

On outflow boundary conditions for CT-based computation of FFR: examination using PET images

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Abstract: CT-based computations of fractional flow reserve (FFR) have been widely utilized for evaluating functional severity of a coronary artery stenosis. Whilst this approach has been successful clinically, assumptions involved in the analysis still need to be investigated for further improvement of predictive accuracy. To better understand the sensitivity of computational FFRs on outflow boundary condition – typically reflecting patient’s own physiology only through anatomical features – FFR computations for 10 patients with different degree of stenosis was conducted. The computations were based on 3D anatomical model reconstructed from CT images and patient-specific in/outflow boundary conditions (BC). Two outflow BCs were considered: (1) conventional morphology-based and (2) PET perfusion-based conditions. The results showed that the FFRs derived from the two boundary conditions agree in general. It was also found that the FFRs computed with the morphology-based BC tend to estimate higher functional severity, especially in patients with reduced vasodilatory response under hyperaemia – an essential physiological condition in FFR measurement. Further investigation was made by varying hyperaemic resistances (30%-90% of the baseline) in the morphology-based BC. The variation of FFR for the varied resistances was narrow for patients with mild stenosis and wider for those who have severe stenosis. This latter approach confirmed that variability of FFR due to outflow condition tends to come from overestimation of vasodilatory response, especially those who have abnormal myocardial perfusion. The results suggest that outflow conditions that are more representative of each patient could be an effective way to improve CT-based FFR computation.

Keywords: Coronary artery stenosis; CT-based FFR computation; outflow boundary conditions; positron emission tomography perfusion imaging

Highlights

- Computational FFR analysis was conducted using outflow conditions based on PET images to reflect myocardial perfusion in CFD.
- FFRs derived using conventional morphology-only-based outflow conditions (MBC) and PET-based conditions (PBC) agreed well.
- The difference of FFRs due to types of boundary condition is minimal for patients with mild stenosis and larger for those having a severe stenosis.
- The MBC with reduced peripheral resistance response generally explains the difference of FFR prediction due to poor myocardial perfusion.
- Additional model for poor vasodilatory response of patients with myocardial disease may improve the accuracy of the CT-based FFR computation.

46 1. Introduction

47 Coronary artery disease (CAD) is the single largest cause of death globally and accounts for 12%
48 of annual deaths in the UK.[1] When coronary arteries are obstructed, it can lead to a mismatch
49 between myocardial demand and supply for oxygen and nutrients, leading to failure of myocardial
50 function, a state known as ischaemia.

51 In the clinical setting, the most common method to evaluate the severity of an obstruction
52 involves X-ray coronary angiography, taking projection images of the arteries to determine the
53 percentage of the diameter of the obstructed artery. However, for obstructions that are of an
54 intermediate level, the predictive power of this approach for ischaemia is low and inter-observer
55 variability is high, in the study by Fischer et al, using 3 observers, agreement on stenosis severity was
56 as low as 55% and the overall concordance with FFR was about 50%.[2–4] This is because the
57 ischaemic consequences of an obstruction does not only depend on the anatomical degree of
58 obstruction, prompting the need of an indicator that assesses its functional severity.[5]

59 It was discovered that the severity of an obstructive stenosis can be determined by the ratio of
60 maximum blood flow distal to a stenotic lesion to normal maximum flow in the myocardium.
61 Practically, the ratio can be calculated by measuring the ratio between the distal and proximal
62 pressures with respect to the stenosis.[6] This ratio is known as Fractional Flow Reserve (FFR). It has
63 been demonstrated in the FAME 1 and FAME 2 trials that FFR is an effective diagnostic indicator in
64 deciding the suitability of stent treatments.[7] FFR is measured during invasive coronary
65 angiography where a pressure wire is inserted as part of catheterization to measure the blood
66 pressure distal and proximal to the obstruction. The measurement takes place when the patient is
67 in induced hyperaemia, under which blood flow across the coronary vasculature is maximized,
68 usually by the administration of vasodilators such as adenosine.[6] The ratio of the pressures is
69 defined as FFR and stenting is recommended when the FFR is lower than 0.8 that means the pressure
70 distal to the obstruction is 80% of the pressure proximal. FFR is currently the gold standard for
71 determining the severity of coronary artery obstructions.[8]

72 Computed Tomography Coronary Angiography (CTCA), has been more recently been pushed
73 to the forefront of CAD diagnosis, due to its relatively lower cost and non-invasive procedure.[9]
74 However, similarly to X-ray coronary angiography, CTCA only depicts the anatomical degree of
75 obstruction rather than the functional severity. To address this limitation, FFRCT, also known as CT-
76 based FFR or CT-FFR has been proposed [10], where 3D coronary artery anatomy obtained from
77 CTCA is used in a computational fluid dynamics (CFD) simulation to calculate pressure in the
78 vasculature. FFR can be obtained subsequently by assessing the computed pressure profile across a
79 stenosis. CT-based FFR analysis for clinical diagnosis is gaining mainstream acceptance with various
80 health governing bodies such as UK National Institute of Care and Excellence (NICE) recommending
81 it as an diagnostic option, and the performance has been demonstrated in various clinical studies.[11–
82 13] Effort has been made to further this technology, mostly to make it simplified for faster
83 computation such as QFR[14], vFFR[15] and for the use with wider variety of imaging modality.
84 Questions always arise however, with any simulation-based “measurements”, regarding model
85 assumptions, sensitivity to boundary conditions and input parameters.

