Can blocking inflammation enhance immunity during ageing?

Emma S Chambers^{1#} and Arne N Akbar¹

1 – Division of Infection and Immunity, The Rayne Building, 5 University Street, University College London, London, WC1E 6EJ

Corresponding author:

Dr Emma Chambers, Division of Infection and Immunity, The Rayne Building, 5 University Street, University College London, London, WC1E 6EJ.

E-mail: <u>emma.chambers@ucl.ac.uk</u>

Tel: 020 31082179

Keywords: Inflammageing; senescence; p38-MAP Kinase; senolytics;

Abbreviations

β-gal	- β-galactosidase
CMV	- Cytomegalovirus
сох	- cyclooxygenase
CRP	- C Reactive protein
DAMPs	- damage associated molecular patterns
EBV	- Epstein Barr Virus
HMGB1	- high mobility group box 1
IL	- Interleukin
LPS	- Lipopolysaccharide
MAP	- mitogen-activated protein
MMP	- matrix metalloproteinases
mTOR	- The mammalian target of rapamycin
p16	- CDK4/6 inhibitor p16INK4A
PD-1	- Programmed cell death protein -1
PDL-1	- Programmed death ligand - 1
PGE ₂	Prostaglandin E2
PRR	- pattern recognition receptor
SASP	- senescence associated secretory phenotype
T _{EMRA}	- T effector memory cells that re-express CD45
TGFβ	- Transforming growth factor
TLR	- Toll-like receptor
TNF	- Tumour necrosis factor
TORC1	- mTOR complex 1
TORC2	- mTOR complex 2

Abstract

Ageing is a global burden and the increase in lifespan does not increase in parallel with health-span. Therefore, older adults are currently living longer with chronic diseases, increased infections and cancer. A characteristic of ageing is the presence of chronic low grade inflammation that is characterised by elevated concentrations of IL-6, TNFα and C-Reactive protein (CRP) that has been termed inflammageing (1). Previous studies have demonstrated that chronic inflammation interferes with T cell response and macrophage function and is also detrimental for vaccine responses. This raises the question of whether therapeutic strategies that reduce inflammation may be useful for improving immunity in older adults. In this review we discuss the potential causes of inflammageing, the cellular source of the inflammatory mediators and the mechanisms by which inflammation that have been used to enhance immunity during ageing.

1. Introduction:

Ageing results in increased susceptibility to infections, reduced vaccine responses and increased susceptibility to cancers (2-4). This is due to changes in both the adaptive and innate immune system which have been reviewed previously (5, 6). The focus of this review is to describe how inflammation and in particular the phenomenon of inflammageing impact on the immune system and to discuss current therapies which are being developed to counteract the inflammatory processes that occur with ageing.

2. Inflammageing

Inflammageing, a term first proposed by Claudio Franceschi, is the state of chronic lowgrade sterile inflammation which is observed with age. It is characterised by high serum concentrations of C Reactive protein (CRP) and other inflammatory mediators such as Interleukin (IL)-6, IL-8 and TNF α (1). The increase in these inflammatory mediators occurs in healthy older adults in the absence of overt inflammatory disease. However, elevated circulating concentrations of IL-6, CRP and Tumour necrosis factor (TNF) α predicts frailty in older subjects (7, 8). Inflammageing is also associated with increased risk of mortality, in healthy and frail older adults (9-12). In addition to elevated IL-6, CRP and TNF α , IL-1 β and inflammasome related genes are also good predictors of all-cause mortality (13). Conversely lower levels of inflammatory cytokines in the peripheral blood correlate with good health outcomes and reduced risk of death of older adults (9).

It has also been shown that inflammation and inflammageing associated cytokines are linked with poor cognitive function and elevated concentrations of plasma IL-6 and CRP are associated with cognitive decline (14-16). Furthermore, inflammageing is associated with chronic diseases such as type 2 diabetes, rheumatoid arthritis and even Alzheimer's disease, however whether this is cause or effect requires further investigation (17).

3. Inflammation and immunity

An acute inflammation is necessary to initiate an immune response against an invading pathogen. After the initial inflammatory response there is a period of resolution which occurs to prevent unnecessary tissue damage and to restore tissue homeostasis (18). However, there is accumulating data showing that chronic inflammation can inhibit immunity *in vivo*. As elevated inflammatory responses are detrimental for vaccine efficacy against influenza (19), yellow fever (20) and hepatitis B. In addition, excessive inflammation in particular TNF α production is linked to decreased killing and clearance of *Streptococcus pneumonia* in

macrophages in an aged mouse model of infection (21). This may occur in part by the induction of premature monocyte egress from the bone marrow by TNF α that impairs their function. Interestingly blockade of TNF α restores monocyte function in old animals, showing that the effect of chronic inflammation can be reversed (22).

Elevated inflammation can also inhibit the response to cutaneous recall antigens *in vivo*. Older humans have decreased response to challenge with antigens such as tuberculin PPD, *candida albicans* antigens and varicella zoster virus (VZV) antigens in the skin compared to young individuals (23). However this was not due to a decrease in the number of circulating or resident memory T cells (23, 24). Instead these subjects exhibit elevated inflammatory responses induced by the injection itself, the extent of which was negatively correlated with their ability to respond to the antigen (25). The temporary inhibition of systemic inflammation with an oral p38 mitogen-activated protein (MAP) kinase inhibitor enhances the response to antigen in older subjects indicating the direct association between excessive inflammation and immune inhibition (26).

4. Source of inflammation

The exact source of elevated inflammation during ageing is may be due to a combination of the following mechanisms that are accentuated in older adults. These include chronic viral infection leading to immune activation, increased inflammatory mediator secretion from visceral fat, increased gut permeability resulting in leakage of bacterial components into the circulation, increase in damage associated molecular patterns (DAMPs), altered immune resolution and accumulation of senescent cells, as shown in Figure 1.

4.1 Chronic viral infections

Chronic infections, which cause a lifelong latent infection, are believed to lead to long term activation of the immune system over time, contributing to inflammageing. The most studied example is cytomegalovirus (CMV) infection that induces a lifelong latent infection after the primary infection, and the virus reactivates periodically and initiates a subclinical immune response. A large proportion of T cells in seropositive older subjects are CMV-specific (27, 28) and these cells are highly differentiated and express senescence-associated markers like CD57 and KLRG1. Furthermore these cells produce high levels of inflammatory cytokines such as IL-2, IFN γ and TNF α after activation (29) that may contribute to inflammageing. Individuals who are CMV seropositive and exhibit elevated CRP levels have increased all-cause mortality as compared to CMV seropositive subjects with low CRP levels

(30). However, the impact of CMV infection on the elevated inflammation in older subjects is controversial (31).

