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SHORT COMMUNICATION

Cholesterol-modifying drugs in COVID-19

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ABSTRACT

Infection with severe acute respiratory syndrom coronavirus 2 (SARS-CoV-2) is more likely to lead to poor outcomes in the elderly and those with cardiovascular disease, obesity or metabolic syndrome. Here, we consider mechanisms by which dyslipidaemia and the use of cholesterol-modifying drugs could influence the virus–host relationship. Cholesterol is essential for the assembly, replication and infectivity of enveloped virus particles; we highlight several cholesterol-modifying drugs with the potential to alter the SARS-CoV-2 life cycle that could be tested in *in vitro* and *in vivo* models. Although cholesterol is an essential component of immune cell membranes, excess levels can dysregulate protective immunity and promote exaggerated pulmonary and systemic inflammatory responses. Statins block the production of multiple sterols, oxy-sterols and isoprenoids, resulting in a pleiotropic range of context-dependent effects on virus infectivity, immunity and inflammation. We highlight antiviral, immunomodulatory and anti-inflammatory effects of cholesterol-modifying drugs that merit further consideration in the management of SARS-CoV-2 infection.

Key words: COVID-19; SARS-CoV-2; viral immunology; T-cells; cholesterol-modifying drugs; inflammation; statins; dyslipidemia.

INTRODUCTION

The coronavirus disease 2019 (COVID-19) pandemic has highlighted the dynamic and variable nature of host–pathogen interactions, with the severity ranging from asymptomatic infection to fatal acute respiratory distress syndrome (ARDS). There is an urgent need to understand the mechanisms underlying the range of outcomes of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection in order to consider repurposing existing drugs to limit morbidity and mortality. Poor outcomes are more frequent in older patients and in those with pre-existing cardiovascular disease, obesity and metabolic syndrome [1–3], implicating hyperlipidaemia as a possible exacerbating factor. The role of diabetes and antihypertensive drugs affecting the renin–angiotensin system (RAS) in these high-risk groups has been considered [4]. However, a sizeable proportion of those infected with SARS-CoV-2 is also taking statins, which are widely prescribed to reduce cardiovascular disease risk, with a good safety profile [5]. COVID-19 patients already taking statins were reported to have a milder course of infection in a care home setting [6]. Retrospective studies have previously found pre-existing statin usage to be associated with an improved outcome of pneumonia due to infections other than SARS-CoV-2 [7–9]. Statins are already being started in those not yet taking them during admission with COVID-19 in some centres [10]. Their use in SARS-CoV-2 infection has been proposed based on anecdotal effects in Ebola, although statin efficacy could not be

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distinguished from that of coadministered angiotensin II blockers, and randomized controlled trials were not performed [11].

Raised cholesterol or cholesterol-modifying drugs may be associated with differential rates of viral infection and outcome through indirect associations with other confounders and/or through direct effects. Here, we review existing mechanistic data that could provide a scientific rationale for hyperlipidaemia and cholesterol-modifying drugs playing a role in the course of SARS-CoV-2 infection. The initial disease outcome is likely to be determined by the balance between the efficiency of SARS-CoV-2 infectivity for tissues expressing the entry receptor angiotensin-converting enzyme 2 (ACE2) [12] (with resultant tissue cytopathology) and timely controlled antiviral immune responses. In contrast, the subsequent pathology seen in those COVID-19 patients progressing to ARDS is characterized by a cytokine storm accompanied by lung and systemic inflammation [13, 14]. We will consider three key categories by which cholesterol and cholesterol-modifying drugs could alter the outcome of SARS-CoV-2 infection: by modulating viral infectivity, antiviral immunity or inflammation.

