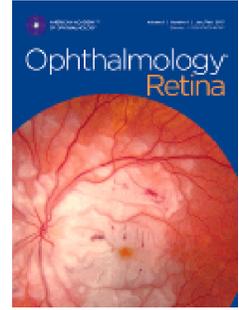


# Journal Pre-proof

Enhanced S-cone syndrome: spectrum of clinical, imaging, electrophysiological and genetic findings in a retrospective case series of 56 patients

Emanuel R. de Carvalho, Anthony G. Robson, Gavin Arno, Camiel Boon, Andrew A. Webster, Michel Michaelides



PII: S2468-6530(20)30286-4

DOI: <https://doi.org/10.1016/j.oret.2020.07.008>

Reference: ORET 847

To appear in: *Ophthalmology Retina*

Received Date: 9 May 2020

Revised Date: 6 July 2020

Accepted Date: 9 July 2020

Please cite this article as: de Carvalho E.R., Robson A.G., Arno G., Boon C., Webster A.A. & Michaelides M., Enhanced S-cone syndrome: spectrum of clinical, imaging, electrophysiological and genetic findings in a retrospective case series of 56 patients, *Ophthalmology Retina* (2020), doi: <https://doi.org/10.1016/j.oret.2020.07.008>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© YEAR Published by Elsevier Inc. on behalf of American Academy of Ophthalmology

- 1 **Enhanced S-cone syndrome: spectrum of clinical, imaging, electrophysiological and**
- 2 **genetic findings in a retrospective case series of 56 patients**

Journal Pre-proof

3 **Enhanced S-cone syndrome: spectrum of clinical, imaging, electrophysiological and**  
4 **genetic findings in a retrospective case series of 56 patients**

5

6 Emanuel R. de Carvalho<sup>a,b</sup>, Anthony G. Robson<sup>a,b</sup>, Gavin Arno<sup>a,c</sup>, Camiel Boon<sup>b,d</sup>, Andrew A.  
7 Webster<sup>a,c</sup>, Michel Michaelides<sup>a,c</sup>

8

9 <sup>a</sup> Moorfields Eye Hospital, City Road, London, United Kingdom

10 <sup>b</sup> Department of Ophthalmology, Amsterdam University Medical Centers, Amsterdam, the  
11 Netherlands

12 <sup>c</sup> UCL Institute of Ophthalmology, University College London, London, United Kingdom

13 <sup>d</sup> Department of Ophthalmology, Leiden University Medical Center, Leiden, the Netherlands

14

15 **Conflict of Interest:** No conflicting relationship exists for any author.

16 **Corresponding Author:**

17 Professor Michel Michaelides, MD, FRCOphth

18 UCL Institute of Ophthalmology, London EC1V 9EL, United Kingdom

19 Email: [michel.michaelides@ucl.ac.uk](mailto:michel.michaelides@ucl.ac.uk)

20 **Financial Support:** Supported by grants from the National Institute for Health Research  
21 Biomedical Research Centre at Moorfields Eye Hospital NHS Foundation Trust and UCL  
22 Institute of Ophthalmology, Fight for Sight, Moorfields Eye Charity, Retina UK, and the  
23 Foundation Fighting Blindness (USA).

24

25 **Running Head:** Clinical Spectrum of Enhanced S-Cone Syndrome

26

27 This article contains additional online-only material. The following should appear online-  
28 only: Supplemental Figures 1 and 2 and Supplemental Tables 1 and 2.

29 **Abstract**

30 **Purpose:** To describe the detailed phenotype, long-term clinical course, clinical variability,  
31 and genotype of patients with Enhanced S-Cone Syndrome (ESCS).

32 **Design:** Retrospective case series.

33 **Participants:** Fifty-six patients with ESCS.

34 **Methods:** Clinical history, examination, imaging and electrophysiological findings of 56  
35 patients (age range 1 – 75 years) diagnosed with ESCS were reviewed. Diagnosis was  
36 established by molecular confirmation of disease-causing variants in the *NR2E3* gene (n = 38)  
37 or by diagnostic full-field electroretinography (ERG) findings (n = 18).

38 **Main outcome measures:** Age at onset of visual symptoms, best-corrected visual acuity  
39 (BCVA), quantitative age-related electrophysiological decline and imaging findings.

40 **Results:** The mean age at onset of visual symptoms was 4.0 years, and median age at  
41 presentation was 20.5 years, with the mean follow-up interval being 6.1 years. Six patients  
42 were assessed once. Disease-causing variants in *NR2E3* were identified in 38 patients. The  
43 mean logMAR BCVA of the better-seeing eye was 0.32 at baseline and 0.39 at follow-up.  
44 BCVA remained stable in the majority of eyes (76%, 76/100), with a mean BCVA change of  
45 0.07 logMAR during follow-up. Nyctalopia was the commonest initial symptom, reported in  
46 92.9% (52/56) of patients. Clinical findings were highly variable, and included foveomacular  
47 schisis (41.1%, 26/56), yellow/white dots (57.1%, 32/56), nummular pigmentation (85.7%,  
48 48/56), torpedo-like lesions (10.7%, 6/56) and circumferential subretinal fibrosis (7.1%,  
49 4/56). Macular and peripheral patterns of autofluorescence were classified as (i) minimal  
50 change, (ii) hypoautofluorescent (mild diffuse; moderate speckled; moderate diffuse;  
51 advanced), or (iii) hyperautofluorescent flecks. One patient had undetectable ERGs;  
52 quantification of the main ERG components in all other patients revealed amplitude and peak  
53 time variability, but with pathognomonic ERG features. The main ERG components showed

54 evidence of age-related worsening over 6.7 decades, at a rate indistinguishable from that seen  
55 in unaffected control subjects. Eighteen sequence variants in *NR2E3* were identified,  
56 including four novel missense changes.

57 **Conclusions** ESCS has a highly variable phenotype with relative clinical and imaging  
58 stability over time. The ERGs have pathognomonic features in most, but quantitative  
59 assessment reveals variability and a normal mean rate of age-related decline, consistent with  
60 largely non-progressive peripheral retinal dysfunction.

61

Journal Pre-proof

**62 Introduction**

63 Enhanced S-Cone Syndrome (ESCS) (OMIM 268100) is an autosomal-recessive retinal  
64 dystrophy caused by disease-causing variants in nuclear receptor subfamily 2, group E,  
65 member 3 (*NR2E3*), a member of the nuclear hormone receptor superfamily of ligand-  
66 modulated transcription factors; also known as photoreceptor-specific nuclear receptor (PNR;  
67 OMIM 604485).<sup>1-4</sup> Goldmann-Favre Syndrome has also been shown to be caused by biallelic  
68 variants in *NR2E3*, rendering distinction between the two entities redundant.<sup>5-9</sup> Similarly,  
69 recessive variants in *NR2E3* have been described in cases of clumped pigmentary retinal  
70 degeneration.<sup>6</sup> A single missense *NR2E3* variant (p.G56R) has also been linked to autosomal  
71 dominant retinitis pigmentosa (OMIM 611131).<sup>10, 11</sup>

72 *NR2E3* was first identified by its homology to *NR2E1*, which acts on cell-fate determination  
73 in *Drosophila* and encodes an orphan receptor of the steroid/thyroid hormone receptor  
74 superfamily of ligand-activated transcription factors.<sup>12, 13</sup> In the eye, *NR2E3* regulates the fate  
75 of retinal progenitor cells during retinogenesis.<sup>14-17</sup> The different cell subtypes in the  
76 vertebrate retina derive from a common population of multipotent progenitors.<sup>18, 19</sup> Cone  
77 primordial cells arise earlier than rod cells.<sup>16, 20</sup> Cellular interactions between cones dictate the  
78 ensuing spatial rearrangement, opsin expression, and ratio of photoreceptor subtypes in the  
79 mature retina. Disease-causing variants in *NR2E3*, expressed in late retinal progenitors and  
80 differentiating photoreceptors in the outer nuclear layer of the retina, disrupt the determination  
81 of photoreceptor cell-fate, affecting the normal ratio and topographical distribution of the  
82 different photoreceptor subtypes in the mature retina.<sup>21, 22</sup> S-cones are expressed earlier than  
83 M (medium wavelength)- and L (long wavelength)- cone photoreceptors and are therefore  
84 regarded as the default primordial cone cells.<sup>22</sup> As a result, in the absence of *NR2E3*, rods  
85 develop into non-functional hybrid photoreceptors and L- and M-cone expression is  
86 suppressed with a concomitant over-expression of ancestral S-cones.<sup>22 15, 16, 20, 23-25</sup>

87 The unique photoreceptor arrangement in patients harboring *NR2E3* variants is responsible for  
88 the increased sensitivity to blue light<sup>26</sup> and is often reflected by pathognomonic full-field  
89 electroretinography (ERG) responses. The dark-adapted (DA) rod-specific dim flash  
90 (DA0.01) ERG is typically undetectable; although detectable responses have been reported in  
91 mild ESCS, which has been suggested to stem from functional dimerization of *NR2E3*  
92 mediated by ligand-binding domain variants.<sup>27-29</sup> Responses in the retina are dominated by  
93 short-wavelength-sensitive mechanisms, leading to a similar simplified and severely delayed  
94 waveform under DA and light-adapted (LA) conditions, with a severely abnormal LA30Hz  
95 flicker ERG.<sup>21, 30-33, 90</sup> Short-wavelength-specific stimulation may elicit a high amplitude  
96 response when compared to those of normal subjects, consistent with the increased number of  
97 S-cone photoreceptors.<sup>23</sup>

98 Previously reported symptoms of patients with ESCS include nyctalopia, variable visual  
99 acuity loss and constricted field of vision.<sup>33-35</sup> The clinical signs encompass a combination of  
100 yellow/white dots, nummular pigmentation at the level of the retinal pigment epithelium  
101 (RPE), especially along the temporal vascular arcades, and variable degrees of foveomacular  
102 schisis.<sup>31, 33, 35-40</sup> Other signs include torpedo-like retinal lesions, cystoid macular edema and  
103 circumferential subretinal fibrosis, the latter thought to occur secondary to choroidal  
104 neovascularization.<sup>41, 42</sup> Although clinical and electroretinographic characteristics are well-  
105 recognised, published analyses are often qualitative and there is a lack of data relating to the  
106 natural history of the disorder.

107 The purpose of the present study was to retrospectively review clinical and  
108 electrophysiological data of a large cohort of patients diagnosed with ESCS to better define  
109 variability of the phenotype, long-term visual outcome, severity and stability of retinal  
110 dysfunction, and the nature of *NR2E3* disease-causing variants.

111

**112 Patients and methods**

113 A cohort of 56 patients with a clinical diagnosis of ESCS were ascertained at Moorfields Eye  
114 Hospital (n = 45) and at the Expertise Center for Hereditary Retinal Diseases of Amsterdam  
115 University Medical Centers/Leiden University Medical Center (n = 11), with a mean follow-  
116 up time of 6.1 years (range 0 – 34 years). All patients were first diagnosed between 1984 and  
117 2018, with the latest examination performed in 2019. A baseline ERG was performed in 31  
118 patients and repeated in 3 patients. The cohort included 3 cases of pseudo-dominance with  
119 consanguineous parents, and 5 sibships (4 sibling pairs, 1 sibling pair with an affected parent,  
120 and 2 pairs of 1 affected parent and 1 affected child). Molecular confirmation of the diagnosis  
121 was established in 43 patients, and 24 of these underwent baseline ERGs. The diagnosis was  
122 established on pathognomonic ERG responses and phenotypical retinal changes in the  
123 remaining 13 patients. The protocol of the study adhered to the tenets of the Declaration of  
124 Helsinki and was approved by the Ethics Committee of all involved institutions. Thirteen  
125 cases were described previously but without detailed ERG quantification and longitudinal  
126 data.<sup>33, 43, 44</sup>

127

**128 Clinical Assessment**

129 Fifty-six patients were ascertained. Six patients were assessed on a single occasion, and all  
130 others on at least 2 occasions. In the latter group, the initial and last visits were taken as  
131 baseline and follow-up examinations, respectively. Follow-up time was determined by the  
132 interval between age at baseline and age at the latest follow-up examination.

133 Color contrast sensitivity was assessed in 12 patients along tritan, protan and deutan axes  
134 using the “ChromaTest”, involving the use of colored optotypes presented on a randomized  
135 luminance noise background.<sup>45, 46</sup> In all patients, a medical history was obtained and a  
136 comprehensive ophthalmologic examination performed which included best-corrected Snellen

137 visual acuity converted to equivalent logarithm of minimal angle of resolution (logMAR) for  
138 the purpose of data analysis.<sup>47</sup> Retinal fundus photographs were obtained by conventional 35°  
139 fundus color photographs (Topcon Europe Medical BV, Capelle aan den IJssel, the  
140 Netherlands) or wide-field confocal scanning laser imaging (Optos PLC, Dunfermline, UK).  
141 Spectral-domain optical coherence tomography (SD-OCT, Heidelberg Engineering,  
142 Heidelberg, Germany) macular scans were performed in all patients.