4.2 Increased visceral fat

Obesity and in particular accumulation of visceral fat is highly associated with inflammatory cytokine production (32). Visceral fat is an inflammatory site, that has an infiltration of mononuclear phagocytes, B cells and T cells which contribute to the production of inflammatory cytokines such as IL-6, IL-1 β and TNF α (33). In young obese individuals, there is an alteration in their circulating leukocyte populations, that resemble the cells found in older adults. There is an increase in circulating end stage senescent-like CD4⁺ and CD8⁺ cells that secrete high levels of inflammatory cytokines after activation and decreased naïve T cells (34). Apart from changes in immune cells resident in fat tissue, there is an increase in visceral adiposity during ageing, due in part to the age related decrease in muscle (35). This increase in visceral fat will contribute to inflammageing due to the inflammatory cytokines produced from the adipocytes themselves (32).

4.3 Gut permeability

Studies performed from aged mouse models have shown that older mice have more permeable intestines with a breakdown in cell-to-cell contacts which leads to leakage of gut contents into the blood stream (21, 36). This results in an increase in bacterial components such as Lipopolysaccharide (LPS) in the circulation which activate circulating mononuclear phagocytes through pattern recognition receptor (PRR) expressed by the monocytes and results in production of inflammatory cytokines such as TNFα and IL-6 (21, 22). In addition there are alterations in the microbiome of older adults that renders them distinct from younger cohorts (37). Ageing is associated with an increase in opportunistic proinflammatory bacteria, termed 'pathobionts', which are normally only observed in low numbers in young guts (38). Older adults with the most evident altered gut microbiome had elevated circulating inflammatory cytokines, implying that inflammageing is linked to alteration in microbiome (39). However, whether this dysbiosis is as a result of altered gut permeability rather than causative of increased inflammation still warrants further investigation. Evidence from an aged drosophila model have shown that microbiome dysbiosis precedes the increased gut permeability observed with age and thus microbiome dysbiosis could be a causative factor in the increase gut permeability seen with age (40).

4.4 Increase in DAMPs

DAMPs are endogenous cellular components which are released at times of injury, stress or cell death. DAMPS can consist of a variety of cellular products including; the S100 family of calcium binding proteins, histones, genomic or mitochondrial DNA or other secreted factors such as ATP, uric acid or heparin sulphate. This is not an exhaustive list and DAMPs have been extensively reviewed previously (41). When DAMPs bind to their PRR receptor there is an increase in inflammatory cytokine production from the cell.

It is proposed that the processes involved in ageing result in increased DAMP production which contribute to inflammageing (42). There is limited human data to support this hypothesis; however in aged murine studies it was observed that there was elevated levels of high mobility group box 1 (HMGB1) has been observed in old mice (43), HMGB1 is an alarmin family member and binds to surface Toll-like receptor (TLR)2 and TLR4 resulting in the production of inflammatory cytokines including IL-6 (44). In addition, NLRP3 inflammasome, an innate immune sensor that is activated in response to an array of DAMPs, was found to be elevated in aged mice. Removing the NLRP3 gene from these mice resulted in a reducing in aged related inflammation; implying DAMPs are involved in the process of inflammageing (45).

4.5 Ineffective immune resolution

After an acute inflammatory response to an infectious agent or traumatic event such as a wound healing response there is a period of immune resolution where the tissue is restored back to its original state (18). Cells involved in resolution of inflammation include mononuclear phagocytes and stromal cells in addition in addition to lipid mediators such as prostaglandins that are involved (18). A recent study has shown that although the onset of acute inflammation between older and younger adults is similar the resolution of the inflammation is impaired in the older adults (46). Indeed there was reduced efferocytosis and clearance of apoptotic neutrophils by the mononuclear phagocytes during the resolution phase of the inflammatory response, which led to a failure to resolve inflammation. This was due in part to reduced expression of TIM-4, a receptor that recognizes apoptotic cells, on mononuclear phagocytes (46). This means that acute inflammatory events are not efficiently resolved in older individuals which could contribute to inflammatory.

4.6 Senescent cell accumulation with age

Cells entering a state of senescence experience irreversible growth arrest that occurs as a result of the irreparable cell damage e.g. DNA damage, telomere erosion or oxidative stress (47). Senescence is a protective process that prevents the proliferation of damaged cells and is viewed as a tumour suppressor mechanism (48, 49). However recent evidence suggests that senescent cells may have beneficial effects and can contribute to wound healing in the skin (50). Senescent cells are characterised by the expression of CDK4/6 cyclin inhibitor p16INK4A (p16) and/or β -galactosidase (β -gal) however this is not an exhaustive list and markers of senescence have been reviewed extensively elsewhere (47).

Ageing is associated with accumulation of senescent cells throughout the body and has been shown to occur in every experimental species and organ studied to date, including mouse and primate models (51-54). In humans senescent cells accumulate in the skin (55, 56) and kidney (57) during ageing. The cell types that are senescent in these tissues include fibroblasts, melanocytes and endothelial cells (50, 56, 58). Senescent cells may accumulate due to long-term exposure to DNA damaging agents such as ultra violet B (UVB) and exposure to pollutants (47). However a recent paper showed that there is reduced elimination of senescent cells during ageing that would also account for their accumulation (59). This reduced clearance may be due in part to the expression of HLA-E by senescent cells that binds to the inhibitory receptor NKG2A expressed on NK and CD8⁺ T cells which inhibits their cytotoxic activity (56). It is possible that other inhibitory receptor/ligand pairs may also be involved in this evasion strategy that enables senescent cell persistence during ageing.