86 A particular challenge is to model the behavior of the microvascular systems – vasodilation
87 during the administration of vasodilators such as adenosine to induce hyperaemia in FFR
88 measurements. Adenosine activates the A2A receptor causing coronary artery vasodilation, leading
89 to 3.5 to 4-fold increase in myocardial flow in healthy humans.[16] The increase in flow under
90 hyperaemia is known as coronary flow reserve (CFR), a functional measurement of coronary health
91 similar but distinct to FFR. It reflects the health of an entire coronary tree including
92 microvasculature whereas FFR only assesses the possibility of epicardial disease. In microvascular
93 ischaemic disease, the vasodilatory response is reduced (a CFR of <2 is common in diseased patients)
94 and in some extreme cases, adenosine causes no change from the resting state ($CFR \leq 1$).[16] In
95 simulation-based measurements, these microvessels are part of the downstream boundary conditions
96 for the outflowing coronary branches, which are typically defined by the resistance of the
97 microvessels. The ability of microvascular dilatation can thus have a significant influence on the flow

98 rate through each coronary branch. As CT cannot resolve microvascular behavior, many
99 simulations resort to using this 4-fold increase of the flow as an assumption to adjust the downstream
100 microvascular resistances for hyperaemic conditions.[10]

101 To tackle this challenge, we integrated CTCA and Positron Emission Tomography (PET)
102 perfusion imaging into CT-based FFR measurements to realise simulations with outflow boundary
103 conditions that are more patient-specific and representative of diseased coronary arteries. PET
104 perfusion is currently the gold standard for assessing myocardial perfusion among available imaging
105 modalities: SPECT, CT perfusion and MR perfusion.[17] Because PET myocardial perfusion imaging
106 is performed on both rest and stressed states (i.e. normal and hyperaemia, respectively), it allows to
107 incorporate a clear picture of how the microvasculature dilates in CT-based FFR simulation as
108 outflow boundary conditions. In this pilot study using 10 patients' data, our aim was to examine the
109 sensitivity of FFR to the types of outflow boundary conditions and evaluate the significance of
110 patient-specific outflow boundary conditions. Additionally, a method to evaluate a potential range
111 of FFR for individual patient is proposed and tested as an alternative method when perfusion is not
112 known from PET or other type of imaging.

113

114 **2. Materials and Methods**

115 In this study, CFD analyses on 3D patient-specific anatomical models were conducted using various
116 outflow boundary conditions: (1) structured tree boundary conditions with typical hyperaemic
117 response (2) structured tree boundary conditions with hyperaemic response that is varied across the
118 possible disease spectrum and (3) perfusion (PET) based boundary conditions.

119

120 *2.1 Patients*

121

122 This study included 10 patients (6 male, 4 female, age: 61.7 ± 12.2 years) of various levels of
123 angiographically determined epicardial stenosis (6 mild, 2 intermediate and 3 severe case). The
124 patients presented chest pain and other symptoms that indicated an intermediate risk of coronary
125 artery disease. All patients underwent 4D CTCA for anatomical assessment and ^{82}Rb PET perfusion
126 imaging to identify ischaemic regions in the myocardium. Demographic details of the patients are
127 summarised in Table 1. The study was carried out in accordance with the recommendations of the
128 South East Research Ethics Research Committee (Aylesford, Kent, UK) with written informed consent
129 from all subjects, in accordance with the Declaration of Helsinki.

130 The acquired ^{82}Rb PET perfusion images were processed in a custom-made MATLAB code to
131 obtain local myocardial flow (details can be found in Supplemental materials A) and also in a clinical
132 software platform (Syngo VB20A HF04) to obtain regional coronary flow reserve to be used as a
133 reference in the analysis.

134

135 *2.2 Image segmentation and meshing*

136

137 CTCA images were segmented using Simpleware ScanIP (Synopsys, CA, USA) to produce 3-D
138 anatomical models of the coronary arteries. Coronary branches were terminated at a diameter size
139 above 2mm due to the resolution limit: 0.488 mm pixel size and 0.625 mm slice thickness. Meshing
140 was performed also using ScanIP, using tetrahedral elements with 6 layers of prism elements along
141 the boundaries, with total element number in the order of 10^6 .

142

143 *2.3 Blood flow computation*

144

145 The blood flow in the anatomical models were computed by numerically solving the incompressible
146 3D Navier-Stokes equations using a commercial package ANSYS CFX 17.0 (ANSYS, Inc.
147 Cannonsburg, USA). The flow was assumed to be laminar and blood was modelled as homogenous
148 and Newtonian fluid with its density and dynamic viscosity 1060 kg/m^3 and 0.004 Pa s , respectively.

149 The vessel wall was approximated as rigid wall, where non-slip boundary conditions were applied,
 150 and cardiac-induced wall motion was not incorporated.

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Table 1: Patient data table.

Patient	Sex	Age	Stenosed branch	Stenosis severity	
				Clinical classification	Diameter reduction on CT
1	M	79	RCA	intermediate	45%
2	M	59	LAD	mild	31%
3	F	64	LAD	severe	76%
4	F	75	LAD	mild	19%
5	F	80	RCA	severe	73%
6	M	51	LAD	trivial to mild	11%
7	F	64	LAD	trivial to mild	9%
8	M	50	RCA	mild	14%
9	M	50	RCA	trivial to mild	4%
10	M	45	LAD	severe	55%
			LCx	intermediate	37%

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RCA: right coronary artery, LAD: left anterior descending (artery), LCx: left circumflex.

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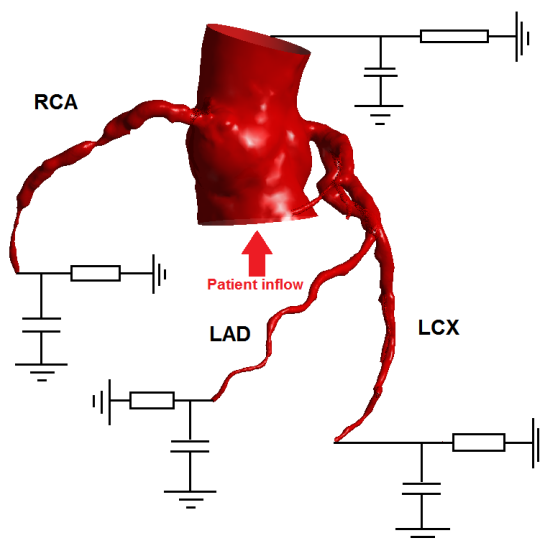
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157

2.4 Inflow boundary conditions

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The inflow into the aorta was set as a steady flow. While this is not representative of ordinary
 159 cardiac function, a detailed comparison between steady and pulsatile flow conditions have
 160 shown that steady flow condition is sufficient in CT-based FFR calculations.[18] Inflows were
 161 set patient specific, calculated based on the difference between segmented ventricular cavity
 162 volume at maximum contraction in systole and maximum dilatation in diastole, multiplied by
 163 the patient's heart rate.



164

165

Figure 1. Schematic showing the outflow boundary conditions. All the boundary conditions are two-element
 166 Windkessel models represented in the circuit diagram.

