- ¹ Enhancing Colorectal Anastomotic Safety
- ² with Indocyanine Green Fluorescence
- ³ Angiography: An update
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25 Abstract

26 Reducing anastomotic leak (AL) continues to be a main focus in colorectal research. Several 27 new technologies have been developed with an aim to reduce this from mechanical devices 28 to advanced imaging techniques. Fluorescence angiography (FA) with indocyanine green 29 (ICG) in colorectal surgery is now a well-established technique and may have a role in 30 reducing AL. By using FA, we are able to have a visual representation of perfusion which aids 31 intraoperative decision making. The main impact is change in the level of bowel transection 32 at the proximal side of an anastomosis and provide a more objective and confident 33 assessment of bowel perfusion. Previous studies have shown that routine FA use is safe and 34 reproducible. Recent results from randomized control trials and meta-analyses show that FA 35 use reduces the rate of anastomotic leak. The main limitation of FA is its lack of ability to 36 quantify perfusion. Novel technologies are being developed that will quantify tissue 37 perfusion and oxygenation. Overall, FA is a safe and feasible technique which may have a 38 role in reducing AL. 39

40 Key Words:

41 Fluorescence angiography, fluorescence imaging, indocyanine green, near infrared,

42 anastomotic leak, colorectal surgery

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57 1. Introduction

58 Despite advances in technology and greater precision in surgical technique, 59 anastomotic leak (AL) continues to be the main concern for patients undergoing colorectal 60 resectional surgery. Reported rates remain between 3-15% depending on the location of the 61 anastomosis with higher rates for left sided or colo-rectal anastomoses [1]. Despite some 62 variability in the exact definition of what constitutes an AL, the generally recognized grading 63 system is that put forward by the International Study Group of Rectal Cancer[2]. It is known 64 that AL causes an increase in patient mortality, morbidity, hospital length of stay, rates of 65 re-operation, permanent stoma and financial burden[3]. Studies have shown that patient 66 specific pre-operative risk factors such as obesity, smoking and chemotherapy increase the 67 risk of AL[4,5]. A Delphi consensus by the Association of Coloproctology of Great Britain and 68 Ireland (ACPGBI) classified risk factors into non-modifiable and modifiable[6]. Separately, 69 identification of intra-operative factors that may pre-dispose to AL are a main focus of 70 research. Intraoperative risk factors can be divided between patient and technical factors. 71 Tumor size, distal location, blood loss, transfusion and duration of surgery > 4 hours have 72 been shown to increase the rate of AL[7].

73 Perfusion of the anastomosis has also been shown to have an effect on healing [8,9]. 74 This is affected by a patient's pre-operative vasculature, the level of resection and surgical 75 technique. One intraoperative factor which surgeons have control over is the level of colonic 76 division and consequently the perfusion to the proximal side of an anastomosis. Several 77 methods have been described to assess blood flow to the anastomosis. The simplest of 78 these is a visual assessment looking for serosal discoloration, pulsatile bleeding at the cut 79 edge of the bowel or flow from the marginal artery[10]. However, this can be inaccurate and 80 provides no indication as to the microperfusion of the colon at the site of anastomosis.

Intra-operative fluorescence angiography (FA) has been shown to assess microperfusion of the colon though this has not been quantified[11]. This process requires the intravenous administration of the fluorophore indocyanine green (ICG) which binds to plasma lipoproteins, therefore remaining within the intravascular space until excretion in bile or urine. When ICG is excited by near infra-red light (NIR) it fluoresces. This fluorescence can be captured with an NIR camera indicating on a conventional screen the location of ICG and thus providing an estimate of tissue perfusion.

- 88 Numerous observational trials have demonstrated safety, feasibility and efficacy in 89 assessing perfusion using ICG with promising results. This purpose of this review is to 90 provide an update in the progress being made in this field.
- 91 2. Search strategy and selection criteria
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An electronic search of PubMed, Embase and the Cochrane library was performed between 93 2005 and 2020 to identify the relevant literature for this review. Medical subject headings 94 95 (MeSH) and text words were searched. The following search terms were used: "anastomotic leak" AND "colorectal"" AND "fluorescence angiography", "fluorescence imaging" or "ICG". 96 97 Peer reviewed papers in the English language available in full were included. Reference lists 98 were reviewed to include any further relevant literature. A systematic review of papers 99 between 2015 and 2020 was performed to identify new clinical research. Comparative 100 studies with an endpoint of anastomotic leak were included. Unmatched observational 101 studies were excluded. These papers formed the basis for this review. Ongoing clinical trials 102 were identified from the searched literature, ClinicalTrials.gov and ISRCTN.