Senescent cells can secrete a range of inflammatory cytokines (such as IL-1 β , IL-6 and TNF α), chemokines (such as CCL2 and IL-8), growth factors (such as fibroblast growth factor) and matrix metalloproteinase (MMP) (such as MMP1 and MMP3). DAMPs such as HMGB1, are also secreted contributing to the inflammatory phenotype of senescent cells (43). This secretion of pro-inflammatory mediators is known as the senescence associated secretory phenotype (SASP). Multiple components of the SASP including CCL2, Transforming growth factor (TGF β) and IL-1 α have the ability to drive senescence in a paracrine manner in nearby non-senescent cells, thus overall increasing the number of senescent cells (60). All components of SASP contribute to the local inflammatory environment and may contribute to the inflammageing phenomenon (61).

Although the majority of senescence research has focussed on cells in tissues, there are also populations of circulating senescent-like leukocytes that accumulate during ageing (62). Examples of senescent-like leukocytes include terminally differentiated CD4⁺ and CD8⁺ T effector memory cells that re-express CD45RA (T_{EMRA}), these cells have low proliferative capacity and secrete inflammatory mediators (63-65). Terminally differentiated NK cells also

accumulate during ageing and these CD16^{dim}KLRG1⁺ cells have increased inflammatory cytokine production (66). All these senescent-like leukocytes in older individuals may also contribute to inflammageing.

5. How does inflammation inhibit immunity?

There are direct effects that inflammation has on immunity, such as the suppressive effect of TNF α on T cell receptor signalling (67) and monocyte phagocytosis (22). There are also other mechanisms by which inflammation inhibits immunity, including increasing the expression of inhibitory receptors, increasing the number and function of Foxp3⁺ T regulatory cells (Tregs) and increasing monocyte infiltration of the tissue.

5.1 Increase of inhibitory ligands and receptors

There is increasing evidence that inflammageing associated cytokines can increase expression of inhibitory ligands on immune cells. An example of this is $TNF\alpha$, which increases expression of Programmed death ligand 1 (PDL-1) on antigen presenting cells such as mononuclear phagocytes (68). PDL-1 binds to Programmed cell death protein-1 (PD-1) that is expressed on T cells, that leads to apoptosis of the cell. The increase in PDL-1 expression is particularly relevant as increased expression of PD-1 on T cells has been shown on in the skin and peripheral blood populations of these cells in older adults that renders them more susceptible to inhibition (24).

5.2 Inflammation effects on Foxp3⁺ Tregs

Tregs are defined by the transcription factor Foxp3, and they play an important role in maintaining immune homeostasis, as in the absence of these cells, there is widespread autoimmune and inflammatory disease which leads to early death (69, 70). There are increased number of Foxp3⁺ Tregs in the skin of older subjects at baseline and in response to antigen that contribute to decreased cutaneous antigen-specific immune responses during ageing (71, 72). It has been proposed that the accumulation of Foxp3⁺ Tregs in the skin of older adults may be due to inflammatory processes since Foxp3⁺ Tregs are recruited to sites of inflammation (73). Interestingly, inflammageing associated cytokines such as TNF α can increase Foxp3⁺ Treg number and induce them to become more suppressive (74). As a result, inflammageing may induce increased numbers and function of Foxp3⁺ Tregs that can inhibit immunity.

5.3 Senescent cells recruit inflammatory cells via SASP

One major component of the SASP is monocyte chemoattractant chemokines such as CCL2 (56). When monocytes are recruited into tissues and exposed to inflammatory signals, they upregulate immune resolution pathways such as CD39/CD73 and PDL-1 which all of which may have a role in inhibiting immunity (75, 76). Indeed in patients with coronary artery disease, their mononuclear phagocytes from the periphery and atherosclerotic plaques have increased expression of PDL-1 which specifically inhibits antigen-specific T cells in a PD-1 dependent manner (75). It has been shown that there is a negative correlation between the number of monocytes recruited to a site of inflammation and cutaneous antigen-specific immunity (26).

5.4 Immunoregulatory SASP components

Not all components of the SASP can be considered directly inflammatory. Indeed, TGFβ an early component of SASP (77), has been shown to have the potential to generate Foxp3⁺ Tregs from CD4⁺ T effector cells (78). Another SASP component is the lipid mediator Prostaglandin E2 (PGE₂), which is a downstream of cyclooxygenase (COX)2 (79, 80). PGE₂ can promote a more tolerogenic environment by increasing production of the immunoregulatory cytokine IL-10 from mononuclear phagocytes as well increasing the number and function of Foxp3⁺ Tregs (81, 82). In addition, PGE₂ has been shown to inhibit the antigen-specific immunity by blocking proliferation of CD8⁺ T cells in response to viral antigens (83, 84).

6. Therapeutic targets of inflammageing

Due to the substantial clinical data linking inflammageing with reduced immunity, health and increased mortality in older adults, it has become a crucial therapeutic target in older adults. The current therapies for reducing inflammation that have been proposed include the removal of senescent cells and mTOR and p38-MAPK Kinase inhibition (Figure 2).

6.1 Senescent cells

As senescent cells are a major contributor to the inflammageing process they are an exciting target for reducing inflammageing. Mouse models have been developed where senescent cells can be specifically removed *in vivo* and these studies showed that these animals have

increased lifespan, improved fitness and reduced fur loss (85, 86). Indeed, removal of senescent cells even after onset of age-related disorders, such as sarcopenia and cataracts, resulted in an attenuation of disease pathology (87). As a result of these exciting murine studies, therapies to remove senescent with drugs termed senolytics, has been an active area of research. Assessments of senescent cell behaviour *in vitro* - identified that anti-apoptotic/pro-survival pathways such as BCL2, p53 and CDKN1A and also Phosphoinositide 3-Kinase δ signalling pathways may represent specific pathways that can be targeted for their elimination (88, 89).

Senolytics that have been tested in aged mouse models include the combination of dasatinib and quercetin that significantly reduced vascular pathologies (90). ABT263, a specific inhibitor for BCL2 and BCL-x, was utilised in an aged mouse model and has resulted in a rejuvenation of hematopoietic stem cells (91). Inhibitors of heat shock protein (HSP)90 have prevented onset of age-related pathologies in mice (92). All these senolytic agents have been shown to significantly reduce the senescent cells in the mice and while they show great promise in mouse models, they have yet to be translated to humans and this is an area of intense investigation.