167

168

169 2.5 Outflow boundary conditions

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171 At each of the outflow boundaries, a two-element Windkessel model was connected to represent the
 172 downstream vasculature as shown in Figure 1. The Windkessel model is a 0D hydraulic-electric
 173 analogue where pressure difference, vascular resistance and compliance corresponds to potential
 174 difference, electrical resistance and capacitance.[19,20] The RC circuit is grounded to represent the
 175 near zero pressure conditions of the capillary bed. The model in practice provides pressure boundary
 176 conditions at each outlet in response to the outflow through the branch from the 3D CFD domain.
 177 Although the simulations were essentially steady state, we ran them as transient simulations with
 178 steady inflow, where compliance helps to stabilize the system and reach a steady state. The actual
 179 procedure to determine compliance values is explained in Section 2.5.1. There are many variations
 180 on the Windkessel model, such as the three-element, four-element and modified Windkessel, they
 181 are mostly used to simulate higher frequency phenomena in the cardiac cycle, which is irrelevant in
 182 a steady inflow simulation.[21]

183 In the baseline state, a reasonable assumption is to assign 5% of the total aortic output to the
 184 coronary arteries.[22] To accomplish this, the resistance at the aortic outlet was tuned such that the
 185 systemic outflow through the aorta corresponds to 95% of the stroke volume. In the hyperaemic state,
 186 however, the proportion of coronary flow to aortic output varies a lot more from patient to patient,
 187 and especially so in patients who suffer from some form of coronary artery disease. This was
 188 accounted for by adjusting peripheral resistance downstream to each branch, without control of the
 189 flow split between systemic and coronary circulations at hyperaemia. In conventional simulations in
 190 studies on coronary artery disease including CT-based FFR calculations, the assumption is made that
 191 the downstream resistance on the coronary branches is decreased to 30% of baseline[23], simulating
 192 the effect of adenosine inducing vasodilation.

193

194 2.5.1 Structured tree model for Windkessel outflow boundary conditions

195

196 To implement the conventional model of peripheral resistance, two parameters need to be
 197 calculated: the downstream resistance and vessel compliance. The resistances were determined using
 198 a structured tree model (Olufsen et al.[24]) to model a typical tree structure of small arteries and
 199 arterioles. This approach has been used effectively to supplement vessel tree structure beyond the
 200 resolution of CT (~0.5 mm) and has demonstrated to produce realistic resistance values for coronary
 201 flow simulations.[25] In practice, the branching structure of the vasculature and diameter of each
 202 segment (Figure 2) were defined using Murray's law[26] and empirical branch ratio 9:6 found in
 203 animal anatomy[24,25] following the equation below.

204

$$d_u^3 = d_{d1}^3 + d_{d2}^3$$

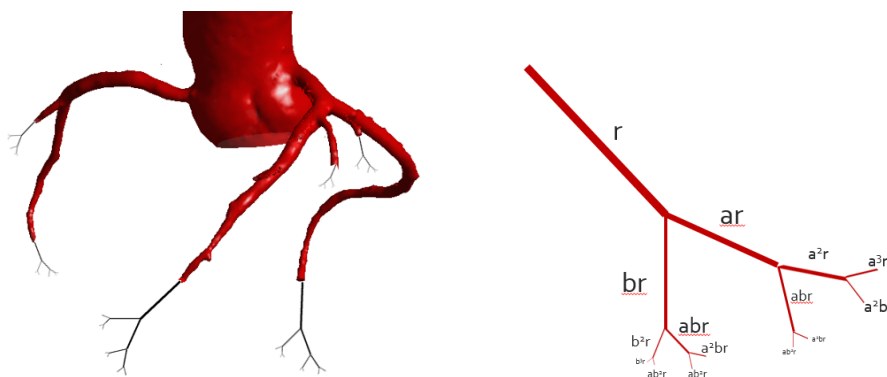
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$$d_{d1} = \frac{a}{b} d_{d2}$$

206 Here, d_u is the diameter of the upstream or parent vessel in a bifurcation, d_{d1} and d_{d2} are
 207 diameters of as the two downstream or daughter vessels. Parameters a and b are branch ratio. The
 208 branching is repeated in a fractal-like manner until the limiting dimensions of an arteriole is reached,
 209 typically 0.05 mm in diameter. Once resistance is determined, vessel compliance was calculated by
 210 setting the time constant ($=1/RC$) equal to 0.063 s following the literature[27], although the compliance
 211 is unlikely to have a drastic effect on the simulation as the system converges to a steady state.
 212 Diameter at the beginning (proximal end) of the tree is equal to the diameter of 3D model at the
 213 peripheral end. In our study, baseline (not hyperaemic) resistance order of magnitude is ~ 100000
 214 $\text{dynes} \cdot \text{s} / \text{cm}^2$

215

216



217

218 **Figure 2:** (Left) Illustration of downstream microvascular trees. (Right) Diagram of structured tree downstream
 219 microvascular structure. r represents the radius of the terminal vessel, and the daughter branches split
 220 asymmetrically in a repeated way, with fractions a and b , with values of 0.9 and 0.6 respectively.[24] This
 221 branching occurs indefinitely until it reaches the minimum radius.

222 2.5.2 Vasodilatory response model with structured tree

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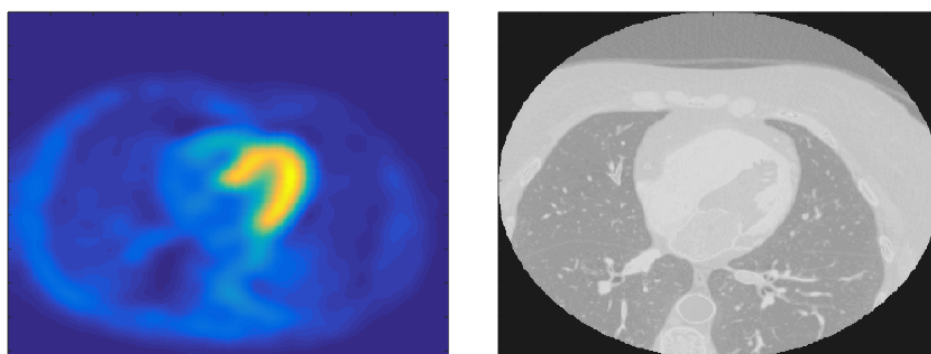
224 To test whether it is appropriate to set hyperaemic resistance to be 30% of baseline resistance and
 225 how sensitive FFR is to this, we performed simulations in which the downstream resistance of the
 226 diseased branch is adjusted to various reduced level (30%, 50%; 70%, 90%) of its baseline value
 227 calculated from the structural tree model. The range reflects the vast majority of patient disease cases,
 228 where microvascular vasodilatory response varies from healthy ideal to ineffectual (virtually no
 229 vasodilation). These conditions are referred as Morphological based Boundary Conditions (MBC)
 230 with their reduced resistance level, e.g. MBC 30%, MBC 50%, etc., later in the manuscript.

