A randomized phase 2 trial of epigenetic priming with guadecitabine and carboplatin in platinum-resistant, recurrent ovarian cancer

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Corresponding author: Daniela Matei, 303 E Superior Street Lurie 4-107, Chicago, IL, 60610. Phone: 312-503-4853 Fax: 312-503-4853 Email: Daniela.matei@northwestern.edu Running title: Guadecitabine and Carboplatin in Ovarian Cancer Word count: 3508 words Clinical Trial Registration: ClinicalTrials.gov NCT01696032

1 ABSTRACT

2 PURPOSE:

- 3 Platinum resistance in ovarian cancer (OC) is associated with epigenetic modifications.
- 4 Hypomethylating agents (HMAs) have been studied as carboplatin re-sensitizing agents in OC.
- 5 This randomized phase 2 trial compared guadecitabine, a second generation HMA, and
- 6 carboplatin (G+C) against second-line chemotherapy in women with measurable or detectable

7 platinum-resistant OC.

8 PATIENTS AND METHODS:

9 Patients received either G+C (guadecitabine 30 mg/m² SC once-daily for 5 days and carboplatin)

10 or treatment of choice (TC; topotecan, pegylated liposomal doxorubicin, paclitaxel, or

11 gemcitabine) in 28-day cycles until progression or unacceptable toxicity. The primary endpoint

12 was progression-free survival (PFS); secondary endpoints were RECIST v1.1 and CA-125

13 response rate, 6-month PFS, and overall survival (OS).

14 RESULTS:

15 Of 100 patients treated, 51 received G+C and 49 received TC, of which 27 crossed over to G+C.

16 The study did not meet its primary endpoint as the median PFS was not statistically different

- between arms (16.3 weeks vs 9.1 weeks in the G+C and TC groups, respectively; P = 0.07).
- 18 However, the 6-month PFS rate was significantly higher in the G+C group (37% vs. 11% in TC
- 19 group; P = 0.003). The incidence of grade 3 or higher toxicity was similar in G+C and TC

20 groups (51% and 49%, respectively), with neutropenia and leukopenia being more frequent in

21 the G+C group.

22 CONCLUSIONS:

- 23 Although this trial did not show superiority for PFS of G+C versus TC, the 6-month PFS
- 24 increased in G+C treated patients. Further refinement of this strategy should focus on
- 25 identification of predictive markers for patient selection.
- 26
- 27

28 TRANSLATIONAL RELEVANCE

- 29 Although women with ovarian cancer (OC) initially respond to platinum-based chemotherapy,
- 30 platinum-resistance commonly develops, leading to fatal outcomes. We set out to determine if
- 31 epigenetic priming with a hypomethylating agent (HMA) prior to carboplatin improved
- 32 progression-free survival (PFS) in platinum-resistant OC when compared with physician's
- 33 choice chemotherapy in a randomized phase 2 trial. The median PFS and overall survival were
- not different, but the 6-month PFS rate was higher in the experimental group. Myelosuppression
- 35 was the main toxicity observed with the experimental regimen and hypomethylating activity was
- 36 measurable in PBMCs. Further development of the strategy will require identification of
- 37 predictive biomarkers for patient selection.

38 INTRODUCTION

39 Advanced stage high-grade serous ovarian cancer (HGSOC), which is distinctively associated with a p53 mutated signature, has a poor estimated five-year survival of 50% (1). Although 40 patients with HGSOC usually respond to initial platinum-based chemotherapy, relapses occur in 41 most, leading to the development of platinum-resistance and subsequent death (2-3). Progression 42 43 of HGSOC to a platinum-resistant state is caused by multiple mechanisms, including aberrant 44 DNA repair responses, alterations in efflux pump proteins, and accumulated genomic and epigenomic modifications which impact the response of cancer cells to DNA damage. Adaptive 45 46 responses include increased DNA methylation and modifications of histone marks (4-5), which 47 cause transcriptional silencing of tumor suppressor genes (TSGs) and other genes required for chemotherapy-induced cell death (6-7). 48

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50 Given preclinical data demonstrating that targeting DNA methylation to re-sensitize HGSOC to 51 platinum is possible (8-11), we hypothesized this approach would restore platinum sensitivity in HGSOC patients (12,13). With early clinical studies demonstrating feasibility of this strategy 52 53 (13-16), we set out to determine whether targeting DNA methylation induces clinically meaningful activity in platinum-resistant HGSOC by conducting a randomized phase 2 trial. The 54 55 objectives were to measure and compare clinical outcomes of a combination regimen of the 56 DNA methyltransferase inhibitor (DNMTI), guadecitabine, and carboplatin, versus FDAapproved physician's choice chemotherapy (liposomal doxorubicin, weekly paclitaxel, 57 58 topotecan, or gemcitabine). Guadecitabine is a dinucleotide linking decitabine to guanosine via a 59 phosphodiester bond. Guadecitabine is resistant to degradation by cytidine deaminase and has a longer half-life compared to other DNMTIs. In a dose-finding phase I trial (17), therapeutic 60