Another potential therapeutic area is to unleash the activity of the individuals own immune system against senescent cells. Since there are strategies that enable the evasion of senescent cells from immune surveillance, preventing this inhibitory axis would facilitate the recognition and removal of senescent cells in old subjects. As the expression of the inhibitory receptor HLA-E prevents NK and CD8⁺ T cells from killing senescent cells (56) the blocking interactions between the negative ligand HLA-E and its receptor NKG2A would be a strategy to enhance senescent cell clearance *in vivo*. An anti-NKG2A monoclonal antibody, Monalizumab, has been developed as a check point inhibitor, that has been shown to enhance anti-tumour immunity via enabling the NK and CD8⁺ T cell to kill tumours cells that also express HLA-E (93, 94). It is possible that Monalizumab may also have the potential to be used as a senolytic agent to remove senescent cells from older adults and this requires further investigation.

6.2 mTOR

The mammalian target of rapamycin (mTOR), is comprised of two distinct protein complexes mTOR complex 1 (TORC1) and mTOR complex 2 (TORC2) that are involved with numerous cellular processes including inflammation. mTOR is involved in many inflammatory process, in particular mTOR signalling is downstream of a number of innate immune cell receptors such as TLRs including TLR4, cytokine receptors such as IL-15 and lipid receptors such as

Prostaglandin receptors, all of which can increase inflammatory mediator production from cells (95). In addition, mTOR has been shown to be a regulator of the SASP in senescent cells via promoting IL-1α production (96). mTOR inhibition, via Rapamycin, has been shown to increase life expectancy (97). However, more recently it has also been used to improve vaccine responses in older adults *in vivo* (98). Mannick *et al* treated older subjects with a specific TORC1 inhibitor called RAD001 prior to Influenza vaccination, they found that there was an enhanced response to vaccination as determined by circulating antibody titres. This improvement in vaccine response was proposed to be due to reduced expression of the inhibitory receptor PD-1 on circulating CD4⁺ and CD8⁺ T cells (98). A subsequent study by the same group, demonstrated that TORC1 inhibitor treatment prior to vaccination also significantly reduced influenza infections in older subjects (99). However, it is not clear if Rapamycin is acting directly on the inflammation in these subjects on some other process to enhance vaccine efficacy.

6.3 p38 MAP Kinase

p38 MAP Kinase has been shown to be a major signalling molecule upstream of SASP production from senescent fibroblast and from CD8⁺ T_{EMRA} cells (100-102). It was shown that a non-specific inflammatory response occurs after mild tissue injury after saline injection in the skin of older but not adults and that this was associated with to p38 MAP Kinase signalling (26). The older subjects also has increased numbers of senescent cells in the skin compared to younger individuals. The observed inflammatory response was reminiscent of inflammageing and correlated negatively with their response to recall antigen challenge (varicella zoster virus antigens) in the skin (26). To test directly if the inflammation observed was responsible for decreasing the immune response, old subjects were pre-treated with an oral p38 MAP Kinase inhibitor (Losmapimod) for four days before injection of the antigen (103). It was found that blocking p38 in vivo significantly increased cutaneous immunity that was associated with an increase in T cell recruitment to the site of antigen challenge (26). Therefore, in addition to senescent cell elimination as a strategy to reduce inflammation, the inflammatory response itself can be manipulated in older individuals with benefit to immunity. It remains to be determined if the inhibition of inflammation may also alleviate other facets of frailty during ageing. However, while short term inhibition would be acceptable, longer term inhibition, especially with p38 MAP Kinase inhibitors is associated with hepatotoxicity (104).

7. Future perspectives

Inflammageing is caused by a combination of age-related defects including increased DAMP production, increased gut permeability, increased visceral fat, chronic infections and increase in senescent cell numbers. Senescent cells contribute to inflammageing due to their SASP production which includes a wide range of inflammatory cytokines and DAMPS. Therefore, strategies to remove the senescent cells from the body are a promising therapeutic target. There has been extensive research in mouse models to show that removal of senescent cells from an old mouse renders the mouse young again. However, what the long-term implications are for removing senescent structural cells, such as fibroblasts, from tissues when they make up a major proportion of the tissue structure needs further investigation. An exciting potential drug candidate to targeting inflammageing is Metformin - which activates the AMP-activated protein kinase signalling pathway and thus blocking inflammatory-cytokine signalling, has been successful used as a long-term therapy in older adults as a first-line therapy for type 2 diabetes, it has been shown to improve cardiovascular health in these individuals (105). However, strategies that target inflammatory signalling pathways using Rapamycin and Metformin have been utilised with some success, but the effect of longer-term inhibition and potential side-effects are not clear at present.

Current therapies that have been developed utilise a short-term inhibition of inflammation to boost immunity without side effects in older individuals may be of benefit to as an adjunct to vaccination and/or anti-tumour therapy. A combination of approaches including one or more of senolytic drug, checkpoint inhibitors (anti-NKG2A) and anti-inflammatory agents may be required for optimal blocking of inflammageing to reduce frailty and enhance immunity in older adults.

References:

1. Franceschi C, Garagnani P, Vitale G, Capri M, Salvioli S. Inflammaging and 'Garbaging'. Trends in endocrinology and metabolism: TEM. 2017;28(3):199-212.

2. Gavazzi G, Krause KH. Ageing and infection. Lancet Infect Dis. 2002;2(11):659-66.

3. Diffey BL, Langtry JA. Skin cancer incidence and the ageing population. Br J Dermatol. 2005;153(3):679-80.

Ciabattini A, Nardini C, Santoro F, Garagnani P, Franceschi C, Medaglini D.
 Vaccination in the elderly: The challenge of immune changes with aging. Semin Immunol.
 2018;40:83-94.

5. Akbar AN, Henson SM, Lanna A. Senescence of T Lymphocytes: Implications for Enhancing Human Immunity. Trends Immunol. 2016;37(12):866-76.

6. Shaw AC, Joshi S, Greenwood H, Panda A, Lord JM. Aging of the innate immune system. Curr Opin Immunol. 2010;22(4):507-13.

7. Cohen HJ, Pieper CF, Harris T, Rao KM, Currie MS. The association of plasma IL-6 levels with functional disability in community-dwelling elderly. J Gerontol A Biol Sci Med Sci. 1997;52(4):M201-8.

8. Ferrucci L, Harris TB, Guralnik JM, Tracy RP, Corti MC, Cohen HJ, et al. Serum IL-6 level and the development of disability in older persons. Journal of the American Geriatrics Society. 1999;47(6):639-46.

