

1 **Cranial integration in the fire salamander, *Salamandra salamandra* (Caudata:**
2 ***Salamandridae*).**

3 Running title: salamander cranial integration

4

5 Margot Bon^{1*†}, Carla Bardua^{1,2*†}, Anjali Goswami¹ Anne-Claire Fabre^{1*}

6 ¹Department of Life Sciences, Natural History Museum, Cromwell Rd, Kensington, London,
7 SW7 5BD, UK

8 ²Department of Genetics, Evolution & Environment, University College London, Gower St,
9 Bloomsbury, London, WC1E 6BT, UK;

10 *corresponding authors: m.bon@nhm.ac.uk, ucbtcb5@ucl.ac.uk, a.fabre@nhm.ac.uk

11 †These authors contributed equally to this work

12 Number of tables: 4

13 Number of figures: 4

14 Supplementary tables: 6

15

16

17

18 **ABSTRACT**

19 Phenotypic integration and modularity are concepts that represent the pattern of
20 connectivity of morphological structures within an organism. Integration describes the
21 coordinated variation of traits, and analyses of these relationships among traits often reveal
22 the presence of modules, sets of traits that are highly integrated but relatively independent
23 of other traits. Phenotypic integration and modularity have been studied at both the
24 evolutionary and static level across a variety of clades, although most studies thus far are
25 focused on amniotes, and especially mammals. Using a high-dimensional geometric
26 morphometric approach, we investigated the pattern of cranial integration and modularity of
27 the Italian fire salamander (*Salamandra salamandra giglioli*). We recovered a highly modular
28 pattern, but this pattern did not support either entirely developmental or functional
29 hypotheses for cranial organisation, possibly reflecting complex interactions amongst
30 multiple influencing factors. We found that size had no significant effect on cranial shape, and
31 that morphological variance of individual modules had no significant relationship with
32 respective degree of within-module integration. The pattern of cranial integration in the fire
33 salamander is similar to that previously recovered for caecilians, with a highly integrated
34 suspensorium and occipital region, suggesting possible conservation of patterns across
35 lissamphibians.

36 **KEYWORDS** Amphibia- crania- disparity- morphology- phenotypic integration

37

38 INTRODUCTION

39 Identifying the primary factors that shape the evolution of organisms is a long-standing
40 interest in biology. Analysing phenotype is a relevant scale to address this question, allowing
41 consideration of both external (e.g. ecological interactions, climate) and internal (i.e. genetic,
42 development) factors (e.g. Goswami *et al.*, 2014; Collar *et al.*, 2010; Moen *et al.*, 2013; Da Silva
43 *et al.*, 2018). However, the types and sources of the data used by studies of external or
44 internal factors are usually different, preventing combined studies of both aspects (Goswami
45 *et al.*, 2014). In 1958, Olson and Miller hypothesized that phenotypic traits are more
46 integrated or correlated (statistically associated) when derived from the same genetic or
47 developmental origin or participating in the same function (Olson & Miller, 1958). Organisms
48 are made of locally integrated units, also defined by Wagner (1996) as modules, which has
49 also been noted as the reason why characters are often easily identifiable across diverse
50 organisms (Wagner & Altenberg, 1996). Traits within a module are highly connected (i.e.
51 integrated) but less connected to traits of other modules. Integration and modularity
52 therefore refer to the pattern and magnitude of connectivity within an organism. It has been
53 suggested that modularity enables sets of traits to evolve independently of other sets of traits,
54 reducing or removing the constraints of high integration, where changes in one trait may
55 negatively impact the function of a closely integrated trait (Wagner, 1996). Empirical studies
56 have focused on this aspect of evolutionary integration and modularity and have showed
57 either positive correlations between the magnitude of integration and rate of evolution or
58 disparity (Goswami *et al.*, 2014; Randau & Goswami, 2017), or that high integration is indeed
59 correlated with low evolutionary rates (Felice *et al.*, 2018). More recent studies have also

60 found no simple linear relationship between strength of integration and either evolutionary
61 rate or disparity (Bardua et al. 2019b, Watanabe et al. 2019).

62

63 Integration and modularity can be explored at the evolutionary level (i.e., at the interspecific
64 level across a whole clade at one ontogenetic stage; Klingenberg 2014), the static level (i.e.,
65 at the intraspecific level, within one species at one ontogenetic stage; Klingenberg 2014) and
66 the ontogenetic level (i.e., within one species across ontogenetic stages; Klingenberg 2014).

67 Study of these three levels reveals insights into the underlying biological processes governing
68 evolution. The static level can be used to infer functional, developmental, and genetic
69 integration (Klingenberg, 2014), all of which mutually influence each other, forming a complex
70 network of interactions (See Fig. 1 from Klingenberg 2008). Functional and genetic modularity
71 are thought to evolve to match, through the creation of a modular 'genotype-phenotype map'
72 (Wagner & Altenberg 1996), and it is hypothesized that developmental pathways evolve so
73 that functional and developmental integration also match ('matching hypothesis', Wagner &
74 Altenberg 1996). Thus, identification of the pattern of static integration can serve as a starting
75 point to then determine functional, genetic and developmental sources of covariation within
76 a structure and how these factors influence evolvability (Klingenberg, 2014). Evolutionary
77 integration reveals the large-scale patterns of change during evolution, which can be driven
78 by both functional (e.g., performance selection) and genetic (e.g., evolution by selection and
79 drift) modularity (Klingenberg, 2008). Concordant patterns within and between species
80 suggest that modularity is affected by common biological processes through evolution (e.g.
81 developmental, genetic), as has been found in compound leaves where development highly
82 modulates any other variation (Klingenberg *et al.*, 2012). Conversely, discrepancy in patterns
83 of integration between evolutionary and static levels have been found in lizards, suggesting

84 that the functional integration pattern at the static level has appeared by adaptation through
85 selection (Urošević *et al.* 2019). Finally, selection can act on any ontogenetic stage and studies
86 have demonstrated that patterns of integration shift through ontogeny (Zelditch, 1988;
87 Zelditch & Carmichael 1989a, 1989b; Willmore *et al.*, 2006; Goswami & Polly 2010a; Goswami
88 *et al.*, 2012; Ackermann, 2005). All three levels of integration and modularity can therefore
89 together aid our understanding of evolution.