61	plasma levels of decitabine persisted beyond 8 hours. This pharmacokinetic profile provides a
62	longer window of exposure to the hypomethylating agent (HMA), potentially exposing more
63	cancer cells undergoing S-phase to the parent drug, decitabine, and promoting hypomethylation.
64	Guadecitabine was shown to exert anti-tumor activity in OC xenografts as a single agent and in
65	combination with carboplatin (11, 18, 19).

66

A recently reported phase 1 trial established the tolerable and biologically active dose of 67 guadecitabine in combination with carboplatin (17). Guadecitabine was tolerable at 30 mg/m² SC 68 69 daily for 5 days prior to carboplatin on Day 8 at an AUC of 4. Each cycle was 28 days and the 70 regimen induced ~20% hypomethylation of long interspersed nuclear elements (LINE-1) in peripheral blood mononuclear cells (PBMCs), indicating biological activity. The phase 1 trial 71 72 reported three patients with partial response (PR) and six patients with stable disease (SD) longer than 3 months (17), providing the rationale for conducting this randomized trial in women with 73 platinum-resistant HGSOC. Here we report clinical outcomes with G+C as compared to 74 physician's choice FDA-approved chemotherapy for OC in this high-need patient population. 75 76

77 METHODS

78 Trial Design and Patient Population:

This was a multicenter, randomized, open-label phase 2 trial conducted at 20 centers in the US,
UK, and Canada. Eligible patients were ≥18 years old with platinum-resistant histologically- or
cytologically-confirmed recurrent high-grade serous, or grade 2-3 endometrioid, mixed cell or
clear cell epithelial OC; primary peritoneal carcinoma (PPC); or fallopian tube (FT) cancer. All

patients were required to have received carboplatin and taxanes. Platinum-resistance was defined 83 as recurrence within 6 months of the last platinum-containing regimen. Patients were required to 84 have either measurable disease according to Response Evaluation Criteria in Solid Tumors 85 (RECIST) v1.1 or detectable disease, defined as baseline values of CA-125 at least twice the 86 87 upper limit of normal and one of the following: (i) ascites and/or pleural effusion attributed to 88 tumor, or (ii) solid and/or cystic abnormalities on radiographic imaging that do not meet RECIST 89 definitions for target lesions. Tumor biopsies, paracentesis, or thoracentesis were performed to 90 recover tumor cells and were required at baseline and on Cycle 2 Day 8, if clinically safe and 91 feasible. Eligible patients had acceptable organ function based on laboratory data, Eastern Cooperative Oncology Group performance status (ECOG PS) of 0 or 1 and were \geq 3 weeks from 92 93 their last therapy. Exclusion criteria included carboplatin hypersensitivity, prior HMA therapy, 94 progression on platinum treatment, left ventricular ejection fraction <50%, grade 2 or greater peripheral neuropathy, known brain metastases, other malignancies, active infections, or life-95 96 threatening illnesses. The trial was conducted in accordance with the International Council for Harmonisation Good Clinical Practice guidelines and applicable local regulatory requirements 97 according to the Declaration of Helsinki. Local Institutional Review Boards and Independent 98 99 Ethics Committees reviewed and approved the protocol and the informed consent form. Patients 100 provided written informed consent before enrollment. The trial is registered on 101 ClinicalTrials.gov as NCT01696032. Trial protocol and amendments are available as 102 Supplements 1 and 2, respectively.

