

Quantitative assessment of experimental ocular inflammatory disease

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Literature survey and drafting of original manuscript and figures (LJB, AW, LBN); Preparation and provision of data (AW, MCYH, JL, DAC); Discussion, editing and critical revision of manuscript (all authors).

Keywords

Uveitis, EAU, OCT, image processing, automated analysis

Abstract

Word count: 203

Ocular inflammation imposes a high medical burden on patients and substantial costs on the health-care systems that mange these often chronic and debilitating diseases. Many clinical phenotypes are recognized and classifying the severity of inflammation in an eye with uveitis is an ongoing challenge. With the widespread application of optical coherence tomography in the clinic has come the impetus for more robust methods to compare disease between different patients and different treatment centers. Models can recapitulate many of the features seen in the clinic, but until recently the quality of imaging available has lagged that applied in humans. In the model experimental autoimmune uveitis (EAU), we highlight three linked clinical states that produce retinal vulnerability to inflammation, all different from healthy tissue, but distinct from each other. Deploying longitudinal, multimodal imaging approaches can be coupled to analysis in the tissue of changes in architecture, cell content and function. This can enrich our understanding of pathology, increase the sensitivity with which the impacts of therapeutic interventions are assessed and address questions of tissue regeneration and repair. Modern image processing, including the application of artificial intelligence, in the context of such models of disease can lay a foundation for new approaches to monitoring tissue health.

Contribution to the field

There is an ongoing need for objective measures of disease, which is especially pressing in chronic persistent conditions such as uveitis, whose severity may fluctuate and whose treatment may extend over many years. A review of different approaches to scoring argues that increased synthesis of information from different modalities has the potential to improve the specificity with which the state of tissue is defined. This would lead to improvements in understanding of the disease process and increased sensitivity to recognizing changes caused by therapeutic intervention.



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29 care systems that mange these often chronic and debilitating diseases. Many clinical phenotypes are

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- 40 tissue regeneration and repair. Modern image processing, including the application of artificial
- 41 intelligence, in the context of such models of disease can lay a foundation for new approaches to
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44 **2** Contribution to the field

45 There is an ongoing need for objective measures of disease, which is especially pressing in chronic

46 persistent conditions such as uveitis, whose severity may fluctuate and whose treatment may extend

47 over many years. A review of different approaches to scoring argues that increased synthesis of

48 information from different modalities has the potential to improve the specificity with which the state

49 of tissue is defined. This would lead to improvements in understanding of the disease process and

- 50 increased sensitivity to recognizing changes caused by therapeutic intervention.
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52 **3** Introduction

53 Ocular inflammation is an important medical concern with a wide range of manifestations from the

54 easily treatable to sight threatening. It arises both as an ocular specific condition and in association

55 with systemic disease and it manifests as more than 30 defined uveitic phenotypes. The pathogenesis

56 is complex and multifactorial and there is a lively debate as to the relative contribution of subclinical

57 infection, autoinflammation and autoimmunity (Lee et al., 2014; Forrester et al., 2018). Conventional

58 approaches to imaging do not distinguish between these different causes.

59 Animal models of uveitis are often autoimmune (e.g. experimental autoimmune uveitis; EAU),

60 inspired in the mouse by early work identifying susceptible strains (Caspi et al., 1988; Caspi, 2010)

- and used widely to probe important aspects of immune function including tolerance (Lin et al., 2005;
- 62 Lee and Taylor, 2015), regulation (Kerr et al., 2008b; Wang et al., 2014), microbiome (Horai et al.,
- 63 2015), lymphocyte dynamics (Boldison et al., 2014) and macrophage/monocyte function (Raveney et
- 64 al., 2009). But other models of ocular inflammation are also important, including endotoxin induced
- 65 uveitis (EIU) (Forrester et al., 1980; Chu et al., 2016a; Bell et al., 2020) and primed mycobacterial
- 66 uveitis (PMU) (Pepple et al., 2015). Ocular infectious disease can also be studied and has proven to
- be an informative model of inflammation (Zinkernagel et al., 2009; Zinkernagel et al., 2012;
- 68 Zinkernagel et al., 2013).