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3. Fluorescence Angiography in Colorectal Surgery

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106 3.1Early Use of Fluorescence Angiography

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Fluorescence angiography (FA) has been used to assess bowel perfusion in colorectal
surgery for more than 15 years. It provides a more objective assessment of perfusion
compared to more traditional, subjective methods described above. Perfusion remains the
most important factor in the healing of bowel anastomoses.

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113 Kudszus et al, began their series in 2003 demonstrating significantly reduced rates of

anastomotic revision in the FA group compared with a retrospective matched control, 3.5%

115 vs 7.5% respectively[12]. This showed a significant difference in the two groups and

116 provided an important first step towards better understanding the role of FA in reducing AL.

117 With the increased availability of CT, we can now use radiologically confirmed anastomotic

118 leak (AL) as an endpoint rather than clinical endpoints such as reoperation.

119 The seminal paper by Jafari et al, the PILLAR II trial, is probably most recognized as 120 the study which proved the feasibility and safety of FA in left-sided colonic and rectal 121 resection[13]. This multi-centered, prospective trial recruited 139 patients across 11 centers 122 in the USA. Importantly, this showed that FA was reproducible across sites as usable images 123 were acquired in 98.6% cases. The use of FA changed the resection level in 6.5% cases, and 124 there were subsequently no leaks in this group. The overall AL rate was low at 1.4% which 125 much reduced compared to the existing literature. In 2018, Ris et al published the results of 126 their multicenter phase II trial from 2013-2016[14]. Much larger than the trials before it, this 127 prospective study recruited 504 patients across 3 tertiary centers. Again, this showed good 128 usability of the technology as NIR images were obtained in all cases. The FA group had an AL 129 rate of 2.4% against 5.8% in an historical unmatched control group. FA led to a change in 130 surgical plan in 5.8% cases, none of which had an AL. Although their series included 131 operations where the anticipated proximal anastomotic perfusion would be a high, such as 132 reversal of Hartmann's or ileo-rectal anastomosis, subgroup analysis for low anterior 133 resection (LAR) showed an AL rate of 3%. They related this to an historical group of LARs 134 which had an AL rate of 10.7%. Although caution must be taken when using historical 135 groups these studies showed that FA was feasible, reproducible and changed intraoperative 136 decision making. It also suggested that its use may reduce the rate of AL.

A systematic review of 5 early studies by Blano-Colino and Epsin-Basany involved
1302 patients[15]. While based on non-randomized retrospective studies it showed a
significant reduction in AL rate when FA was used in patients undergoing surgery for
colorectal cancer (OR 0.35; CI 0.16-0.74; p-0.006). In particular there was significant
reduction in the AL rate in a less heterogenous sub-group, patients undergoing rectal cancer
resection, 1.1% FA vs 6.1% non-FA (p=0.02).

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145 3.2 Recent Trials using Fluorescence Angiography

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147 Since this period there have been 8 published comparative studies, two of which are

randomized control trials (RCTs). There is a wide variation in these studies as some include

any colonic resection and others solely low anterior resection with TME (3/8). The trial

150 protocols did differ in their administration of ICG with doses varying widely.

151 2 studies specifically looked at the use of FA in patients undergoing laparoscopic LAR. In 2017 Boni et al showed a reduction in AL for LAR with TME using FA in 42 patients against 152 153 a retrospective matched cohort (0% vs 5%)[16]. These results were reproduced by Mizrahi 154 et al in 2018[17]. In this study 30 patients undergoing LAR were evaluated against a 155 comparable historical group. 4 patients (13.3) had their surgical plan changed after FA 156 assessment. Their study had no leaks in the FA group and 2 (6.7%) in the comparative group. 157 These studies demonstrate that FA use may be of benefit in a patient group more at risk of AL. The authors from both studies concluded that the use of FA was safe though a 158 159 randomized study was is needed.