103 Randomization, Trial Intervention and Clinical Outcomes:

104 Eligible subjects were randomly assigned (1:1) to receive a 28-day treatment cycle of either a

105 G+C combination treatment (guadecitabine 30 mg/m² SC once-daily on Days 1–5 and

106	carboplatin IV AUC 4 on Day 8), or treatment choice (TC) of topotecan IV $(3.5-4.0 \text{ mg/m}^2/\text{wk})$
107	administered on Days 1, 8 and 15), pegylated liposomal doxorubicin IV (PLD; 40–50 mg/m^2
108	administered on Day 1), paclitaxel IV (60-80 mg/m ² /wk administered on Days 1, 8, 15 and 22),
109	or gemcitabine IV ($800-1000 \text{ mg/m}^2$ administered on Days 1, 8 and 15); treatment choice in the
110	TC arm was at the investigator's discretion. Randomization was stratified by number of prior
111	chemotherapies and by treatment center using an unblinded approach using a centralized web-
112	based system. Concomitant medications and therapies were allowed, as deemed necessary for
113	supportive care and safety of subjects; administration of other anti-cancer agents was not
114	permitted. Treatment in both arms continued until disease progression or unacceptable toxicity.
115	If the investigator decided to stop carboplatin treatment after 4 or more cycles, guadecitabine
116	could be continued until progression or initiation of an alternative anti-cancer treatment.
117	Crossover from the TC arm to the G+C arm was permitted after evidence of disease progression
118	in the standard therapy arm.

119 The primary endpoint was PFS. Secondary efficacy endpoints included objective response rate 120 (ORR: defined as complete response [CR] and partial response [PR] based on both measurable 121 and evaluable disease), PFS at 6 months, clinical benefit rate (CBR: defined as CR+ PR + stable 122 disease for at least 3 months), proportion of patients with CA-125 reduction of at least 50%, 123 duration of response (DOR), and overall survival (OS); in subjects crossing over from the TC to 124 the G+C arm, ORR was measured. Response was assessed using RECIST v1.1 for patients with 125 measurable disease (20), and modified Rustin criteria for patients with detectable disease 126 according to CA-125 criteria (21-22). Tumor measurements were obtained by CT or MRI at 127 screening, after every 2 cycles for the first six cycles, and every three months until progression.

128 Safety was assessed by subject-reported and investigator-observed adverse event (AE) recording, 129 along with physical examination, 12-lead electrocardiograms, hematology, chemistry, and 130 urinalysis with each cycle. There was a 30-day (+5 day) safety visit after the last treatment. AEs 131 were graded by Common Terminology Criteria for Adverse Events (CTCAE) v4.0. Treatment-132 emergent AEs (TEAEs) were defined as events that first occurred or worsened after the first dose 133 of trial drug given on the first day of the first treatment cycle until 30 days after the last dose of 134 treatment. Related serious AEs (SAEs) that occurred more than 30 days after the last dose were 135 also considered TEAEs; AEs occurring after the start of an alternative anti-cancer treatment were 136 not considered TEAEs. Patients lost to follow-up were included in statistical analyses to the point of their last evaluation.

137 point of their last evaluation.

138 Exploratory pharmacodynamic endpoints included quantitative analysis of LINE-1 methylation 139 in PBMCs and tumor DNA, and of selected gene promoters in tumor tissue. Blood samples for 140 methylation assays were collected weekly during Cycle 1 and on Day 1 and Day 8 thereafter. 141 Global DNA methylation was evaluated by sodium bisulfite pyrosequencing for LINE-1 CpGs 142 using PyroMark Q24 as previously described (17). Ascites, pleural fluid, or fresh tumor biopsies 143 were obtained at screening and on Day 8 of Cycle 2 for assessment of methylation of selected 144 genes listed in the supplementary information (Supplementary Table S1). DNA was extracted 145 from tumor biopsies or ascites using DNeasy Blood & Tissue Kit (Qiagen, Netherlands) and 146 LINE-1 and specific gene pyrosequencing was performed at EpigenDx Inc (Hopkinton, MA).

147 Statistical Design and Analyses:

148 It was estimated a sample size of ≥96 patients randomized 1:1 into two treatment arms would
149 provide approximately 80% power to detect a difference between the two PFS curves (median

150 PFS of 15 vs. 28 weeks for the TC and G+C arms) at 5% significance level using a two-sided log-rank test, assuming uniform accrual of subjects over 12 months, a 24-month trial duration 151 152 and an exponential distribution of the PFS endpoint. PFS, OS, and 95% confidence intervals 153 (CIs) were evaluated using the Kaplan-Meier method. PFS and OS were compared using the log-154 rank test, while ORR and CBR were compared using Fisher's exact test. Subjects still alive with 155 no progression and those who withdrew were censored on the date of the last adequate tumor, 156 CA-125, or clinical progression assessment. Subjects initiating subsequent anti-cancer therapy, 157 including those who crossed over, were censored accordingly, but prior to the initiation. Survival 158 time was censored on the last date the subject was known to be alive or lost to follow-up before 159 reaching the event of death. Efficacy and safety data for subjects who crossed over were 160 tabulated separately once guadecitabine was first administered. All analyses are descriptive and 161 inferential statistical tests and CIs were two-sided with alpha equal to 0.05 unless otherwise specified. The database was locked for analysis on July 7, 2016 with mature PFS data; 97 of the 162 163 100 treated patients progressed or did not survive and all patients discontinued protocol therapy at this time (Figure 1). LINE-1 and gene-specific methylation level differences before and after 164 G+C treatment were determined using paired t-tests. SAS version 9.3 was used for all statistical 165 166 analyses.