143 The patterns of macular and peripheral fundus autofluorescence (FAF) were assessed in 49  
144 pairs of eyes. Macular FAF images were obtained using a confocal scanning laser  
145 ophthalmoscope with blue light excitatory beam (Spectralis, Heidelberg Engineering). When  
146 available, peripheral FAF was analyzed with wide-field Optos imaging with green light  
147 excitatory beam. Specific macular and peripheral FAF patterns were classified as: (i) no  
148 change; (ii) hypoautofluorescence - minimal change pattern, mild diffuse, moderate speckled,  
149 moderate diffuse (mid-peripheral half-ring or ring  $\leq 5000\mu\text{m}$  widest diameter), moderate  
150 diffuse  $> 5000\mu\text{m}$  hypoautofluorescence, nummular (patchy); and (iii) hyperautofluorescent  
151 flecks.

152

### 153 **Electrophysiology**

154 A total of 32 patients underwent electrophysiological assessment at Moorfields Eye Hospital  
155 (age range 6 – 73 years at the time of testing). The electrophysiological assessment included  
156 full-field ERG and pattern electroretinography (PERG), incorporating the minimum standards  
157 of the International Society for Clinical Electrophysiology of Vision (ISCEV)<sup>48, 49</sup> and  
158 recorded using gold foil corneal electrodes. Additionally, 28 patients underwent short-  
159 wavelength flash ERG (S-cone ERG),<sup>50</sup> obtained using a blue stimulus (445 nm, 80 cd/m<sup>2</sup>;  
160 stimulus duration 5 ms) on a constant orange background (620 nm, 560 cd/m<sup>2</sup>).<sup>33</sup> S-cone ERG  
161 peak times were measured to the second or single positive peak; amplitudes were measured

162 from baseline to the single or second positive peak<sup>50</sup> or, if an early negative trough was  
163 present, as a trough to peak amplitude to better characterize the magnitude of responses.

164 The patient data were compared with the control (normative) electrophysiological data  
165 obtained from 160 healthy subjects (age range 10 – 79 years) which included validated  
166 recordings for DA0.01 (n = 117), DA 10.0 (n = 141), LA 3.0 30 Hz (n = 131), and the LA 3.0  
167 (single flash cone) ERG (n = 109).

168 The amplitude and peak time ratios between the LA 3.0 ERG a-wave and LA 30 Hz were  
169 calculated for each patient and these and other main ERG components compared with age  
170 and the control data.

171 A total of 3 patients seen at the Amsterdam University Medical Centers underwent baseline  
172 electrophysiological assessment. Electrophysiological data concerning these patients were  
173 excluded from analysis, given that flash ERGs were performed according to older or  
174 abbreviated protocols using silver thread electrodes, precluding comprehensive ERG  
175 phenotyping and direct comparison with ISCEV-standard recordings.

176

### 177 **Genetic screening**

178 Patients were screened for disease-causing variants by direct sequencing of all 8 exons and  
179 intron-exon boundaries of *NR2E3*. Subsequently, available relatives also underwent  
180 sequencing. Genomic DNA was isolated from peripheral blood lymphocytes using a kit  
181 (Gentra Puregene blood extraction kit; Qiagen). DNA was amplified using specifically  
182 designed primers by polymerase chain reaction, and the polymerase chain reaction fragments  
183 were sequenced using standard protocols (details are available from the author on request).

184 The likely pathogenicity of novel missense variants was assessed using the predictive  
185 algorithms of InterVar [according to the guidelines by the American College of Medical  
186 Genetics and Genomics (ACMG) and the Association for Molecular Pathology (AMP) for

187 interpretation of causality of sequence variants, <http://wintervar.wglab.org>], PROVEAN  
188 (Protein Variation Effect Analyzer (<http://provean.jcvi.org/index.php>)<sup>52-54</sup> and PolyPhen-2  
189 (<http://genetics.bwh.harvard.edu/pph2>).<sup>55</sup> Information regarding the domain structure of  
190 NR2E3 was retrieved using UniProtKB - Q9Y5X4 (NR2E3\_HUMAN). To predict the  
191 consequences of the ligand-binding domain missense mutations in the 3-dimensional space,  
192 we analyzed the crystal structure of apo NR2E3 ligand-binding domain with pdb code 4LOG,  
193 retrieved from the SWISS-MODEL server.<sup>56-59</sup>

194

### 195 **Statistical Analysis**

196 The mean, standard error of mean, median, standard deviation and range were used as  
197 appropriate. Best-corrected visual acuities (BCVA) were ascertained and converted to  
198 logarithm of the minimum angle of resolution (logMAR) scale for statistical analysis.<sup>47</sup> Mean  
199 BCVA change over follow-up was calculated per each eye, right and left, using the related  
200 samples Wilcoxon signed rank test with  $P < 0.05$  deemed clinically significant. Variability  
201 between BCVA in the right and left eye recordings at baseline and follow-up was assessed  
202 using the paired samples Wilcoxon signed rank test, with  $P < 0.05$  deemed clinically  
203 significant. Age was correlated with BCVA at baseline and follow-up applying a Spearman  
204 correlation model with  $P < 0.05$  deemed clinically significant. The relationship between  
205 visual acuity and electrophysiological responses was assessed by multiple linear regression  
206 analysis and  $P < 0.05$  was considered statistically significant. Pearson correlation coefficients  
207 were calculated to compare the PERG P50 measure of macular function with central visual  
208 acuity and age. Photopic ERG a-wave / 30Hz flicker ratios were compared with unpaired  
209 Mann Whitney t-test. Statistical analyses were performed using IBM SPSS ver. 25.0 (IBM  
210 Corp., Armonk, NY, USA) and GraphPad Prism 8 (GraphPad Software, San Diego, CA,  
211 USA).

212

213 **Results**214 **Clinical Findings**

215 Fifty-six patients including 33 female (59%) and 23 male (41%) were included. Thirty  
216 patients had European ethnic origin (54%), 20 had Middle Eastern ethnic origin (36%), 4 had  
217 South Asian ethnic origin (7%) and 2 had Black ethnic origin (3%). Nyctalopia was reported  
218 as the first symptom in 52 patients (93%), with or without reduced central visual acuity. The  
219 remaining 4 patients described reduced central acuity without nyctalopia as their initial  
220 symptom. At presentation, a manifest squint was observed in 9 patients and nystagmus was  
221 recorded in 3 patients. Refractive assessment was conducted in 21 patients; hyperopia in 12  
222 (57.1%), myopia in 6 (28.5%) and plano in 3 (14.2%). Color contrast sensitivity was tested in  
223 9 patients; 6 patients had relative sparing of the tritan axis and moderately elevated protan and  
224 deutan thresholds, 1 patient had non-specific dyschromatopsia, and 2 had normal results.

225 The median age at onset of visual symptoms was 4.0 years (range, 0 – 27 years). The median  
226 age of presentation to the eye clinic (baseline) and follow-up were 20.5 years (range, 1 – 75  
227 years) and 33.0 years (range, 2 – 81 years), respectively. The mean follow-up interval was 6.1  
228 years (range, 0 – 34 years). Six patients were assessed once. Twenty-eight patients (50%)  
229 presented before 21 years of age. Thirteen patients presented after 50 years of age (23%).

230 The mean logMAR BCVA of the better-seeing eye at baseline and at follow-up were 0.32  
231 [standard error of mean (SEM) 0.045; range, 0.0 – 1.77] and 0.39 (SEM 0.054; range, 0.0 –  
232 1.60), respectively (Table 1). No clinically significant difference was found between BCVA  
233 in the right and left eyes ( $P = 0.14$  for baseline BCVA,  $P = 0.22$  for follow-up BCVA).

234 Overall, mean logMAR BCVA change was 0.01 logMAR (SEM 0.05), although this included  
235 cases with short follow-up time (< 6 years). In the group with extended follow-up time ( $\geq 6$   
236 years,  $n = 21$ ) mean logMAR BCVA change was 0.12 (SEM 0.09, Figure 1). A score of

237 BCVA severity was attributed for each eye and progression over time was assessed for each  
238 eye separately. Severity was graded as very mild ( $\log\text{MAR BCVA} \leq 0.1$ ,  $n = 7$ , 12.5%), mild  
239 ( $\leq 0.3$ ,  $n = 21$ , 37.5%), moderate ( $\leq 0.6$ ,  $n = 12$ , 21.4%), severe ( $\leq 1$ ,  $n = 11$ , 19.6%) and very  
240 severe ( $> 1.0$ ,  $n = 5$ , 9%). No progression was observed in 79 of 100 eyes. Ten eyes progressed  
241 from very mild and mild severity scores to moderate severity (10%). Five eyes progressed  
242 from mild and moderate severity scores to severe (5%), and six eyes progressed from severe  
243 to very severe (6%). Poor visual acuity ( $\log\text{Mar BCVA} > 0.6$ ) was observed in 16 patients at  
244 baseline (28.5%, median age 29.0 years, range, 0.5 – 51 years) and in 18 patients at last  
245 follow-up (36%). In two patients, BCVA loss in one eye could be directly attributed to other  
246 significant ophthalmic events, namely retinal detachment and dense amblyopia. In five other  
247 patients, BCVA loss could be partly ascribed to concurrent ophthalmic pathology. In one  
248 patient with undetectable cone and rod ERG responses, optic disc pallor was observed at  
249 initial presentation. The patient was assessed by neuro-ophthalmology and no underlying  
250 neuro-ophthalmic aetiology was found for the optic neuropathy. In 6 patients with  
251 documented BCVA worsening over time, no other unrelated significant ophthalmic events  
252 were reported. The majority of patients (13/18) with severe visual outcomes had moderate to  
253 advanced foveomacular schisis with accompanying or ensuing macular atrophy in 8 patients.  
254 Three patients presented with advanced macular atrophy at initial visit. Cystoid macular  
255 edema was confirmed on fluorescein angiography in one patient, and diagnosed based on  
256 structural OCT appearance in three other patients with severe visual outcomes. Treatment  
257 with oral carbonic anhydrase inhibitors was attempted in 4 patients, with a positive  
258 anatomical response attained in two patients, albeit with no visual improvement noted post-  
259 treatment. One patient was diagnosed with congenital nystagmus at birth, with delayed visual  
260 development that might have contributed to poor visual outcome. There was no significant

261 correlation between PERG responses and clinical severity. Clinical findings are summarized  
262 in Tables 1, 2 and 3.

263

#### 264 **Clinical and Fundus Autofluorescence Features**

265 Clinical notes, imaging data and color fundus photographs were reviewed in all patients.

266 In most cases, yellow-white dots and/or nummular pigmentation were observed, located  
267 within the vascular arcades in 16 patients (28.6%), and in the mid-peripheral retina, outside  
268 the vascular arcades in 49 patients (87.5%, Figure 2A-E). Nummular pigmentation,  
269 characterized by deep round-shaped pigmentation at the level of the RPE, was usually located  
270 in the mid-peripheral retina, along the vascular arcades and often associated with RPE atrophy  
271 and end-stage hypoautofluorescence (Figure 4J). This was the most common clinical finding,  
272 present in 48 patients (85.7%), followed by yellow-white dots which were seen in 32 patients  
273 (57.1%). The combined presence of nummular pigmentation with yellow dots was observed  
274 in 26 patients (46.4%). Two patients developed clumped and nummular pigmentary changes  
275 in an area of the mid-peripheral retina with yellow-white dots (Figure 2K, L).

276 Other clinical findings included vitreous opacities (21.4%), peripheral torpedo-like lesions  
277 (10.7%, Figure 2F), circumferential subretinal fibrosis (7.1%, Figure 2G), optic disc pallor  
278 (5.4%), and recurring vitreous hemorrhages secondary to pre-retinal neovascularization in one  
279 patient (1.8%, Figure 2I). Color fundus photographs and optical coherence tomography scans  
280 of a representative group of patients depict the above-mentioned clinical features in Figures 2  
281 and 3.

282 Foveomacular schisis was identified in 23 of 56 patients (41%, Figure 3B and 3C). Two  
283 patients developed giant foveomacular schisis (Figure 3D), which evolved into advanced  
284 macular atrophy in one eye (Figure 3F and 4E). In all other patients, OCT appearances were  
285 relatively stable. The schitic cavities were located at the level of the inner nuclear layer,

286 characterized by a round shape, and in the outer nuclear layer (ONL) where they appeared  
287 elongated in a stellate-like configuration (Figure 3C). A total of 16 patients (28.6%,) were  
288 diagnosed with presumed cystoid macular edema (CME), based on structural appearance on  
289 SD-OCT. Four cases were treated with oral carbonic anhydrase inhibitors. Resolution of CME  
290 was attained in two patients, albeit with no visual gain, including in the single patient where,  
291 prior to treatment, leakage had been demonstrated on fluorescein angiography.

292 The majority of patients (55%) presented with minimal autofluorescent changes at the level of  
293 the macula (minimal change pattern), combined with the presence of hyperautofluorescent  
294 flecks in some cases. End-stage macular atrophy with secondary severe macular  
295 hypoautofluorescence was observed in one patient and this was preceded by giant  
296 foveomacular schisis. The hyperautofluorescent flecks correlated with the presence of  
297 yellow/white dots (Figure 4K and 4L). In the peripheral retina, moderate decrease in  
298 autofluorescence was observed in the majority of patients (31.7%), usually combined with  
299 patchy severe hypoautofluorescence, the latter corresponding to the presence of nummular  
300 pigmentary deposition. A strong half-ring of pronounced hyperautofluorescence along the  
301 temporal macular rim was observed in all cases presenting with peripheral half-ring  
302 hypoautofluorescence. In 5 patients, the peripheral FAF pattern progressed from moderate  
303 decrease in autofluorescence to patchy decrease in autofluorescence, with documented  
304 progression of pigmentary changes. Clinical FAF patterns are shown in Figure 4 and  
305 summarized in Supplemental Table 1 (available at <https://www.opthalmologyretina.org>),  
306 where the mean BCVA is presented in relation to the pattern of macular FAF.