90

91 Within tetrapods (limbed vertebrates), evolutionary, ontogenetic, and static modularity have
92 been investigated across a range of taxa. Evolutionary modularity has been investigated in
93 mammals, where many studies found as many as six-modules in the cranium (e.g., Cheverud
94 1995; Goswami, 2006; Porto *et al.*, 2009; Goswami & Polly, 2010b), influenced by both
95 development and function. Six-module cranial organisation was also recovered at the static
96 and the ontogenetic levels, for example in Macaques (Goswami & Finarelli, 2016), suggesting
97 that modularity in the mammal cranium is constrained by both functional and developmental
98 influences. Within archosaurs, patterns of evolutionary modularity are generally conserved,
99 with highly modular systems recovered across the crania of birds, non-avian dinosaurs and
100 crocodylomorphs (Felice & Goswami, 2018; Felice *et al.*, 2019). Alternatively, the avian skull
101 has also been found to be highly integrated at the evolutionary level (Klingenberg & Marugán-
102 Lobón, 2013). Across squamates, Watanabe *et al.* (2019) found that the cranium comprised
103 nine modules in snakes and ten modules in lizards, revealing highly modular structures
104 influenced by functional constraints across these clades. In contrast, at the static level,
105 Urošević *et al.* (2012) found that the head of the common lizard (*Podarcis muralis*) is a highly
106 integrated structure. Patterns of cranial integration across lacertids (lizards) have been found

107 to vary across evolutionary and static levels, with patterns at the evolutionary level more
108 influenced by development and patterns at the static level more influenced by function,
109 which suggests that functional modularity is adaptive and appeared through selection
110 (Urošević *et al.* 2019). In *Anolis* lizards, patterns of static modularity have also been found to
111 vary across species; a three-module cranial organisation is supported in some *Anolis* (lizards)
112 species but no support for modular organisation was found in other *Anolis* species. Thus,
113 similar to lacertids (Urošević *et al.*, 2019), patterns of cranial integration in *Anolis* lizards may
114 be evolutionary flexible and influenced by unique functional pressures acting on the diverse
115 cranial shapes (Sanger *et al.*, 2012). Across amniotes, studies of evolutionary and static
116 modularity have therefore recovered a range of modular organisations, from fully integrated
117 to highly modular structures, although differences in results may partially reflect differences
118 in type of data collected (Felice *et al.*, 2018; Goswami *et al.*, 2019).

119
120 Relatively few studies have focused on lissamphibians (Anura, Caudata and Gymnophiona).
121 The large range of ecologies and developmental histories across lissamphibians, along with
122 their impressive cranial diversity, suggest patterns of cranial integration in this clade may be
123 complex. However, studies of European newts at the static level have found that the skull is
124 highly integrated with no distinct modules (Ivanović *et al.*, 2005; Ivanović & Kalezić, 2010), a
125 result mirrored at the evolutionary level across the myobatrachid frog family (Vidal-García &
126 Keogh 2017). Interestingly, static integration of the cranium is lower for paedomorphic forms
127 of two European newts compared with fully metamorphosed forms, which can be explained
128 by ontogenetic integration, where integration increases post-metamorphosis (Ivanović *et al.*,
129 2005). Mixed support was found for a range of three to five module models across different
130 toad species of the *Rhinella granulosa* complex (Simon & Marroig, 2017), and for caecilians,

131 a two-module model was found as best supported for the skull at both the evolutionary and
132 static level (Sherratt, 2011). However, comparison of patterns of integration across clades is
133 hindered by the range of data types, from linear distances (e.g., Simon & Marroig, 2017) to
134 landmarks (e.g., Sherratt, 2011), and some methods prohibit the exploration of multiple
135 models of modularity. Recent advancements in morphometric methods may facilitate the
136 detection of a finer-scale modular signal. Recent high-dimensional studies across caecilians
137 (Bardua *et al.*, 2019b) and within two caecilian species (Marshall *et al.*, 2019) have both
138 recovered similarly highly modular cranial structures, suggesting possible conservation of
139 modularity across static and evolutionary levels. Thus investigation into finer-scale patterns
140 of static and evolutionary integration for additional lissamphibian groups may reveal whether
141 this pattern is conserved across Lissamphibia.

142

143 Caudata (salamanders) comprises 10 ecologically and morphologically diverse families and
144 displays a tremendous range of life cycle strategies (e.g., Bonett, 2018; Bonett & Blair, 2017;
145 Bonett *et al.*, 2013; Ledbetter & Bonett, 2019). Extensive phenotypic variation can also extend
146 intraspecifically in salamanders, as in the fire salamander *Salamandra salamandra* (Linnaeus,
147 1758) which exhibits a range of morphologies, colour patterns and reproductive strategies
148 depending on the subspecies (Sparreboom, 2014; Beukema *et al.*, 2016). Taxonomy of the
149 fire salamander is repeatedly being revised, but *S. salamandra* currently comprises at least 10
150 subspecies (Sparreboom, 2014). Across these subspecies, the fire salamander is polymorphic
151 in its mode of reproduction, with most populations ovoviviparous but some viviparous
152 (Dopazo & Alberch, 1994; Alcobendas *et al.*, 1996; Buckley *et al.*, 2007). The high flexibility in
153 development in this species suggests developmental influences on cranial integration may be
154 variable, and provides an interesting opportunity for investigating developmental and

155 functional hypotheses of static modularity. The skull is suitable for investigating patterns of
156 integration, as it is a highly developmentally complex structure (Hanken & Hall, 1993), given
157 the different embryonic origins (neural crest and paraxial mesoderm) and types of ossification
158 (endochondral and intramembranous) across the cranial regions. In addition, the transition
159 from larval to adult morphology for metamorphic subspecies involves deep osteological
160 remodelling in the skull, including the resorption of bones (e.g., vomer, and palatine portion
161 of the palatopterygoid), and the late development of the maxilla and prefrontal (Rose,
162 2003). The cranium of *Salamandra salamandra* comprises 15 bones, although three of them
163 fuse in one single complex- the prootic and opisthotic fuse to form the otic capsule, which
164 fuses posteriorly with the exoccipital and form the occipito-otic bone (Rose, 2003). Apart from
165 three endochondral bones (the occipito-otic complex, the orbitosphenoid and the quadrate),
166 all bones form by intramembranous ossification (Rose, 2003). The skull is also functionally
167 complex (Moore, 1981; Hanken & Hall, 1993), playing a major role in foraging and feeding, as
168 well as protecting the brain and housing the organs that perceive the main senses (hearing,
169 sight, smell and taste). The skull is therefore an ideal system for investigating fine-scale
170 patterns of integration.

171

172 Here we investigate patterns of morphological integration and modularity in the skull of the
173 Italian subspecies of fire salamander [*Salamandra salamandra giglioli* (Eiselt & Lanza, 1956)],
174 which undergoes metamorphosis (Seidel & Gerhardt, 2016). We directly compare eight
175 different modular structures based on hypothesized functional and developmental
176 relationships between cranial regions. We also determine whether integration constrains or
177 facilitates morphological diversity (disparity) by quantifying the relationship between within-
178 module variance and within-module magnitude of integration. Combined, these analyses

179 allow us to assess static (intraspecific) modularity and integration within a population, for
180 qualitative comparison to previous studies of modularity and integration within
181 lissamphibians [e.g., in caecilians (Bardua *et al.*, 2019b; Marshall *et al.*, 2019), frogs and toads
182 (Simon & Marroig, 2017; Vidal-García & Keogh 2017) and salamanders and newts (Ivanović *et*
183 *al.*, 2005; Ivanović & Kalezić, 2010)] and amniotes. This study thus adds to a rich and increasing
184 pool of understanding the evolution and significance of phenotypic integration and
185 modularity for shaping organismal variation at micro- and macroevolutionary scales.