MODULATION OF SARS-CoV-2 INFECTION

Viruses have complex interactions with their host; cholesterol biosynthesis pathways are essential for the assembly, replication and infectivity of enveloped virus particles [15]. Cholesterol-modifying drugs could exert antiviral effects by altering target cell membrane cholesterol through reduced systemic absorption or synthesis or by direct antiviral activity. In silico prediction studies for drug repurposing suggested the potential for rosuvastatin, a member of the statin family, to bind to SARS-CoV-2 [16], whereas another study failed to report such an interaction [17]. A recent in vitro study investigating the SARS-CoV-2 protein-protein interactome identified scavenger receptor class B type 1 (SR-B1), a cholesterol trafficking receptor, as a potential drug target [18] and blockade of SR-BI inhibited SARS-CoV-2 infectivity in an in vitro study [19]. It is worth noting that SR-B1 is an essential cofactor for hepatitis C virus (HCV) infection and an SR-B1 inhibitor was well tolerated in an openlabel clinical Phase Ib trial in HCV [20].

Lipid rafts, cholesterol-enriched microdomains in the cell membrane, play a role in coronavirus (CoV) entry into cells [21-23] and specifically in the activity of SARS-CoV fusion peptide [24]. Increasing total cellular cholesterol levels promoted ACE2 recruitment to lipid rafts and enhanced SARS-CoV-2 pseudovirus entry [25]. Conversely, Takano et al. showed a role for the cholesterol transport inhibitor U18666A in reducing feline CoV infection [26]. U18666A inhibited both cholesterol biosynthesis and functional activity of the cholesterol transporter Niemann-Pick C1 (NPC1), a receptor that has been implicated in the life cycle of several viruses [26-28], highlighting a need for further studies to evaluate the role of NPC1 in SARS-CoV-2 infection. A recent screen of existing Food and Drug Administration (FDA)-approved drugs for potential activity against SARS-CoV-2 identified cepharanthine as a potent inhibitor of virus-cell attachment [29], with a previous report suggesting it has broad antiviral activity against CoV [30]. This natural compound targets several aspects of cell metabolism including cholesterol trafficking [31]. Statins reduce intracellular and extracellular cholesterol by targeting 3-hydroxy-3-methylglutaryl-Coenzyme A (HMG-CoA) reductase, the rate-limiting first step in the cholesterol biosynthesis (mevalonate) pathway; they therefore also inhibit the synthesis of multiple other biologically active sterols, oxysterols and key isoprenoid intermediates [32,

33]. Their inhibition of protein prenylation via the reduction of isoprenoid intermediates could provide an additional mechanism to inhibit SARS-CoV-2 infectivity (reviewed by Marakasova et al. [34]).

Much of our current knowledge on coronavirus biology was acquired using mouse hepatitis virus (MHV) as a model system. Early studies on the viral encoded structural proteins showed that the MHV spike protein (S) is synthesized as a precursor and intracellular proteolytic cleavage events expose the fusion peptide necessary for viral entry into cells [35]. The SARS-CoV-2 spike protein encodes a furin cleavage site in the S1/S2 junction that differs from SARS-CoV and other closely related bat viruses [36], which may influence particle infectivity. The SARS-CoV-2 spike protein shows a 10- to 20-fold higher affinity for ACE2 than SARS-CoV, which may explain its higher transmissibility [12]. Statins were reported to increase cardiac ACE2 expression in a rabbit model of atherosclerosis [37]; however, further reports in rodent models show a negligible effect of statins on ACE2 expression levels [38, 39]. It will be important to evaluate the impact of statins on ACE2 expression in human lung epithelia as this may impact on viral entry and enhance tissue tropism in early infection (while also promoting anti-inflammatory effects through ACE2 discussed below). However, it is worth noting that coronavirus tropism is not solely defined by the spike protein, with the nucleocapsid, replicase and accessory genes also playing a role (reviewed by Weiss and Leibowitz [40]). Hence one should take care to infer viral pathogenesis from spike protein-ACE2 affinities alone and await further studies on SARS-CoV-2 tissue tropism.

MODULATION OF ANTIVIRAL IMMUNITY

Distinguishing protective from pathogenic immune responses to SARS-CoV-2 will aid the development of successful vaccines and other immunotherapeutic approaches; it will also allow the identification of biomarkers to detect patients mounting effective responses that limit the severity of natural infection. Although these immune correlates are still being defined for SARS-CoV-2, we can speculate how cholesterol may impact on protective immune responses, based on related studies. The complex effects of sterols, and the drugs modulating them, on different components of the immune system have been reviewed previously [33, 41–43]; here we will highlight some relevant examples.