307

### 308 **Electrophysiological Findings**

309 The PERG P50 component (Supplemental Figure 1, available at  
310 <https://www.opthalmologyretina.org>) was undetectable (n = 11), delayed and reduced (n =

311 11), delayed and of normal amplitude ( $n = 3$ ; see Figure 5 for an example) or normal ( $n = 3$ ).  
312 The threshold values for the PERG P50 minimum amplitude/maximum peak time are  
313 presented in Supplemental Table 2 (available at <https://www.opthalmologyretina.org>). There  
314 was no correlation between the PERG P50 amplitude and visual acuity in right ( $r = -0.20$ ,  $P =$   
315  $0.277$ ;  $n = 31$ ) or left eyes ( $r = -0.18$ ,  $P = 0.326$ ;  $n = 31$ ). There was significant negative  
316 correlation between age and PERG P50 amplitude for right ( $r = 0.65$ ,  $P < 0.05$ ;  $n = 36$ ) and  
317 left eyes ( $r = 0.6$ ;  $P < 0.05$ ) and positive correlation between age and P50 peak time for right  
318 ( $r = 0.6$ ;  $P < 0.05$ ;  $n = 23$ ) and left eyes ( $r = 0.55$ ;  $P < 0.05$ ).

319 The full-field ERG waveforms were undetectable in one patient with a genetically confirmed  
320 diagnosis. All other individuals that underwent ISCEV-standard testing ( $n = 31$ ) had  
321 pathognomonic ERG abnormalities, as described below (typical recordings are shown in  
322 Figure 5). The DA3.0, DA10.0 and LA3.0 ERGs had a similar simplified and delayed  
323 waveform shape. The ranges of full-field ERG component amplitudes and peak times are  
324 compared with those in a control group in Figures 6 and 7. The rod-specific (DA0.01) ERG  
325 was undetectable in all but one patient (age 14 years) with a detectable but subnormal  
326 response (reduction 58% compared with the mean for the control group). The DA10.0 ERG a-  
327 and b-wave mean amplitudes were reduced by 52% and 63% respectively and mean a- and b-  
328 wave peak times were 16 ms and 14 ms longer respectively compared with those for the  
329 control group (Figure 6).

330 The LA 30Hz ERG and LA3.0 ERG a- and b-wave amplitudes were on average 93%, 28%  
331 and 69% lower respectively and mean peak times 14 ms, 8 ms and 20 ms greater respectively  
332 compared with mean values for the control group (Figure 7). The LA30Hz flicker ERG was  
333 smaller than the LA3.0 ERG a-wave in the majority ( $n = 48$  eyes of 27 subjects) and of equal  
334 amplitude to the LA3.0 ERG a-wave in others ( $n = 9$  eyes of 7 subjects) including the 4 eyes  
335 with the smallest detectable responses. The mean amplitude ratio between the LA3.0 ERG a-

336 wave and LA30Hz ERG was 1.86 ( $n = 30$ ; 44.53% coefficient of variation,  $SD = 0.9$ , Figure  
337 8) in the ESCS cohort and 0.37 ( $n = 111$ , 24.38% coefficient of variation,  $SD = 0.09$ ) in the  
338 healthy controls. The peak time ratio between the LA3.0 ERG a-wave and LA30Hz ERG was  
339 0.55 ( $n = 30$ , 14.02 % coefficient of variation,  $SD = 0.08$ , Figure 8) in the ESCS cohort ( $P =$   
340 0.0001) and 1.85 ( $n = 42$ , 5.0 % coefficient of variation,  $SD = 0.09$ ) in the healthy controls.

341 The mean S-cone ERG amplitude in the ESCS patients was greater (mean 81  $\mu V$ , median 54  
342  $\mu V$ ,  $n = 28$ , mean age 27 years) than in the control group (mean 43.35  $\mu V$ , median 43  $\mu V$ ,  $n =$   
343 51, mean age 29 years, Figure 9A) and peak times were severely delayed (mean peak time  
344 difference (ESCS - control) = 28.3 ms, Figure 9B). S-cone ERGs in ESCS were largest in  
345 some of the children and young adults but there was no significant correlation between  
346 amplitude or peak time and age ( $r^2 = 0.06$  and  $r^2 = 0.001$  respectively,  $P > 0.05$ , Figures  
347 9A,9B). In ESCS there was significant correlation between the S-cone ERG and LA3 ERG b-  
348 wave amplitudes ( $r^2 = 0.56$ ,  $P < 0.001$ ) and peak times ( $r^2 = 0.34$ ,  $P < 0.05$ , Figures 9C, 9D).

349 Plots of the major ISCEV-standard ERG component amplitudes and peak times against age  
350 are shown in Figures 6 and 7 . There was evidence of age related ERG reduction in the DA  
351 and LA ERGs at a rate that was indistinguishable from that seen in healthy subjects, over 6.7  
352 decades (LA3.0 ERG a-wave,  $r^2 = 0.22$ ,  $P = 0.006$ ; LA3.0 ERG b-wave,  $r^2 = 0.22$ ,  $P = 0.007$ ;  
353 DA10 a-wave,  $r^2 = 0.17$ ,  $P = 0.02$  and DA10 b-wave,  $r^2 = 0.16$ ,  $P = 0.03$ ). The peak times of  
354 the major ERG components in ESCS showed high stability with increasing age, as in the  
355 control group.

356 The patient with compound heterozygous changes in *NR2E3* (c.119-2A>C and the novel  
357 p.L303P) had a particularly severe clinical phenotype with early onset of visual symptoms,  
358 severely reduced visual acuity (1.0 logMAR), sensory nystagmus and giant foveomacular  
359 schisis which evolved into end-stage macular atrophy (Supplemental Figure 2, available at  
360 <https://www.ophtalmologyretina.org>). His ERGs, performed at the age of 53, differed from

361 all other patients, characterized by undetectable DA and LA ERGs and an undetectable  
362 PERG.

363 Five individuals underwent follow-up ERG testing after intervals of 4, 6, 9, 10 and 17 years.  
364 The mean annual rate of ERG reduction (averaged between eyes) for DA10 ERG a- and b-  
365 wave amplitudes was 1.6% (range 0-6.0 %) and 3.9% (range 0-6.2 %) respectively; for LA3  
366 ERG a- and b-wave amplitudes the mean rate of reduction was 3.4% (range 2.0-4.7 %) and  
367 1.3% (range 0-2.0 %) respectively.

368

### 369 **Molecular Genetics**

370 Forty-one out of 56 patients underwent screening of the nine coding exons of *NR2E3*.  
371 Disease-causing variants were identified in 41 subjects. Twenty-four subjects were  
372 homozygous and 17 had compound heterozygous variants. Eighteen sequence variants were  
373 identified, including four novel missense variants (p.F71L, p.R247W, p.L303P, p.R309Q)  
374 (Table 4). The other reported variants encompassed two splice acceptor variants in intron 1  
375 (c.119-2A>C<sup>22</sup> and c.119-3C>G<sup>33</sup>), ten missense mutations (p.R76Q<sup>22</sup>, p.C83Y<sup>60</sup>, p.A102D<sup>44</sup>,  
376 p.R104W<sup>22</sup>, p.G216S, p.R104Q<sup>27</sup>, p.R311Q<sup>16</sup>, p.A256E<sup>5</sup>, p.V342A<sup>44</sup>, p.L371W<sup>61</sup>), one  
377 frameshift mutation (p.P399Qfs\*3<sup>44</sup>) and a 9-bp deletion leading to deletion of 3 amino acid  
378 residues (p.C67\_G69del<sup>16</sup>).

379 The p.G216S substitution (c.646G>A; exon 5) was found as a homozygous change in one  
380 patient. This variant, predicted to be benign, is rare in gnomAD, but the amino acid change is  
381 not predicted to be damaging by any of the in silico tools utilised. This, however, may be  
382 irrelevant to causality. The variant introduces an exonic splice acceptor site: TGCGGCC >  
383 tgcagCC (human splice finder [HSF] score: 91.23, nnsplce score 0.97) into exon 5 which is  
384 likely to lead to an out of frame deletion of the 5' 77bp of exon 5 and thus may represent a  
385 loss of function allele. Without mRNA analysis of relevant patient tissue, it is not possible to

386 determine if this predicted splice altering effect is indeed occurring in vivo or if any normally  
387 spliced transcript would escape and produce functional protein but we are of the opinion that  
388 the case for causality is sufficient for this rare variant.

389 Four novel disease-causing variants were identified. These are likely to be pathogenic given  
390 that all are located within highly conserved domains critical to protein function, and all are  
391 rare or absent from control datasets. The p.F71L substitution (c.211T>C; exon 2) was found  
392 as a heterozygous change in one patient. The p.R247W substitution (c.739C>T; exon 5) was  
393 found as a heterozygous change in one patient. The p.L303P substitution (c.908T>C; exon 5)  
394 was found as a heterozygous change in one patient. The p.R309Q substitution (c.926G>A;  
395 exon 6) was found in a homozygous state in two affected siblings. The Phe71, Arg247,  
396 Leu309 and Arg309 are highly conserved across NR2E3 orthologues. NR2E3 has the  
397 evolutionarily conserved modular structure of nuclear receptors, namely a highly conserved  
398 DNA-binding domain that specifically binds to consensus binding sites located in promoters  
399 of target genes, and a ligand-binding domain.<sup>12, 13, 62</sup> Three of the afore-mentioned novel  
400 mutations, p.R247W, p.L303P and p.R309Q, are located in the evolutionary-conserved ligand  
401 binding domain of NR2E3, in helices 4 and 7, causing a rearrangement of the bulky side  
402 chains and loss of some hydrogen bonds, suggesting a reduction of protein stability (Figure  
403 10). The p.F71L is located in the evolutionary-conserved DNA binding domain of NR2E3.<sup>59</sup>  
404 <sup>63</sup> Definite confirmation of the pathogenicity of the four novel variants remains dependent on  
405 functional studies that would assess the effects of these sequence variants with regards to  
406 NR2E3 stability, targeting, and ability to interact reversibly and effectively with DNA or  
407 ligands.

408

409 **Discussion**

410 This study describes the largest cohort of patients diagnosed with ESCS. We characterize the  
411 clinical variability, and describe molecular characteristics, including four novel variants in  
412 *NR2E3*. Detailed quantification of the electrophysiological findings characterizes the  
413 phenotypic variability of pathognomonic ERG features and assesses the relative stability of  
414 macular and retinal dysfunction over 6 decades, pertinent to possible future interventional  
415 studies.

416 ESCS is characterized by early onset of visual symptoms. In this cohort, all but two patients  
417 experienced symptoms in the first two decades of life, with the majority presenting in  
418 childhood. Nyctalopia, with or without reduced central visual acuity, was the most frequently  
419 described initial complaint. Hyperopia with a variable degree of astigmatism was the most  
420 common refractive error, in accordance with other reports.<sup>5, 33, 64</sup>

421 Sparing along the tritan axis was demonstrated in six patients that underwent color contrast  
422 sensitivity testing, suggesting preservation of short-wavelength discrimination. This is also  
423 consistent with the high amplitude S-cone ERGs seen in many individuals and with high  
424 correlation between S-cone ERGs and LA3 ERGs, likely having identical S-cone-opsin-  
425 mediated origins.

426 Visual function was highly variable amongst patients, ranging from normal to severely  
427 reduced (2.0 logMAR). It is noteworthy that in most patients, BCVA remained relatively  
428 stable throughout follow-up with no clinical progression observed in 79 of 100 eyes. The  
429 slight deterioration in BCVA with increasing age may be ascribed to the expected age-related  
430 decline in the general population. In two cases, poor visual outcome was related to non-  
431 dystrophic significant ophthalmic events (retinal detachment and dense amblyopia). Poorer  
432 visual outcomes were associated with the presence of moderate to advanced (giant)  
433 foveomacular schisis, but no other association was found, neither with age at onset of visual  
434 symptoms, nor with genotype or electrophysiological responses. There was also a high degree

435 of interocular symmetry, which could enable the use of the contralateral eye as a valid  
436 untreated control in future therapeutic trials in which one eye received treatment.

437 In the family demonstrating a pseudo-dominance pattern, clinical severity was highly  
438 variable. While the father was found to have severely reduced BCVA in both eyes when first  
439 assessed at the age of 17, his children had mildly reduced BCVA when tested at an equivalent  
440 age. Interestingly, clinical presentation was not only variable within the same family, but also  
441 observed in patients from different families harboring the same variant, suggesting that  
442 modifier genes (and environmental factors) may modulate disease outcome.<sup>26, 65</sup>

443 One patient developed bilateral non-diabetic pre-retinal neovascularization and a  
444 midperipheral vasoproliferative lesion in one eye which led to recurrent vitreous hemorrhages  
445 (Figure 2I). This is an unusual finding and it remains unanswered whether this is related to the  
446 underlying retinal dystrophy. Choroidal neovascularization (CNV), on the other hand, has  
447 been previously described in patients with ESCS. Asymptomatic development of CNV has  
448 also been linked to the presence of torpedo-like lesions and circumferential subretinal fibrosis,  
449 both infrequent findings in ESCS.<sup>40, 66-68</sup>

450 Clinical appearance was highly variable, however, three consistent clinical signs were  
451 observed in a large proportion of patients, yellow/white dots, nummular pigmentation at level  
452 of the RPE, and foveomacular schisis. In the appropriate clinical context, the presence of  
453 these combined features should raise the strong possibility of ESCS.