186

187 MATERIAL AND METHODS

188 STUDY SPECIMENS

189 The sample analysed in this study includes 40 specimens of the subspecies *Salamandra*
190 *salamandra gigliolii* (Table S1). None of the cloacal regions of the specimens displayed an
191 obvious swelling that could differentiate males from females (Brizzi & Calloni, 1992), thus sex
192 information was not available for these specimens. All the specimens were preserved in
193 alcohol in the collections of the Natural History Museum (NHM), London. The specimens were
194 imaged using micro CT-scanning (Nikon Metrology X-Tek HMX ST 225) at the NHM and
195 resulting tomographs were further segmented in Avizo Lite v.9.3 (FEI, Hillsboro, OR, USA) to
196 obtain 3D models of the crania. Because this study focuses on cranial morphology, the
197 reconstructed meshes were processed in Geomagic Wrap (3D Systems) to remove vertebral
198 elements and the mandible from each cranium, which could hinder the access to the surface
199 of interest. The right half of each skull was then prepared for surface analyses using Geomagic
200 Wrap, by smoothing noise introduced from scanning and removing holes that could prevent
201 the acquisition of surface details (Bardua *et al.*, 2019a). The quality of the left-hand side of

202 the skull was favored in two specimens, for which the skull was thus medially mirrored with
203 the “Mirror” function in Geomagic Wrap.

204

205 **MORPHOMETRIC DATA**

206 To characterise the shape of the regions of interest and assess how they correlate, a
207 landmarking procedure was performed using Checkpoint (Stratovan, Davis, CA, USA) by the
208 same user to keep the placement of landmark consistent and to avoid user bias. Eighty-five
209 anatomical landmarks were manually placed on the right-hand side of each skull, defining 20
210 regions in total (Fig. 1 and Table 1) that were identifiable in all specimens of the dataset. These
211 landmarks were homologous across all specimens. Regions delineate bones, or sub-regions
212 of bones when the bone is anatomically polyvalent (e.g. with ventral and dorsal surfaces), and
213 thus potentially modular due to differing functional pressures. The regions are delimited with
214 sliding semi-landmarks (‘curves’) in between landmarks (Fig. 1 and Table S2). Eighty-five
215 curves in total were drawn over the skull. These curves were then resampled in R (R Core
216 Development Team, 2019) to 689 curve semilandmarks to ensure that they were equidistant
217 and that they capture shape optimally (Botton-Divet *et al.*, 2016). To capture surface
218 information from the regions, 375 sliding surface points were placed over the crania following
219 a semi-automatic procedure using the R (R Core Development Team, 2019) package *Morpho*
220 v.2.6 (Schlager, 2017). First, one specimen of the dataset was defined as the template and
221 surface points were manually placed onto each region. Then, surface points were semi-
222 automatically projected from this template onto each specimen with the “placePatch”
223 function in *Morpho* v.2.6 (Schlager, 2017). Finally, all curve and surface points were slid to
224 become geometrically homologous, minimizing the bending energy criterion, using the

225 'slider3d' function in *Morpho* v.2.6 (Schlager, 2017). A detailed description of this method is
226 available in Bardua *et al* (2019a).

227

228 Following sliding, morphometric data were subjected to Procrustes analysis to remove the
229 non-shape aspects of isometric size, rotation, and translation (Rohlf & Slice, 1990). However,
230 because alignment of only one side of a bilateral structure can have a negative impact on the
231 Procrustes alignment (Cardini, 2016), we first mirrored morphometric data to produce a fully
232 bilateral configuration using the "mirrorfill" function in *paleomorph* v.0.1.4 (Lucas &
233 Goswami, 2017). We then performed Procrustes alignment with the "gpagen" function in
234 *geomorph* 3.0.4 (Adams *et al.*, 2019). Finally, the mirrored side was removed from the
235 resulting Procrustes shape coordinates, leaving the right-hand side coordinates only for
236 further analysis.

237

238 **ALLOMETRY**

239 Allometry corresponds to the impact of size on shape (Klingenberg, 2016). In our study, cranial
240 size was measured as the centroid size (Klingenberg, 2016), which was calculated with the
241 "gpagen" function in *geomorph* 3.0.4 (Adams *et al.*, 2019) during the Procrustes alignment
242 (see Supplementary Table S3). Allometry was assessed with a regression of the shape data on
243 log-transformed centroid size, using the "procD.allometry" function in *geomorph* v.3.0.4
244 (Adams *et al.*, 2019).

245

246 **MODULARITY AND INTEGRATION**

247 The eight hypothetical patterns of modularity tested in this study are outlined below. Most
248 simply, we tested for a fully integrated cranium, constrained by its highly diverse functions

249 (Fig. 2A and Table 2). The cranium could also be divided into two modules based on the two
250 types of ossification (endochondral and dermal), resulting in two different developmental
251 modules (Fig. 2B and Table 2). We also tested two functional hypotheses that were previously
252 tested for caecilians in the literature (Bardua *et al.*, 2019b ; Marshall *et al.*, 2019), in which
253 the skull is either dorsoventrally divided (Fig. 2C and Table 2) or partitioned into four
254 functional modules (Fig. 2D and Table 2). Since bones have different ossification sequences,
255 which could influence skull variation and evolution, we hypothesised that cranial modules are
256 defined by their time of ossification (Fig. 2E and Table 2) as previously explored in the
257 literature (Ivanović & Kalezić, 2010), resulting in four different modules: early, mid, late or
258 metamorphosis modules. We further partitioned and tested a six-module model (Fig. 2F and
259 Table 2) similar to the model found in mammals (Goswami, 2006). Each bone is an identifiable
260 unit that originates from an independent signal, so we also hypothesised that bones comprise
261 their own modules (Fig. 2G and Table 2). Finally, because some bones form distinct regions
262 that are implied in different functions (e.g. the *pars facialis* and the *pars palatina* of the
263 maxilla) and are thus potentially functionally modular, we further hypothesised that the
264 cranium could be highly partitioned into 20 functional modules (Fig. 2H and Table 2).

265 Modularity was investigated with two different approaches. Firstly, we conducted EMMLi
266 ('Evaluating Modularity with Maximum Likelihood') analysis. For this, congruence coefficients
267 were calculated with the "EMMLi" function from the *EMMLi* v. 0.0.3 R package (Goswami *et*
268 *al.*, 2017). Robustness of the results was tested with a random subsampling down to 10% of
269 the full data, and EMMLi ran iteratively 100 times, using the "subSampleEMMLi" function
270 from the *EMMLi* v. 0.0.3 R package. Mean results from the 100 subsamples were then
271 compared to the analysis of the full dataset.