9. Harris TB, Ferrucci L, Tracy RP, Corti MC, Wacholder S, Ettinger WH, Jr., et al. Associations of elevated interleukin-6 and C-reactive protein levels with mortality in the elderly. Am J Med. 1999;106(5):506-12.

Bruunsgaard H, Ladelund S, Pedersen AN, Schroll M, Jorgensen T, Pedersen BK.
 Predicting death from tumour necrosis factor-alpha and interleukin-6 in 80-year-old people.
 Clin Exp Immunol. 2003;132(1):24-31.

 Giovannini S, Onder G, Liperoti R, Russo A, Carter C, Capoluongo E, et al.
 Interleukin-6, C-reactive protein, and tumor necrosis factor-alpha as predictors of mortality in frail, community-living elderly individuals. Journal of the American Geriatrics Society.
 2011;59(9):1679-85.

12. de Gonzalo-Calvo D, de Luxan-Delgado B, Martinez-Camblor P, Rodriguez-Gonzalez S, Garcia-Macia M, Suarez FM, et al. Chronic inflammation as predictor of 1-year hospitalization and mortality in elderly population. Eur J Clin Invest. 2012;42(10):1037-46.

13. Furman D, Chang J, Lartigue L, Bolen CR, Haddad F, Gaudilliere B, et al. Expression of specific inflammasome gene modules stratifies older individuals into two extreme clinical and immunological states. Nature medicine. 2017;23(2):174-84.

14. Lin T, Liu GA, Perez E, Rainer RD, Febo M, Cruz-Almeida Y, et al. Systemic Inflammation Mediates Age-Related Cognitive Deficits. Front Aging Neurosci. 2018;10:236.

15. Schram MT, Euser SM, de Craen AJ, Witteman JC, Frolich M, Hofman A, et al. Systemic markers of inflammation and cognitive decline in old age. Journal of the American Geriatrics Society. 2007;55(5):708-16.

16. Weaver JD, Huang MH, Albert M, Harris T, Rowe JW, Seeman TE. Interleukin-6 and risk of cognitive decline: MacArthur studies of successful aging. Neurology. 2002;59(3):371-8.

17. Freund A, Orjalo AV, Desprez PY, Campisi J. Inflammatory networks during cellular senescence: causes and consequences. Trends Mol Med. 2010;16(5):238-46.

18. Gilroy D, De Maeyer R. New insights into the resolution of inflammation. Semin Immunol. 2015;27(3):161-8.

19. Parmigiani A, Alcaide ML, Freguja R, Pallikkuth S, Frasca D, Fischl MA, et al. Impaired antibody response to influenza vaccine in HIV-infected and uninfected aging women is associated with immune activation and inflammation. PLoS One. 2013;8(11):e79816.

20. Muyanja E, Ssemaganda A, Ngauv P, Cubas R, Perrin H, Srinivasan D, et al. Immune activation alters cellular and humoral responses to yellow fever 17D vaccine. J Clin Invest. 2014;124(7):3147-58.

Thevaranjan N, Puchta A, Schulz C, Naidoo A, Szamosi JC, Verschoor CP, et al.
Age-Associated Microbial Dysbiosis Promotes Intestinal Permeability, Systemic
Inflammation, and Macrophage Dysfunction. Cell Host Microbe. 2017;21(4):455-66 e4.

22. Puchta A, Naidoo A, Verschoor CP, Loukov D, Thevaranjan N, Mandur TS, et al. TNF Drives Monocyte Dysfunction with Age and Results in Impaired Anti-pneumococcal Immunity. PLoS Pathog. 2016;12(1):e1005368.

23. Agius E, Lacy KE, Vukmanovic-Stejic M, Jagger AL, Papageorgiou AP, Hall S, et al. Decreased TNF-alpha synthesis by macrophages restricts cutaneous immunosurveillance by memory CD4+ T cells during aging. The Journal of experimental medicine. 2009;206(9):1929-40.

24. Vukmanovic-Stejic M, Sandhu D, Seidel JA, Patel N, Sobande TO, Agius E, et al. The Characterization of Varicella Zoster Virus-Specific T Cells in Skin and Blood during Aging. J Invest Dermatol. 2015;135(7):1752-62.

25. Jacinto TA, Meireles GS, Dias AT, Aires R, Porto ML, Gava AL, et al. Increased ROS production and DNA damage in monocytes are biomarkers of aging and atherosclerosis. Biol Res. 2018;51(1):33.

26. Vukmanovic-Stejic M, Chambers ES, Suarez-Farinas M, Sandhu D, Fuentes-Duculan J, Patel N, et al. Enhancement of cutaneous immunity during aging by blocking p38 mitogen-activated protein (MAP) kinase-induced inflammation. J Allergy Clin Immunol. 2018;142(3):844-56.

27. Khan N, Shariff N, Cobbold M, Bruton R, Ainsworth JA, Sinclair AJ, et al. Cytomegalovirus seropositivity drives the CD8 T cell repertoire toward greater clonality in healthy elderly individuals. Journal of immunology. 2002;169(4):1984-92.

28. Hadrup SR, Strindhall J, Kollgaard T, Seremet T, Johansson B, Pawelec G, et al. Longitudinal studies of clonally expanded CD8 T cells reveal a repertoire shrinkage predicting mortality and an increased number of dysfunctional cytomegalovirus-specific T cells in the very elderly. Journal of immunology. 2006;176(4):2645-53.

29. Riddell NE, Griffiths SJ, Rivino L, King DC, Teo GH, Henson SM, et al. Multifunctional cytomegalovirus (CMV)-specific CD8(+) T cells are not restricted by telomere-related senescence in young or old adults. Immunology. 2015;144(4):549-60.

30. Simanek AM, Dowd JB, Pawelec G, Melzer D, Dutta A, Aiello AE. Seropositivity to cytomegalovirus, inflammation, all-cause and cardiovascular disease-related mortality in the United States. PLoS One. 2011;6(2):e16103.

31. Bartlett DB, Firth CM, Phillips AC, Moss P, Baylis D, Syddall H, et al. The age-related increase in low-grade systemic inflammation (Inflammaging) is not driven by cytomegalovirus infection. Aging Cell. 2012;11(5):912-5.

32. Ellulu MS, Patimah I, Khaza'ai H, Rahmat A, Abed Y. Obesity and inflammation: the linking mechanism and the complications. Arch Med Sci. 2017;13(4):851-63.