454 The yellow/white dots are often characterized by an increase in autofluorescence signal and  
455 present in both the macula and midperiphery at the level of the RPE. Histological analysis of  
456 autofluorescent white dots seen across the retina of the *rd7* mouse, which harbors a  
457 homozygous deletion in *NR2E3*, showed that the autofluorescence signal arose mostly from  
458 macrophages, which were associated with whorls and rosettes of dysplastic photoreceptors in

459 the outer nuclear layer.<sup>69</sup> Further *in vivo* studies are warranted to ascertain the exact origin of  
460 the hyperautofluorescent dots observed in ESCS patients.

461 Nummular pigmentary deposition alone is not specific to ESCS, and has been described in  
462 other retinal dystrophies such as Bardet-Biedl syndrome,<sup>70</sup> *CRBI*-associated early-onset  
463 severe retinal dystrophy,<sup>71</sup> retinitis pigmentosa with preserved para-arteriolar RPE (*RP12*,  
464 associated with *CRBI*)<sup>72</sup> and thioridazine retinopathy.<sup>73</sup> Whenever present, nummular  
465 pigmentary deposition was associated with disorganization of the neurosensory retina,  
466 including marked loss of the ellipsoid zone and absence of autofluorescence, in keeping with  
467 previous reports.<sup>23, 33, 35</sup> In some patients with mid-peripheral, nummular pigmentation,  
468 clumped pigmentary deposition was observed. The presence of yellow/white dots has been  
469 proposed as a harbinger of more marked pigmentary changes, developing early in the  
470 pathogenesis of the disease, followed by the development of nummular and clumped  
471 pigmentary deposition at a later stage.<sup>5</sup> Corroborating this assumption, documented  
472 progression of pigmentary changes over time was observed in two patients with extended  
473 follow-up. The development of pigmentary changes occurred independent of age. In our  
474 cohort, clumped or nummular pigmentary deposition, although skewed towards older  
475 subjects, was present in 11 young patients (age  $\leq 20$ ) and absent in 4 older patients (age  $> 20$ ),  
476 corroborating the high variability of clinical phenotype.

477 The scarcity of fundus fluorescein angiography in the diagnosis of CME poses an important  
478 limitation, as we are unable to confirm this solely based on SD-OCT structural appearance. It  
479 is possible that the presumed CME documented in many patients represents a variant of  
480 foveomacular schisis that mimics the appearance of cystoid spaces. A positive anatomical  
481 response to carbonic anhydrase inhibitors was observed in solely two patients although this  
482 did not translate into a significant gain in subjective and objective visual function.

483 Notwithstanding, poorer visual outcomes were associated with macular changes, namely

484 foveomacular schisis, presumed CME and macular atrophy, rendering prevention and  
485 treatment of maculopathy an invaluable target in future treatment strategies.

486 Pattern ERGs ranged from undetectable (indicating severe macular dysfunction) in a large  
487 minority to normal (3 of 28 cases), with a higher incidence of P50 delay (Figure 5) than in  
488 many other forms of maculopathy. The PERG P50 did not correlate with BCVA, highlighting  
489 the value of objective assessment of macular function, likely to be of relevance in the  
490 selection of candidates considered amenable to possible future therapeutic interventions.

491 All but 3 eyes of 2 molecularly confirmed patients had pathognomonic ERG features  
492 consistent with ESCS. The full-field ERG findings in the large majority quantified the  
493 magnitude, severity and variability of pathognomonic ERG abnormalities, pertinent to  
494 diagnostic accuracy and precise phenotyping. The rod-system specific (DA0.01) ERG was  
495 undetectable in all but one individual, consistent with a lack of rod function, and the delayed  
496 and simplified stronger flash (DA3.0 and DA10.0) ERGs had qualitative similarities to the  
497 LA3.0 ERG. In any healthy (control) subject, the LA30Hz ERG peak to peak amplitude is  
498 greater than the LA3.0 ERG a-wave and smaller than the LA3 ERG b-wave.<sup>72,73</sup> The LA30Hz  
499 ERG is smaller than the LA3 single-flash cone ERG a-wave in ESCS, as previously reported,  
500 and relating to the minimal contribution of the (relatively slow) S-cone system to the 30Hz  
501 flicker response,<sup>59,60</sup> The current study highlights both the variability and high specificity of  
502 this feature; the LA3 a-wave to LA30Hz ERG amplitude ratio was never less than 1.0 and the  
503 lowest ratios (1.0) included cases with grossly reduced ERGs associated with a lower  
504 signal:noise ratio. Furthermore, relatively increased sensitivity to short-wavelength  
505 stimulation was observed, as demonstrated by large, delayed and simplified S-cone ERG  
506 responses or S-cone ERGs that were larger than the corresponding LA3 ERGs. The S-cone  
507 ERGs are not required for the diagnosis of ESCS, but the high correlation with the LA 3.0  
508 ERGs, is consistent with both having the same S-cone-dominated origin.

509 All but 3 eyes of 2 molecularly confirmed patients had the above-mentioned pathognomonic  
510 ERG features, in accordance with previous reports demonstrating that solely patients found to  
511 harbor mutations in *NR2E3* have pathognomonic ERG responses when compared to patients  
512 with retinal dystrophies unrelated to *NR2E3*.<sup>5</sup> Thus, in our cohort, the presence of clinical  
513 features consistent with ESCS alongside typical ERG responses were deemed diagnostic for  
514 ESCS, irrespective of molecular confirmation.

515 A comparison of multiple ERG component amplitudes with age, suggests a low mean rate of  
516 reduction over more than 6 decades, with no evidence of worsening beyond that explained by  
517 age. This finding highlights the relative stability of peripheral retinal function in most ESCS  
518 patients, and may be an important prognostic consideration for retention of peripheral retinal  
519 function. Marked inter-subject variability is evident with some younger adults showing  
520 markedly reduced ERG amplitudes, highlighting the importance of detailed phenotyping and  
521 need to manage cases individually. Peak times of the main ERG components are delayed but  
522 show a similar high level of stability to that in the control group. Longitudinal ERG data were  
523 available in three patients, showing relatively stable responses in two patients and mild  
524 reduction of both LA and DA function in one.

525 Four evolutionary conserved domains are identified in the *NR2E3* protein, shared by the  
526 nuclear hormone receptor family; the highly variable A/B domain, a terminal DNA binding  
527 domain, a flexible hinge region and the ligand-binding and dimerization domain in the C  
528 terminus.<sup>74</sup> Most mutations are located within the DNA binding domain and the ligand-  
529 binding domain.<sup>2, 3, 5, 16, 26</sup>

530 Genetic heterogeneity occurs in ESCS. Autosomal recessive variants in the neural retina  
531 leucine zipper (*NRL*) gene have been identified in patients presenting with an ESCS-like  
532 phenotype.<sup>7, 75-78</sup> This gene had been proposed as a possible candidate following the  
533 phenotypical characterization of the *Nrl*<sup>-/-</sup> mouse which revealed a complete loss of rod

534 function and super-normal cone function, driven by over-expressed S-cones.<sup>79</sup> Expression of  
535 *NR2E3* is almost absent in the *Nrl*<sup>-/-</sup>, implying that *NR2E3* is completely dependent of *NRL*  
536 expression.<sup>18</sup> *NRL* encodes a basic-motif leucine zipper DNA binding protein that interacts  
537 with the paired-type homeobox transcription factor cone-rod homeobox (*CRX*) and *NR2E3*,  
538 driving the differentiation of post-mitotic photoreceptors into the rod lineage.<sup>76-78, 80-82</sup>.

539 The function of genetic modifiers of *NR2E3*, such as the nuclear hormone receptor *Nr1d1*  
540 (*Rev-erba*), has been explored as a therapeutic option in the *NR2E3*-associated retinal disease,  
541 *rd7*, mouse model.<sup>26, 69, 83</sup> Delivery of the *Nr1d1* gene restored the retinal topography of the  
542 *NR2E3*<sup>rd7/rd7</sup> neonates, and re-regulated the expression of key genes involved in  
543 phototransduction.<sup>26</sup> Future studies will need to assess whether this approach would be suited  
544 for patients with advanced disease.

545 The present study describes the detailed clinical, imaging, molecular and electrophysiological  
546 findings in a cohort of 56 patients with ESCS, which, to the best of our knowledge, is the  
547 largest cohort to date. Four novel *NR2E3* variants are identified. The data quantify diagnostic  
548 ERG criteria and phenotypic spectrum, with evidence to suggest relative stability of  
549 peripheral retinal function over more than 6 decades, and additional evidence suggesting that  
550 central visual function remains relatively stable in the majority of patients, which is  
551 invaluable for counseling on prognosis. Any future intervention directed at preventing visual  
552 decline in ESCS will need to address its impact on the development of macular complications,  
553 namely foveomacular schisis and macular atrophy, which are largely responsible for the poor  
554 visual outcome observed in a subset of affected patients.

555

556

557 **References**

- 558 1. Kobayashi M, Takezawa S, Hara K, et al. Identification of a photoreceptor cell-specific  
559 nuclear receptor. *Proc Natl Acad Sci U S A* 1999;96(9):4814-9.
- 560 2. Schorderet DF, Escher P. NR2E3 Mutations in Enhanced S-Cone Sensitivity Syndrome  
561 (ESCS), Goldmann-Favre Syndrome (GFS), Clumped Pigmentary Retinal Degeneration (CPRD), and  
562 Retinitis Pigmentosa (RP). *Hum Mutat* 2009;30(11):1475-85.
- 563 3. Coppieters F, Leroy BP, Beysen D, et al. Recurrent mutation in the first zinc finger of the  
564 orphan nuclear receptor NR2E3 causes autosomal dominant retinitis pigmentosa. *Am J Hum Gen*  
565 2007;81(1):147-57.
- 566 4. Gerber S, Rozet JM, Takezawa SI, et al. The photoreceptor cell-specific nuclear receptor gene  
567 (PNR) accounts for retinitis pigmentosa in the Crypto-Jews from Portugal (Marranos), survivors from  
568 the Spanish Inquisition. *Hum Genet* 2000;107(3):276-84.
- 569 5. Sharon D, Sandberg MA, Berson EL, Dryja TP. Shared mutations in NR2E3 in enhanced S-  
570 cone syndrome, Goldmann-Favre syndrome, and clumped pigmentary retinal degeneration. *Invest*  
571 *Ophthalmol Vis Sci* 2002;43:U172-U.
- 572 6. Sharon D, Sandberg MA, Caruso RC, et al. Shared mutations in NR2E3 in enhanced S-cone  
573 syndrome, Goldmann-Favre syndrome, and many cases of clumped pigmentary retinal degeneration.  
574 *Arch Ophthalmol* 2003;121(9):1316-23.
- 575 7. Acar C, Mears A, Yashar B, et al. Mutation screening of patients with Leber congenital  
576 amaurosis or the enhanced S-cone syndrome reveals a lack of sequence variations in the NRL gene.  
577 *Mol Vis* 2003;9(3-4):14-7.
- 578 8. Jacobson SG, Roman AJ, Roman MI, et al. Relatively Enhanced S-Cone Function in the  
579 Goldmann-Favre Syndrome. *Am J Ophthalmol* 1991;111(4):446-53.
- 580 9. Jacobson SG, Roman AJ, Roman MI, et al. A Pattern of Retinal Dysfunction That Links the  
581 Goldmann-Favre Syndrome with the Enhanced S (Blue) Cone Syndrome. *Invest Ophthalmol Vis Sci*  
582 1991;32(4):912-.
- 583 10. Chavala SH, Sari A, Lewis H, et al. An Arg311Gln NR2E3 mutation in a family with classic  
584 Goldmann-Favre syndrome. *Br J Ophthalmol* 2005;89(8):1065-6.
- 585 11. Gire AI, Sullivan LS, Bowne SJ, et al. The Gly56Arg mutation in NR2E3 accounts for 1-2%  
586 of autosomal dominant retinitis pigmentosa. *Mol Vis* 2007;13(220-22):1970-5.
- 587 12. Kobayashi M, Hara K, Yu RT, Yasuda K. Expression and functional analysis of Nr2e3, a  
588 photoreceptor-specific nuclear receptor, suggest common mechanisms in retinal development between  
589 avians and mammals. *Dev Gen Evol* 2008;218(8):439-44.
- 590 13. Kobayashi M, Takezawa S, Hara K, et al. Identification of a photoreceptor cell-specific  
591 nuclear receptor. *PProc Natl Acad Sci U S A* 1999;96(9):4814-9.
- 592 14. Hood DC, Cideciyan AV, Roman AJ, Jacobson SG. Enhanced S-Cone-Syndrome - Evidence  
593 for an Abnormally Large Number of S-Cones. *Vis Res* 1995;35(10):1473-81.
- 594 15. Haider N, Naggert JK, Nishina PM. The transcription factor NR2E3 suppresses cone cell  
595 proliferation. *Invest Ophthalmol Vis Sci* 2005;46.
- 596 16. Haider NB, Jacobson SG, Cideciyan AV, et al. Mutation of a nuclear receptor gene, NR2E3,  
597 causes enhanced S cone syndrome, a disorder of retinal cell fate. *Nature Genet* 2000;24(2):127-31.
- 598 17. Haider NB, Naggert JK, Nishina PM. Elucidating the function of NR2E3 through  
599 identification of interacting factors. *Invest Ophthalmol Vis Sci* 2004;45:U631-U.
- 600 18. Cepko CL. The Determination of Rod and Cone Photoreceptor Fate. *Annu Rev Vis Sci, Vol 1*  
601 2015;1:211-34.
- 602 19. Wang S, Cepko CL. Photoreceptor Fate Determination in the Vertebrate Retina. *Invest*  
603 *Ophthalmol Vis Sci* 2016;57(5).
- 604 20. Haider NB, Demarco P, Nystuen AM, et al. The transcription factor Nr2e3 functions in retinal  
605 progenitors to suppress cone cell generation. *Vis Neurosci* 2006;23(6):917-29.