272 Secondly, we conducted Covariance Ratio (CR) analysis (Adams, 2016) using the
273 “modularity.test” function from the *geomorph* v.3.0.4 R package. While EMMLi is a model
274 selection approach that compares different models of modular organisation and outputs the
275 most likely model along with its between and within module correlations, covariance ratio
276 analysis is a hypothesis testing method that calculates covariance ratio between the different
277 regions in one specific model. The ratio is the total between-module covariance over the
278 within-module covariance (Adams, 2016), meaning that a ratio of one reflects a lack of
279 modularity. Therefore, support for the preferred model from EMMLi analysis was assessed by
280 calculating the covariance ratio of that model, to confirm that both methods supported
281 similar patterns of trait integration and modularity. We further conducted both analyses with
282 a landmark-only dataset for comparison.

283

284 **MORPHOLOGICAL VARIANCE**

285 To discern how one module varies from one skull to another, we computed the shape variance
286 (the Procrustes distance between shapes and the mean shape of a module) of each module
287 with the “morphol.disparity” function from the *geomorph* v.3.0.4 R package. To compare the
288 variances between modules, each variance was corrected by dividing by the total number of
289 landmarks defined in the module concerned.

290 To investigate the relationship between shape variance and the magnitude of within-module
291 integration, we computed a linear regression of corrected variances on within-module
292 correlation.

293

294 RESULTS

295 ALLOMETRY

296 The regression of shape on log-transformed centroid size was not significant ($R^2 = 0.037$, $p =$
297 0.125), suggesting that there is no significant effect of size on shape in this cranial structure.
298 Therefore, we did not apply allometric corrections in further analyses.

299

300 MODULARITY AND INTEGRATION

301 The most supported model by EMMLi for the complete dataset is the maximal partitioning
302 modular pattern (Table 3), comprising 20 modules (Fig. 3). However, since we could not test
303 all possible models of modularity, and because previous analyses have suggested that EMMLi
304 has a tendency to pick the most-parametrized model with semilandmark data, we explored
305 the correlation (ρ) values among and within modules to assess possible further groupings
306 of modules into larger modules, following the method previously described (Bardua *et al.*,
307 2019b; Felice & Goswami, 2018; Marshall *et al.*, 2019). Modules were hence further grouped
308 when the between-module estimated correlation was within 0.1 of the smallest within-
309 module trait correlation of the module pair under question. Therefore, we grouped 11 regions
310 into four larger modules, as follows: the squamosal, the pterygoid, the jaw joint and the
311 quadrate were grouped into one jaw suspensorium module, the ventral and dorsal regions of
312 the premaxilla formed a second module, the ventral and dorsal regions of the maxilla a third
313 module, and the ventral and dorsal part of the occipital were grouped with the occipital
314 condyle (Fig. 3 and Table 3). The average random 10% subsampling of the data resulted in a
315 near-identical pattern of trait integration (Supplementary Table S4) compared with the full

316 data, confirming the robustness of our results. Through this approach we recovered a 13-
317 module model as best supported.

318

319 EMMLi analysis of the landmark-only dataset supported the 15-module model (which
320 subdivided the cranium according to osteological units). However, further assessment of the
321 within- and between-module rho values revealed that no hypothesized cranial module stood
322 out as distinct, as within- and between-module correlations were similar and low, below 0.3
323 in most cases (See Supplementary Table S5). Within-module correlations were considerably
324 lower in the landmark-only dataset compared with the complete shape dataset, whereas
325 between-module correlations were similar across both datasets. Landmark-only analysis
326 therefore suggested the cranium of the Italian fire salamander was only weakly modular, with
327 no cranial regions standing out as highly integrated, distinct modules.

328

329 **COVARIANCE RATIO**

330 Covariance Ratio analysis for the full shape data with the most parametrized model was
331 significant ($CR = 0.69$, $p = 0.01$) and revealed similar patterns of modularity (Table 3) to those
332 recovered from EMMLi, with CR values closer to 1 for regions related to the jaw suspensorium
333 and occipital. Therefore, we find that the regions of the jaw suspensorium (pterygoid,
334 squamosal, quadrate and jaw joint) and the regions of the occipital (dorsal and ventral sides
335 and condyle) are relatively more integrated, in concordance with EMMLi analysis, but the
336 dorsal and ventral sides of the premaxilla are relatively less integrated ($CR = 0.7$).

337 Covariance ratio analysis of the landmark-only dataset found significant support for the 15-
338 module model ($CR = 0.85$, $p = 0.01$). Covariance ratio values within this model were generally
339 higher than CR values from the full shape dataset, indicating weaker evidence of modularity

340 with the landmark-only dataset. Moreover, many CR values approached or exceeded one for
341 several pairs of hypothesized modules, indicating that, despite the significant support for an
342 overall pattern of modularity, CR analysis does not show strong support for most of the
343 hypothesized cranial modules when quantified by landmarks alone (Table S6).

344

345 **MORPHOLOGICAL DISPARITY**

346 Cranial modules within the jaw suspension (pterygoid, squamosal, lateral surface of the
347 quadrate and jaw joint bones) exhibited the highest disparity after correcting for landmark
348 number, and the orbitosphenoid exhibited the lowest disparity (Table 4).

349 The linear regression of Procrustes variances of the modules on their respective within-
350 module correlations (Fig. 4 and Table 4) was not significant (multiple $R^2 = 0.008$, adjusted R^2
351 $= -0.081$, $p = 0.763$). Repeating this regression with the original 20 cranial regions revealed a
352 similar, non-significant relationship (multiple $R^2 = 0.008$, adjusted $R^2 = -0.047$, $p = 0.706$) (See
353 Supplementary Table S7).

354

355 **DISCUSSION**

356 This study comprehensively sampled the cranial morphology of the Italian fire salamander
357 using a high-dimensional approach, and found a complex pattern of modularity, where the
358 Italian fire salamander cranium comprises 13 modules. This pattern of trait integration was
359 supported from EMMLi analysis with both full data and data subsampled to 10%, as well as
360 with CR analysis. The fire salamander skull therefore comprises multiple, semi-independent
361 regions, with fine-scale, localised variation in functional or developmental influences. The
362 modules we found correspond primarily to individual osteological units, as well as a functional

363 module corresponding to the jaw suspensorium (including the pterygoid, quadrate, jaw joint
364 surface of the quadrate and squamosal regions). The influences of type and timing of
365 ossification do not appear to drive the overall pattern of integration, possibly because the
366 superimposition of multiple sources of covariance can confound each other (Hallgrímsson *et*
367 *al.*, 2009). However, because genetic, developmental and functional modularity are thought
368 to evolve to match (Wagner & Altenberg 1996), other developmental or genetic influences
369 than those tested here may act as processes driving the pattern of modularity that we
370 identify. Identifying the pattern of static integration can therefore help identify the drivers of
371 this pattern (i.e. functional or developmental processes) by narrowing possible factors to
372 those related to the specific patterns observed. The static pattern can also be compared with
373 patterns of integration at the evolutionary and ontogenetic level for determining the factors
374 driving the morphological evolution of salamanders, and for understanding whether our
375 observed pattern of static integration, affected by both development and functional
376 pressures, is conserved through evolution.