231

232 2.5.3 PET-based myocardial perfusion outflow boundary conditions

233

234 Because the CT and PET scans were not taken simultaneously though sequentially, co-registration is
 235 needed for the PET images to align with the correct myocardial region in the CT image. The PET
 236 images were first oversampled to produce the same pixel sizes as the CT images. Typically, the spatial
 237 resolution of CT was 0.5 mm/pixel (slice thickness 0.6mm) and that of PET image was 3 mm/pixel.
 238 The images were then aligned using a 5-point iterative closest point algorithm in Matlab, where 5
 239 anatomical landmarks, such as the apex and basal end of the interventricular septum, are identified
 240 manually in both image sets and referred in the alignment. Co-registration uncertainty was calculated
 241 through 3 repeated attempts, which was found to be 1.74 ± 0.40 mm within the transversal image plane
 242 and 2.71 ± 0.55 mm out-of-plane (i.e. in the axis normal to the imaging plane).



243

244 Figure 3: Typical PET image (left) and CT image (right) showing the long-axis slice of the heart. The colours in
 245 the PET image represents the spatial map of radiation intensity, corresponding to the myocardial perfusion.
 246 Here, the area of yellow-green indicates strong signal, depicting myocardium. The grayscale in the CT image
 247 corresponds to Hounsfield unit, the bright areas indicate regions of higher radiodensity.

248

249 A PET-perfusion-based boundary condition was developed to contrast the structured tree
250 method which prescribes downstream resistances only based on the size of the of the branch terminus
251 in the 3D model. The perfusion-based boundary condition (PBC) refers to the local perfusion
252 quantitatively, based on the PET image intensity (representing perfusion in ml/100ml/min [16]), in
253 the region supplied by each branch. Local perfusion was quantified by placing a sampling sphere (20
254 mm diameter) at the end of each branch in 3D model, and the special average of perfusion was
255 calculated, excluding image pixels with its value lower than 10 ml/100ml/min in order to eliminate
256 the space outside the myocardium in the sphere. The peripheral resistances downstream to each of
257 the branches are then determined such that the flow split through each branch corresponds to the
258 split in the PET-based measurement. Thus, in this approach, the resistance does not depend on the
259 terminal branch size. Here, as in the MBC, the coronary outflow is assumed to be 5% of the total aortic
260 output. A more detailed description of this process is presented in Supplemental material A.

261 Each patient has two sets of PET perfusion images, taken during the rest state (baseline) and
262 during the hyperaemic state. The patient-specific and spatially local hyperaemic response can be
263 calculated and implemented in the model, as a reduction of resistance so as to replicate the increased
264 flow at hyperaemia in each branch. As the result, the total hyperaemic coronary outflow as a
265 proportion of cardiac output is unbound and reflect the wide spectrum between patients.

266

267 *2.6 Computational schemes*

268

269 The governing equations are discretized in space using element-based finite volume method,
270 where volume and surface integrations are performed at the Gaussian integration points on each
271 element/face using tri-linear shape function interpolating nodal values of velocity and pressure in 3D
272 within each element. The time integration was performed using 2nd order backward Euler scheme.
273 Stabilization of the advection term is achieved by adaptive 2nd order upwinding scheme in which 1st
274 order upwinding is blended with the 2nd order scheme in reference to the local flow velocity.

275 The simulations were carried out in quasi-steady condition, i.e. transient simulations were
276 conducted with steady inflow boundary condition. This was required to account for the transient
277 response of the downstream impedance. Here, the time step and convergence criteria were set to was
278 set to 0.001 s and 1.0×10^{-5} , respectively. Sensitivity tests of the computational results to both mesh
279 and time step size were carried out such that the pressure drop across a stenosis computed with the
280 finally-chosen mesh and time step is less than 1% of difference compared to a mesh with doubled
281 number of elements. Computations were conducted using 2 cores on standard desktop workstations
282 (Intel Core i7 6700K 4GHz, 16GB RAM, 4 cores and Intel Xeon E5-2670 2.6GHz, 128GB RAM, 32
283 cores).

284

285 *2.7 Calculation of FFR*

286

287 Monitor points were placed at the coronary ostium and in the coronary artery at a point
288 approximately 4 cm distal to the stenosis. FFR standards in invasive measurements specify at least
289 2-3 cm distally, 4 cm was chosen to be consistent and to ensure the minimum possible FFR (i.e. largest
290 pressure drop in the vessel) is captured.[28] When the simulation has converged, the pressure distal
291 to the stenosis divided by the pressure at the coronary ostium produces the final CT-based FFR value.

292

293 **3. Results**

294 Typical examples of the computational results, in terms of pressure distributions along the coronary
295 vessel tree, are shown in Figure 4. Wide spectrum of anatomical variations and some different levels
296 of pressure drop across the tree can be observed. The pressure drop from the aorta to the end of
297 branches is in general larger for the models with PBC.

298 A quantitative comparison of FFR values across the stenosis is presented in Table 1, including
299 those with all the peripheral resistance variations of MBC. FFR range is from 0.64 (Patient 5 with MBC
300 30%) to 0.99 (Patients 6 and 7 with PBC). In reference to the cut-off value of CT-based FFR (0.80 [29]),
301 the range of FFR in this study reflects the wide range of disease state included in the study.