160 Losurdo et al used a propensity score-matching (PSM) system in their series to try 161 and mitigate the inherent bias from the heterogeneity within their cohort of patients 162 undergoing laparoscopic left sided colonic or colorectal resection, including patients with 163 handsewn coloanal anastomosis[18]. Cases converted to open were excluded. Before 164 matching statistically fewer patients in the FA group underwent reoperation for AL. A 1:1 165 PSM system grouped 75 patients from each cohort. This score accounted for tumor stage, 166 co-morbidities and baseline demographics. After matching there was a significant reduction 167 in AL within the FA group, 9.3 vs 16.3% (p=0.058). A multicenter study by Watanabe et al 168 used PSM in patients undergoing LAR[19]. 211 patients were matched in each group, FA and 169 non-FA. Their study found a significant reduction in Clavien-Dindo (CD) Grade II and III 170 anastomotic leakage.

171 At the time of this review there have been 2 RCTs looking at FA and AL. De Nardi et al published the first RCT in patients undergoing left sided or rectal resection[20]. In this 172 173 multi-center trial 252 patients were randomized and after exclusions there were 118 174 patients in the study group. 11% patients in the study group had a change of surgical plan 175 due to FA. The study did not show a significant difference in AL between groups. However, 176 the leak rate was lower in the study group and the authors concluded that FA was a safe 177 adjunct that was not time consuming or detrimental. Alekseev et al published the results of the FLAG trial, a second RCT focused on patients undergoing anterior rection with stapled 178 179 end-to-end colorectal anastomosis[21]. They included both open and laparoscopic 180 approaches, 380 patients were randomized. This trial demonstrated a significant reduction 181 in the AL rate when using FA (9.1% vs 16.3% p=0.04). It is worth noting that there was a 182 comparatively high AL rate in patients undergoing LAR without FA, 25.7% (FA group 14.4%

p=0.04). Additionally, there was a slightly higher, but non-specific, reoperation rate in the FA
group (3.7% vs 2.1% p=0.38). This study demonstrates that FA has a role but that it is mainly
limited to low colorectal anastomoses.

186 In 2020 Chan et al published a systematic review of 20 studies including the above 187 RCTs[22]. 5498 patients were included in the meta-analysis. This showed that FA decreased AL with an odds ratio (OR) of 0.46 (95% CI 0.34-0.62; p<0.0001). Although largely based on 188 189 retrospective studies a subgroup analysis of 4 prospective trials confirmed this result (OR 190 0.49 95% CI 0.3-0.81; p=0.005). Furthermore, this study confirmed that patients undergoing 191 LAR for rectal cancer with colorectal anastomosis may benefit from ICG. Arezzo et al 192 published their meta-analysis containing individual participant data from 9 trials involving 193 1,330 patients[23]. Their results showed a significant reduction in the rate of AL in the FA 194 group compared with standard care 4.2% vs 11.3% respectively (p=<0.001). Additionally, risk 195 of AL was found to be significantly lower with anastomoses <6cm from the anal verge and in 196 patients with BMI >25.

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198 3.3 Ongoing trials

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There is only 1 current randomized control trial investigating FA and AL. The IntAct trial is a
multi-center European RCT currently recruiting[24]. They aim to randomize 880 patients.
This will be the largest trial of its kind and is focused on patients undergoing laparoscopic or
robotic surgery for rectal cancer. An additional sub-study intervention will look at CT
perfusion scanning aiming to investigate the link between pre-operative vascular anatomy
and AL.

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207 4. Challenges and Skepticism

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209 Whilst current research is yielding promising results there are still some challenges 210 to be overcome. Although studies produce can reproduce fluorescence, there is a broad 211 range in the dose of ICG administered and the timing to assessment of the bowel. A recent 212 Delphi Consensus Conference of international experts across surgical specialties, including 213 colorectal, agreed that both dose administered and timing to assessment was important (89.5% and 89.5% consensus)[25]. A recent review of protocols recommends a dose of
2.5mg as multiple studies have had good results at lower dosages[26]. This correlates with
work undertaken in esophagogastric anastomoses[26]. Although there is a very low risk of
anaphylaxis to ICG, current studies in colorectal surgery use concentrations well below that
which is known to cause toxicity[27]. The European Association for Endoscopic Surgery
(EAES) technology committee are preparing a consensus conference for fluorescence and
we await the results of this later in 2021.

