of gray matter atrophy in dementia with Lewy bodies: a voxel-based morphometry study. *Int Psychogeriatr*, 24(4), 532-540. doi:10.1017/S1041610211002171
- Wessel, J. R., & Aron, A. R. (2013). Unexpected events induce motor slowing via a brain mechanism for action-stopping with global suppressive effects. *J Neurosci*, *33*(47), 18481-18491. doi:10.1523/JNEUROSCI.3456-13.2013
- Wessel, J. R., Conner, C. R., Aron, A. R., & Tandon, N. (2013). Chronometric electrical stimulation of right inferior frontal cortex increases motor braking. *J Neurosci*, 33(50), 19611-19619. doi:10.1523/JNEUROSCI.3468-13.2013
- Wessel, J. R., Jenkinson, N., Brittain, J. S., Voets, S. H., Aziz, T. Z., & Aron, A. R. (2016). Surprise disrupts cognition via a fronto-basal ganglia suppressive mechanism. *Nat Commun*, *7*, 11195. doi:10.1038/ncomms11195
- Wills, J., Jones, J., Haggerty, T., Duka, V., Joyce, J. N., & Sidhu, A. (2010). Elevated tauopathy and alpha-synuclein pathology in postmortem Parkinson's disease brains with and without dementia. *Exp Neurol*, 225(1), 210-218. doi:10.1016/j.expneurol.2010.06.017
- Zarranz, J. J., Alegre, J., Gomez-Esteban, J. C., Lezcano, E., Ros, R., Ampuero, I., . . . de Yebenes, J. G. (2004). The new mutation, E46K, of alpha-synuclein causes Parkinson and Lewy body dementia. *Ann Neurol*, *55*(2), 164-173. doi:10.1002/ana.10795