33. Frasca D, Blomberg BB, Paganelli R. Aging, Obesity, and Inflammatory Age-Related Diseases. Front Immunol. 2017;8:1745.

34. Spielmann G, Johnston CA, O'Connor DP, Foreyt JP, Simpson RJ. Excess body mass is associated with T cell differentiation indicative of immune ageing in children. Clin Exp Immunol. 2014;176(2):246-54.

35. Hunter GR, Gower BA, Kane BL. Age Related Shift in Visceral Fat. Int J Body Compos Res. 2010;8(3):103-8.

36. Kim KA, Jeong JJ, Yoo SY, Kim DH. Gut microbiota lipopolysaccharide accelerates inflamm-aging in mice. BMC Microbiol. 2016;16:9.

37. Claesson MJ, Cusack S, O'Sullivan O, Greene-Diniz R, de Weerd H, Flannery E, et al. Composition, variability, and temporal stability of the intestinal microbiota of the elderly.Proc Natl Acad Sci U S A. 2011;108 Suppl 1:4586-91.

38. Rampelli S, Candela M, Turroni S, Biagi E, Collino S, Franceschi C, et al. Functional metagenomic profiling of intestinal microbiome in extreme ageing. Aging (Albany NY).
2013;5(12):902-12.

 Claesson MJ, Jeffery IB, Conde S, Power SE, O'Connor EM, Cusack S, et al. Gut microbiota composition correlates with diet and health in the elderly. Nature.
 2012;488(7410):178-84.

40. Clark RI, Salazar A, Yamada R, Fitz-Gibbon S, Morselli M, Alcaraz J, et al. Distinct Shifts in Microbiota Composition during Drosophila Aging Impair Intestinal Function and Drive Mortality. Cell Rep. 2015;12(10):1656-67.

41. Bianchi ME. DAMPs, PAMPs and alarmins: all we need to know about danger. J Leukoc Biol. 2007;81(1):1-5.

42. Feldman N, Rotter-Maskowitz A, Okun E. DAMPs as mediators of sterile inflammation in aging-related pathologies. Ageing Res Rev. 2015;24(Pt A):29-39.

43. Davalos AR, Kawahara M, Malhotra GK, Schaum N, Huang J, Ved U, et al. p53dependent release of Alarmin HMGB1 is a central mediator of senescent phenotypes. J Cell Biol. 2013;201(4):613-29.

44. Park JS, Gamboni-Robertson F, He Q, Svetkauskaite D, Kim JY, Strassheim D, et al. High mobility group box 1 protein interacts with multiple Toll-like receptors. Am J Physiol Cell Physiol. 2006;290(3):C917-24.

45. Youm YH, Grant RW, McCabe LR, Albarado DC, Nguyen KY, Ravussin A, et al. Canonical NIrp3 inflammasome links systemic low-grade inflammation to functional decline in aging. Cell Metab. 2013;18(4):519-32.

46. Roel P. H. De Maeyer RCvdM, Rikah Louie, Olivia Bracken, Oliver P. Devine, Daniel
R. Goldstein, Mohib Uddin, Arne N. Akbar & Derek W. Gilroy. Blocking elevated p38 MAPK
restores efferocytosis and inflammatory resolution in the elderly. Nature Immunology.
2020;in press.

47. Gorgoulis V, Adams PD, Alimonti A, Bennett DC, Bischof O, Bishop C, et al. Cellular Senescence: Defining a Path Forward. Cell. 2019;179(4):813-27.

48. Xue W, Zender L, Miething C, Dickins RA, Hernando E, Krizhanovsky V, et al. Senescence and tumour clearance is triggered by p53 restoration in murine liver carcinomas. Nature. 2007;445(7128):656-60.

49. Lujambio A, Akkari L, Simon J, Grace D, Tschaharganeh DF, Bolden JE, et al. Noncell-autonomous tumor suppression by p53. Cell. 2013;153(2):449-60.

50. Demaria M, Ohtani N, Youssef SA, Rodier F, Toussaint W, Mitchell JR, et al. An essential role for senescent cells in optimal wound healing through secretion of PDGF-AA. Dev Cell. 2014;31(6):722-33.

51. Herbig U, Ferreira M, Condel L, Carey D, Sedivy JM. Cellular senescence in aging primates. Science. 2006;311(5765):1257.

52. Biran A, Zada L, Abou Karam P, Vadai E, Roitman L, Ovadya Y, et al. Quantitative identification of senescent cells in aging and disease. Aging Cell. 2017;16(4):661-71.

53. Krishnamurthy J, Torrice C, Ramsey MR, Kovalev GI, Al-Regaiey K, Su L, et al. Ink4a/Arf expression is a biomarker of aging. J Clin Invest. 2004;114(9):1299-307.

54. Burd CE, Sorrentino JA, Clark KS, Darr DB, Krishnamurthy J, Deal AM, et al. Monitoring tumorigenesis and senescence in vivo with a p16(INK4a)-luciferase model. Cell. 2013;152(1-2):340-51.

55. Ressler S, Bartkova J, Niederegger H, Bartek J, Scharffetter-Kochanek K, Jansen-Durr P, et al. p16INK4A is a robust in vivo biomarker of cellular aging in human skin. Aging Cell. 2006;5(5):379-89.

56. Pereira BI, Devine OP, Vukmanovic-Stejic M, Chambers ES, Subramanian P, Patel N, et al. Senescent cells evade immune clearance via HLA-E-mediated NK and CD8(+) T cell inhibition. Nat Commun. 2019;10(1):2387.

57. Melk A, Schmidt BM, Takeuchi O, Sawitzki B, Rayner DC, Halloran PF. Expression of p16INK4a and other cell cycle regulator and senescence associated genes in aging human kidney. Kidney Int. 2004;65(2):510-20.

58. Victorelli S, Lagnado A, Halim J, Moore W, Talbot D, Barrett K, et al. Senescent human melanocytes drive skin ageing via paracrine telomere dysfunction. The EMBO journal. 2019:e101982.

59. Karin O, Agrawal A, Porat Z, Krizhanovsky V, Alon U. Senescent cell turnover slows with age providing an explanation for the Gompertz law. Nat Commun. 2019;10(1):5495.