221 A further challenge is that whilst FA with ICG can provide a visual estimation of 222 microperfusion, there is no standard method of quantifying this. This is perhaps the biggest 223 hurdle at the present time. The rationale behind using FA is to be able to provide a 224 reproducible and objective method of perfusion assessment. On the surface it may seem 225 like ICG fulfils these criteria but in practice, the operating team still have to subjectively 226 decide whether the fluorescent signal is strong enough to justify creation of the 227 anastomosis or that the transection point should be revised more proximally. Recent work 228 from Soares et al, have shown variability in users relating to specialty and experience [28]. 229 Further, it is not known how the intensity of fluorescence correlates with microperfusion at 230 tissue level. Several studies have modelled colonic perfusion patterns by measuring 231 fluorescence intensity and time of onset[29,30]. This has been achieved in real time for 232 intraoperative use[29]. A retrospective video analysis study showed that slow perfusion was 233 an independent risk factor for AL[30]. However, parameter based models vary and are 234 difficult to reproduce. Park et al generated an artificial intelligence (AI) model which was 235 more accurate in retrospectively predicting the risk of AL compared with parametric 236 models[31]. Further work is required to ascertain specific, generalizable cut off levels for 237 intensity and time of onset that may influence intraoperative decision making.

238 Though FA can give an estimation of perfusion it cannot quantify oxygen delivery to 239 the tissues. Hyperspectral imaging (HIS) uses a sensor to capture electromagnetic waves at a 240 spectrum beyond visible light, and in greater detail. Reconstructed false color images 241 provide a visual representation of tissue oxygen saturation. This technology is non-invasive 242 and can accurately identify the margin of perfusion[32]. This has been shown to be 243 comparable to FA[33]. Moreover, Clancy et al have demonstrated in patients that there is a 244 strong correlation between high fluorescent intensity and oxygen saturation. Although, 245 these methods require calibration and are not widely available they likely to be the main

focus of tissue perfusion assessment going forward providing simultaneous optical andbiological imaging patterns.

248 While the discussed techniques can give an estimate of perfusion at the time of 249 anastomosis there is currently no reliable measure in the post-operative period. Recognition 250 of patients in whom the anastomosis is failing due to ischemia may allow early intervention. 251 Cahill et al have used an AI model to accurately identify tumors from their perfusion 252 patterns using FA[34]. Development of this technology can lead to real-time assessment of 253 bowel perfusion at the anastomosis. By knowing how our post-operative treatment regimen 254 affects anastomotic perfusion we may be able to specifically tailor patient management.

255 Lastly, if we can reduce the rate or accurately predict AL then we can allow FA to 256 have a greater impact on in other areas of our intraoperative decision making. Spinelli et al 257 have used FA to guide vascular ligation when forming an ileal pouch[35]. By using FA they 258 were able to confidently ligate the ileocolic vessels more proximally where required, giving 259 more length for the pouch. There were no anastomotic leaks. It may be that we can make 260 further decisions such as whether or not to create a defunctioning stoma. FA influenced this 261 decision in a pilot by Ris et al[36]. Stomas are known to add to patient financial burden and 262 reduce quality of life[37]. Conversely, if we can measure perfusion at the anastomosis post-263 operatively then we may be able to identify the patient group that benefits most from early 264 stoma reversal which has been shown to reduce costs and increasing quality of life[38]. 265

266 5. Conclusion

267 Fluorescence angiography in colorectal surgery is a safe and reproducible technique. There 268 is increasingly strong evidence that the use of FA reduces the AL rate. In particular, this may 269 be of greatest benefit in patients undergoing LAR where the AL rate is known to be the highest. Although further randomized studies are needed, we conclude that, where 270 271 available, routine use of FA is not to the detriment of the patient and often influences 272 surgical decision making. This may reduce the overall rate of AL and moderate the need for 273 defunctioning stoma. A comprehensive protocol is required to establish a standard 274 technique across all centers using FA. Ultimately, a way to quantify microperfusion is 275 needed and this should be a focus of research.