377

378 An absence of distinct developmental modules in the fire salamander cranium is consistent
379 with a previous study of the alpine newt skull (Ivanović & Kalezić, 2010). These results may
380 reflect confounding or overlapping developmental and environmental interactions through
381 ontogeny, or cranial partitioning may have been too simple given the complexity of
382 developmental influences acting on the skull. However, Ivanović & Kalezić (2010) found no
383 support for any hypothesized modular structure, in contrast to the highly modular pattern
384 that we recover in the fire salamander skull. This large discordance may suggest patterns of
385 integration may be highly flexible within different salamander species. Integration can be
386 flexible even within species with different morphs, and can vary through ontogeny (Ivanović

387 *et al.*, 2005), suggesting patterns of integration may be flexible according to life history.
388 However, differences may also be largely attributable to differences in data type and
389 analyses. Ivanović & Kalezić (2010) used two-dimensional landmark data and consequently
390 could not capture shape information for the orbitosphenoid and some bones of the jaw
391 suspension (quadrate and squamosal). In addition, testing hypotheses comprising two to four
392 modules prevented the detection of finer-scale patterns of integration and thus hinders direct
393 comparison between our study and that of Ivanović & Kalezić (2010). Comparison of
394 integration patterns within different subspecies of fire salamander varying in developmental
395 strategy may reveal whether integration patterns are flexible and influenced by
396 developmental strategy.

397

398 In contrast, comparing our results to studies implementing a similarly high-dimensional
399 approach reveals instead a surprisingly conserved pattern of integration, particularly within
400 Lissamphibia. The 13-module model we recover in the fire salamander cranium is very similar
401 to the 12- and 13- module models identified intraspecifically within caecilians (Marshall *et*
402 *al.*, 2019), and to the 10-module model found across the caecilian clade (Bardua *et al.*, 2019b).
403 This similarity is notable given the osteological differences between salamanders and
404 caecilians, which hinders direct comparison. Like Marshall *et al.* (2019), we find that the
405 parasphenoid (analogous to the caecilian ventral os basale) and the vomer form two
406 independent modules, probably due to functional decoupling, the parasphenoid being the
407 floor of the braincase and the vomer being part of the palate (Rose, 2003). Independent
408 palatal and braincase floor regions have also been found across bird and squamate crania
409 (Felice & Goswami, 2018; Watanabe *et al.*, 2019), although across the caecilian clade these
410 regions formed one module (Bardua *et al.*, 2019b). We also find the three regions comprising

411 the occipital bone form one distinct module, despite their functional differences (braincase
412 protection and connection to the vertebral column). A distinct occipital module is also found
413 in caecilian (Bardua *et al.*, 2019b; Marshall *et al.*, 2019), bird (Felice & Goswami, 2018), non-
414 avian dinosaur and crocodylomorph (Felice *et al.*, 2019) crania. The modular structure
415 identified across the fire salamander skull is more strongly concordant with the pattern
416 recovered within caecilians (Bardua *et al.*, 2019b; Marshall *et al.*, 2019) than amniotes,
417 suggesting a possible divergence in modularity patterns between amniotes and amphibians.
418 The similarity of integration patterns between salamanders and caecilians suggests
419 conservation of modularity despite the great diversity of developmental histories and
420 ecologies across these two amphibian clades.

421

422 We recover a highly integrated jaw suspensorium module (quadrate, pterygoid and
423 squamosal) within the fire salamander skull, likely driven by constraints from feeding
424 mechanics. A highly integrated jaw suspensorium region has also been found across and
425 within caecilians, comprising two strongly correlated modules (quadrate-squamosal and
426 pterygoid) (Bardua *et al.*, 2019b; Marshall *et al.*, 2019). In addition, across birds, the quadrate
427 and pterygoid form a module, although the squamosal belongs to the cranial vault (Felice &
428 Goswami, 2018). This jaw joint region of both caecilians and birds exhibits coordinated
429 movement through kinesis, which may be driving its strong integration (Felice & Goswami,
430 2018; Bardua *et al.*, 2019b). The highly integrated jaw suspensorium module of the fire
431 salamander may be due to tightly linked, functional constraints in this region related to
432 changing feeding requirements through ontogeny. Fire salamanders experience disparate
433 selective pressures through ontogeny, from larval to adult stages, related to changes in both
434 environment and diet (from water to land), and thus in their feeding mechanism (Shaffer &

435 Lauder, 1988; Wainwright & Reilly, 1994). The highly integrated jaw joint region shared across
436 fire salamanders, caecilians, and birds suggests feeding mechanics plays a large role in shaping
437 the pattern of integration for these clades.

438

439 Data type can have a strong impact on studies of integration. Here, whilst our full landmark
440 and semilandmark dataset recovers a highly modular structure, our landmark-only dataset
441 instead suggest a weakly integrated model, with little to no support for many of the
442 hypothesized cranial modules (despite both methods supporting an overall highly-modular
443 pattern for the cranium). This present study, along with previous comparisons of landmark
444 and semilandmark data to landmark-only data, thus reveals how landmark-only data may
445 exaggerate between-region trait correlations and understate within-region trait correlations
446 (Bardua *et al.*, 2019b; Marshall *et al.*, 2019) resulting in landmark-only datasets recovering
447 weaker support for modular structure. This is because landmark-only datasets suffer from
448 boundary bias (Goswami *et al.*, 2019) and do not capture shape information along curves and
449 across surfaces, meaning the shape data are not fully representative of a structure. Analyses
450 using semilandmarks may have the opposite effect, due to non-independence of
451 semilandmarks, but it has been demonstrated in multiple studies that they better capture the
452 morphology of complex structures (Watanabe, 2018; Bardua *et al.*, 2019b; Goswami *et al.*,
453 2019). While all methods suffer from biases and artefacts, the improved characterisation of
454 shape achievable with a full landmark and semilandmark dataset likely better reflects the
455 structure of the cranium and the interactions among its regions.

456 We found no significant relationship between integration and shape variance, and thus no
457 support for the hypothesis that integration facilitates or constrains morphological variance in
458 the fire salamander skull. Our results may thus suggest that integration has limited or variable

459 influence on morphological diversification. No significant relationship between integration
460 and variance was also recovered intraspecifically within caecilians (Marshall *et al.*, 2019).
461 Similarly, the crania of domestic dogs display a conserved pattern of integration despite a high
462 cranial variance across the entire order (Drake & Klingenberg, 2010). Studies at the
463 macroevolutionary scale have thus far found contrasting patterns, suggesting the relationship
464 between integration and variance is highly flexible across clades, or that the relationship
465 cannot be expressed linearly. Whilst no significant linear relationship between integration
466 and variance was found across caecilians (Bardua *et al.*, 2019), high levels of integration are
467 associated with low morphological disparity in the crania of birds and mammals (Felice *et al.*,
468 2018; Goswami *et al.*, 2014), and conversely, the modular fins of ray-finned fish are highly
469 variable (Larouche *et al.*, 2018). More likely, whether integration promotes or constrains
470 morphological evolution of a module may depend heavily on the alignment of the direction
471 that it facilitates in the morphospace and the direction of selection (Hansen *et al.*, 2011;
472 Goswami *et al.*, 2014; Felice *et al.*, 2018). The heterogeneity of results across studies so far
473 therefore suggests that the relationship between integration and disparity may be complex,
474 with no simple relationship between the two metrics in many, if not most, cases.