Cholesterol is an essential component of cell membranes and lipid rafts that facilitate immune synapse formation and downstream signalling [44-46]. Intracellular levels of cholesterol are tightly regulated, with the transcription factors liver X receptor (LXR) and sterol regulatory element-binding protein (SREBP) serving as metabolic checkpoints regulating the shift between resting and proliferating T cells [47, 48]. However, cholesterol metabolism and the mevalonate pathway are not only important for effector cells but can potently drive the proliferation of regulatory T cells, enhancing and stabilizing their suppressive capacity [49, 50]. Mice fed a high cholesterol diet upregulate the inhibitory ligand PD-L1 on B cells, which can suppress follicular helper T-cell differentiation to limit the adaptive response [51]. High cholesterol in the tumour environment can suppress CD8⁺ T cells, promoting the transcription of exhaustion markers like PD-1 through the ER stress sensor Xbox-binding protein 1 (XBP1) [52] and inhibiting interleukin-9 (IL-9) expression through LXR sumoylation [53]. Esterified cholesterol accumulates in neutral lipid droplets, an excess of which can be cytotoxic and limit immune cell functionality (as

exemplified by natural killer cells [54]). Thus, several lines of evidence suggest that disrupted cholesterol homeostasis could alter the balance of effector and inhibitory immune responses to limit effective antiviral control.

By blocking cholesterol synthesis, statins may be able to reverse some of the immunosuppressive effects driven by excess cholesterol described above, to promote functional antiviral immune responses [32, 33, 52]. In addition, a number of studies have underscored the immunomodulatory potential of statins beyond their lipid-lowering capacity [32]. By blocking HMG-CoA reductase, statins also reduce synthesis of a number of other relevant mediators including oxysterols, that have multiple immunomodulatory roles [33] and 25-hydroxycholesterol, involved in the Type I interferon (IFN) antiviral response [55]. The statin-induced block in 25-hydroxycholesterol synthesis has also been shown to limit the switching of human CD4⁺T cells from production of the antiviral/pro-inflammatory cytokine IFN- γ to the immunoregulatory IL-10 [56], suggesting they could enhance the antiviral capability of CD4⁺ T cells while limiting their suppressive potential.

A mechanism underlying several of the immunomodulatory effects of statins is the reduced isoprenylation of small GTPases [57, 58]. Of particular relevance to the quest for a vaccine to protect against SARS-CoV-2, lipophilic statins can act as vaccine adjuvants in animal models by inhibiting prenylation of Rab5a in antigen-presenting cells; this arrested endosomal maturation and enhanced antigen retention for presentation, thereby boosting germinal centre B-cell responses for high-affinity antibodies as well as improving CD8⁺ T-cell responses [59]. The adjuvant potential of statins was supported by a small placebocontrolled prospective interventional study showing increased antibody titres to pneumococcal vaccines in healthy subjects given the lipophilic atorvastatin [60]. However, observational studies in elderly people taking long-term statins have shown mixed effects on vaccine responses to influenza [61], raising the possibility that their adjuvant capacity is lost with chronic use.

MODULATION OF INFLAMMATION

There is a delicate balance between the protective antiviral immune responses considered above and uncontrolled bystander responses that drive the cytokine storm and inflammation characterizing the ARDS phase of SARS-CoV-2 infection. Since cholesterol has been reported to induce inflammatory signalling in macrophages and CD11c+ cells [42, 62], pre-existing hypercholesterolaemia may alter the pathological response to SARS-CoV-2. Oxidized low-density lipoprotein (LDL) and cholesterol crystals can activate NOD-, LRR- and pyrin domain-containing protein 3 (NLRP3) inflammasomes to activate IL-1 β and IL-18 production [63]. Accumulation of unesterified cholesterol in macrophages is associated with the production of the proinflammatory cytokines tumour necrosis factor-alpha (TNF-α) and IL-6 [64] that early data suggest are a feature of SARS-CoV-2 immunopathogenesis [13, 14]. Interestingly, cholesterol constitutes the major neutral lipid in lung surfactant, raising the possibility that hypercholesterolaemia can dysregulate the protective properties of surfactant in alveolar spaces [65]. Lipidladen macrophages (foam cells), a common feature in lung pathology, are more likely to accumulate in hypercholesterolaemia [65] and would be expected to predispose to an exaggerated inflammatory response to SARS-CoV-2.

