

Cerebrospinal fluid folate, ascorbate, and tetrahydrobiopterin deficiency in superficial siderosis: a new potential mechanism of neurological dysfunction?

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Dear Editor,

Superficial siderosis of the central nervous system (CNS) is a disabling but rare disease caused by chronic bleeding in the subarachnoid space of the brain and spinal cord [1,2,3]. This chronic slow leakage of red blood cells into the CSF, causes an accumulation of haem. In response, microglia and Bergmann glial cells release hemoxygenase-12 [**Fig 1, Panel A (i)**] which breaks down haem into free iron (ferrous iron, Fe^{2+}) and biliverdin. Ferritin binds the free iron to form hemosiderin. Deposits of hemosiderin can be detected by MRI, allowing in vivo diagnosis of superficial siderosis [4]. After prolonged low level bleeding (sometimes over decades) it is presumed that the homeostatic system of the glial cells is overwhelmed, leading to the release of potentially neurotoxic free iron species into the CSF. Free iron is a potent oxidant and can produce free radicals which damage cellular components including cofactors and vitamins, e.g. tetrahydrobiopterin (BH_4) and 5-methyltetrahydrofolate (5MTHF) [5,6].

Case report

In view of their lability, we assessed CSF BH_4 plus its oxidised catabolite, dihydrobiopterin (BH_2) and 5MTHF in a 34 year old female patient who had presented with progressive balance and hearing impairment, and a myelopathy. MRI showed low signal on blood-sensitive sequences over the surface of the brainstem, cerebellum, craniocervical junction and spinal cord [**Fig 1, Panel B**], leading to a diagnosis of superficial siderosis of the CNS. Because of the key cofactor role BH_4 plays in monoamine metabolism, we quantified dopamine and serotonin metabolites (homovanillic acid (HVA) and 5-hydroxyindoleacetic acid (5HIAA)). Additionally we determined ascorbate status because it is a key antioxidant and a cofactor for the downstream metabolism of dopamine to noradrenaline.

CSF ferritin was **126** ng/ml (reference range <16 ng/ml), consistent with active or recent haemorrhage into the subarachnoid space. BH₄ (<**4** nmol/L (reference range 9-39 nmol/L), ascorbate (**26** nmol/L (reference range 118-246 nmol/L) and 5MTHF (<**10** nmol/L (reference range 46-160 nmol/L) concentrations were all markedly below their respective reference ranges. BH₂, an oxidation product of BH₄ catabolism, was elevated (**15.9** nmol/L (reference range 0.4-13.9 nmol/L).

Dopamine turnover and/or availability appeared increased (HVA **566** nmol/L (reference range 71-565 nmol/L), which was further supported by the elevated HVA to 5HIAA ratio (**4.4** (reference range 1-3.7)). In contrast, serotonin metabolism was not affected (5HIAA 130 nmol/L (reference range 58-220 nmol/L).

Previous studies have described CSF pterin and monoamine analysis in other neurological diseases, including Parkinson's disease [5], cerebrovascular diseases and subarachnoid haemorrhage [6]. These studies have described a deficit of tetrahydrobiopterin/total biopterin in both Parkinson's disease and subarachnoid haemorrhage [6] but have not analysed CSF ascorbate or folate. To the best of our knowledge, this is the first report of the CSF status of BH₄, 5MTHF, HVA, 5-HIAA and ascorbate in superficial siderosis of the CNS. Our findings are consistent with oxidative stress, due to free iron species, and raise the possibility that this is a potential mechanism of neurological disability in superficial siderosis. Both BH₄ and 5MTHF are highly susceptible to oxidative catabolism *in vitro* [**Fig 1, Panel A (ii)**][7,8]. A reduced level of BH₄, via oxidative breakdown, is supported by the increased concentration of one of its oxidation products, BH₂. Our finding of a very low CSF ascorbate provides further evidence for oxidative stress, as this antioxidant may become depleted by increased generation of reactive oxygen species [8]. The markedly low 5MTHF CSF levels reported here might also be expected to impair metabolic pathways that are dependent upon folate cofactors including the one-carbon metabolic pathways, responsible for the methylation of DNA, RNA

and proteins. Further work is therefore required to ascertain the potential biochemical effects of this deficiency in superficial siderosis.

Both ascorbate and 5MTHF are vitamins and must be constantly replaced by dietary intake.

In contrast, BH₄ levels can be maintained within the cell by *de novo* synthesis or recycling.

Although CSF BH₄ levels were low in this patient, the CSF profile reported here does not

suggest a deficiency of BH₄ within dopaminergic or serotonergic cells as HVA and the

HVA:5-HIAA ratio were elevated. The synthesis of noradrenaline [**Fig 1, Panel A (iii)**] from

dopamine requires the enzyme dopamine-β-hydroxylase (DβH), for which ascorbate is an

essential cofactor. Decreased ascorbate due to iron-mediated oxidative stress exceeding

dietary replenishment, may thus lead to reduced dopamine-β-hydroxylase activity, leading to

an accumulation of the substrate dopamine, and increased turnover to HVA. It should be

noted that, although serotonin and its metabolites are generally considered to be relatively

labile in aqueous solutions and in some biological matrices, it has been shown that 5HIAA is

stable in 'control' CSF for up to 48 hours at room temperature [9]. This may explain why

serotonin turnover is not affected in this patient. However, it is possible that excessive iron

accumulation could eventually result in degradation of other labile species in the CNS, such

as serotonin and its metabolites.