167 **RESULTS**

One hundred and three patients with HGSOC, FT cancer, or PPC were enrolled and randomized (52 G+C, 51 TC) and 100 received treatment (51 G+C, 49 TC; Figure 1). Baseline characteristics are summarised in Table 1 and were well balanced between the two arms in terms of age, performance status, prior therapy, and ethnicity. More patients randomized to the G+C arm had PPC compared to those randomized to TC (10 vs. 0). Most subjects were white, with a median

173	age of 62 years, and all received prior platinum-based therapy (Table 1). Of the patients
174	randomized to TC, 11 received weekly paclitaxel, 15 received liposomal doxorubicin, 20
175	received topotecan, and 3 received gemcitabine. Patients in the G+C arm received more
176	treatment cycles than subjects in the TC arm (median of 4.0 vs. 2.0 cycles, respectively), with
177	59% of subjects in the G+C arm receiving at least 3 cycles of treatment and 37% receiving at
178	least 6 cycles of treatment vs. 47% and 31% of subjects in the TC arm, respectively. Fifty-five
179	percent of patients from the TC arm crossed over to G+C arm following progression (Figure 1).
180	Disease progression was the most common reason for discontinuing treatment (~80% of patients
181	in each group; Figure 1). The most common TEAEs occurring in more than 5% of the trial
182	population are reported in Table 2. AE frequencies between the two arms were similar, but
183	neutropenia, diarrhea, nausea and vomiting were more common in the G+C arm (Tables 2 and
184	3).

185 The median duration of PFS in the G+C arm was 16.2 weeks compared to 9.1 weeks in TC arm 186 (P=0.07; Figure 2A and Table 4). The 6-month PFS rate was 37% in the G+C arm (95% CI, 187 [0.24; 0.50]) compared to 11% in the TC arm (95% CI, [0.04; 0.22]; p=0.003) and did not meet 188 the pre-specified criterion for superiority (HR 0.686, 95% CI, [0.456; 1.030]; Figure 2 and Table 189 4). There was no difference between the two arms in OS (43 and 40 weeks in the G+C and TC 190 arms, respectively; Figure 2B and Table 4), OS survival rate at 6 months (0.72 and 0.67 in the 191 G+C and TC arms, respectively; Table 4), overall response rate (ORR; 16% and 8% in the G+C 192 and TC arms, respectively; Table 4), or clinical benefit response by RECIST v1.1 or CA-125 193 (Table 4, Supplementary Table S2). Twenty-seven patients from the TC arm crossed over post-194 progression into the G+C arm and received a median of 3 cycles (14 subjects received \geq 3 cycles

195	and 5 subjects received \geq 6 cycles) with a CA-125 response being confirmed in 6 of 21 evaluable
196	subjects (29%). Patient disposition and outcomes are included in Supplementary Table S3.
197	To determine the biological activity of the G+C regimen, LINE1 methylation was assessed in
198	PBMCs from 48 patients randomized to the G+C arm. Similar to the first stage of this trial (17),
199	LINE1 hypomethylation approximated 20% (C1D8 vs. C1D1; range +15% to -55%;
200	Supplementary Figure S1A) (17). In 15 patients who continued treatment beyond 2 cycles and
201	for whom PBMCs were available, LINE1 hypomethylation observed during Cycle 1 was
202	maintained or increased during subsequent cycles (Supplementary Figure S1B), indicating that
203	G+C maintains its biological effects throughout treatment. Correlation between clinical response
204	and pharmacodynamic effects as measured by LINE-1 hypomethylation in PBMCs was not
205	observed. Promoter methylation of selected genes representing TSGs (23-24) or tumor antigens
206	known to be methylated in OC (25-26) was measured in bisulfite-converted DNA obtained from
207	paired tumor biopsies on C1D1 and C2D8 (n = 8 paired specimens). Treatment-induced
208	hypomethylation of MAGE-A2 and MAGE-A3 promoters in tumor DNA was significant
209	(Supplementary Figure S1C). A non-significant decrease in promoter CpG methylation was also
210	observed for LINE-1 and for the tumor antigens NY-ESO-1 and MAGE-A11, but not for the
211	TSGs RASSF1A, MLH1 and BRCA1 (data not shown) or for the differentiation associated gene
212	HOXA11. Taken together, these results provide evidence that G+C treatment exerts in vivo
213	hypomethylating activity detectable in PBMCs and tumors.