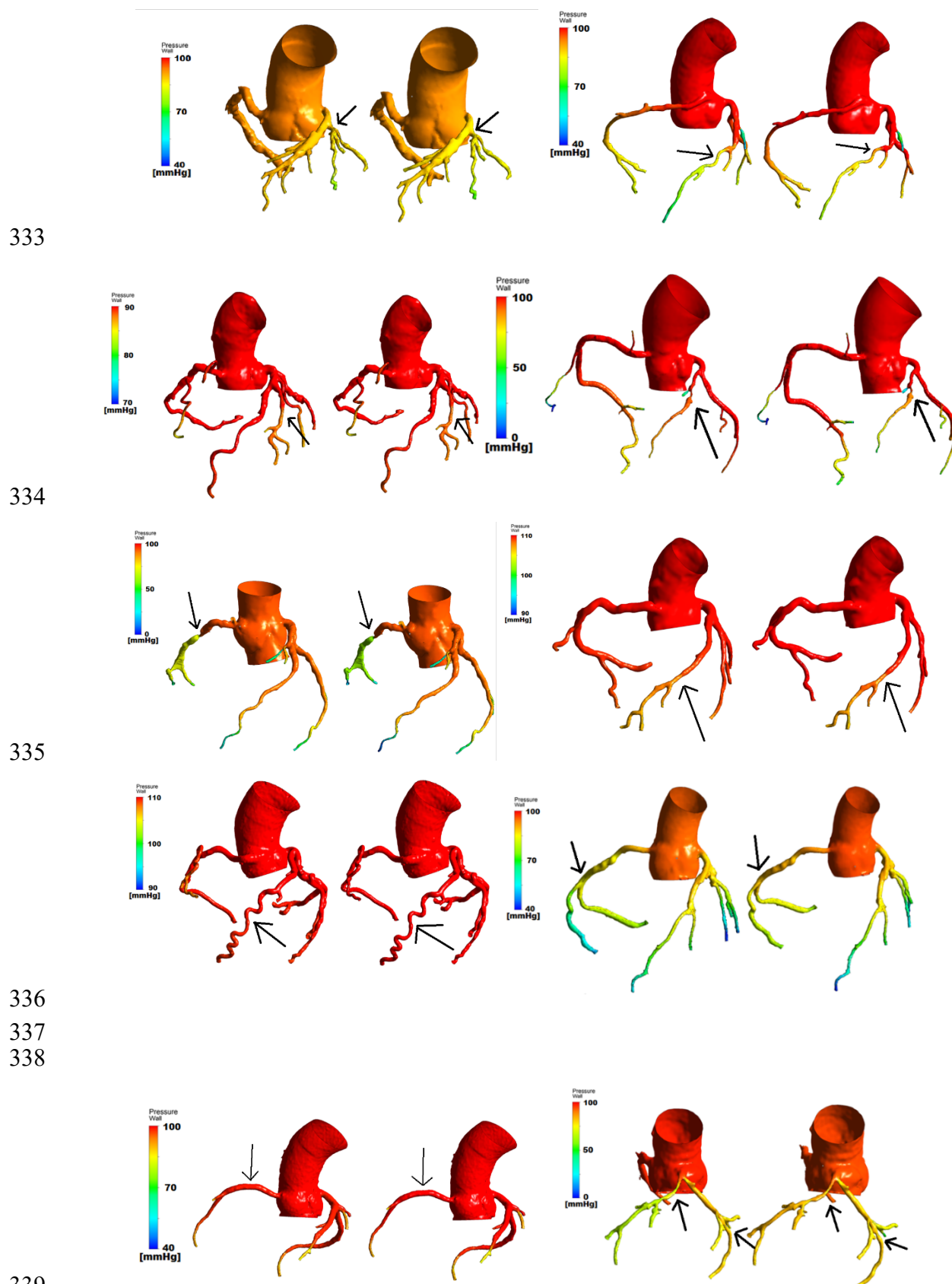
302 The FFR with PBC and 'the standard (i.e. 30%)' MBC are in general correlated well ($r = 0.68$).
303 The correlation is higher for the patients with high FFR values ($FFR \gg 0.80$), i.e. patients with
304 relatively minor or insignificant stenosis. The PBC tends to result in FFRs that are higher than the
305 ones with the conventional boundary condition (MBC 30%). However, this is not true for Patients 3,
306 9 and 10. Patient 3 is the most extreme case where the FFR with PBC (0.76) indicates a different
307 diagnostic result than the conventional MBC 30% ($FFR=0.81$), straddling across the standard cut-off
308 value of 0.80.[29]

309 The FFR values obtained with the variable peripheral resistances in MBC show a clear trend of
310 high FFR for high resistance (i.e. smaller degree of resistance reduction – close to the baseline)
311 consistently across the patients. This was expected, based on a principle of fluid mechanics; low
312 peripheral resistance invites higher flow to the branch which results in a larger pressure drop thus
313 smaller FFR.

314 The flow rates through different branches are summarised in Table 3, both in absolute value and
315 proportion to the total coronary flow. A comparison between Tables 2 and 3 indicates that the flow
316 rate is a strong determinant of FFR, with higher flow rates corresponding to lower FFR (more severe
317 stenosis). Considering Poiseuille's law, the flow rate increase should linearly be related to the
318 pressure drop ($1 - FFR$ gives the pressure drop as a proportion of proximal pressure), which is also
319 true in these vessels. Additionally, the MBCs with various level of peripheral resistance demonstrate
320 a strong association between the flow rate and the percentage of coronary flow that is distributed to
321 the stenotic branch. In general, the flow through the stenosed branch is lower with PBC than with
322 MBC but for Patients 3, 9 and 10 (LCx), where the PBC-based FFR was lower than that of MBC 30%,
323 the flow with PBC is indeed higher.

324 Figure 5 is a graphical representation of Table 2, showing FFRs obtained with PBC and MBC
325 with the range of reduced peripheral resistance. Here, the range of FFR with the variable MBC is
326 examined in a different way to illustrate more fundamental principle underpinning their
327 relationship. The patients are reordered in reference to FFR with MBC 30% so that the trend is clearly
328 visible. For patients with an FFR close to or below the cutoff of 0.8, the divergence in FFR values
329 between PBC and MBC 30% is more significant. The range of FFR for the various peripheral resistance
330 is also larger for the lower overall FFR. On the other hand, the range for patients having high overall
331 FFR is small, only 0.01 for Patients 4, 7 and 9.

332



340 **Figure 4:** Comparison of pressure profiles obtained from PET-based boundary conditions (left of
 341 pairs) versus conventional morphology-based boundary conditions (right of pairs). Patient 1-10
 342 from top left to the bottom right. Arrows indicate focal stenosis.