Statins have the potential to limit inflammation by altering a number of mediators from the cholesterol biosynthesis pathway. These anti-inflammatory properties are thought to be central to their protective effects in cardiovascular disease beyond the simple reduction in cholesterol [32]. The many other ways in which statins can exert anti-inflammatory effects include their capacity to stabilize endothelial leakage, limit leukocyte transmigration and increase local nitric oxide [32]. These are likely to be relevant to the pathology of SARS-CoV-2, which can include endothelial cell infection and endotheliitis in several organs [66]. If the published data suggesting that statins can increase the expression of ACE2 in animal models [37] hold out *in vivo* in humans, their impact on the RAS axis would provide a further mechanism by which they could exert an antiinflammatory effect, with ACE2 converting the proinflammatory angiotensin II into angiotensin (1–7), which act on the Mas receptor to attenuate inflammation through nitric oxide [67].

However, the pleiotropic effects of statins make it difficult for *in vitro* and *in vivo* models to predict the full complexity of their action in different disease states in humans. For example, an intervention study of rosuvastatin in healthy normocholesterolaemic subjects resulted in a small increase in circulating inflammatory cytokines [68], whereas pravastatin treatment of subjects with hypercholesterolaemia reduced their production of IL-6 and TNF- α following *in vitro* stimulation with lipopolysaccharide (LPS) [69]. Of note, data from several large randomized trials testing the addition of statins for ARDS [70–72] (of non-SARS-CoV-2 aetiologies) showed no overall benefit or capacity to combat rising levels of IL-18 [73], suggesting they are unlikely to exert useful anti-inflammatory activity if started in the advanced stages of COVID-19.

CONCLUSIONS

Several lines of evidence provide a rationale for high cholesterol predisposing to a worse outcome in SARS-CoV-2 infection, by dysregulating protective immunity and promoting exaggerated pulmonary and systemic inflammatory responses. The capacity of statins to reduce lipids, boost protective immune responses and exert anti-inflammatory properties could all contribute to a potential benefit during SARS-CoV-2 infection.

However, further studies are required to fully understand the impact of hyperlipidaemia and cholesterol-modifying drugs on the clinical course of SARS-CoV-2 infection, with careful adjustment for potential confounders. Statin compliance, shown to be very low in some studies [74], should ideally be assessed and the use of lipophilic and hydrophilic statins examined separately since their differential cell permeability can alter their mechanisms of action [43]. Genetic polymorphisms influencing cholesterol uptake and efflux transporters may also need to be considered [75]. Of particular note, the acute phase response in severe viral infections can be accompanied by transient dyslipidaemia [76] that may not accurately reflect pre-illness levels. This is exemplified by studies in hospitalized patients with SARS-CoV-2, revealing striking reductions in plasma LDL and high-density lipoprotein (HDL) levels, that were associated with disease severity and only returned to the normal range 2 weeks after recovery (in one patient who was followed longitudinally) [77, 78]. Whether these transient reductions and changes in oxidized lipoproteins contribute to disease pathogenesis, as suggested in HIV [79], remains to be investigated. However, these data imply that reliable assessment of associations between dyslipidaemia and SARS-CoV-2 outcome will require data on lipid levels measured prior to, as well as during, the illness.