- 606 21. Hood DC, Cideciyan AV, Roman AJ, Jacobson SG. Enhanced S cone syndrome: evidence for  
607 an abnormally large number of S cones. *Vision Res* 1995;35(10):1473-81.
- 608 22. Haider NB, Jacobson SG, Cideciyan AV, et al. Mutation of a nuclear receptor gene, NR2E3,  
609 causes enhanced S cone syndrome, a disorder of retinal cell fate. *Nat Genet* 2000;24(2):127-31.
- 610 23. Milam AH, Rose L, Cideciyan AV, et al. The nuclear receptor NR2E3 plays a role in human  
611 retinal photoreceptor differentiation and degeneration. *Proc Natl Acad Sci U S A* 2002;99(1):473-8.
- 612 24. Haider NB, Mollema N, Gaule M, et al. Nr2e3-directed transcriptional regulation of genes  
613 involved in photoreceptor development and cell-type specific phototransduction. *Exp Eye Res*  
614 2009;89(3):365-72.
- 615 25. Corbo JC, Cepko CL. A hybrid photoreceptor expressing both rod and cone genes in a mouse  
616 model of enhanced S-cone syndrome. *PLoS Genet* 2005 Aug;1(2):e11.
- 617 26. Cruz NM, Yuan Y, Leehy BD, et al. Modifier Genes as Therapeutics: The Nuclear Hormone  
618 Receptor Rev Erb Alpha (Nr1d1) Rescues Nr2e3 Associated Retinal Disease. *PLoS One* 2014;9(1).
- 619 27. Hayashi T, Gekka T, Goto-Omoto S, et al. Novel NR2E3 mutations (R104Q, R334G)  
620 associated with a mild form of enhanced S-cone syndrome demonstrate compound heterozygosity.  
621 *Ophthalmology* 2005;112(12):2115-22.
- 622 28. Lam BL, Goldberg JL, Hartley KL, Stone EM. Atypical mild enhanced S-Cone syndrome  
623 with novel compound heterozygosity of the NR2E3 gene. *Am J Ophthalmol* 2007;144(1):157-9.
- 624 29. von Alpen D, Tran HV, Guex N, et al. Differential Dimerization of Variants Linked to  
625 Enhanced S-Cone Sensitivity Syndrome (ESCS) Located in the NR2E3 Ligand-Binding Domain. *Hum*  
626 *Mut* 2015;36(6):599-610.
- 627 30. Jacobson SG, Marmor MF, Kemp CM, Knighton RW. SWS (blue) cone hypersensitivity in a  
628 newly identified retinal degeneration. *Invest Ophthalmol Vis Sci* 1990;31(5):827-38.
- 629 31. Marmor MF, Jacobson SG, Foerster MH, et al. Diagnostic clinical findings of a new syndrome  
630 with night blindness, maculopathy, and enhanced S cone sensitivity. *Am J Ophthalmol*  
631 1990;110(2):124-34.
- 632 32. Greenstein VC, Zaidi Q, Hood DC, et al. The enhanced S cone syndrome: an analysis of  
633 receptor and post-receptor changes. *Vision Res* 1996;36(22):3711-22.
- 634 33. Audo I, Michaelides M, Robson AG, et al. Phenotypic variation in enhanced S-cone  
635 syndrome. *Invest Ophthalmol Vis Sci* 2008;49(5):2082-93.
- 636 34. Corbo JC, Cepko CL. A hybrid photoreceptor expressing both rod and cone genes in a mouse  
637 model of enhanced S-cone syndrome. *PLoS Genet* 2005;1(2):140-53.
- 638 35. Audo IS, Neveu MM, Robson AG, et al. Characterization of enhanced S-cone Syndrome  
639 (ESCS). *Invest Ophthalmol Vis Sci* 2004;45:U579-U.
- 640 36. Jurklies B, Weismann M, Kellner U, et al. Clinical findings in autosomal recessive enhanced  
641 S-cone sensitivity syndrome. *Ophthalmology* 2001;98(3):285-93.
- 642 37. Khan AO, Aldahmesh M, Meyer B. The enhanced S-cone syndrome in children. *Br J*  
643 *Ophthalmol* 2007;91(3):394-6.
- 644 38. Marmor MF. A teenager with nightblindness and cystic maculopathy: enhanced S cone  
645 syndrome (Goldmann-Favre syndrome). *Doc Ophthalmol* 2006;113(3):213-5.
- 646 39. Wang NK, Fine HF, Chang S, et al. Cellular origin of fundus autofluorescence in patients and  
647 mice with a defective NR2E3 gene. *Br J Ophthalmol* 2009;93(9):1234-40.
- 648 40. Yzer S, Barbazetto I, Allikmets R, et al. Expanded Clinical Spectrum of Enhanced S-Cone  
649 Syndrome. *JAMA Ophthalmol* 2013;131(10):1324-30.
- 650 41. Yzer S, Barbazetto I, Allikmets R, et al. Expanded clinical spectrum of enhanced S-cone  
651 syndrome. *JAMA Ophthalmol* 2013;131(10):1324-30.
- 652 42. Zerbib J, Blanco Garavito R, Gerber S, et al. Retinochoroidal Anastomosis Associated with  
653 Enhanced S-Cone Syndrome. *Retin Cases Brief Rep* 2017.
- 654 43. Vaclavik V, Chakarova C, Bhattacharya SS, et al. Bilateral giant macular schisis in a patient  
655 with enhanced S-cone syndrome from a family showing pseudo-dominant inheritance. *Br J*  
656 *Ophthalmol* 2008;92(2):299-300.
- 657 44. Hull S, Arno G, Sergouniotis PI, et al. Clinical and Molecular Characterization of Enhanced S-  
658 Cone Syndrome in Children. *JAMA Ophthalmol* 2014;132(11):1341-9.

- 659 45. Arden G, Gunduz K, Perry S. Color-Vision Testing with a Computer-Graphics System -  
660 Preliminary-Results. *Doc Ophthalmol* 1988;69(2):167-74.
- 661 46. Arden GB, Wolf JE. Colour vision testing as an aid to diagnosis and management of age  
662 related maculopathy. *Br J Ophthalmol* 2004;88(9):1180-5.
- 663 47. Holladay JT. Proper method for calculating average visual acuity. *J Refract Surg*  
664 1997;13(4):388-91.
- 665 48. Bach M, Brigell MG, Hawlina M, et al. ISCEV standard for clinical pattern  
666 electroretinography (PERG): 2012 update. *Doc Ophthalmol* 2013;126(1):1-7.
- 667 49. McCulloch DL, Marmor MF, Brigell MG, et al. ISCEV Standard for full-field clinical  
668 electroretinography (2015 update). *Doc Ophthalmol* 2015;130(1):1-12.
- 669 50. Perlman I, Kondo M, Chelva E, et al. ISCEV extended protocol for the S-cone ERG. *Doc*  
670 *Ophthalmol* 2020;140(2):95-101.
- 671 51. Li Q, Wang K. InterVar: Clinical Interpretation of Genetic Variants by the 2015 ACMG-AMP  
672 Guidelines. *Am J Hum Gen* 2017;100(2):267-80.
- 673 52. Choi Y, Chan AP. PROVEAN web server: a tool to predict the functional effect of amino acid  
674 substitutions and indels. *Bioinformatics* 2015;31(16):2745-7.
- 675 53. Choi Y, Sims GE, Murphy S, et al. Predicting the functional effect of amino acid substitutions  
676 and indels. *PLoS One* 2012;7(10):e46688.
- 677 54. Ng PC, Henikoff S. SIFT: predicting amino acid changes that affect protein function. *Nucleic*  
678 *Acids Res* 2003;31(13):3812-4.
- 679 55. Adzhubei IA, Schmidt S, Peshkin L, et al. A method and server for predicting damaging  
680 missense mutations. *Nat Methods* 2010;7(4):248-9.
- 681 56. Bienert S, Waterhouse A, de Beer TAP, et al. The SWISS-MODEL Repository-new features  
682 and functionality. *Nucleic Acids Res* 2017;45(D1):D313-D9.
- 683 57. Guex N, Peitsch MC, Schwede T. Automated comparative protein structure modeling with  
684 SWISS-MODEL and Swiss-PdbViewer: A historical perspective. *Electrophoresis* 2009;30:S162-S73.
- 685 58. Waterhouse A, Bertoni M, Bienert S, et al. SWISS-MODEL: homology modelling of protein  
686 structures and complexes. *Nucleic Acids Res* 2018;46(W1):W296-W303.
- 687 59. Tan MHE, Zhou XE, Soon FF, et al. The Crystal Structure of the Orphan Nuclear Receptor  
688 NR2E3/PNR Ligand Binding Domain Reveals a Dimeric Auto-Repressed Conformation. *PLoS One*  
689 2013;8(9).
- 690 60. Rocha-Sousa A, Hayashi T, Gomes NL, et al. A novel mutation (Cys83Tyr) in the second zinc  
691 finger of NR2E3 in enhanced S-cone syndrome. *Graefes Arch Clin Exp Ophthalmol*.  
692 2011;249(2):201-8.
- 693 61. Ripamonti C, Aboshiha J, Henning GB, et al. Vision in Observers With Enhanced S-Cone  
694 Syndrome: An Excess of S-Cones but Connected Mainly to Conventional S-Cone Pathways. *Invest*  
695 *Ophthalmol Vis Sci* 2014;55(2):963-76.
- 696 62. Chen JC, Rattner A, Nathans J. The rod photoreceptor-specific nuclear receptor Nr2e3  
697 represses transcription of multiple cone-specific genes. *J Neurosci* 2005;25(1):118-29.
- 698 63. Roduit R, Escher P, Schorderet DF. Mutations in the DNA-Binding Domain of NR2E3 Affect  
699 In Vivo Dimerization and Interaction with CRX. *PLoS One* 2009;4(10).
- 700 64. Marmor MF, Jacobson SG, Foerster MH, et al. Diagnostic Clinical Findings of a New  
701 Syndrome with Night Blindness, Maculopathy, and Enhanced S-Cone Sensitivity. *Am J*  
702 *Ophthalmol* 1990;110(2):124-34.
- 703 65. Escher P, Gouras P, Roduit R, et al. Mutations in NR2E3 Can Cause Dominant or Recessive  
704 Retinal Degenerations in the Same Family. *Hum Mutat* 2009;30(3):342-51.
- 705 66. Bertoli F, Pignatto S, Rizzetto F, Lanzetta P. A 5-Year-Old Case of Choroidal  
706 Neovascularization in Enhanced S-Cone Syndrome Treated with Ranibizumab. *Case Rep Ophthalmol*  
707 2018;9(3):510-5.
- 708 67. Broadhead GK, Grigg JR, McCluskey P, et al. Bevacizumab for choroidal neovascularisation  
709 in enhanced S-cone syndrome. *Docum Ophthalmol* 2016;133(2):139-43.
- 710 68. Sambricio J, Tejada-Palacios P, Barcelo-Mendiguchia A. Choroidal neovascularization, outer  
711 retinal tubulation and fundus autofluorescence findings in a patient with enhanced S-cone syndrome.  
712 *Clin Exp Ophthalmol* 2016;44(1):69-71.