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Table 2. CT-based FFR values obtained using various outflow boundary conditions. PBC: PET-based boundary condition, MBC: morphology-based boundary condition. The percentage values indicate reduced resistance level to account for hyperaemic flow increase and reduction to 30% (MBC 30%) is the conventional assumption.

Patient	PBC	MBC (30%) "Conventional"	MBC (50%)	MBC (70%)	MBC (90%)
1	0.91	0.89	0.91	0.92	0.92
2	0.90	0.83	0.88	0.92	0.93
3	0.76	0.81	0.83	0.84	0.85
4	0.97	0.96	0.96	0.97	0.97
5	0.71	0.64	0.74	0.75	0.75
6	0.99	0.96	0.96	0.98	0.99
7	0.99	0.97	0.98	0.98	0.98
8	0.85	0.82	0.85	0.86	0.88
9	0.96	0.98	0.98	0.98	0.99
10 (LAD)	0.73	0.56	0.68	0.74	0.78
10 (LCx)	0.74	0.77	0.85	0.88	0.88

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Table 3. Flow rates through stenosis and proportion of that to the overall coronary flow (%), obtained using various outflow boundary conditions.

Patient	PBC, [ml/s]	MBC (30%), [ml/s]	MBC (50%), [ml/s]	MBC (70%), [ml/s]	MBC (90%), [ml/s]
1	3.98 (48%)	4.69 (56%)	3.78 (45%)	3.39 (41%)	3.26 (39%)
2	3.41 (40%)	5.54 (64%)	4.59 (53%)	2.72 (32%)	2.50 (29%)
3	2.78 (29%)	2.23 (28%)	2.07 (26%)	1.82 (23%)	1.74 (22%)
4	2.61 (32%)	3.45 (38%)	2.87 (32%)	2.57 (29%)	2.42 (27%)
5	1.39 (20%)	1.72 (22%)	1.28 (16%)	1.26 (16%)	1.20 (15%)
6	2.19 (33%)	5.18 (68%)	4.37 (57%)	2.79 (37%)	1.45 (19%)
7	1.34 (17%)	2.86 (22%)	2.35 (18%)	2.11 (16%)	1.98 (15%)
8	1.69 (16%)	2.14 (21%)	1.71 (17%)	1.52 (15%)	1.32 (13%)
9	5.01 (55%)	4.50 (53%)	4.12 (48%)	3.90 (46%)	2.56 (30%)
10 (LAD)	2.24 (21%)	3.83 (34%)	2.82 (25%)	2.17 (19%)	1.91 (17%)
10 (LCx)	1.22 (11%)	1.09 (9.5%)	0.71 (6.2%)	0.58 (5.1%)	0.56(4.9%)

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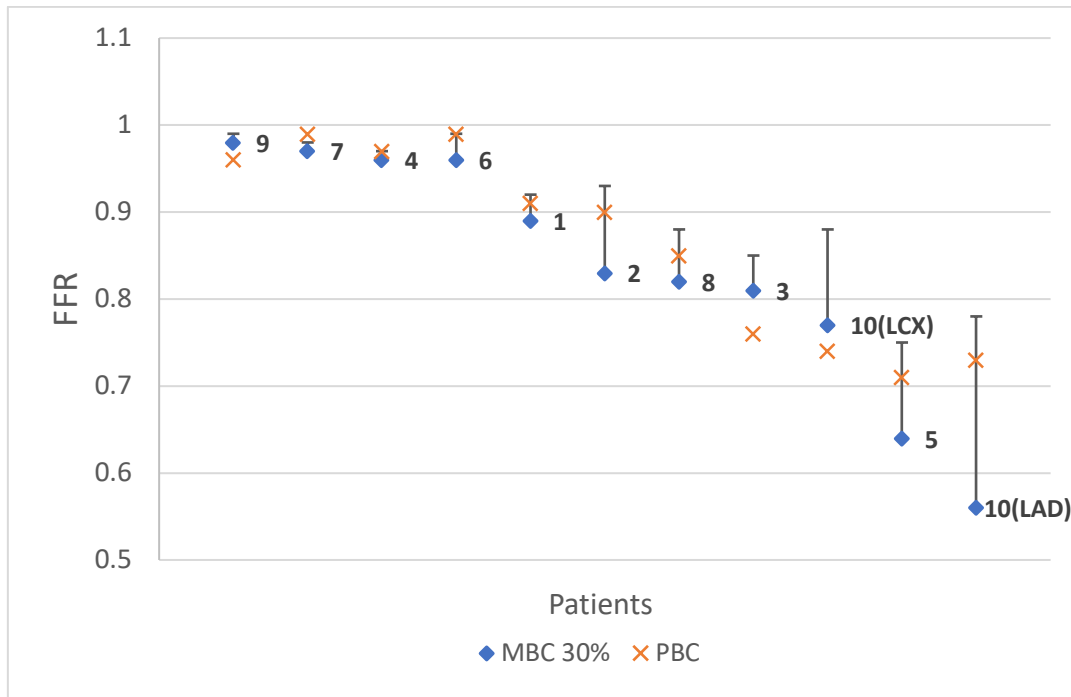
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Figure 5: The FFRs of patients, reordered in reference to the value of FFR calculated with MBC 30%. The bars indicate the range of FFRs obtained using the various MBCs, with the conventional (MBC 30%) marked as a blue diamond, and the PBC marked as orange cross. Patient numbers are shown on the plot as reference.

371 4 4. Discussion

372 Although coronary artery flow computations have relatively long history and boundary conditions
373 have always been a point of discussion, to the authors' knowledge, there was no study utilising
374 myocardial perfusion as patient-specific outflow conditions. The perfusion data was used to examine
375 the impact of resistance reduction during hyperaemia, and an attempt was made to characterise an
376 uncertainty range of FFR due to non-ideal response to vasodilator by the vasculature peripheral to
377 the stenosis. Uncertainty of FFR computation has been studied in terms of the sensitivity of FFR to
378 imaging and segmentation uncertainty[29], but not the hyperaemic response to adenosine.

379 The results show a high correlation of FFR values computed using the two types of outflow
380 boundary conditions. This is not surprising for the patients with relatively minor stenosis because
381 there is no significant pressure drop across the stenosis anyways. Therefore, even with different flow
382 distributions across the branches of the coronary arteries obtained via the varied outflow boundary
383 conditions, the pressure drop across the stenosis for those patients was not significantly altered (e.g.
384 Patient 9, FFR range: 0.98-0.99 for various MBCs).