In conclusion, carefully conducted analysis of available patient data will contribute to further assessment of the relevance of dyslipidaemia and recommendations on the future use of cholesterol-modifying drugs in the setting of SARS-CoV-2. The therapeutic potential of statins across the spectrum, from vaccine trials in healthy individuals to early and advanced phases of SARS-CoV-2, needs further evaluation. We have highlighted several other cholesterol-modifying drugs of potential relevance to SARS-CoV-2 outcome. These, along with statins, merit testing in primary human epithelial cultures and animal models replicating SARS-CoV-2 for their potential range of antiviral, immunomodulatory and anti-inflammatory effects.

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AUTHORS' CONTRIBUTION

All authors carried out literature searches, reviewed published work and contributed to the writing of the manuscript. All authors checked and approved the final version.

CONFLICT OF INTEREST STATEMENT

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DATA AVAILABILITY

No new data were generated or analysed in support of this research.

REFERENCES

- Zhou F, Yu T, Du R et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet* 2020;395:1054–62.
- Bornstein SR, Dalan R, Hopkins D et al. Endocrine and metabolic link to coronavirus infection. Nat Rev Endocrinol 2020;16: 297–8.
- Docherty AB, Harrison EM, Green CA et al. Features of 20 133 UK patients in hospital with covid-19 using the ISARIC WHO Clinical Characterisation Protocol: prospective observational cohort study. BMJ 2020;369. doi:10.1136/bmj.m1985.
- Vaduganathan M, Vardeny O, Michel T et al. Renin–angiotensin–aldosterone system inhibitors in patients with Covid-19. N Engl J Med 2020. doi:10.1056/NEJMsr2005760.
- Adhyaru BB, Jacobson TA. Safety and efficacy of statin therapy. Nat Rev Cardiol 2018;15:757–69.
- De Spiegeleer A, Bronselaer A, Teo JT et al. The effects of ARBs, ACEIs and statins on clinical outcomes of COVID-19 infection among nursing home residents. *medRxiv* 2020: 2020.05.11.20096347.

- Frost FJ, Petersen H, Tollestrup K et al. Influenza and COPD mortality protection as pleiotropic, dose-dependent effects of statins. Chest 2007;131:1006–12.
- 8. Mortensen EM, Nakashima B, Cornell J et al. Population-based study of statins, angiotensin II receptor blockers, and angiotensin-converting enzyme inhibitors on pneumoniarelated outcomes. Clin Infect Dis 2012;55:1466–73.
- Douglas I, Evans S, Smeeth L. Effect of statin treatment on short term mortality after pneumonia episode: cohort study. BMJ 2011;342:d1642.
- 10. Bloom PP, Meyerowitz EA, Reinus Z et al. Liver biochemistries in hospitalized patients with COVID-19. Hepatology. doi:10.1002/hep.31326.
- 11. Fedson DS, Opal SM, Rordam OM. Hiding in plain sight: an approach to treating patients with severe COVID-19 infection. *mBio* 2020;11:e00398–20.
- 12. Hoffmann M, Kleine-Weber H, Schroeder S et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell* 2020;**181**: 271–80.e8.
- 13. Cao X. COVID-19: immunopathology and its implications for therapy. Nat Rev Immunol 2020;20:269–70.
- 14. Feldmann M, Maini RN, Woody JN et al. Trials of anti-tumour necrosis factor therapy for COVID-19 are urgently needed. *Lancet* 2020. doi:10.1016/S0140-6736(20)30858-8.
- 15. Heaton NS, Randall G. Multifaceted roles for lipids in viral infection. Trends Microbiol 2011;19:368–75.
- 16. Farag, A, Wang, P, Boys IN et al. Identification of atovaquone, ouabain and mebendazole as FDA approved drugs Tar-geting SARS-CoV-2 (Version 4). 2020. doi:10.26434/chemrxiv. 12003930.v4.
- 17. Zhou Y, Hou Y, Shen J et al. Network-based drug repurposing for novel coronavirus 2019-nCoV/SARS-CoV-2. Cell Discov 2020;6:1–18.
- Gordon DE, Jang GM, Bouhaddou M et al. A SARS-CoV-2 protein interaction map reveals targets for drug repurposing. Nature 2020:1–13.
- 19. Wei C, Wan L, Zhang Y *et al*. Cholesterol Metabolism–Impact for SARS-CoV-2 infection prognosis, entry, and antiviral therapies. *medRxiv* 2020:2020.04.16.20068528.
- 20. Rowe IA, Tully DC, Armstrong MJ et al. Effect of scavenger receptor class B type I antagonist ITX5061 in patients with hepatitis C virus infection undergoing liver transplantation. *Liver Transpl* 2016;**22**:287–97.
- 21. Guo H, Huang M, Yuan Q et al. The important role of lipid raftmediated attachment in the infection of cultured cells by coronavirus infectious bronchitis virus beaudette strain. *PLoS One* 2017;**12**:e0170123.
- 22.Li G-M, Li Y-G, Yamate M et al. Lipid rafts play an important role in the early stage of severe acute respiratory syndromecoronavirus life cycle. Microbes Infect 2007;9:96–102.
- Jeon JH, Lee C. Cholesterol is important for the entry process of porcine deltacoronavirus. Arch Virol 2018;163: 3119–24.
- 24. Meher G, Bhattacharjya S, Chakraborty H. Membrane cholesterol modulates oligomeric status and peptide-membrane interaction of severe acute respiratory syndrome coronavirus fusion peptide. J Phys Chem B 2019;**123**:10654–62.
- 25. Wang H, Yuan Z, Pavel MA *et al*. The role of high cholesterol in age-related COVID19 lethality. *bioRxiv* 2020:2020.05.09. 086249.
- 26. Takano T, Endoh M, Fukatsu H et al. The cholesterol transport inhibitor U18666A inhibits type I feline coronavirus infection. *Antiviral Res* 2017;**145**:96–102.