475

476 With a high dimensional approach, we analysed the cranial organisation of the Italian fire
477 salamander (*Salamandra salamandra giglioli*). The fire salamander's cranium is highly
478 modular, comprising 13 modules, and this modular organisation is highly similar to that
479 recovered in caecilians in terms of both the number and pattern of modules (Bardua *et al.*,
480 2019b; Marshall *et al.*, 2019). This result suggests a possible conservation of the pattern of
481 integration across lissamphibian crania. We found no support for purely developmental or
482 functional hypotheses of modular organisation, which suggests that the cranial modular

483 pattern in the fire salamander is complex and originates from a mixture of both functional
484 and developmental constraints. Cranial size had little effect on cranial shape in this species,
485 and morphological disparity had no significant relationship with within-module integration,
486 contrary to hypotheses that integration may facilitate or constrain morphological variation.
487 The study of cranial integration of salamanders at the evolutionary level would complement
488 this present study on static integration, providing a more comprehensive understanding of
489 the intrinsic factors shaping the evolution of the salamander skull.

490 ACKNOWLEDGMENTS

491

492 We thank Jeffrey W. Streicher at the NHM for giving us access to the collection and helping
493 us with the choice of specimens used in this study. We also thank Vincent Fernandez and Brett
494 Clark for providing training for CT-scanning at the NHM. Finally, we would like to thank three
495 anonymous reviewers for their helpful and insightful comments. This work was funded by the
496 European Research Council (grant STG-2014–637171 to AG).

497 CONFLICTS OF INTEREST

498

499 The authors declare no conflicts of interest.

500 REFERENCES

501 **Ackermann R. 2005.** Ontogenetic integration of the hominoid face. *Journal of Human Evolution* **48**:
502 175–197.

503 **Adams DC. 2016.** Evaluating modularity in morphometric data: challenges with the RV coefficient
504 and a new test measure (P Peres-Neto, Ed.). *Methods in Ecology and Evolution* **7**: 565–572.

505 **Adams DC, Collyer ML, Kaliontzopoulou A. 2019.** *Geomorph: Software for geometric morphometric*
506 *analyses*. R package version 3.1.0. <https://cran.r-project.org/package=geomorph>.

507 **Alcobendas M, Dopazo H, Alberch P. 1996.** Geographic variation in allozymes of populations of
508 *Salamandra salamandra* (Amphibia: Urodela) exhibiting distinct reproductive modes. *J Evolution Biol*
509 **9:** 83–102.

510 **Bardua C, Felice RN, Watanabe A, Fabre A-C, Goswami A. 2019a.** A practical guide to sliding and
511 surface semilandmarks in morphometric analyses. *Integrative Organismal Biology*. obz016

512 **Bardua C, Wilkinson M, Gower DJ, Sherratt E, Goswami A. 2019b.** Morphological evolution and
513 modularity of the caecilian skull. *BMC Evolutionary Biology* **19:** 30.

514 **Beukema W, Niecieza AG, Lourenço A, Velo-Antón G. 2016.** Colour polymorphism in *Salamandra*
515 *salamandra* (Amphibia: Urodela), revealed by a lack of genetic and environmental differentiation
516 between distinct phenotypes. *Journal of Zoological Systematics and Evolutionary Research* **54:** 127–
517 136.

518 **Bonett RM. 2018.** Heterochrony. In: Nuño de la Rosa L and Müller GB, eds. *Evolutionary*
519 *Developmental Biology*. Cham: Springer. 1-14.

520 **Bonett RM, Blair AL. 2017.** Evidence for complex life cycle constraints on salamander body form
521 diversification. *Proceedings of the National Academy of Sciences* **114:** 9936-9941.

522 **Bonett RM, Steffen MA, Lambert SM, Wiens JJ, Chippindale PT. 2013.** Evolution of paedomorphosis
523 in plethodontid salamanders: ecological correlates and re-evolution of metamorphosis. *Evolution* **68:**
524 466-482.

525 **Botton-Divet L, Cornette R, Fabre A-C, Herrel A, Houssaye A. 2016.** Morphological analysis of long
526 bones in semi-aquatic mustelids and their terrestrial relatives. *Integrative and Comparative Biology*
527 **56:** 1298-1309.

528 **Brizzi R, Calloni C. 1992.** Male cloacal region of the spotted salamander, *Salamandra salamandra*
529 *gigliolii* (Amphibia, Salamandridae). *Bolletino di zoologia* **59:** 377–385.

530 **Buckley D, Alcobendas M, García-París M, Wake MH. 2007.** Heterochrony, cannibalism, and the
531 evolution of viviparity in *Salamandra salamandra*. *Evolution & Development* **9:** 105–115.

532 **Cardini A. 2016.** Lost in the other half: Improving accuracy in geometric morphometric analyses of
533 one side of bilaterally symmetric structures. *Systematic Biology* **65**: 1096-1106.

534 **Cheverud JM. 1995.** Morphological integration in the saddle-back tamarin cranium. *American*
535 *Naturalist* **145**: 63–89.

536 **Collar DC, Schulte JA, O’Meara BC, Losos JB. 2010.** Habitat use affects morphological diversification
537 in dragon lizards. *Journal of Evolutionary Biology* **23**: 1033–1049.

538 **Da Silva FO, Fabre A-C, Savriama Y, Ollonen J, Mahlow K, Herrel A, Müller J, Di-Poï N. 2018.** The
539 ecological origins of snakes as revealed by skull evolution. *Nature Communications* **9**:376.

540 **Dopazo HJ, Alberch P. 1994.** Preliminary results on optional viviparity and intrauterine siblicide in
541 *Salamandra salamandra* populations from northern Spain. *Mertensiella* **4**:125-138.

542 **Drake AG, Klingenberg CP. 2010.** Large-Scale Diversification of Skull Shape in Domestic Dogs:
543 Disparity and Modularity. *The American Naturalist* **175**: 289-301.

544 **Felice RN, Goswami A. 2018.** Developmental origins of mosaic evolution in the avian cranium.
545 *Proceedings of the National Academy of Sciences* **115**: 555-560.

546 **Felice RN, Randau M, Goswami A. 2018.** A fly in a tube: Macroevolutionary expectations for
547 integrated phenotypes. *Evolution* **72**: 2580-2594.

548 **Felice RN, Watanabe A, Cuff AR, Noirault E, Pol D, Witmer LM, Norell MA, O’Connor PM, Goswami**
549 **A. 2019.** Evolutionary Integration and Modularity in the Archosaur Cranium. *Integrative and*
550 *Comparative Biology* **59**: 371–382.

551 **Goswami A. 2006.** Morphological integration in the carnivoran skull. *Evolution* **60**: 170-180.