- 713 69. Haider NB, Naggert JK, Nishina PM. Excess cone cell proliferation due to lack of a functional  
714 NR2E3 causes retinal dysplasia and degeneration in rd7/rd7 mice. *Hum Mol Genet* 2001;10(16):1619-  
715 26.
- 716 70. Mockel A, Perdomo Y, Stutzmann F, Letsch J, Marion V, Dollfus H. J Retinal dystrophy in  
717 Bardet-Biedl syndrome and related syndromic ciliopathies. *Prog Retin Eye Res.* 2011 Jul;30(4):258-  
718 74.
- 719 71. Talib M, van Schooneveld MJ, van Genderen MM, et al. Genotypic and Phenotypic  
720 Characteristics of CRB1-Associated Retinal Dystrophies A Long-Term Follow-up Study.  
721 *Ophthalmology* 2017;124(6):884-95.
- 722 72. den Hollander AI, ten Brink JB, de Kok YJ, et al. Mutations in a human homologue of  
723 *Drosophila crumbs* cause retinitis pigmentosa (RP12). *Nat Genet* 1999;23(2):217-21.
- 724 73. Marmor MF. Is Thioridazine Retinopathy Progressive - Relationship of Pigmentary Changes  
725 to Visual Function. *Br J Ophthalmol* 1990;74(12):739-42.
- 726 74. Mollema N, Haider NB. Focus on Molecules: Nuclear hormone receptor Nr2e3: Impact on  
727 retinal development and disease. *Exp Eye Res* 2010;91(2):116-7.
- 728 75. Ben-Yosef T, Newman H, Braverman I, et al. A combination of oculopharyngeal muscular  
729 dystrophy and a variant of enhanced S-cone syndrome in Bukharan Jews due to linked mutations in  
730 PABPN1 and NRL. *Invest Ophthalmol Vis Sci* 2016;57(12).
- 731 76. Littink KW, Stappers PTY, Riemsdag FCC, et al. Autosomal Recessive NRL Mutations in  
732 Patients with Enhanced S-Cone Syndrome. *Genes* 2018;9(2).
- 733 77. Newman H, Blumen SC, Braverman I, et al. Homozygosity for a Recessive Loss-of-Function  
734 Mutation of the NRL Gene Is Associated With a Variant of Enhanced S-Cone Syndrome. *Invest*  
735 *Ophthalmol Vis Sci* 2016;57(13):5361-71.
- 736 78. Wright AF, Reddick AC, Schwartz SB, et al. Mutation analysis of NR2E3 and NRL genes in  
737 Enhanced S Cone Syndrome. *Hum Mutat* 2004;24(5):439.
- 738 79. Mears AJ, Kondo M, Swain PK, et al. Nrl is required for rod photoreceptor development. *Nat*  
739 *Genet* 2001;29(4):447-52.
- 740 80. Oh ECT, Cheng H, Hao H, et al. Rod differentiation factor NRL activates the expression of  
741 nuclear receptor NR2E3 to suppress the development of cone photoreceptors. *Brain Res*  
742 2008;1236:16-29.
- 743 81. Farjo Q, Jackson A, PiekeDahl S, et al. Human bZIP transcription factor gene NRL: Structure,  
744 genomic sequence, and fine linkage mapping at 14q11.2 and negative mutation analysis in patients  
745 with retinal degeneration. *Genomics* 1997;45(2):395-401.
- 746 82. Swaroop A, Xu JZ, Pawar H, et al. A Conserved Retina-Specific Gene Encodes a Basic Motif  
747 Leucine Zipper Domain. *Proc Natl Acad Sci U S A* 1992;89(1):266-70.
- 748 83. Mollema NJ, Yuan Y, Jelcick AS, et al. Nuclear Receptor Rev-erb Alpha (Nr1d1) Functions in  
749 Concert with Nr2e3 to Regulate Transcriptional Networks in the Retina. *PLoS One* 2011;6(3)
- 750

751

## 752 FIGURE 1

753 (A) Plot of best-corrected visual acuity (BCVA, LogMar) of the right eye at baseline and last follow-  
754 up visit against patient's age. (B) Plot of BCVA (LogMar) in the right eye as a function of period of  
755 follow-up time per individual patient. (C) Plot of BCVA change in the right eye ( $BCVA_{FU} -$   
756  $BCVA_{baseline}$ ) as function of follow-up time ( $y = \text{years}$ ) and (D) age at baseline.

757

## 758 FIGURE 2

759 Phenotypical variation of Enhanced S-Cone Syndrome in individual patients (numbered). (A)  
760 Nummular pigmentary deposition in the mid-peripheral retina. (B) Circumscribed area of nummular  
761 pigmentary deposition with halo of atrophy in inferior peripheral retina. (C) Nummular pigmentary  
762 deposition, yellow/white dots and clumped pigmentary changes in the mid-peripheral retina. (D)  
763 Yellow/white dots along vascular arcades, with increased fundus autofluorescence inside the vascular  
764 arcades, sparing the central macula. (E) Magnified view of nummular pigmentary deposition, yellow-  
765 white dots and clumped pigmentary changes in mid-peripheral retina. (F) Torpedo-like lesion in  
766 peripheral retina. (G) Subretinal fibrosis and spectral-domain optical coherence (SD-OCT)  
767 tomography across lesion (marked) showing a large subretinal hyper-reflective deposit. (H) Magnified  
768 view of yellow-white dots with early pigmentary hyperplastic changes. (I) Retinal angioma in patient  
769 with bilateral pre-retinal non-diabetic neovascularization. (J) Maculopathy, characterized by patchy  
770 atrophic macular changes, more visible on FAF. (K) Color fundus photograph of the right peripheral  
771 retina of patient 11 at baseline (right image) and 17 years later, at last follow-up (left image, year of  
772 OCT acquisition marked in left bottom corner). At baseline, retinal sclerosed vessels and yellow-white  
773 dots are seen which progressed to nummular and clumped pigmentary deposition as observed in the  
774 follow-up photograph of the same area. (L) Color fundus photograph of right superior vascular arcade  
775 in patient 15 at baseline (right image) and 11 years later, at last follow-up (left image, year of OCT  
776 acquisition marked in left bottom corner). A well-defined area of yellow-white dots is observed at  
777 baseline which developed into clumped pigmentary deposition, shown in the follow-up image.

778

## 779 FIGURE 3

780 Variation of optical coherence tomography features of Enhanced S-Cone Syndrome in individual  
781 patients (numbered). (A) Preserved foveal architecture and outer retinal atrophy, with loss of the  
782 ellipsoid zone. (B) Foveomacular schisis. (C) Magnified view of area outlined in (B), with pseudo-  
783 color representation of the the schitic cavities (round shape, in red) at the level of the inner nuclear

784 layer (round shape, in red) and at the level of the outer nuclear layer (elongated shape, in blue). (D)  
785 End-stage giant foveomacular schisis. (E) Disorganization of retinal layers in atrophic area of mid-  
786 peripheral retina. (F) Macular atrophy.

787

788 FIGURE 4

789 Macular and peripheral fundus autofluorescence (FAF) patterns in Enhanced S-Cone Syndrome in  
790 individual patients (numbered). (A) Minimal change macular FAF pattern. (B) (A) Minimal change  
791 macular FAF pattern with hyperautofluorescent flecks. (C) Mild diffuse macular  
792 hypoautofluorescence. (D) Moderate speckled macular hypoautofluorescence with increased para-  
793 macular FAF. (E) Severe end-stage macular hypoautofluorescence. (F) Peripheral  
794 hyperautofluorescent flecks. (G) Moderate diffuse (mid-peripheral half-ring or ring < 5000  $\mu\text{m}$  widest  
795 diameter) peripheral hypoautofluorescence with half-ring of pronounced hyperautofluorescent ring  
796 along the temporal macular rim. (H) Near-peripheral moderate diffuse hypoautofluorescence with  
797 patchy advanced hypoautofluorescence. (I) Moderate diffuse peripheral hypoautofluorescence (> 5000  
798  $\mu\text{m}$ ). (J) Advanced peripheral hypoautofluorescence. (K) Colour fundus photograph and related  
799 autofluorescence image showing the correspondence between yellow-white dots and  
800 hyperautofluorescent flecks. (L) Wide-field autofluorescence image in control subject. The macula  
801 was defined as the region encompassing 5.5 mm from the temporal margin of the optic nerve head and  
802 the mid-periphery as 3 mm around the macula.

803

804 FIGURE 5

805 Full-field ERG and pattern ERG (PERG) recordings from the right (RE) and left (LE) eye of a patient  
806 with Enhanced S-Cone Syndrome are compared with recordings from a representative unaffected  
807 control subject (N). ERGs include the dark-adapted (DA) ERGs (flash strengths 0.01 and 10.0  $\text{cd.s/m}^2$ ;  
808 DA 0.01 and DA 10.0) and light-adapted (LA) ERGs for a flash strength of 3.0  $\text{cd.s/m}^2$  (LA 3.0; 30Hz  
809 and 2Hz). The PERG is recorded to an alternating chequerboard. There is a 20ms pre-stimulus delay in  
810 single flash ERG recordings, with the exception of the S-cone ERG. Broken lines replace blink  
811 artefacts occurring after ERG b-waves, for clarity. Patient responses are superimposed to demonstrate  
812 reproducibility. In this patient the PERG P50 component is delayed but of normal amplitude. The  
813 DA0.01 ERG is undetectable. The single flash DA 3.0, DA 10.0, LA3.0 ERGs have similarly  
814 simplified and severely delayed waveforms, qualitatively comparable in shape to the S-cone ERG and  
815 consistent with generation by the same (S-cone) mechanism. The S-cone ERG is delayed and grossly  
816 enlarged. The LA30 Hz ERG is smaller than the LA 3 ERG a-wave whereas in the typical normal

817 subject, the LA30Hz ERG amplitude is between that of the LA3 a- and b-waves. Measurements of the  
818 main ERG components are compared with the control range in supplementary table O.

819

820 FIGURE 6

821 The main dark-adapted (DA) full-field ERG component amplitudes and peak times in each eye in the  
822 Enhanced S-Cone Syndrome (ESCS) cohort (filled circles and squares) and in healthy controls (grey  
823 circles) are plotted against age (in years) at the time of testing, illustrating the severity and range of  
824 ERG abnormality in the ESCS group. Data are shown for the DA strong flash (DA10) ERG a-wave  
825 amplitude (A) and peak time (B) and for the b-wave amplitude (C) and peak time (D). Regression  
826 analysis shows a similar, statistically significant ( $P < 0.05$ ) age-related reduction in amplitudes for both  
827 control (broken lines) and ESCS (solid lines) groups.

828

829 FIGURE 7

830 The main light-adapted (LA) full-field ERG component amplitudes and peak times in each eye in the  
831 Enhanced S-Cone Syndrome (ESCS) cohort (filled circles and squares) and in healthy controls (grey  
832 circles) are plotted against age (in years) at the time of testing, illustrating the severity and range of  
833 ERG abnormality in the ESCS group. Data are shown for the LA30 Hz flicker ERG amplitude (A) and  
834 peak time (B) and for the single flash cone (LA3) ERG a-wave amplitude (C) and peak time (D) and  
835 for the LA3 ERG b-wave amplitude (E) and peak time (F). Regression analysis shows a similar,  
836 statistically significant ( $P < 0.05$ ) age-related reduction in amplitudes for both control (broken lines)  
837 and ESCS (solid lines) groups.

838

839 FIGURE 8

840 Comparison of amplitude and peak time ratios between the LA3.0 ERG a-wave and LA30Hz ERG in  
841 the Enhanced S-Cone Syndrome (ESCS) cohort (filled circles and squares) and healthy controls (open  
842 grey circles and squares). The horizontal bars show the mean  $\pm$  1 standard deviation (SD) for  
843 amplitudes in the ESCS group. The LA30Hz ERG has an amplitude greater than the LA3 ERG a-wave  
844 in all control subjects. In ESCS the LA30Hz ERG amplitude is equal or smaller than the LA3 ERG a-  
845 wave, resulting in a ratio greater or equal to 1. \*\*\*  $P = 0.0001$

846

## 847 FIGURE 9

848 The S-cone ERG component amplitudes (A) and peak times (B) are shown for the Enhanced S-Cone  
849 Syndrome (ESCS) cohort (n = 28, filled circles) and a healthy control group (n = 51, open grey circles)  
850 for comparison, plotted against age (in years). S-cone ERG amplitudes were measured from the early  
851 negative trough to maximum peak, or in the absence of an early trough from baseline to the peak of  
852 the positive polarity S-cone ERG component. The largest S-cone ERGs were seen in some of the  
853 younger ESCS individuals (solid regression line shows a negative slope) but there was no age-related  
854 statistically significant differences. Comparison of S-cone ERG amplitudes (C) and peak times (D)  
855 with those for LA3 ERG b-waves are shown for ESCS and control groups, and illustrate high positive  
856 correlation in the ESCS group, consistent with S-cone and LA3 ERGs being dominated by abnormal  
857 S-cone-opsin-mediated activity. All data relate to right eye recordings.

858

## 859 FIGURE 10

860 (A). Stereo representation of the NR2E3 ligand-binding domain (LBD) monomer (pdb code 4LOG),  
861 starting at residue 217, and locations of the novel missense NR2E3 LBD mutations mapped on the  
862 receptor. (B-D) Predicted effect of the mutations (p.R247W, p.L303P, p.R309Q) leading to a  
863 rearrangement of the bulky side chains and loss of hydrogen bonds.