385 On the other hand, the hyperaemic condition is shown to have a strong influence on the FFR
386 calculations for more severe stenosis such as Patients 3, 5 and 10. The pressure drop ΔP across a flow
387 limiting pipe is related to its flow rate Q and resistance R (in this case, the stenosis): $\Delta P \approx Q \times R$, using
388 Poiseuille's law. An increase in flow rate for a given stenosis increases the pressure drop, and
389 Similarly, a more severe stenosis (i.e. increased resistance) increases the pressure drop, therefore
390 lowering FFR. This is a simple fluid mechanical principle behind the differences caused in FFR.
391 The same principle also implies that a reduced flow rate in a stenosis would result in a higher FFR,
392 i.e. indication for a less functional severity. A reduced flow can occur if there is any disease in the
393 peripheral vascular bed (microvascular disease), which could elevate FFR. From diagnostic point of
394 view, this may appear to be false negative scenario, i.e. diseased patient seen as healthy. However,

395 the purpose of FFR is to detect a focal flow-limiting stenosis, and a relatively high FFR indicating the
396 stenosis being no flow-limiting in such a scenario still provides a valid indication.

397 The simulations using the PET perfusion-based boundary conditions take into account the
398 patient-specific distribution of the flow across the different branches and the change in flow rate from
399 normal physiological state to hyperaemia. The FFR obtained using the PBCs are in general higher
400 than the FFRs with conventional boundary conditions. This is because the hyperaemic response in
401 reality is generally less than 4 with a wide range standard deviation of 0.9)[16] and the PET perfusion
402 boundary conditions reflect that. Clinically-evaluated CFRs of our patients' diseased vessels are
403 2.12 ± 1.28 , indicating that the conventional assumption, i.e. $CFR=4$, is indeed an overestimate.
404 However, in Patients 3 and 10, the FFR values obtained using PBC are lower than those using MBC.
405 Even though the conventional boundary conditions make the assumption of the ideal vasodilatory
406 condition ($4x$ the baseline flow), the baseline flow for the diseased branch may be higher in the model
407 with PBC than with MBC, and therefore even with a reduced vasodilatory condition with PBC ($<4x$),
408 the absolute hyperaemic flow across the diseased branch can be higher than that of the conventional
409 model as shown in Table 3, causing a larger pressure drop and hence lower FFR. This is indeed true
410 for those 2 patients, suggesting that the myocardium downstream to those vessels are still healthy
411 despite the stenosis in its upstream, making these cases as illustrative examples of 'flow-limiting'
412 stenosis that should ideally be detected by FFR. The flow through the diseased branch of those
413 patients are underestimated with the conventional MBC, thus potentially causing false negative –
414 indeed, the FFR of MBCs are higher than the cutoff 0.8 and that with PBC is lower. To make the
415 analysis framework more accurate, this group of patients need to be looked at for further
416 characterisation.

417 The PBC-based FFRs generally fall within the range of FFR calculated using the varied MBC
418 (30%-90%). The patients where the PET perfusion model predicts an FFR that is outside the bounds
419 all have a higher flow through the stenosed branch compared to the computations with MBC. The
420 method of ranging the hyperaemic response to obtain a lower and upper bound of FFR could
421 therefore be useful approach to have a 'confidence interval' of FFR calculation. From clinical
422 diagnostic point of view, it is particularly concerning when the range of FFR for a particular patient's
423 stenosis straddles the cutoff value 0.9. The largest variance in FFR can be found in Patient 10,
424 specifically with the LAD (0.56-0.78), however the bounds do not cross over the cut off and therefore
425 it is unlikely to affect clinical decisions. For the LCx of Patient 10, while the variance is not as high,
426 the range is across the clinical cutoff of 0.8 (0.74 – 0.88).

427 It should be noted that PET perfusion or other similar myocardial perfusion imaging generally
428 supplants FFR measurement obtained both invasively or non-invasively.[31] It is rare in the clinic
429 that both of those are performed before a diagnostic judgement is made, due to the cost and time it
430 requires. The comparison in this study is not to suggest that a perfusion-based CT-FFR model
431 should be adopted as clinical practice, but rather verifying the flow condition estimation in the
432 downstream of the coronary arteries using purely morphological methods. That being said, there
433 have been clinical studies that implement myocardial perfusion imaging in cases where CT-FFR
434 provided an ambiguous indication, and has shown a noticeable improvement in diagnostic
435 accuracy.[32]

436 Among patients having stable angina, 65% of women and 32% of men have no obstructive CAD
437 (stenosis $<50\%$).[33] A significant portion of those patients suffer from coronary microvascular
438 dysfunction which can be ischaemia without a focal stenosis[33]. This means that a significant
439 number of patients would not be best served by current CT-based FFR techniques that make the
440 assumption of ideal downstream microvascular health. Because the assumption uses the maximal
441 possible vasodilation, for patients whose vasodilatory response is impaired, the simulation would
442 overestimate the flow passing through a stenosis, producing a lower FFR than the true value,
443 potentially leading to false positive diagnosis and hence an unnecessary invasive revascularization.

444 Although the FAME trials – the original and main clinical trial for FFR – suggest a strict 0.8
445 cutoff, research has since suggested that clinicians should be cognizant of the biological variability of
446 FFR measurements, where repeated measurements will produce different values and possibly

447 different diagnostic outcomes. [34] It has been suggested that there should be a gray zone around
448 the cutoff, considered to be between 0.75 and 0.85, with particular caution given to values between
449 0.77 and 0.83, where the clinician will need to consider other patient metrics before deciding
450 treatment.[34]. This study has examined one major consideration that can have a drastic effect on CT-
451 based FFR: the vasodilatory capacity of the patient's microvessels. The PET perfusion-based model
452 has shown that the majority of patient's microvascular health is between ideal and diseased, based
453 on their CFR. In the clinic, additional assessment of microvascular health through testing or
454 identifying risk factors such as diabetes, age, sex could be used to inform CT-based FFR
455 measurements directly, producing a possibly more reliable FFR value, and if that isn't available, a
456 similar approach to this study where a band of FFR values can be produced to identify the likelihood
457 that a stenosis falls below the cutoff.
458