552 **Goswami A, Finarelli JA. 2016.** EMMLi: A maximum likelihood approach to the analysis of
553 modularity. *Evolution* **70**: 1622-1637.

554 **Goswami A, Lucas T, Sivasubramaniam P, Finarelli J. 2017.** *EMMLi: A Maximum Likelihood Approach*
555 *to the Analysis of Modularity*. R package version 0.0.3. <https://cran.r-project.org/package=EMMLi>

556 **Goswami A, Polly PD. 2010a.** Methods for Studying Morphological Integration and Modularity. *The*
557 *Paleontological Society Papers* **16**: 213–243.

558 **Goswami A, Polly PD. 2010b.** The Influence of Modularity on Cranial Morphological Disparity in
559 Carnivora and Primates (Mammalia) (AW Shingleton, Ed.). *PLoS ONE* **5**: e9517.

560 **Goswami A, Polly PD, Mock OB & Sánchez-Villagra MR. 2012.** Shape, variance and integration
561 during craniogenesis: contrasting marsupial and placental mammals. *Journal of Evolutionary Biology*
562 **25**: 862–872.

563 **Goswami A, Smaers JB, Soligo C, Polly PD. 2014.** The macroevolutionary consequences of
564 phenotypic integration. *Philosophical Transactions of the Royal Society of London B* **369**: 20130254.

565 **Goswami A, Watanabe A, Felice RN, Bardua C, Fabre A-C, Polly PD. 2019.** High-Density
566 Morphometric Analysis of Shape and Integration: The Good, the Bad, and the Not-Really-a-Problem.
567 *Integrative and Comparative Biology* icz120.

568 **Hallgrímsson B, Jamniczky H, Young NM, Rolian C, Parsons TE, Boughner JC, Marcucio RS. 2009.**
569 Deciphering the Palimpsest: Studying the Relationship Between Morphological Integration and
570 Phenotypic Covariation. *Evolutionary Biology* **36**: 355–376.

571 **Hanken J, Hall BK. 1993.** *The skull. Volume 2 - Patterns of Structural and Systematic Diversity.*
572 University of Chicago Press: Chicago.

573 **Hansen TF, Pélabon C, Houle D. 2011.** Heritability is not Evolvability. *Evolutionary Biology* **38**: 258.

574 **Ivanović A, Kalezić ML. 2010.** Testing the hypothesis of morphological integration on a skull of a
575 vertebrate with a biphasic life cycle: a case study of the alpine newt. *Journal of Experimental Zoology*
576 *Part B: Molecular and Developmental Evolution* **314B**: 527-538.

577 **Ivanović A, Kalezić ML, Aleksić I. 2005.** Morphological integration of cranium and postcranial
578 skeleton during ontogeny of facultative paedomorphic European newts (*Triturus vulgaris* and *T.*
579 *alpestris*). *Amphibia-Reptilia* **26**: 485-495.

580 **Klingenberg CP. 2008.** Morphological Integration and Developmental Modularity. *Annual Review of*
581 *Ecology, Evolution, and Systematics* **39**: 115–132.

582 **Klingenberg CP. 2014.** Studying morphological integration and modularity at multiple levels:
583 concepts and analysis. *Philosophical transactions of the Royal Society B: Biological sciences* **369**:
584 20130249.

585 **Klingenberg CP. 2016.** Size, shape, and form: concepts of allometry in geometric morphometrics.
586 *Development Genes and Evolution* **226**: 113-137.

587 **Klingenberg CP, Duttke S, Whelan S, Kim M. 2012.** Developmental plasticity, morphological
588 variation and evolvability: a multilevel analysis of morphometric integration in the shape of
589 compound leaves: Morphometric integration in compound leaf shape. *Journal of Evolutionary*
590 *Biology* **25**: 115–129.

591 **Klingenberg CP, Marugán-Lobón J. 2013.** Evolutionary covariation in geometric morphometric data:
592 Analyzing integration, modularity, and allometry in a phylogenetic context. *Systematic Biology* **62**:
593 591-610.

594 **Larouche O, Zelditch ML, Cloutier R. 2018.** Modularity promotes morphological divergence in ray-
595 finned fishes. *Scientific Reports* **8**: 7278.

596 **Ledbetter NM, Bonett RM. 2019.** Terrestriality constrains salamander limb diversification:
597 Implications for the evolution of pentadactyly. *Journal of Evolutionary Biology* **32**: 642-652.

598 **Lucas T, Goswami A. 2017.** *paleomorph: Geometric morphometric tools for paleobiology*. R package
599 version 0.1.4. <https://cran.r-project.org/package=paleomorph>.

600 **Marshall AF, Bardua C, Gower DJ, Wilkinson M, Sherratt E, Goswami A. 2019.** High-density three-
601 dimensional morphometric analyses support conserved static (intraspecific) modularity in caecilian
602 (Amphibia: Gymnophiona) crania. *Biological Journal of the Linnean Society* **126**: 721:742.

603 **Moen DS, Irschick DJ, Wiens JJ. 2013.** Evolutionary conservatism and convergence both lead to
604 striking similarity in ecology, morphology and performance across continents in frogs. *Proceedings of*
605 *the Royal Society B: Biological Sciences* **280**: 20132156.

606 **Moore WJ. 1981.** *The mammalian skull*. Cambridge: Cambridge University Press.

607 **Oslon EC, Miller RL. 1958.** *Morphological integration*. University of Chicago Press: Chicago, IL, USA.

608 **Porto A, de Oliveira FB, Shirai LT, De Conto V, Marroig G. 2009.** The Evolution of Modularity in the
609 Mammalian Skull I: Morphological Integration Patterns and Magnitudes. *Evolutionary Biology* **36**:
610 118-135.

611 **R Core Development Team. 2019.** R: A language and environment for statistical computing. R
612 Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.

613 **Randau M, Goswami A. 2017.** Unravelling intravertebral integration, modularity and disparity in
614 Felidae (Mammalia). *Evolution and Development* **19**: 85-95.

615 **Rohlf FJ, Slice D. 1990.** Extensions of the procrustes method for the optimal superimposition of
616 landmarks. *Systematic Zoology* **39**: 40-59.

617 **Rose CS. 2003.** The Developmental Morphology of Salamander Skulls. In: Heatwole H and Davies M,
618 eds. *Amphibian Biology, Vol. 5 Osteology*: Surrey Beatty & Sons.

619 **Sanger TJ, Mahler DL, Abzhanov A, Losos JB. 2012.** Roles for modularity and constraint in the
620 evolution of cranial diversity among *Anolis* lizards. *Evolution* **66**: 1525-1542.

621 **Schlager S. 2017.** Morpho and Rvcg – Shape Analysis in R. In: Zheng G, Li S and Székely G, eds.
622 *Statistical Shape and Deformation Analysis*: Academic Press. 217–256.

623 **Seidel U, Gerhardt P. 2016.** *The genus Salamandra: history, biology, systematics, captive breeding*.
624 Frankfurt am Main: Edition Chimaira.