214

215 **DISCUSSION**

216 This is the first randomized study comparing a regimen of G+C to standard of care 217 chemotherapy for recurrent platinum-resistant OC. Although the 6-month PFS rate was higher in 218 the G+C arm than the TC arm, the study did not meet its primary endpoint in this heavily pre-219 treated population. These results are comparable with previous single-arm phase 2 studies using 220 an epigenetic priming with decitabine (13-14) or 5-azacitadine (15) prior to carboplatin. Those 221 trials used repetitive low doses of DNMTIs, which is similar to the strategy employed with this 222 class of HMAs in hematological malignancies (27-28). The repetitive administration of the HMA 223 increases drug exposure of cells undergoing S-phase and incorporation of the nucleoside 224 analogue into the replicating DNA, trapping DNMTs and inhibiting *de novo* methylation.

225

226 In contrast, a previous trial conducted by the Scottish Gynecological Trials Group that used 227 bolus administration of decitabine on Day 1 prior to administration of carboplatin a week later 228 was prematurely closed due to high hematological toxicity and indicated lower efficacy of the 229 combination regimen compared to carboplatin alone (29). This trial reported reduction in 230 efficacy with the addition of decitabine to patients with partially platinum sensitive recurrence 231 when given in conjunction with carboplatin (29). Whether the difference in administration (bolus 232 vs. low-dose repetitive administration) was solely responsible for the differences in levels of 233 clinical activity remains unknown. The clinical efficacy differences with this trial may be 234 attributable to the Scottish trial's inclusion of less heavily pre-treated subjects who retained 235 partial platinum sensitivity. Since increased DNA methylation is observed in advanced bladder 236 cancer, colon cancer, cholangiocarcinoma, and germ cell tumors (30), DNMTI-induced

237	sensitization to platinum or to chemotherapy is also explored in these settings with early
238	promising results having been reported recently in colon cancer (31).

239

240 The G+C regimen had myelosuppression as the main toxicity. Prolonged neutropenia required 241 growth factor support in >80% of the patient population to maintain the intended every-4-week 242 administration of the combination. However, infections were rare and no episodes of neutropenic 243 sepsis were recorded. Hypersensitivity and other adverse infusion reactions were observed in 9 244 (18%) and 8 (15%) patients in the G+C arm compared with 6% in the TC arm in this trial, which 245 is concordant with similar observations from prior trials of DNMTIs and carboplatin (13, 29). This is most likely due to increased exposure to platinum therapy in the experimental arm, but it 246 247 is also possible HMA treatment may augment type II allergic reactions.

248

249 The study has few limitations. While all patients in this trial had platinum-resistant disease, 250 platinum-refractory disease was excluded. Given that carboplatin was not included among the 251 potential control regimens, and could conceivably induce clinical benefit in selected patients, this 252 trial cannot exclude the activity of single-agent carboplatin in this population. Additionally, 253 topotecan administration in the TC arm followed a weekly administration schedule. While this 254 schedule was favored among treating oncologists due to its favorable toxicity profile and early 255 reports of activity (32), the regimen was subsequently shown to induce a decreased response rate 256 compared to the schedule using daily administration for 5 days, although OS was not affected 257 (33). Chemotherapy with bevacizumab became FDA-approved and an accepted standard for 258 patients with platinum resistant OC after results of Aurelia trial were reported (34), which 259 occurred after the inception of this protocol. Of note is that prior therapy with bevacizumab was

260	not excluded, and 33 patients enrolled in this trial had received bevacizumab. The shorter median
261	PFS observed in the control group of this study (~2 months) compared to the Aurelia trial (3.4
262	months; 34) reflects a more heavily pre-treated group patients included here (mean of 3-4 prior
263	regimens) for whom limited treatment options currently exist.
264	
265	High-quality nucleic acids were extracted from tumor biopsies from 40 subjects at baseline and
266	from 8 patients after two cycles of G+C. The precise mechanism by which G+C induces anti-
267	tumor responses remains unknown. Our tissue- and cell-based analyses showed a number of
268	genes and pathways involved in DNA repair and response to chemotherapy (e.g., DOK2,
269	miR193a, 14-3-3 σ , RASSF1A) are silenced through promoter methylation and re-expressed after
270	guadecitabine treatment (35). Using overexpression or knock-down strategies, we have shown
271	some of these pathways restore platinum sensitivity in OC cell lines and xenografts (10, 35). It is
272	likely that not one gene, but a more global genomic program is reactivated in response to DNA
273	hypomethylation, allowing tumor cells to undergo apoptosis in response to chemotherapy. Since
274	preclinical models show that guadecitabine selectively eliminates chemotherapy-resistant OC
275	stem cells (11) by inducing a cellular differentiation program, the G+C regimen may exert anti-
276	tumor activity through multiple mechanisms. The low number of post-treatment biopsies
277	collected in the trial limits the strength of the conclusions we can draw regarding the
278	mechanisms elicited by this HMA in vivo.
279	
280	This randomized trial demonstrated that epigenetic priming in combination with carboplatin did