- 27.Gong X, Qian H, Zhou X et al. Structural Insights into the Niemann-Pick C1 (NPC1)-mediated cholesterol transfer and ebola infection. Cell 2016;**165**:1467–78.
- Stoeck IK, Lee J-Y, Tabata K et al. Hepatitis C virus replication depends on endosomal cholesterol homeostasis. J Virol 2018; 92:e01196–17.
- 29. Ohashi H, Watashi K, Saso W et al. Multidrug treatment with nelfinavir and cepharanthine against COVID-19. bioRxiv 2020: 2020.04.14.039925.
- 30. Kim DE, Min JS, Jang MS et al. Natural bis-benzylisoquinoline alkaloids-tetrandrine, fangchinoline, and cepharanthine, inhibit human coronavirus OC43 infection of MRC-5 human lung cells. *Biomolecules* 2019;**9**:696.
- 31. Bailly C. Cepharanthine: an update of its mode of action, pharmacological properties and medical applications. Phytomedicine Int J Phytother Phytopharm 2019;62:152956.
- 32. Jain MK, Ridker PM. Anti-inflammatory effects of statins: clinical evidence and basic mechanisms. Nat Rev Drug Discov 2005;4:977–87.
- 33. Spann NJ, Glass CK. Sterols and oxysterols in immune cell function. Nat Immunol 2013;14:893–900.
- 34. Marakasova ES, Eisenhaber B, Maurer-Stroh S et al. Prenylation of viral proteins by enzymes of the host: virusdriven rationale for therapy with statins and FT/GGT1 inhibitors. Bioessays 2017;**39**:1700014.
- 35. Frana MF, Behnke JN, Sturman LS *et al*. Proteolytic cleavage of the E2 glycoprotein of murine coronavirus: host-dependent differences in proteolytic cleavage and cell fusion. *J* Virol 1985;**56**:912–20.
- 36. Coutard B, Valle C, de Lamballerie X et al. The spike glycoprotein of the new coronavirus 2019-nCoV contains a furin-like cleavage site absent in CoV of the same clade. Antiviral Res 2020;176:104742.
- 37. Tikoo K, Patel G, Kumar S et al. Tissue specific up regulation of ACE2 in rabbit model of atherosclerosis by atorvastatin: Role of epigenetic histone modifications. Biochem Pharmacol 2015; 93:343–51.
- 38. Shin YH, Min JJ, Lee J-H et al. The effect of fluvastatin on cardiac fibrosis and angiotensin-converting enzyme-2 expression in glucose-controlled diabetic rat hearts. *Heart Vessels* 2017;**32**:618–27.
- 39. Min JJ, Shin B-S, Lee J-H et al. Effects of pravastatin on Type 1 diabetic rat heart with or without blood glycemic control. J Diabetes Res 2018;2018:1067853.
- Weiss SR, Leibowitz JL. Coronavirus pathogenesis. Adv Virus Res 2011;81:85–164.
- Fessler MB. The intracellular cholesterol landscape: dynamic integrator of the immune response. Trends Immunol 2016;37: 819–30.
- 42. Tall AR, Yvan-Charvet L. Cholesterol, inflammation and innate immunity. Nat Rev Immunol 2015;15:104–16.
- Parihar SP, Guler R, Brombacher F. Statins: a viable candidate for host-directed therapy against infectious diseases. Nat Rev Immunol 2019;19:104–17.
- 44. Bromley SK, Burack WR, Johnson KG et al. The immunological synapse. Annu Rev Immunol 2001;**19**:375–96.
- 45. Sezgin E, Levental I, Mayor S et al. The mystery of membrane organization: composition, regulation and roles of lipid rafts. Nat Rev Mol Cell Biol 2017;**18**:361–74.
- 46. Bietz A, Zhu H, Xue M et al. Cholesterol metabolism in T cells. Front Immunol 2017;**8**:1664.
- 47. Bensinger SJ, Bradley MN, Joseph SB et al. LXR signaling couples sterol metabolism to proliferation in the acquired immune response. *Cell* 2008;**134**:97–111.