Pt	Family ID	Age Onset (y)	BL	FU	Length of FU (y)	LogMar		Variants identified
						BL	FU	
1	NA	0	49	63	14	1/1	1/1	c.119-2A>C (Hom)
2	19824	4	11	16	5	0.1/0.6	0.1/0.6	c.119-2A>C / c.1194delT; p.P399Qfs*3
3	19824	0	9	14	5	0.5/0.5	0.4/0.3	c.119-2A>C / c.1194delT; p.P399Qfs*3
4	19940	3	15	17	2	0.17/0.6	0.86/1.2	c.305C>A; p.A102D / c.767C>A; p.A256E
5	20195	3	8	14	6	0.7/0.7	1.6/1.6	c.119-2A>C / c.1194delT; p.P399Qfs*3
6	20766	4	44	49	5	0.12/0	0.17/0.17	<b>c.211T&gt;C; p.F71L</b> / c.932G>A; p.R311Q
7	22497	5	8	11	3	0.26/0.36	0.6/0.5	NA
8	16200	6	34	48	14	0.77/0.6	1.77/1	c.119-2A>C (Hom)
9	23115	2	7	8	2	0.3/0.5	0.3/0.17	c.119-2A>C / c.1025T>C; p.V342A
10	27357	6	2.5	20	14	0.17/0.17	0.17/0.18	c.311G>A; p.R104Q (Hom)
11	15494	10	17	50	33	1.77/1.77	1.3/1.3	c.311G>A; p.R104Q (Hom)
12	27357	5	5	18	13	0.19/0.19	0/0	c.311G>A; p.R104Q (Hom)
13	3006	0	32	46	14	0.3/0.17	0.47/0.47	c.119-2A>C / c.767C>A; p.A256E
14	4644	1	13	34	21	0/0.47	0.5/0.47	c.119-2A>C (Hom)
15	16337	20	25	43	18	0.47/0.77	0.47/0.77	c.119-2A>C / c.932G>A; p.R311Q
16	15128	0	0.5	34.5	34	0.77/0.77	1.47/1	c.119-3C>G (Hom)
17	NA	6	21	35	14	0.17/2	0.17/2	NA
18	18491	20	27	35	8	0.17/0.17	0.17/0.18	c.932G>A; p.R311Q / c.1112T>G; p.L371W
19	NA	12	34	37	3	0.3/0.17	0.3/0	NA
20	20091	0	40	42	2	0.6/0.6	1/1	NA
21	18411	27	43	44	1	1/0.77	1/0.7	NA
22	NA	5	12	No FU	No FU	0.9/0.77	No FU	NA
23	20907	0	5	7.5	2.5	0.3/0.3	0.3/0.19	NA
24	19784	0	5	1	6	0.14/0	0/0	NA
25	18115	4	44	47	3	0.19/0.19	0.19/0.19	c.646G>A; p.G216S (Hom)
26	18758	4	72	81	9	0.19/0.2	0.3/0.3	c.305C>A; p.A102D (Hom)
27	NA	4	12	15	3	0.12/0	0/0	NA
28	22924	4	11	12	1	0/0	0/0	c.932G>A; p.R311Q / c.747+1G>C
29	27135	5	35	36	1	0/0.17	0.17/0.3	NA
30	22633	4	15	17	2	0.12/0	0.12/0.22	c.932G>A; p.R311Q (Hom)
31	19530	4	14	16	2	0/0	0.04/0.06	NA
32	23064	5	5	7	2	0.3/0.2	0.12/0.12	c.310C>T; p.R104W (Hom)
33	19530	4	8	10	2	0.04/0.02	0.18/0.2	NA
34	24703	12	19	21	2	0.47/1.17	0.30/0.80	c.310C>T; p.R104W (Hom)
35	19668	5	5	12	7	0.6/0.4	0.3/0.2	c.311G>A; p.R104Q / c.767C>A; p.A256E
36	18880	0	46	54	8	1/1	1/1	c.119-2A>C / <b>c.908T&gt;C; p.L303P</b>
37	NA	20	40	45	5	0.5/0.8	0.6/1	NA
38	17494	4	20	24	4	0/0	0.1/0.2	NA
39	NA	3	11	21	10	0.3/0.3	0.4/0.4	c.119-2A>C (Hom)
40	NA	11	44	No FU	No FU	0.3/0.1	No FU	c.932G>A; p.R311Q (Hom)
41	NA	4	33	44	11	0.8/1.1	0.22/1.7	c.119-2A>C (Hom)
42	NA	5	21	27	6	0.1/0.22	0.1/0.5	c.119-2A>C / c.932G>A; p.R311Q
43	NA	4	4	10	6	0.3/0.3	0.22/0.3	c.200_208del9del; p.C67_G69del (Hom)
44	NA	3	31	No FU	No FU	0.4/0.5	No FU	c.932G>A; p.R311Q / <b>c.739C&gt;T; p.R247W</b>
45	NA	3	11	12	4	0.22/0.1	0.22/0.1	c.932G>A; p.R311Q (Hom)
46	NA	5	46	No FU	No FU	1/0.5	No FU	c.932G>A; p.R311Q (Hom)
47	NA	5	75	76	1	0.3/2	0.4/2	c.119-2A>C / c.227G>A; p.R76Q
48	25595	12	35	36	1	0.47/0.17	0.47/0.17	c.932G>A; p.R311Q (Hom)
49	25690	3	33	No FU	No FU	0.3/0.8	No FU	c.248G>A; p.C83Y (Hom)
50	23064	8	6	2	8	0.3/0.2	0.06/0.06	c.310C>T; p.R104W (Hom)
51	25574	10	49	50	1	0.3/0.3	0.5/0.2	c.119-2A>C (Hom)
52	247	10	38	54	4.5	0.6/0.2	1/0.5	c.119-2A>C / c.932G>A; p.R311Q
53	NA	16	16	No FU	No FU	0/0	No FU	<b>c.926G&gt;A; p.R309Q</b> (Hom)
54	NA	25	29	No FU	No FU	0/0	No FU	<b>c.926G&gt;A; p.R309Q</b> (Hom)
55	26532	0	32	33	1	0.6/0.5	0.6/0.6	c.119-2A>C (Hom)
56	22052	0	51	55	4	1/1	1/1	c.119-2A>C (Hom)

Table 1. Clinical Data and Molecular Genetic Status of 56 Patients with Enhanced S-Cone Syndrome.

Abbreviations: BL = baseline; FU = follow-up; LogMAR = logarithm of minimal angle of resolution; Hom = homozygous variant; NA = not available; Pt = patient; VA = visual acuity. Putative novel changes are shown in bold.

Pt	Onset (y)	Length of FU (y)	LogMar		Progression	Macular changes				Other ophthalmic pathology
			BL	FU		Foveomacular schisis	Macular oedema	CAI treatment	Macular atrophy	
1	0	14	1/1	1/1	N	N	N	N	Y	Senile cataracts
4	3	2	0.17/0.6	0.86/1.2	Y	Y	N	N	N	<b>Squint, amblyopia</b>
5	3	6	0.7/0.7	1.6/1.6	Y	N	Y	Y (no response)	N	
8	6	14	0.77/0.6	1.77/1	Y	Y	Y	Y (response)	Y	
11	10	33	1.77/1.77	1.3/1.3	N	Y	Y	Y (no response)	Y	
15	20	18	0.47/0.77	0.47/0.77	N	Y	Y	Y (response)	N	
16	0	34	0.77/0.77	1.47/1	Y	Y	N	N	Y (in one eye)	Congenital nystagmus
17	6	14	0.17/2	0.17/2	N	N	N	N	Y (in one eye)	<b>Amblyopia, retinal detachment (age 6)</b>
20	0	2	0.6/0.6	1/1	Y	N	N	N	N	Senile cataracts
21	27	1	1/0.77	1/0.7	N	Y	N	N	Y	
22	5	No FU	0.9/0.77	No FU	NA	Y	N	N	N	TED
34	12	2	0.47/1.17	0.30/0.80	N	Y	N	N	Y	
36	0	8	1/1	1/1	N	Y	N	N	Y	ERM, Optic nerve pallor
37	20	5	0.5/0.8	0.6/1	Y	Y	N	N	Y	
41	4	11	0.8/1.1	0.22/1.7	N	Y	Y	N	N	
46	5	No FU	1/0.5	No FU	NA	Y	N	N	N	
49	3	No FU	0.3/0.8	No FU	NA	N	N	N	Y	
56	0	4	1/1	1/1	N	Y	N	N	Y	

Table 2. Clinical characteristics of patients with severe visual impairment (LogMAR BCVA > 0.6). Significant ophthalmic events that have contributed to poor visual acuity in one eye are highlighted in bold. Abbreviations: CAI = Carbonic anhydrase inhibitors; ERM = Epiretinal membrane; FU = Follow.up; N = No; NA = Not applicable; Pt = Patient; TED = Thyroid Eye Disease; Y = Yes

<b>Pt (n)</b>	56	
<b>Age at presentation (median. range)</b>	4	0-27
<b>Age at first visit (median. range)</b>	20.5	1-75
<b>Age at last FU (median. range)</b>	33	2-81
<b>Years of FU (mean. range)</b>	6.1	0-34
<b>BCVA (better-seeing eye) at presentation (mean. SEM)</b>	0.32	0.0-1.77
<b>BCVA (better-seeing eye) at last visit (mean. SEM)</b>	0.39	0.0-1.6
<b>BCVA reduction (mean. SEM)</b>	0.07	0.04
<b>Gender</b>		
Female gender (n. percent)	33	58.9%
Male gender (n. percent)	23	41.1%
<b>Ethnicity</b>		
White (n. percent)	30	53.6%
Non-White (n. percent)	26	46.4%
<b>Refraction (n. percent)</b>		
Plano	4	7.1%
Myopia	5	8.9%
Hyperopia	12	21.4%
NA	35	62.5%
<b>First symptom/sign (n. percent)</b>		
Nyctalopia	52	92.9%
Squint	9	16.1%
Nystagmus	3	5.4%
<b>Clinical signs (n. percent)</b>		
Optic nerve pallor	3	5.4%
Macular edema (based on structural OCT appearance)	16	28.5%
Foveomacular schisis	23	41.1%
Nummular pigmentation	48	85.7%
Yellow dots	32	57.1%
Circumferential subretinal fibrosis	4	7.1%
Torpedo-like lesions	6	10.7%
Vitreous opacities	12	21.4%
Preretinal neovascularization	1	1.8%

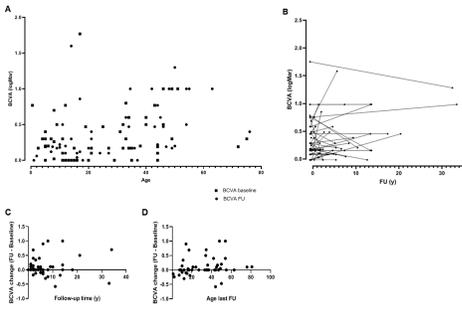
Table 3. Clinical characteristics of the Enhanced S-Cone Syndrome Cohort. Abbreviations: FU = follow-up; NA = not specified; Pt = patient; SEM = standard error of mean

Exon	Nucleotide Substitution and Amino Acid Change	Previous Report	InterVar Prediction	PROVEAN Prediction	Index	PolyPhen 2 Prediction	Hum Var Score (0-1)	Allelic Frequency
IVS1	c.119-2A>C	<sup>22</sup>			NA		NA	0.0005031
IVS1	c.119-3C>G	<sup>33</sup>			NA		NA	Not reported on gnomAD
2	c.211T>C; p.F71L	Novel	Uncertain significance	Deleterious	-5.218	PRD	1.00	Not reported on gnomAD
2	c.200_208del9del; p.C67_G69del	<sup>22</sup>		Deleterious	-26.485		NA	0.00001347
2	c.227G>A; p.R76Q	<sup>22</sup>	Uncertain significance	Deleterious	-3.343	PRD	1.00	0.0002140
3	c.248G>A; p.C83Y	<sup>83</sup>	Uncertain significance	Deleterious	-9.185	PRD	1.00	0.00001308
3	c.305C>A; p.A102D	<sup>44</sup>	Likely pathogenic	Deleterious	-4.862	PRD	0.99	0.00002792
3	c.310C>T; p.R104W	<sup>22</sup>	Uncertain significance	Deleterious	-6.962	PRD	1.00	0.00001964
5	c.646G>A; p.G216S	<sup>22</sup>	Uncertain significance	Neutral	0.491	Benign	0	0.00003611
5	c.311G>A; p.R104Q	<sup>27</sup>	Likely pathogenic	Deleterious	-3.533	PRD	1.00	0.00001964
5	c.739C>T; p.R247W	Novel	Uncertain significance	Deleterious	-7.733	PRD	1.00	Not reported on gnomAD
6	c.908T>C; p.L303P	Novel	Uncertain significance	Deleterious	-6.552	PRD	1.00	Not reported on gnomAD
6	c.932G>A; p.R311Q	<sup>22</sup>	Likely pathogenic	Neutral	-1.831	PSD	0.627	0.0004071
6	c.767C>A; p.A256E	<sup>84</sup>	Likely pathogenic	Deleterious	-3.659	PRD	0.998	0.00003715
6	c.926G>A; p.R309Q	Novel	Likely pathogenic	Deleterious	-3.520	PRD	0.959	0.00001528
7	c.1025T>C; p.V342A	<sup>85</sup>	Uncertain significance	Deleterious	-3.677	PRD	0.996	Not reported on gnomAD
8	c.1194delT; p.P399Qfs*3	<sup>85</sup>			NA		NA	Not reported on gnomAD
8	c.1112T>G; p.L371W	<sup>86</sup>	Uncertain significance	Deleterious	-5.107	PRD	1.00	Not reported on gnomAD

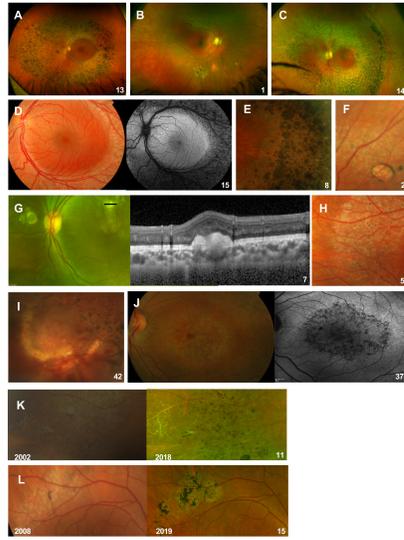
Table 4. NR2E3 variants. gnomAD = Genome Aggregation Database; Hum Var Score = human variation score; InterVar = Clinical Interpretation of Genetic Variants by the 2015 ACMG-AMP Guidelines, which categorizes causality of variants as pathogenic, likely pathogenic, uncertain significance, likely benign, and benign [<http://wintervar.wglab.org>. Accessed June 12, 2020]. NA = not applicable; PRD = probably damaging; PSD = Possibly damaging; PROVEAN = Protein Variation Effect Analyzer [<http://provean.jcvi.org/index.php>. Accessed February 15, 2019]. Variants with a score equal to or below -2.5 are considered "deleterious". Variants with a score above -2.5 are considered "neutral". Polyphen 2 (vision 2.1) appraises mutations qualitatively as benign, possibly damaging or probably damaging based on the model's false positive rate [<http://genetics.bwh.harvard.edu/pph2/>. Accessed February 28, 2020]. HumanVar-trained model of Polyphen 2 was selected,

since diagnostics of mendelian diseases requires distinguishing mutations with drastic effects from all the remaining human variation, including abundant mildly deleterious alleles.