276 Conflict of Interest

277 Pampiglione, T and Chand, M declare no conflict of interest

278 Author Statement

- 279 Pampiglione, T: Conceptualization, Methodology, Writing original draft preparation.
- 280 Chand, M: Conceptualization, Methodology, Writing reviewing and editing

281 References

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- M. Frasson, B. Flor-Lorente, J.L. Ramos Rodríguez, P. Granero-Castro, D. Hervás, M.A.
 Alvarez Rico, M.J.G. Brao, J.M. Sánchez González, E. Garcia-Granero, Risk Factors for
 Anastomotic Leak After Colon Resection for Cancer, Annals of Surgery. 262 (2015)
 321–330. https://doi.org/10.1097/SLA.00000000000973.
- [2] N.N. Rahbari, J. Weitz, W. Hohenberger, R.J. Heald, B. Moran, A. Ulrich, T. Holm,
 W.D. Wong, E. Tiret, Y. Moriya, S. Laurberg, M. den Dulk, C. van de Velde, M.W.
 Büchler, Definition and grading of anastomotic leakage following anterior resection of
 the rectum: A proposal by the International Study Group of Rectal Cancer, Surgery.
 147 (2010) 339–351. https://doi.org/10.1016/j.surg.2009.10.012.
- S.Q. Ashraf, E.M. Burns, A. Jani, S. Altman, J.D. Young, C. Cunningham, O. Faiz, N.J.
 Mortensen, The economic impact of anastomotic leakage after anterior resections in
 English NHS hospitals: are we adequately remunerating them?, Colorectal Disease. 15
 (2013) e190–e198. https://doi.org/10.1111/codi.12125.
- V.C. Nikolian, N.S. Kamdar, S.E. Regenbogen, A.M. Morris, J.C. Byrn, P.A. Suwanabol,
 D.A. Campbell, S. Hendren, Anastomotic leak after colorectal resection: A populationbased study of risk factors and hospital variation, in: Surgery (United States), Mosby
 Inc., 2017: pp. 1619–1627. https://doi.org/10.1016/j.surg.2016.12.033.
- A. Fawcett, M. Shembekar, J.S. Church, R. Vashisht, R.G. Springall, D.M. Nott,
 Smoking, hypertension, and colonic anastomotic healing; a combined clinical and
 histopathological study, Gut. 38 (1996) 714–718.
- 303 https://doi.org/10.1136/gut.38.5.714.
- Anastomotic Leakage Working Group ASGBI, F. McDermott, S. Arora, J. Smith, R.J.C.
 Steele, G. Carlson, D.C. Winter, Prevention, Diagnosis and Management of Colorectal
 Anastomotic Leakage, ASGBI, 2016. www.cla.co.uk]. (accessed January 4, 2021).
- F.D. McDermott, A. Heeney, M.E. Kelly, R.J. Steele, G.L. Carlson, D.C. Winter,
 Systematic review of preoperative, intraoperative and postoperative risk factors for
 colorectal anastomotic leaks, British Journal of Surgery. 102 (2015) 462–479.
 https://doi.org/10.1002/bjs.9697.
- 311 [8] C.K. Enestvedt, S.K. Thompson, E.Y. Chang, B.A. Jobe, Clinical review: Healing in
 312 gastrointestinal anastomoses, part II, Microsurgery. 26 (2006) 137–143.
 313 https://doi.org/10.1002/micr.20198.
- 314 [9]M. Rutegård, J. Rutegård, Anastomotic leakage in rectal cancer surgery: The role of315blood perfusion, World J Gastrointest Surg. 7 (2015) 289–292.
- 316 https://doi.org/10.4240/wjgs.v7.i11.289.