The CSF findings of this patient with superficial siderosis support a role for oxidative stress

in this disease. This is a plausible pathogenic mechanism of neurological disability that

merits further exploration. In addition, central 5MTHF and ascorbate status might be

potentially treatable with supplementation, although caution is required since folate

supplementation can aggravate B12 deficiency and ascorbate might act as a pro-oxidant as

well as an antioxidant [10]. Whether this treatment strategy can help reduce progression of

this disabling disease will require controlled clinical trials.

Appendix 1

Authorship

Jack Belsten – responsible for biochemical analysis, interpretation of data and drafted and revised manuscript for intellectual content.

David Werring – responsible for acquisition and interpretation of clinical data and drafted and revised manuscript for intellectual content.

Howell Jones – responsible for acquisition and interpretation of clinical data and revised the manuscript for intellectual content.

Simon Heales – responsible for study design, interpretation of data and drafted and revised manuscript for intellectual content.

Simon Pope - responsible for biochemical analysis, interpretation of data and drafted and revised manuscript for intellectual content.

References

1. Fearnley JM, Stevens JM, Rudge P. Superficial siderosis of the central nervous system. *Brain* 1995; 118(pt 4):1051–1066.
2. Wilson D, Chatterjee F, Farmer SF, Rudge P, McCarron MO, Cowley P, Werring DJ. Infratentorial Superficial Siderosis: Classification, Diagnostic Criteria, and Rational Investigation Pathway. *Ann Neurol*. 2017; Mar;81(3):333-343. doi: 10.1002/ana.24850. Epub 2017 Jan 28.
3. Leussink V, Flachenecker P, Brechtelsbauer D, Bendszus M, Sliwka U, Gold R, Becker G. Superficial siderosis of the central nervous system: Pathogenetic heterogeneity and therapeutic approaches. *Acta Neurologica Scandinavica*, 2003; 107(1), 54-61.
4. Kumar N. Neuroimaging in Superficial Siderosis: An In-Depth Look. *American Journal Of Neuroradiology* 2010; 31(1), 5-14.
5. Fujishiro K, Hagihara M, Takahashi A, Nagatsu T. Concentrations of neopterin and biopterin in the cerebrospinal fluid of patients with Parkinson's disease. *Biochem Med Metab Biol*. 1990 Oct;44(2):97-100.
6. Candito M, Nagatsu T, Chambon P, Chatel M. High-performance liquid chromatographic measurement of cerebrospinal fluid tetrahydrobiopterin, neopterin, homovanillic acid and 5-hydroxyindoleacetic acid in neurological diseases. *J Chromatogr B Biomed Appl*. 1994 Jul 1;657(1):61-6.

7. Heales SJ, Blair JA, Meinschad C, Ziegler I. Inhibition of monocyte luminol-dependent chemiluminescence by tetrahydrobiopterin, and the free radical oxidation of tetrahydrobiopterin, dihydrobiopterin and dihydroneopterin. *Cell Biochem Funct.* 1988; Jul;6(3):191-5.
8. Aylett S, Neergheen V, Hargreaves IP, Eaton S, Land JM, Rahman S, Heales SJ. Levels of 5-methyltetrahydrofolate and ascorbic acid in cerebrospinal fluid are correlated: Implications for the accelerated degradation of folate by reactive oxygen species. *Neurochemistry International*, 2013; 63(8), 750-755.
9. [Langlais](#) PJ, Bird ED, McEntee WJ. Stability of monoamine metabolites in human cerebrospinal fluid. *Ann Neurol.* 1982 Jul;12(1):48-51.
10. Duarte TL, Lunec J. Review: When is an antioxidant not an antioxidant? A review of novel actions and reactions of vitamin C. *Free Radic Res.* 2005; Jul;39(7):671-86.

Figure legend

Figure 1.

Panel A

(i) The production of hemosiderin and free iron from a dural leak in the central nervous system (CNS).

(ii) The iron based Fenton chemistry for the production of the hydroxyl radical from hydrogen peroxide.

(iii) The metabolism of serotonin, dopamine and noradrenaline. BH_4 is a cofactor for the synthesis of dopamine and serotonin. Ascorbate is a cofactor for DBH in the conversion of dopamine to noradrenaline. Dopamine and serotonin are metabolised to HVA and 5HIAA, respectively. HVA and 5HIAA can be used as markers of dopamine and serotonin turnover.

Panel B

Axial T2-weighted MRI showing low signal consistent with haemosiderin deposition around the superior cerebellar vermis and brainstem (white arrows).

BH_2 : dihydrobiopterin, BH_4 : tetrahydrobiopterin, 5HIAA: 5-hydroxyindoloacetic acid, HVA: homovanillic acid, DBH: dopamine- β -Hydroxylase.