60. Acosta JC, Banito A, Wuestefeld T, Georgilis A, Janich P, Morton JP, et al. A complex secretory program orchestrated by the inflammasome controls paracrine senescence. Nat Cell Biol. 2013;15(8):978-90.

61. Lasry A, Ben-Neriah Y. Senescence-associated inflammatory responses: aging and cancer perspectives. Trends Immunol. 2015;36(4):217-28.

62. Liu Y, Sanoff HK, Cho H, Burd CE, Torrice C, Ibrahim JG, et al. Expression of p16(INK4a) in peripheral blood T-cells is a biomarker of human aging. Aging Cell. 2009;8(4):439-48.

63. Di Mitri D, Azevedo RI, Henson SM, Libri V, Riddell NE, Macaulay R, et al. Reversible senescence in human CD4+CD45RA+CD27- memory T cells. Journal of immunology. 2011;187(5):2093-100.

64. Henson SM, Franzese O, Macaulay R, Libri V, Azevedo RI, Kiani-Alikhan S, et al. KLRG1 signaling induces defective Akt (ser473) phosphorylation and proliferative dysfunction of highly differentiated CD8+ T cells. Blood. 2009;113(26):6619-28.

65. Tilly G, Doan-Ngoc TM, Yap M, Caristan A, Jacquemont L, Danger R, et al. IL-15 Harnesses Pro-inflammatory Function of TEMRA CD8 in Kidney-Transplant Recipients. Front Immunol. 2017;8:778. 66. Muller-Durovic B, Lanna A, Covre LP, Mills RS, Henson SM, Akbar AN. Killer Cell Lectin-like Receptor G1 Inhibits NK Cell Function through Activation of Adenosine 5'-Monophosphate-Activated Protein Kinase. Journal of immunology. 2016;197(7):2891-9.

67. Cope AP, Liblau RS, Yang XD, Congia M, Laudanna C, Schreiber RD, et al. Chronic tumor necrosis factor alters T cell responses by attenuating T cell receptor signaling. The Journal of experimental medicine. 1997;185(9):1573-84.

68. Lim SO, Li CW, Xia W, Cha JH, Chan LC, Wu Y, et al. Deubiquitination and Stabilization of PD-L1 by CSN5. Cancer Cell. 2016;30(6):925-39.

69. Bennett CL, Christie J, Ramsdell F, Brunkow ME, Ferguson PJ, Whitesell L, et al. The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. Nat Genet. 2001;27(1):20-1.

70. Brunkow ME, Jeffery EW, Hjerrild KA, Paeper B, Clark LB, Yasayko SA, et al. Disruption of a new forkhead/winged-helix protein, scurfin, results in the fatal lymphoproliferative disorder of the scurfy mouse. Nat Genet. 2001;27(1):68-73.

71. Vukmanovic-Stejic M, Agius E, Booth N, Dunne PJ, Lacy KE, Reed JR, et al. The kinetics of CD4+Foxp3+ T cell accumulation during a human cutaneous antigen-specific memory response in vivo. J Clin Invest. 2008;118(11):3639-50.

72. Vukmanovic-Stejic M, Sandhu D, Sobande TO, Agius E, Lacy KE, Riddell N, et al. Varicella zoster-specific CD4+Foxp3+ T cells accumulate after cutaneous antigen challenge in humans. Journal of immunology. 2013;190(3):977-86.

73. Korn T, Muschaweckh A. Stability and Maintenance of Foxp3(+) Treg Cells in Nonlymphoid Microenvironments. Front Immunol. 2019;10:2634.

74. Chen X, Baumel M, Mannel DN, Howard OM, Oppenheim JJ. Interaction of TNF with TNF receptor type 2 promotes expansion and function of mouse CD4+CD25+ T regulatory cells. Journal of immunology. 2007;179(1):154-61.

75. Watanabe R, Shirai T, Namkoong H, Zhang H, Berry GJ, Wallis BB, et al. Pyruvate controls the checkpoint inhibitor PD-L1 and suppresses T cell immunity. J Clin Invest. 2017;127(7):2725-38.

76. Cohen HB, Briggs KT, Marino JP, Ravid K, Robson SC, Mosser DM. TLR stimulation initiates a CD39-based autoregulatory mechanism that limits macrophage inflammatory responses. Blood. 2013;122(11):1935-45.

77. Campisi J, d'Adda di Fagagna F. Cellular senescence: when bad things happen to good cells. Nat Rev Mol Cell Biol. 2007;8(9):729-40.

78. Chen W, Jin W, Hardegen N, Lei KJ, Li L, Marinos N, et al. Conversion of peripheral CD4+CD25- naive T cells to CD4+CD25+ regulatory T cells by TGF-beta induction of transcription factor Foxp3. The Journal of experimental medicine. 2003;198(12):1875-86.

79. Dagouassat M, Gagliolo JM, Chrusciel S, Bourin MC, Duprez C, Caramelle P, et al. The cyclooxygenase-2-prostaglandin E2 pathway maintains senescence of chronic obstructive pulmonary disease fibroblasts. American journal of respiratory and critical care medicine. 2013;187(7):703-14.

80. Kabir TD, Leigh RJ, Tasena H, Mellone M, Coletta RD, Parkinson EK, et al. A miR-335/COX-2/PTEN axis regulates the secretory phenotype of senescent cancer-associated fibroblasts. Aging (Albany NY). 2016;8(8):1608-35.

81. Baratelli F, Lin Y, Zhu L, Yang SC, Heuze-Vourc'h N, Zeng G, et al. Prostaglandin E2 induces FOXP3 gene expression and T regulatory cell function in human CD4+ T cells. Journal of immunology. 2005;175(3):1483-90.

82. Sharma S, Yang SC, Zhu L, Reckamp K, Gardner B, Baratelli F, et al. Tumor cyclooxygenase-2/prostaglandin E2-dependent promotion of FOXP3 expression and CD4+ CD25+ T regulatory cell activities in lung cancer. Cancer Res. 2005;65(12):5211-20.

83. Okano M, Sugata Y, Fujiwara T, Matsumoto R, Nishibori M, Shimizu K, et al. E prostanoid 2 (EP2)/EP4-mediated suppression of antigen-specific human T-cell responses by prostaglandin E2. Immunology. 2006;118(3):343-52.