- P.G. Horgan, T.F. Gorey, Operative assessment of intestinal viability, Surgical Clinics
 of North America. 72 (1992) 143–155. https://doi.org/10.1016/S00396109(16)45632-X.
- D.S. Keller, T. Ishizawa, R. Cohen, M. Chand, Indocyanine green fluorescence imaging
 in colorectal surgery: overview, applications, and future directions, The Lancet
 Gastroenterology and Hepatology. 2 (2017) 757–766. https://doi.org/10.1016/S2468 1253(17)30216-9.
- S. Kudszus, C. Roesel, A. Schachtrupp, J.J. Höer, Intraoperative laser fluorescence
 angiography in colorectal surgery: A noninvasive analysis to reduce the rate of
 anastomotic leakage, Langenbeck's Archives of Surgery. 395 (2010) 1025–1030.
 https://doi.org/10.1007/s00423-010-0699-x.
- M.D. Jafari, S.D. Wexner, J.E. Martz, E.C. McLemore, D.A. Margolin, D.A. Sherwinter,
 S.W. Lee, A.J. Senagore, M.J. Phelan, M.J. Stamos, Perfusion assessment in
 laparoscopic left-sided/anterior resection (PILLAR II): A multi-institutional study,
 Journal of the American College of Surgeons. 220 (2015) 82-92.e1.
 https://doi.org/10.1016/j.jamcollsurg.2014.09.015.
- F. Ris, E. Liot, N.C. Buchs, R. Kraus, G. Ismael, V. Belfontali, J. Douissard, C.
 Cunningham, I. Lindsey, R. Guy, O. Jones, B. George, P. Morel, N.J. Mortensen, R.
 Hompes, R.A. Cahill, Multicentre phase II trial of near-infrared imaging in elective
 colorectal surgery, British Journal of Surgery. 105 (2018) 1359–1367.
 https://doi.org/10.1002/bjs.10844.
- R. Blanco-Colino, E. Espin-Basany, Intraoperative use of ICG fluorescence imaging to
 reduce the risk of anastomotic leakage in colorectal surgery: a systematic review and
 meta-analysis, Techniques in Coloproctology. 22 (2018) 15–23.
 https://doi.org/10.1007/s10151-017-1731-8.
- L. Boni, A. Fingerhut, A. Marzorati, S. Rausei, G. Dionigi, E. Cassinotti, Indocyanine
 green fluorescence angiography during laparoscopic low anterior resection: results of
 a case-matched study, Surgical Endoscopy. 31 (2017) 1836–1840.
 https://doi.org/10.1007/s00464-016-5181-6.
- I. Mizrahi, M. Abu-Gazala, A.S. Rickles, L.M. Fernandez, A. Petrucci, J. Wolf, D.R.
 Sands, S.D. Wexner, Indocyanine green fluorescence angiography during low anterior
 resection for low rectal cancer: results of a comparative cohort study, Techniques in
 Coloproctology. 22 (2018) 535–540. https://doi.org/10.1007/s10151-018-1832-z.
- P. Losurdo, T.C. Mis, D. Cosola, L. Bonadio, F. Giudici, B. Casagranda, M. Bortul, N. de
 Manzini, Anastomosis Leak: Is There Still a Place for Indocyanine Green Fluorescence
 Imaging in Colon-Rectal Surgery? A Retrospective, Propensity Score-Matched Cohort
 Study, Surgical Innovation. 0 (2020) 155335062097525.
- 354 https://doi.org/10.1177/1553350620975258.
- J. Watanabe, A. Ishibe, Y. Suwa, H. Suwa, M. Ota, C. Kunisaki, I. Endo, Indocyanine
 green fluorescence imaging to reduce the risk of anastomotic leakage in laparoscopic
 low anterior resection for rectal cancer: a propensity score-matched cohort study,
 Surgical Endoscopy. 34 (2020) 202–208. https://doi.org/10.1007/s00464-019-067519.
- P. de Nardi, U. Elmore, G. Maggi, R. Maggiore, L. Boni, E. Cassinotti, U. Fumagalli, M.
 Gardani, S. de Pascale, P. Parise, A. Vignali, R. Rosati, Intraoperative angiography with
 indocyanine green to assess anastomosis perfusion in patients undergoing