459 5. Limitations

460 The main limitation of this study is that the CT-based FFR values for the patients were not
461 validated against invasive FFR, the gold standard for FFR. This preliminary study was designed as
462 a sensitivity test, examining the variability of FFR in simulation-based CT-FFR with various outflow
463 boundary conditions. Additionally, diffuse disease, where the narrowing isn't focal but spread
464 along an artery, is also a known cause of ischaemia, and the pressure drops can be as severe as those
465 of focal stenoses, however revascularization of diffuse disease has shown mixed results and therefore
466 FFR is not applicable[35]. Model assumptions, not only with boundary conditions, are inevitable in
467 computational analysis. We chose to simulate the flows with steady flow condition using Newtonian
468 approximation of the blood. As discussed in method section, these are not deemed significant in FFR
469 computation but would carry more importance in analyses of stented segment with potentially larger
470 flow recirculation, and/or of patients with cardiac rhythm disorders. Lastly, the finding from our
471 study still need to be confirmed with a larger number of patients, which is planned for future.
472

473 6. Conclusions

474 In this study, a series of computational FFR analysis was conducted using various outflow boundary
475 conditions to investigate their impact on the FFR derivation. The FFRs computed with a conventional
476 morphology-based and the novel PET-based outflow boundary conditions agreed in general.
477 However, the models with PET-based condition revealed that there are cases in which conventional
478 boundary condition overestimate the functional severity of a stenosis, potentially placing the patient
479 in different diagnostic category. The derivation of a potential range of FFR a patient might have, by
480 varying peripheral resistance over a physiologically possible range, indeed indicated that the
481 overestimation of vasodilatory response is likely reason behind the overestimation of functional
482 severity. These results indicate that, although perfusion data such as PET images are not always
483 available in clinics, a better estimation of outflow boundary condition reflecting the physiological
484 state of downstream coronary vasculature could improve the CT-based estimation of FFR.
485

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494 **Supplemental material A**

495

496 The PET perfusion measurement works as follows:

497 1. Generate a sphere of 20 mm diameter around the terminal end of a coronary artery branch (this has to be
 498 located using the CT image during segmentation) in the 3D model. The sphere in general overlaps
 499 significantly with the myocardial region supplied by the chosen coronary branch.

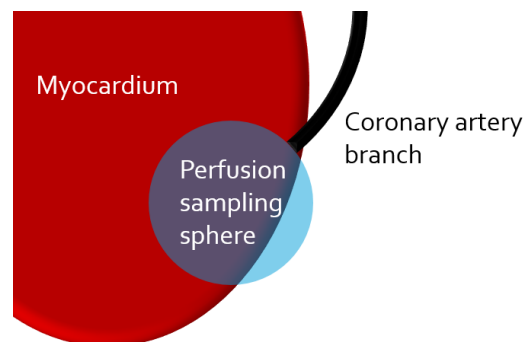
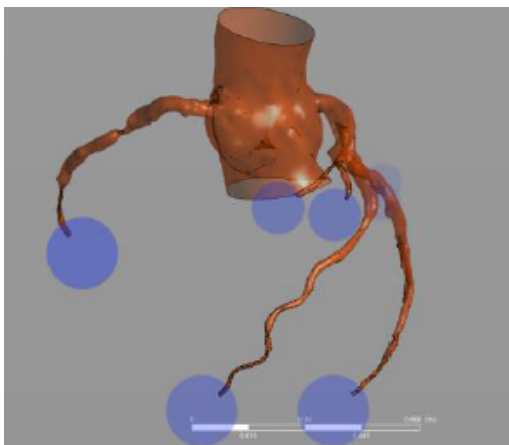
500 2. Sample the sphere for perfusion values above 10 ml/100ml/min (a normal value of perfusion is
 501 between 50 – 300ml/100ml/min[16]), effectively ruling out the spaces captured in the sphere where it
 502 is outside the myocardium.

503 3. Calculate the spatial mean perfusion of the myocardial region within the sphere, this value is used to
 504 represent the flow capacity of a coronary branch.

505 4. In the baseline (resting state) simulation, the 5% of aortic output that is allocated to the coronaries is
 506 divided by the various branches via their flow capacities, that are implemented in the form of
 507 downstream microvascular resistance.

508 5. In the hyperaemic (stressed state) simulation, the magnitude decrease in resistance of each branch is
 509 derived from the change in perfusion that is observed between the rest and stressed states.

510 The 20mm sphere size was chosen to be large enough to mitigate the problems that may arise from the
 511 registration error of 2-3mm, mis-identification of the location of the terminal end (usually due to CT
 512 resolution terminating branch far too proximal), and small enough that the regional perfusion supplied by
 513 different branches do not overlap significantly.



514

515

516 **Figure A1:** Perfusion sampling spheres at the downstream terminus of coronary artery branches (left). The
 517 overlapping volume between sampling sphere and myocardium is used to quantify the local perfusion (right).

518

519 **Supplemental material B**

520 Patient 10 has two significant stenosis on two separate branches, the LAD and LCx. In order to
 521 examine potential interaction of the flow in the two branches under the varied peripheral resistances,
 522 we varied the hyperaemic condition first in each of the LAD and LCx individually, and then both
 523 LAD/LCx simultaneously. The result is summarised in Table B1, it is observed that though the FFR
 524 values one branch are affected by the varying outflow condition of other branches, the effect is minor.

525 Therefore, the effect of varying peripheral resistance in two stenosed branches can practically be seen
 526 as independent.
 527

528 **Table B1.** CT-based FFR values for Patient 10 when each diseased vessel's hyperaemic
 529 condition were varied individually and in combination.

530

Branch with varied BC	MBC 30%	MBC 50%	MBC 70%	MBC 90%
LAD	(LAD) 0.56 (LCx) 0.77	(LAD) 0.68 (LCx) 0.77	(LAD) 0.74 (LCx) 0.76	(LAD) 0.78 (LCx) 0.76
LCx	(LAD) 0.56 (LCx) 0.77	(LAD) 0.56 (LCx) 0.85	(LAD) 0.56 (LCx) 0.88	(LAD) 0.55 (LCx) 0.88
LAD+LCx	(LAD) 0.56 (LCx) 0.77	(LAD) 0.65 (LCx) 0.86	(LAD) 0.73 (LCx) 0.89	(LAD) 0.76 (LCx) 0.90

531

532

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