84. Chen JH, Perry CJ, Tsui YC, Staron MM, Parish IA, Dominguez CX, et al. Prostaglandin E2 and programmed cell death 1 signaling coordinately impair CTL function and survival during chronic viral infection. Nature medicine. 2015;21(4):327-34.

85. Baar MP, Brandt RMC, Putavet DA, Klein JDD, Derks KWJ, Bourgeois BRM, et al. Targeted Apoptosis of Senescent Cells Restores Tissue Homeostasis in Response to Chemotoxicity and Aging. Cell. 2017;169(1):132-47 e16.

86. Baker DJ, Childs BG, Durik M, Wijers ME, Sieben CJ, Zhong J, et al. Naturally occurring p16(Ink4a)-positive cells shorten healthy lifespan. Nature. 2016;530(7589):184-9.
87. Baker DJ, Wijshake T, Tchkonia T, LeBrasseur NK, Childs BG, van de Sluis B, et al. Clearance of p16Ink4a-positive senescent cells delays ageing-associated disorders. Nature.

2011;479(7372):232-6.

Zhu Y, Tchkonia T, Pirtskhalava T, Gower AC, Ding H, Giorgadze N, et al. The
Achilles' heel of senescent cells: from transcriptome to senolytic drugs. Aging Cell.
2015;14(4):644-58.

89. Zhu Y, Doornebal EJ, Pirtskhalava T, Giorgadze N, Wentworth M, Fuhrmann-Stroissnigg H, et al. New agents that target senescent cells: the flavone, fisetin, and the BCL-XL inhibitors, A1331852 and A1155463. Aging (Albany NY). 2017;9(3):955-63.

90. Roos CM, Zhang B, Palmer AK, Ogrodnik MB, Pirtskhalava T, Thalji NM, et al. Chronic senolytic treatment alleviates established vasomotor dysfunction in aged or atherosclerotic mice. Aging Cell. 2016;15(5):973-7. 91. Chang J, Wang Y, Shao L, Laberge RM, Demaria M, Campisi J, et al. Clearance of senescent cells by ABT263 rejuvenates aged hematopoietic stem cells in mice. Nature medicine. 2016;22(1):78-83.

92. Fuhrmann-Stroissnigg H, Ling YY, Zhao J, McGowan SJ, Zhu Y, Brooks RW, et al.
Identification of HSP90 inhibitors as a novel class of senolytics. Nat Commun.
2017;8(1):422.

93. Andre P, Denis C, Soulas C, Bourbon-Caillet C, Lopez J, Arnoux T, et al. Anti-NKG2A mAb Is a Checkpoint Inhibitor that Promotes Anti-tumor Immunity by Unleashing Both T and NK Cells. Cell. 2018;175(7):1731-43 e13.

94. van Montfoort N, Borst L, Korrer MJ, Sluijter M, Marijt KA, Santegoets SJ, et al.
NKG2A Blockade Potentiates CD8 T Cell Immunity Induced by Cancer Vaccines. Cell.
2018;175(7):1744-55 e15.

95. Weichhart T, Hengstschlager M, Linke M. Regulation of innate immune cell function by mTOR. Nature reviews Immunology. 2015;15(10):599-614.

96. Laberge RM, Sun Y, Orjalo AV, Patil CK, Freund A, Zhou L, et al. MTOR regulates the pro-tumorigenic senescence-associated secretory phenotype by promoting IL1A translation. Nat Cell Biol. 2015;17(8):1049-61.

97. Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, et al. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. Nature.
2009;460(7253):392-5.

98. Mannick JB, Del Giudice G, Lattanzi M, Valiante NM, Praestgaard J, Huang B, et al. mTOR inhibition improves immune function in the elderly. Sci Transl Med. 2014;6(268):268ra179.

99. Mannick JB, Morris M, Hockey HP, Roma G, Beibel M, Kulmatycki K, et al. TORC1 inhibition enhances immune function and reduces infections in the elderly. Sci Transl Med.

2018;10(449).

100. Alimbetov D, Davis T, Brook AJ, Cox LS, Faragher RG, Nurgozhin T, et al. Suppression of the senescence-associated secretory phenotype (SASP) in human fibroblasts using small molecule inhibitors of p38 MAP kinase and MK2. Biogerontology. 2016;17(2):305-15.

101. Callender LA, Carroll EC, Beal RWJ, Chambers ES, Nourshargh S, Akbar AN, et al. Human CD8(+) EMRA T cells display a senescence-associated secretory phenotype regulated by p38 MAPK. Aging Cell. 2018;17(1).

102. Freund A, Patil CK, Campisi J. p38MAPK is a novel DNA damage responseindependent regulator of the senescence-associated secretory phenotype. The EMBO journal. 2011;30(8):1536-48. 103. Akbar AN, Reed JR, Lacy KE, Jackson SE, Vukmanovic-Stejic M, Rustin MH. Investigation of the cutaneous response to recall antigen in humans in vivo. Clin Exp Immunol. 2013;173(2):163-72.

104. Sweeney SE. The as-yet unfulfilled promise of p38 MAPK inhibitors. Nat Rev Rheumatol. 2009;5(9):475-7.

105. Valencia WM, Palacio A, Tamariz L, Florez H. Metformin and ageing: improving ageing outcomes beyond glycaemic control. Diabetologia. 2017;60(9):1630-8.



Figure 1: Potential mechanisms of inflammageing

Schematic representation of the proposed causes of inflammageing observed in the old. Increased visceral fat with associated increased leukocyte infiltration; increased DAMP production which bind to TLRs; increased senescent cell production with production of SASP which also includes DAMPs; increased gut permeability and LPS leakage and subsequent TLR activation; and finally chronic viral infection which lead to chronic immune activation. The SASP secreted from the senescent cells also has the capability to drive senescence in nearby cells, increasing the number of cells overall. All of these outcomes result in the production of inflammatory cytokines and the subsequent on set of inflammageing.



Figure 2: Schematic of the therapeutic targets of inflammageing

Schematic representation of the current therapeutic targets of inflammageing. In humans the two main targets to date have been the use of mTOR blockade to enhance vaccine responses and p38 MAP Kinase blockade to enhance antigen-specific cutaneous immunity. In mouse models senolytics have been shown to be a very promising target and are as yet untried in humans. The most untested approach is the use of inhibitory ligand blockade to enhance senescent cell clearance via the hosts own immune system.