- laparoscopic colorectal resection: results of a multicenter randomized controlled trial,
 Surgical Endoscopy. 34 (2020) 53–60. https://doi.org/10.1007/s00464-019-06730-0.
- M. Alekseev, E. Rybakov, Y. Shelygin, S. Chernyshov, I. Zarodnyuk, A study
 investigating the perfusion of colorectal anastomoses using fluorescence
 angiography: results of the FLAG randomized trial, Colorectal Disease. 22 (2020)
 1147–1153. https://doi.org/10.1111/codi.15037.
- 369 [22] D.K.H. Chan, S.K.F. Lee, J.J. Ang, Indocyanine green fluorescence angiography
 370 decreases the risk of colorectal anastomotic leakage: Systematic review and meta371 analysis, Surgery (United States). 168 (2020) 1128–1137.

372 https://doi.org/10.1016/j.surg.2020.08.024.

- A. Arezzo, M.A. Bonino, F. Ris, L. Boni, E. Cassinotti, D.C.C. Foo, N.F. Shum, A.
 Brolese, F. Ciarleglio, D.S. Keller, R. Rosati, P. de Nardi, U. Elmore, U. Fumagalli
 Romario, M.D. Jafari, A. Pigazzi, E. Rybakov, M. Alekseev, J. Watanabe, N. Vettoretto,
 R. Cirocchi, R. Passera, E. Forcignanò, M. Morino, Intraoperative use of fluorescence
 with indocyanine green reduces anastomotic leak rates in rectal cancer surgery: an
 individual participant data analysis, Surgical Endoscopy. 34 (2020) 4281–4290.
 https://doi.org/10.1007/s00464-020-07735-w.
- G. Armstrong, J. Croft, N. Corrigan, J.M. Brown, V. Goh, P. Quirke, C. Hulme, D.
 Tolan, A. Kirby, R. Cahill, P.R. O'Connell, D. Miskovic, M. Coleman, D. Jayne, IntAct:
 intra-operative fluorescence angiography to prevent anastomotic leak in rectal cancer
 surgery: a randomized controlled trial, Colorectal Disease : The Official Journal of the
 Association of Coloproctology of Great Britain and Ireland. 20 (2018) O226–O234.
 https://doi.org/10.1111/codi.14257.
- 386 [25] F. Dip, Ã. Luigi Boni, M. Bouvet, T. Carus, ô Michele Diana, jj Jorge Falco, Ã.C. 387 Geoffrey Gurtner, yy Takeaki Ishizawa, zz Norihiro Kokudo, zz Emanuele Lo Menzo, 388 P.S. Low, J. Masia, ôô Derek Muehrcke, jjjj A. Francis Papay, Ã. Carlo Pulitano, yyy 389 Sylke Schneider-Koraith, zzz Danny Sherwinter, G. Spinoglio, ôôô Laurents Stassen, 390 jjjjjj Yasuteru Urano, ôôôô Alexander Vahrmeijer, Ã. Eric Vibert, yyyy Jason Warram, 391 zzzz D. Steven Wexner, K. White, R.J. Rosenthal, Consensus Conference Statement on 392 the General Use of Near-Infrared Fluorescence Imaging and Indocyanine Green 393 Guided Surgery Results of a Modified Delphi Study, Annals of Surgery. Ahead of Print 394 (2020). https://doi.org/10.1097/SLA.000000000004412.
- L. van Manen, H.J.M. Handgraaf, M. Diana, J. Dijkstra, T. Ishizawa, A.L. Vahrmeijer,
 J.S.D. Mieog, A practical guide for the use of indocyanine green and methylene blue in
 fluorescence-guided abdominal surgery, Journal of Surgical Oncology. 118 (2018)
 283–300. https://doi.org/10.1002/jso.25105.
- R. Alford, H.M. Simpson, J. Duberman, G.C. Hill, M. Ogawa, C. Regino, H. Kobayashi,
 P.L. Choyke, Toxicity of Organic Fluorophores Used in Molecular Imaging: Literature
 Review, Molecular Imaging. 8 (2009) 7290.2009.00031.
- 402 https://doi.org/10.2310/7290.2009.00031.

403 [28] Soares et al, Unpublished Work, 2021.