- 48.Kidani Y, Elsaesser H, Hock MB et al. Sterol regulatory element-binding proteins are essential for the metabolic programming of effector T cells and adaptive immunity. Nat Immunol 2013;14:489–99.
- 49. Timilshina M, You Z, Lacher SM et al. Activation of mevalonate pathway via LKB1 is essential for stability of Treg cells. *Cell Rep* 2019;**27**:2948–61.e7.
- 50.Zeng H, Yang K, Cloer C *et al.* mTORC1 couples immune signals and metabolic programming to establish T reg-cell function. *Nature* 2013;**499**:485–90.
- 51. Nus M, Sage AP, Lu Y et al. Marginal zone B cells control the response of follicular helper T cells to a high-cholesterol diet. Nat Med 2017;**23**:601–10.
- 52. Ma X, Bi E, Lu Y et al. Cholesterol induces CD8+ T cell exhaustion in the tumor microenvironment. *Cell Metab* 2019;**30**: 143–56.e5.
- 53. Ma X, Bi E, Huang C *et al*. Cholesterol negatively regulates IL-9–producing CD8+ T cell differentiation and antitumor activity. *J Exp Med* 2018;**215**:1555–69.
- 54. Michelet X, Dyck L, Hogan A *et al*. Metabolic reprogramming of natural killer cells in obesity limits antitumor responses. Nat Immunol 2018;**19**:1330–40.
- 55. Cyster JG, Dang EV, Reboldi A et al. 25-Hydroxycholesterols in innate and adaptive immunity. Nat Rev Immunol 2014;**14**:731–43.
- 56. Perucha E, Melchiotti R, Bibby JA et al. The cholesterol biosynthesis pathway regulates IL-10 expression in human Th1 cells. Nat Commun 2019;**10**:1–13.
- 57. Thurnher M, Gruenbacher G. T lymphocyte regulation by mevalonate metabolism. Sci Signal 2015;8:re4.
- 58. Greenwood J, Steinman L, Zamvil SS. Statin therapy in autoimmunity: from protein prenylation to immunomodulation. Nat Rev Immunol 2006;6:358–370.
- 59.Xia Y, Xie Y, Yu Z *et al*. The mevalonate pathway is a druggable target for vaccine adjuvant discovery. *Cell* 2018;**175**: 1059–73.e21.
- 60. Wildes TJ, Grippin A, Fasanya H et al. Effect of atorvastatin on humoral immune response to 23-valent pneumococcal polysaccharide vaccination in healthy volunteers: the StatVax randomized clinical trial. *Vaccine* 2019;**37**:1313–24.
- 61. Black S, Nicolay U, Del Giudice G et al. Influence of statins on influenza vaccine response in elderly individuals. J Infect Dis 2016;**213**:1224–8.
- 62.Ito A, Hong C, Oka K et al. Cholesterol accumulation in CD11c+ immune cells is a causal and targetable factor in autoimmune disease. *Immunity* 2016;**45**:1311–26.
- 63. Duewell P, Kono H, Rayner KJ et al. NLRP3 inflammasomes are required for atherogenesis and activated by cholesterol crystals. Nature 2010;**464**:1357–61.
- 64.Li Y, Schwabe RF, DeVries-Seimon T et al. Free cholesterolloaded macrophages are an abundant source of tumor necrosis factor-alpha and interleukin-6: model of NF-kappaB- and map kinase-dependent inflammation in advanced atherosclerosis. J Biol Chem 2005;**280**:21763–72.
- 65.Fessler MB. A new frontier in immunometabolism. Cholesterol in lung health and disease. Ann Am Thorac Soc 2017;**14**:S399–405.
- 66. Varga Z, Flammer AJ, Steiger P et al. Endothelial cell infection and endotheliitis in COVID-19. *Lancet* 2020;**395**:1417–8.
- 67. South AM, Tomlinson L, Edmonston D et al. Controversies of renin–angiotensin system inhibition during the COVID-19 pandemic. Nat Rev Nephrol 2020:1–3.
- 68.Karmaus PWF, Shi M, Perl S et al. Effects of rosuvastatin on the immune system in healthy volunteers with normal serum cholesterol. JCI Insight 2019;4:e131530.