281 not increase PFS compared to standard chemotherapy, but improved 6-month PFS in platinum-

resistant OC. Although these results do not support development of this strategy for an

- unselected population, they suggest a subgroup of patients might have benefitted from G+C
- treatment. Future studies should focus on developing predictive markers to enrich a patient
- 285 population more likely to benefit from the use of HMAs.
- 286
- 287

288 DISCLOSURE OF POTENTIAL CONFLICTS OF INTEREST

289 Angeles Alvarez Second reported being paid for consulting or participating in an advisory role 290 for Alexion, Aravive, AstraZeneca, Clovis, Janssen/Johnson & Johnson, Mesano, Myriad, 291 Roche/Genentech, and Tesaro, and received research funding from Amgen, AbbVie, Amgen, 292 Astellas Pharma Inc., Astex Pharmaceuticals Inc., AstraZeneca, Boerhinger Ingelheim, Bristol 293 Myers Squibb, Eisai, Exelixis, Endocyte, Roche/Genentech, Incyte, Merck, PharmaMar, Prima 294 Biomed, and Tesaro. Sarah Blagden reported serving in a consultant or advisory role for 295 Novartis, Octimet and Roche, receiving travel, accommodation and expense reimbursement from 296 NuCana, BioMed and Tesaro, receiving research funding from NuCana, BioMed, Sierra 297 Oncology, Incyte, DesigneRx and Tesaro, and holds patents or receives royalties from RNA 298 Guardian Ltd. Susana Banerjee reported receiving honoraria from AstraZeneca and Tesaro, 299 serving in a consultant or advisory role for AstraZeneca, Tesaro, Clovis, Seattle Genetics, and 300 receiving research funding from AstraZeneca. John Nemunaitis disclosed employment with 301 Gradulis, leadership roles with Gradulis and Symvivo. He has stock or other ownership interest 302 to disclose with Gradulis, received honoraria from AstraZeneca, has consulted for AstraZeneca 303 and Symvivo, participated in a speaker's bureau for AstraZeneca, received research funding from 304 Gradulis, holds patents or receives royalties from Gradulis, receives travel, accommodations, or 305 expenses from AstrazZeneca, Symvivo and Gradulis, and been paid to provide expert testimony 306 on behalf of Foundation Medicine. Hal Hirte reported receiving honoraria from AstraZeneca, 307 Merck and Roche. Diane Provencher reported consulting and advising AstraZeneca. Benjamin 308 Schwartz reported receiving honoraria from NOVADAQ. Patricia Braly reported participating in 309 a speakers' bureau for Myriad, Invitae, Tesaro, AstraZeneca, Clovis and Roche, and receiving 310 research funding from Tesaro, AstraZeneca, Merck, Janssen, Pharma Mar and Xenetic. Geoffrey

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318	AUTHORS' CONTRIBUTIONS:
319	Conception and design: Matei, Nephew, Azab, Oganesian, Naim, Hao, Keer.
320	Development of methodology: Matei, Nephew, Azab, Oganesian, Naim, Hao, Keer.
321	Acquisition of data (provided animals, acquired and managed patients, provided
322	facilities, etc.): All authors.
323	Analysis and interpretation of data (e.g., statistical analysis, biostatistics,
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325	Writing, review, and/or revision of the manuscript: All authors.
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REFERENCES

1. Society AC. Cancer Facts and Figures 2018. Atlanta, GA, American Cancer Society.

2. Liu CM. Cancer of the ovary. *New Engl J Med*. 2005;352:1268-9; author reply 1268-9.

3. Sandercock J, Parmar MK, Torri V, Wian W. First-line treatment for advanced ovarian cancer: paclitaxel, platinum and the evidence. *Br J Cancer* 2002;87:815-24.

4. Watts GS, Futscher BW, Holtan N, Degeest K, Domann, FE, Rose SL. DNA methylation changes in ovarian cancer are cumulative with disease progression and identify tumor stage. *BMC Med Genomics* 2008;1:47.