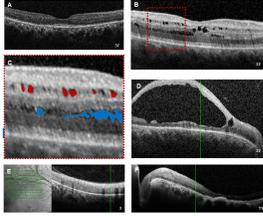
Journal Pre-proof



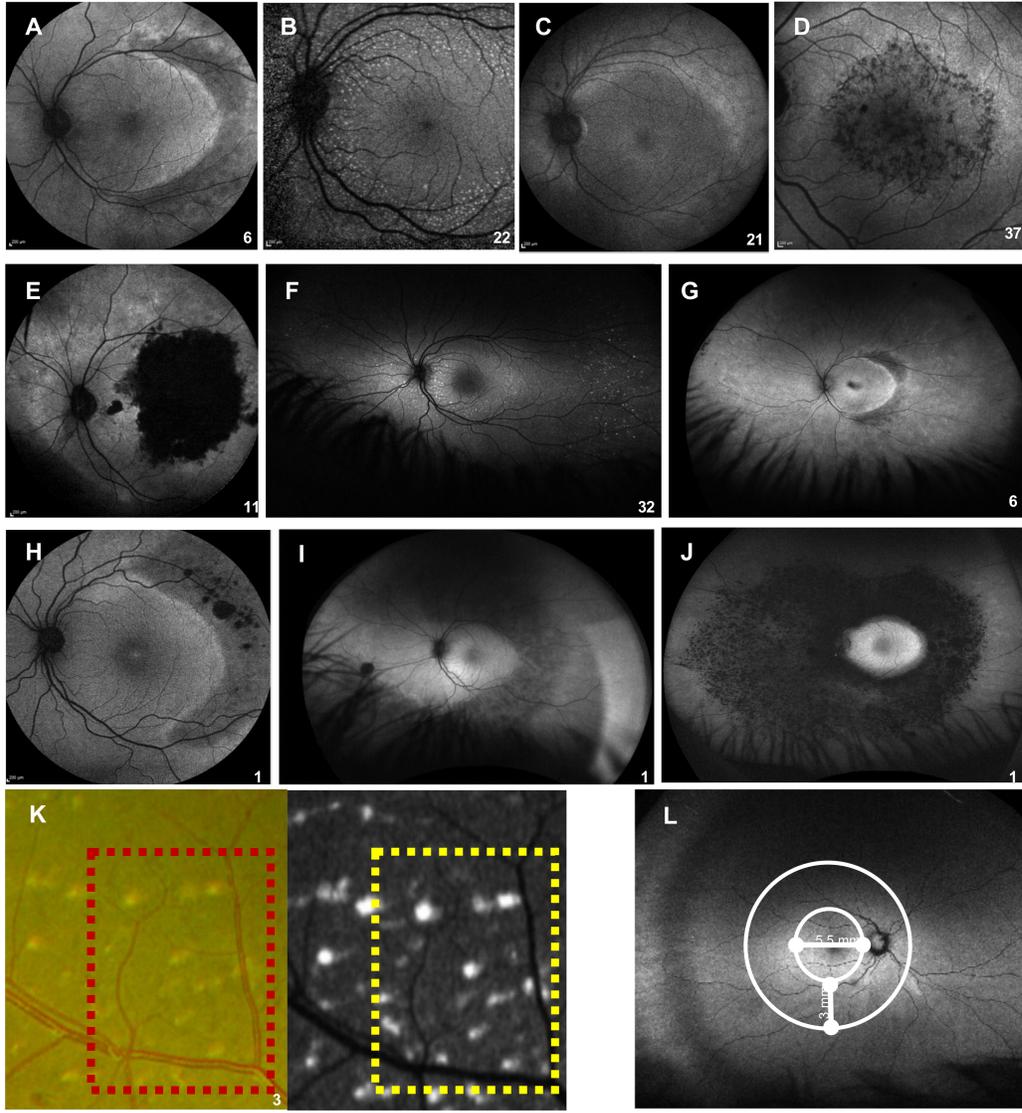
Journal Pre-proof

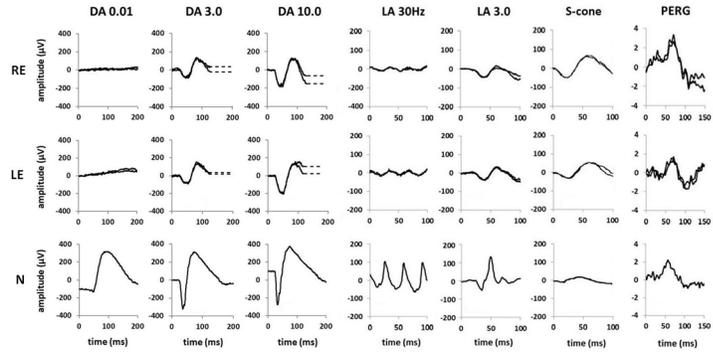


Journal Pre

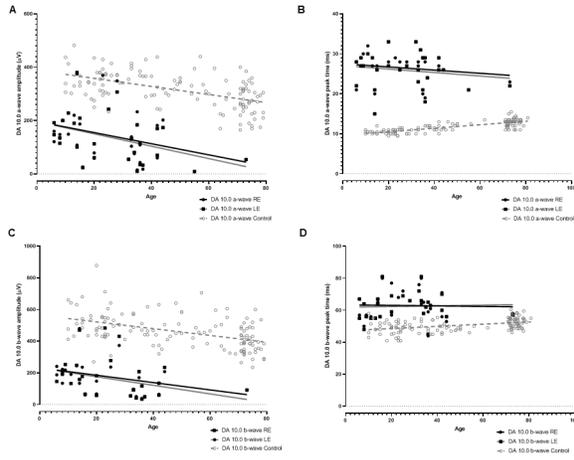


Journal Pre-proof

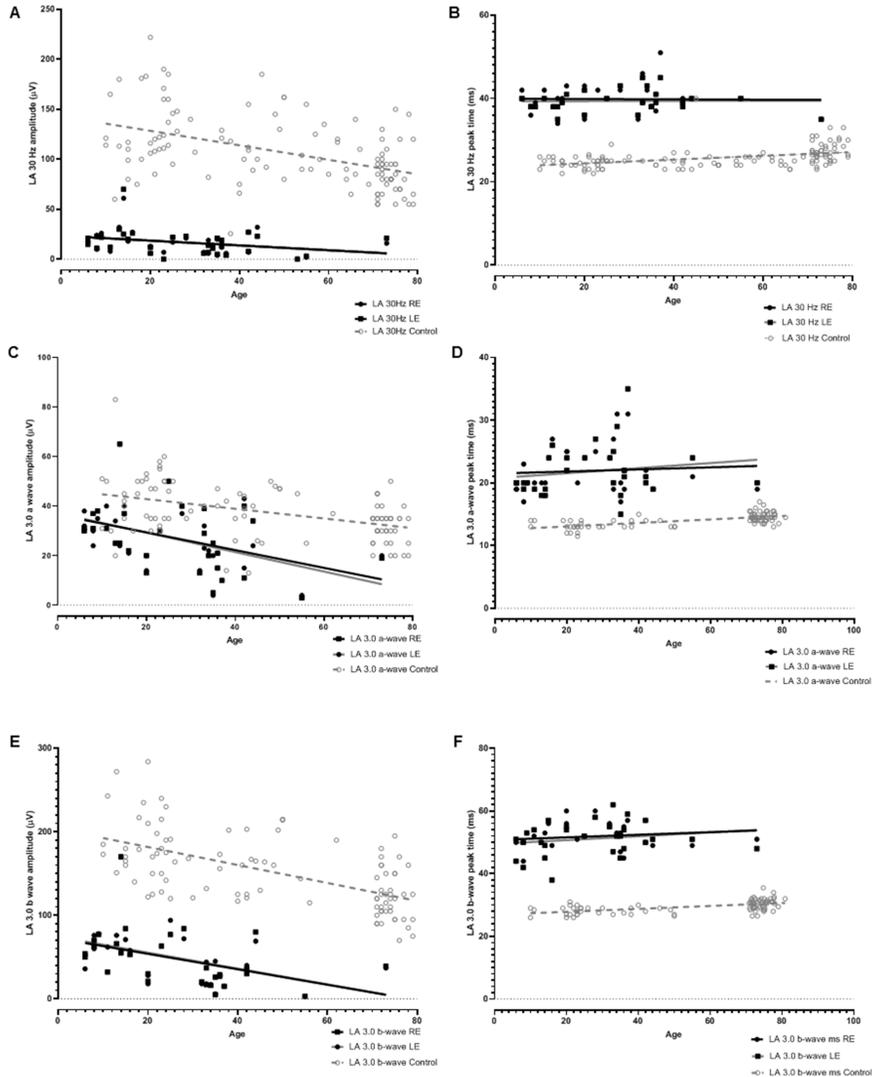


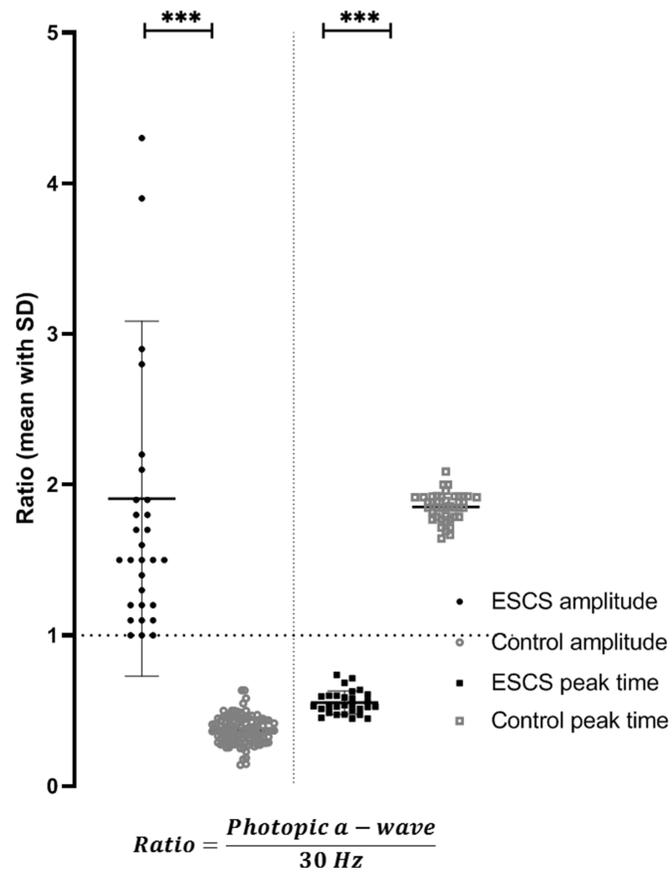


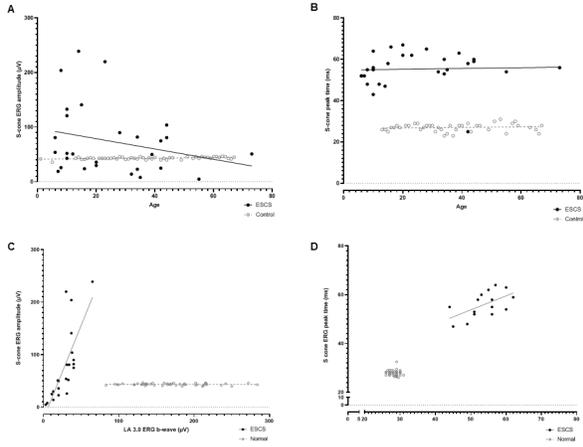
Journal Pre-proof



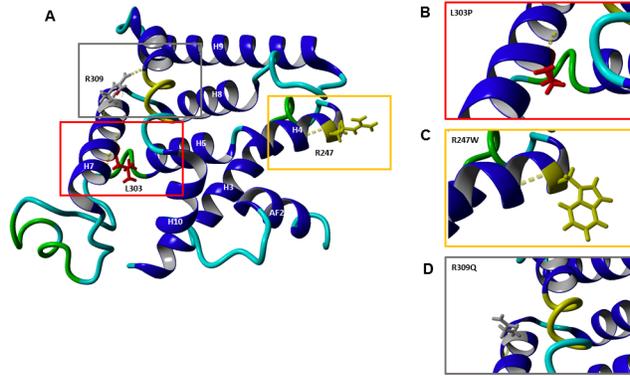
Journal Pre-proof







Journal Pre-proof



Journal Pre-proof

**Précis**

Enhanced S-Cone Syndrome, caused by mutations in the *NR2E3* gene, has a variable clinical phenotype and typical electrophysiological responses that are relatively stable over time.

Journal Pre-proof