625 **Shaffer HB, Lauder GV. 1988.** The ontogeny of functional design: metamorphosis of feeding
626 behaviour in the tiger salamander (*Ambystoma tigrinum*). *Journal of Zoology* **216**: 437–454.

627 **Sherratt E. 2011.** Evolution of the Caecilian Skull. Manchester, UK: The University of Manchester.

628 **Simon MN, Marroig G. 2017.** Evolution of a complex phenotype with biphasic ontogeny:
629 Contribution of development versus function and climatic variation to skull modularity in toads.
630 *Ecology and Evolution* **7**: 10752-10769.

631 **Sparreboom M. 2014.** *Salamanders of the Old World: the salamanders of Europe, Asia and Northern*
632 *Africa*. Zeist, The Netherlands: KNNV Publishing.

633 **Urošević A, Ljubisavljević K, Jelić D, Ivanović A. 2012.** Variation in the cranium shape of wall lizards
634 (Podarcis spp.): effects of phylogenetic constraints, allometric constraints and ecology. *Zoology* **115**:
635 207-216

636 **Urošević A, Ljubisavljević K, Ivanović A. 2019.** Multilevel assessment of the Lacertid lizard cranial
637 modularity. *Journal of Zoological Systematics and Evolutionary Research* **57**: 145-158.

638 **Vidal-García, M., Keogh, J.S. 2017.** Phylogenetic conservatism in skulls and evolutionary lability in
639 limbs - morphological evolution across an ancient frog radiation is shaped by diet, locomotion and
640 burrowing. *BMC Evolutionary Biology*. **17**:165.

641 **Wagner GP. 1996.** Homologues, natural kinds and the evolution of modularity. *American Zoology* **36**:
642 36-43.

643 **Wagner GP, Altenberg L. 1996.** Perspective: complex adaptations and the evolution of evolvability.
644 *Evolution* **50**: 967-976.

645 **Wainwright PC, Reilly SM. 1994.** *Ecological morphology: integrative organismal biology*. Chicago:
646 University of Chicago Press.

647 **Watanabe A. 2018.** How many landmarks are enough to characterize shape and size variation? *PLoS*
648 *One* **13**: e0198341.

649 **Watanabe A, Fabre A-C, Felice RN, Maisano JA, Müller J, Herrel A, Goswami A. 2019.**
650 Ecomorphological diversification in squamates from conserved pattern of cranial integration.
651 *Proceedings of the National Academy of Sciences* **116**: 14688-14697.

652 **Willmore KE, Leamy L, Hallgrímsson B. 2006.** Effects of developmental and functional interactions
653 on mouse cranial variability through late ontogeny. *Evolution & Development* **8**:550–67.

654 **Zelditch ML. 1988.** Ontogenetic variation in patterns of phenotypic integration in the laboratory rat.
655 *Evolution* **42**:28–41.

656 **Zelditch ML, Carmichael AC. 1989a.** Growth and intensity of integration through postnatal growth in
657 the skull of *Sigmodon fulviventer*. *Journal of Mammalogy* **70**:477–84.

658 **Zelditch ML, Carmichael AC. 1989b.** Ontogenetic variation in patterns of developmental and
659 functional integration in skulls of *Sigmodon fulviventer*. *Evolution* **43**:814–24.

660

661

662

663 **Figure captions**

664

665 **Figure 1.** Landmark and semi-landmark positions on the cranium, in (A) ventral, (B) dorsal and
666 (C) lateral views, shown on a Fire salamander *Salamandra salamandra gigliolii* (NHM
667 1911.2.22.62). Points are colour-coded as follows: landmarks (red), curve semilandmarks
668 (yellow) and surface semilandmarks (blue).

669

670 **Figure 2.** Hypotheses of modular patterns tested in EMMLi analysis. All of the landmarks and
671 semi landmarks from each region were assigned to hypothesised modules and color-coded
672 accordingly. **A**, no module model (no modular organisation); **B**, Developmental module model
673 (purple: dermal, blue: endochondral); **C**, Dorso-ventral module model (blue: dorsal, yellow:
674 ventral); **D**, 4 functional module model (blue: snout, yellow: braincase, green: jaw suspension,
675 orange: occipital-otic complex); **E**, time of ossification module model (blue: early
676 development, green: mid-development, light pink: late development, yellow:
677 metamorphosis); **F**, 7 functional module model (navy blue: snout, light blue: floor of the
678 braincase, light pink: jaw suspension, yellow: skull roof, magenta: palatine, green: occipital-
679 otic complex); **G**, bone module model (frontal: dark blue, maxilla: blue, nasal: yellow;
680 occipital: peach, occipital condyle: grey, orbitosphenoid: black, otic: green khaki,
681 parasphenoid: green, parietal: violet, prefrontal: red, premaxilla: brown, pterygoid: egg shell,
682 quadrate: cyan, squamosal: gold, vomer: hot pink); **H**, Functionally partitioned module model
683 (frontal: dark blue, jaw joint: grey, maxilla: blue, maxilla ventral: cyan, nasal: yellow; occipital
684 dorsal: egg shell, occipital ventral: black, occipital condyle: peach, orbitosphenoid: light
685 orange, otic dorsal: light blue, otic ventral: black, parasphenoid: green, parietal: dark violet,
686 prefrontal: orange, premaxilla dorsal: dark gold, pterygoid: golden brown, quadrate: light

687 violet, squamosal: green khaki , vomer: magenta). For further details about the results on
688 modularity, please see Table 3, 4 and S3-S5).

689

690 **Figure 3.** The thirteen-module model identified with EMMLi analysis. Top left, network graph
691 of the 20 cranial regions defined in this study, coloured in the thirteen modules identified
692 from EMMLi analysis. The network displays a right-hand side lateral view of the skull. The
693 thickness of a line and the size of a circle respectively correspond to the strength of
694 integration between and within modules. At the right side of the figure, the resulting thirteen
695 modules are visualised on a specimen (*Salamandra salamandra gigliolii* NHM 1911.2.22.62)
696 in lateral, ventral and dorsal views. The thirteen modules correspond to the following regions:
697 occipital in light blue, dorsal (OccD) and ventral (OccV) part of the occipital bone and condyle
698 (OC); dorsal otic (oticD) in yellow and ventral otic (OticV) in light brown; jaw suspension
699 module with squamosal (Sq), quadrate (Qd), jaw joint (JJ) and pterygoid (Pt) in red;
700 parasphenoid (Psph) in black; vomer (Vo) in magenta; orbitosphenoid (Osph) in orange;
701 prefrontal (Prf) in green; Maxilla in blue with the dorsal (MaxD) and ventral (MaxV) sides;
702 premaxilla in brown with dorsal (PmxD) and ventral (PmxV) sides; nasal (Nas) in pale pink;
703 frontal (Fr) in dark blue; Parietal (Par) in dark purple.

704

705 **Figure 4.** Linear regression of Procrustes variances corrected for landmark number on
706 respective within-module integration. The relationship between the two variables is not
707 significant ($p = 0.76$).

708

709