- S. Hayami, K. Matsuda, H. Iwamoto, M. Ueno, M. Kawai, S. Hirono, K. Okada, M.
 Miyazawa, K. Tamura, Y. Mitani, Y. Kitahata, Y. Mizumoto, H. Yamaue, Visualization
 and quantification of anastomotic perfusion in colorectal surgery using near-infrared
 fluorescence, Techniques in Coloproctology. 23 (2019) 973–980.
- 408 https://doi.org/10.1007/s10151-019-02089-5.

- 409 [30] G.M. Son, M.S. Kwon, Y. Kim, J. Kim, S.H. Kim, J.W. Lee, Quantitative analysis of
 410 colon perfusion pattern using indocyanine green (ICG) angiography in laparoscopic
 411 colorectal surgery, Surgical Endoscopy. 33 (2019) 1640–1649.
 412 https://doi.org/10.1007/s00464-018-6439-y.
- [31] S.-H. Park, H.-M. Park, K.-R. Baek, H.-M. Ahn, I.Y. Lee, G.M. Son, Artificial intelligence
 based real-time microcirculation analysis system for laparoscopic colorectal surgery,
 World Journal of Gastroenterology. 26 (2020) 6945–6962.
- 416 https://doi.org/10.3748/wjg.v26.i44.6945.
- B. Jansen-Winkeln, N. Holfert, H. Köhler, Y. Moulla, J.P. Takoh, S.M. Rabe, M.
 Mehdorn, M. Barberio, C. Chalopin, T. Neumuth, I. Gockel, Determination of the
 transection margin during colorectal resection with hyperspectral imaging (HSI),
 International Journal of Colorectal Disease. 34 (2019) 731–739.
 https://doi.org/10.1007/s00384-019-03250-0.
- B. Jansen-Winkeln, I. Germann, H. Köhler, M. Mehdorn, M. Maktabi, R. Sucher, M.
 Barberio, C. Chalopin, M. Diana, Y. Moulla, I. Gockel, Comparison of hyperspectral
 imaging and fluorescence angiography for the determination of the transection
 margin in colorectal resections—a comparative study, International Journal of
 Colorectal Disease. (2020). https://doi.org/10.1007/s00384-020-03755-z.
- R.A. Cahill, D.F. O'shea, M.F. Khan, H.A. Khokhar, J.P. Epperlein, P.G. mac Aonghusa,
 R. Nair, S.M. Zhuk, Artificial intelligence indocyanine green (ICG) perfusion for
 colorectal cancer intra-operative tissue classification, British Journal of Surgery.
 Ahead of Print (2021). https://doi.org/10.1093/bjs/znaa004.
- 431 [35] A. Spinelli, M. Carvello, P.G. Kotze, A. Maroli, I. Montroni, M. Montorsi, N.C. Buchs,
 432 F. Ris, Ileal pouch–anal anastomosis with fluorescence angiography: a case-matched
 433 study, Colorectal Disease. 21 (2019) codi.14611. https://doi.org/10.1111/codi.14611.
- 434 [36] F. Ris, R. Hompes, C. Cunningham, I. Lindsey, R. Guy, O. Jones, B. George, R.A. Cahill,
 435 N.J. Mortensen, Near-infrared (NIR) perfusion angiography in minimally invasive
 436 colorectal surgery, Surgical Endoscopy. 28 (2014) 2221–2226.
- 437 https://doi.org/10.1007/s00464-014-3432-y.
- K.P. Nugent, P. Daniels, B. Stewart, R. Patankar, C.D. Johnson, Quality of life in stoma
 patients, Diseases of the Colon and Rectum. 42 (1999) 1569–1574.
 https://doi.org/10.1007/BF02236209.
- J. Park, E. Angenete, D. Bock, A. Correa-Marinez, A.K. Danielsen, J. Gehrman, E.
 Haglind, J.E. Jansen, S. Skullman, A. Wedin, J. Rosenberg, Cost analysis in a
 randomized trial of early closure of a temporary ileostomy after rectal resection for
- 444 cancer (EASY trial), Surgical Endoscopy. 34 (2020) 69–76.
- 445 https://doi.org/10.1007/s00464-019-06732-y.
- 446
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- 449 450