5. Barton CA, Hacker NF, Clark SJ, O'Brien PM. DNA methylation changes in ovarian cancer: implications for early diagnosis, prognosis and treatment. *Gynecol Oncol* 2008;109:129-39.

6. Balch C, Fang F, Matei DE, Huang TH, Nephew KP. Minireview: epigenetic changes in ovarian cancer. *Endocrinology* 2009;150:4003-11.

7. Barton CA, Clark SJ, Hacker NF, O'Brien PM. Epigenetic markers of ovarian cancer. *Adv Exp Med Biol* 2008;622:35-51.

8. Plumb JA, Finn PW, Williams RJ, Bandara MJ, Watkins CJ, La Thangue NB, *et al.* Pharmacodynamic response and inhibition of growth of human tumor xenografts by the novel histone deacetylase inhibitor PXD101. *Mol Cancer Ther* 2003;2:721-8.

9. Plumb JA, Strathdee G, Sludden J, Kaye SB, Brown R. Reversal of drug resistance in human tumor xenografts by 2'-deoxy-5-azacytidine-induced demethylation of the hMLH1 gene promoter. *Cancer Res* 2000;60:6039-44.

10. Fang F, Munck J, Tang J, Taverna P, Wang Y, Miller DF, *et al.* The novel, small-molecule DNA methylation inhibitor SGI-110 as an ovarian cancer chemosensitizer. *Clin Cancer Res* 2014;20:6504-16.

11. Wang Y, Cardenas H, Fang F, Condello S, Taverna P, Segar M, *et al.* Epigenetic targeting of ovarian cancer stem cells. *Cancer Res* 2014;74:4922-36.

12. Fang F, Zuo Q, Pilrose J, Wang Y, Shen C, Li M, *et al.* Decitabine reactivated pathways in platinum resistant ovarian cancer. *Oncotarget* 2014;5:3579-89.

13. Matei D, Fang F, Shen C, Schilder J, Arnold A, Zeng Y, *et al.* Epigenetic resensitization to platinum in ovarian cancer. *Cancer Res* 2012;72:2197-205.

14. Fang F, Balch C, Schilder J, Breen T, Zhang S, Shen C, *et al.* A phase 1 and pharmacodynamic study of decitabine in combination with carboplatin in patients with recurrent, platinum-resistant, epithelial ovarian cancer. *Cancer* 2010;116:4043-53.

15. Fu S, Hu W, Iyer R, Kavanagh JJ, Coleman RL, Levenback CF, *et al.* Phase 1b-2a study to reverse platinum resistance through use of a hypomethylating agent, azacitidine, in patients with platinum-resistant or platinum-refractory epithelial ovarian cancer. *Cancer* 2011;117:1661-9.

16. Appleton K, Mackay HJ, Judson I, Plumb JA, McCormick C, Strathdee G, *et al.* Phase I and pharmacodynamic trial of the DNA methyltransferase inhibitor decitabine and carboplatin in solid tumors. *J Clin Oncol* 2007;25:4603-9.

17. Matei D, Ghamande S, Roman L, Alvarez Secord A, Nemunaitis J, Markham MJ, *et al.* A Phase I Clinical Trial of Guadecitabine and Carboplatin in Platinum-Resistant, Recurrent Ovarian Cancer: Clinical, Pharmacokinetic, and Pharmacodynamic Analyses. *Clin Cancer Res* 2018;24:2285-93.

18. Wang Y, Zong X, Mitra S, Mitra AK, Matei D, Nephew KP. IL-6 mediates platinum-induced enrichment of ovarian cancer stem cells. *JCI Insight* 2018; 3:doi:10.1172/jci.insight.122360.

19. Alvero AB, Chen R, Fu HH, Montagna M, Schwartz PE, Rutherford T, *et al.* Molecular phenotyping of human ovarian cancer stem cells unravels the mechanisms for repair and chemoresistance. *Cell Cycle* 2009;8:158-66.

20. Eisenhauer EA, Therasse P, Bogaerts J, Schwartz LH, Sargent D, Ford R, *et al.* New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). *Eur J Cancer* 2009;45:228-47.

21. Rustin GJ. Use of CA-125 to assess response to new agents in ovarian cancer trials. *J Clin Oncol* 2003;21:187s-193s.

22. Rustin GJ, Bast RC, Jr., Kelloff GJ, Barrett JC, Carter SK, Nisen PD, *et al.* Use of CA-125 in clinical trial evaluation of new therapeutic drugs for ovarian cancer. *Clin Cancer Res* 2004;10:3919-26.