- Rosenson RS, Tangney CC, Casey LC. Inhibition of proinflammatory cytokine production by pravastatin. Lancet 1999;353: 983–4.
- 70. McAuley DF, Laffey JG, O'Kane CM et al. Simvastatin in the acute respiratory distress syndrome. N Engl J Med 2014;371: 1695–703.
- 71. Papazian L, Roch A, Charles P-E et al. Effect of statin therapy on mortality in patients with ventilator-associated pneumonia: a randomized clinical trial. JAMA 2013;**310**: 1692–700.
- 72. National Heart, Lung, and Blood Institute ARDS Clinical Trials Network, Truwit JD, Bernard GR, Steingrub J et al. Rosuvastatin for sepsis-associated acute respiratory distress syndrome. N Engl J Med 2014;370:2191–200.
- 73. Rogers A, Guan J, Trtchounian A et al. Association of elevated plasma interleukin-18 level with increased mortality in a clinical trial of statin treatment for acute respiratory distress syndrome. Crit Care Med 2019;47:1089–96.

- 74. Colantonio LD, Rosenson RS, Deng L et al. Adherence to statin therapy among US adults between 2007 and 2014. *J Am Heart* Assoc 2019;**8**:e010376.
- 75.Guan Z-W, Wu K-R, Li R et al. Pharmacogenetics of statins treatment: efficacy and safety. J Clin Pharm Ther 2019;44: 858–67.
- 76. Cabana VG, Siegel JN, Sabesin SM. Effects of the acute phase response on the concentration and density distribution of plasma lipids and apolipoproteins. J Lipid Res 1989;30:39–49.
- 77. Hu X, Chen D, Wu L et al. Low serum cholesterol level among patients with COVID-19 infection in Wenzhou, China. SSRN Electron J 2020. doi:10.2139/ssrn.3544826.
- 78. Wei X, Zeng W, Su J et al. Hypolipidemia is associated with the severity of COVID-19. J Clin Lipidol 2020. doi:10.1016/ j.jacl.2020.04.008.
- 79. Kelesidis T, Jackson N, McComsey GA *et al*. Oxidized lipoproteins are associated with markers of inflammation and immune activation in HIV-1 infection. AIDS 2016;**30**:2625–33.