23. Ibanez de Caceres I, Battagli C, Esteller M, Herman JG, Dulaimi E, Edelson MI, *et al.* Tumor cell-specific BRCA1 and RASSF1A hypermethylation in serum, plasma, and peritoneal fluid from ovarian cancer patients. *Cancer Res* 2004;64:6476-81.

24. Imura M, Yamashita S, Cai LY, Furuta JI, Wakabayashi M, Yasugi T, *et al.* Methylation and expression analysis of 15 genes and three normally-methylated genes in 13 Ovarian cancer cell lines. *Cancer Lett* 2006;241:213-20.

25. Odunsi K, Matsuzaki J, James SR, Mhawech-Fauceglia P, Tsuji T, Miller A, *et al.* Epigenetic potentiation of NY-ESO-1 vaccine therapy in human ovarian cancer. *Cancer Immunol Res* 2014;2:37-49.

26. Zhang W, Barger CJ, Link PA, Mhawech-Fauceglia P, Miller A, Akers SN, *et al.* DNA hypomethylation-mediated activation of Cancer/Testis Antigen 45 (CT45) genes is associated with disease progression and reduced survival in epithelial ovarian cancer. *Epigenetics* 2015;10:736-48.

27. Issa JP, Gharibyan V, Cortes J, Jelinek J, Morris G, Verstovsek S, *et al.* Phase II study of low-dose decitabine in patients with chronic myelogenous leukemia resistant to imatinib mesylate. *J Clin Oncol* 2005;23:3948-56.

28. Issa JJ, Roboz G, Rizzieri D, Jabbour E, Stock W, O'Connell C, *et al.* Safety and tolerability of guadecitabine (SGI-110) in patients with myelodysplastic syndrome and acute myeloid leukaemia: a multicentre, randomised, dose-escalation phase 1 study. *Lancet Oncol* 2015;16:1099-110.

29. Glasspool RM, Brown R, Gore ME, Rustin GJ, McNeish IA, Wilson RH, *et al.* A randomised, phase II trial of the DNA-hypomethylating agent 5-aza-2'-deoxycytidine (decitabine) in combination with carboplatin vs carboplatin alone in patients with recurrent, partially platinum-sensitive ovarian cancer. *Br J Cancer* 2014;110:1923-9.

30. Albany C, Hever-Jardine MP, von Herrmann KM, Yim CY, Tam J, Warzecha JM, *et al.* Refractory testicular germ cell tumors are highly sensitive to the second generation DNA methylation inhibitor guadecitabine. *Oncotarget* 2017;8:2949-59.

31. Lee V, Wang JS, Zahurak ML, Gootjes E, Verheul HM, Parkinson R, *et al.* A phase I trial of a guadecitabine (SGI-110) and irinotecan in metastatic colorectal cancer patients previously exposed to irinotecan. *Clin Cancer Res* 2018;24:6160-7.

32. Levy T, Inbar M, Menczer J, Grisaru D, Glezerman M, Safra T. Phase II study of weekly topotecan in patients with recurrent or persistent epithelial ovarian cancer. *Gynecol Oncol* 2004;95:686-90.

33. Sehouli J, Stengel D, Harter P, Kurzeder C, Belau A, Bogenrieder T, *et al.* Topotecan Weekly Versus Conventional 5-Day Schedule in Patients With Platinum-Resistant Ovarian Cancer: a randomized multicenter phase II trial of the North-Eastern German Society of Gynecological Oncology Ovarian Cancer Study Group. *J Clin Oncol* 2011;29:242-8.

34. Pujade-Lauraine E, Hilpert F, Weber B, Reuss A, Poveda A, Kristensen G, *et al.* Bevacizumab combined with chemotherapy for platinum-resistant recurrent ovarian cancer: The AURELIA open-label randomized phase III trial. *J Clin Oncol* 2014;32:1302-8.

35. Fang F, Cardenas H, Huang H, Jiang G, Perkins SM, Zhang C, *et al.* Genomic and Epigenomic Signatures in Ovarian Cancer Associated with Re-sensitization to Platinum Drugs. *Cancer Res* 2018;78:631-644.

FIGURE LEGENDS

Figure 1. Disposition of subjects in the trial. AUC indicates the target area under the concentration-versus-time curve.

Figure 2. Survival of subjects assigned to G+C arm versus TC arm. A: Kaplan-Meier estimates of progression-free survival with the G+C treatment and TC regimens. B: Kaplan-Meier estimates of overall survival with the G+C treatment and TC regimens. For subjects in the TC group who crossed over to receive G+C, OS time was censored at the crossover time point.