- 69 Over the last 15 years, techniques for imaging the mouse retina have advanced substantially, first
- 70 with fundal photography, acquired by topical endoscopic fundal imaging (TEFI) (Paques et al., 2007;
- 71 Copland et al., 2008; Xu et al., 2008) facilitating clinical grading by individuals blinded to the origin
- of the images. Then followed by adaptation of clinical tools (Chu et al., 2016a) and development of
- the Micron system for imaging rodent eyes (Phoenix technologies, CA). These advances have made
- 74 acquisition of experimental image data more accessible and routine (Chu et al., 2013; Chu et al.,
- 75 2016a; Zhong et al., 2016; Chen and Caspi, 2019a). The application of optical coherence tomography
- (OCT) to the mouse eye adds new information on changes deep in the tissue. The eye offers unique
 advantages for imaging studies of the autoimmune process in a target tissue, permitting serial
- 77 advantages for imaging studies of the autominume process in a target tissue, permitting serial assessment, and sophisticated quantification of different parameters of inflammation that go beyond
- 79 more general clinical scores used in models such as experimental autoimmune encephalomyelitis.
- 80 Advances in image processing that have been developed in patient populations, can also find
- 81 application in experimental studies. There is potential for automatic segmentation of structures (in
- 82 which the boundaries between, for example, different layers of the retina are identified in an
- 83 unsupervised process), quantification of infiltration and disease classification by machine learning,
- 84 which can be used to support unsupervised clinical assessment (Abramoff et al., 2010;
- 85 Anantrasirichai et al., 2014). This is seen in the recent application of deep learning to EAU (Sun et
- 86 al., 2020). Alternative powerful technologies are also available; using bioluminescent reporters, can
- 87 delineate sequential cell population specific patterns of infiltration (Gutowski et al., 2017; John et al.,
- 88 2020), and multi-optical imaging approaches can produce data on phenotype and the spatial
- 89 relationship between different cell types (Radtke et al., 2020). Objective measurements, that provide
- a more granular multi-modal analysis of the state of the tissue, can then form the basis for
- 91 quantifying the impact of treatment on ocular disease not limited to a single time-point but integrated
- 92 across a longer disease course.
- 93

94 **4** Ocular tissue and inflammation

EAU is often studied with a focus on the acute inflammation that occurs with the explosive influx of
immune cells that flood into the tissue in the first wave of clinical disease. But it has been apparent
for a number of years (Shao et al., 2006; Kerr et al., 2008a) that it can also be used to develop

98 insights into the processes of persistent disease and tissue remodeling. For example, memory cells

- 99 that reside in the bone marrow are implicated in chronic retinal degeneration (Oh et al., 2011) and
- persistent inflammation can lead to retinal angiogenesis (Chen et al., 2012). In both mouse
 (Kielczewski et al., 2016) and human (Epps et al., 2020), chronic disease can drive the development
- 102 of ectopic lymphoid like structures and is accompanied by changes in the other lymphocyte
- populations and vascular remodeling (Chen et al., 2012; Boldison et al., 2014). The ocular tissue can
- 104 therefore exist in a minimum of four well demarcated states (Fig. 1).
- 105 ____> Fig 1 here

106 Healthy tissue resists insult and maintains normal visual function. In the EAU model, there are a

107 minimum of three non-healthy states, which correlate with changes in immune cell content and

108 vascular function (Kerr et al., 2008a). Vulnerable tissue may be in the prodromal phase of EAU, at

- 109 peak of disease, with active infiltration by many different leukocytes, or vulnerable but to a greater or
- 110 lesser extent recovered, which state is described as post-peak. It is possible to observe experimentally
- 111 that the pre-peak state can resolve to a state of health, or progress to peak disease. Tissue can reach

- 112 peak disease from either the pre-peak state or as a relapse from the post-peak state (Diedrichs-
- 113 Möhring et al., 2018). But it is unknown whether from peak or post-peak, tissue can ever return to a
- 114 healthy state. In the broader context, a useful framework for these changes is found in the extensive
- 115 literature describing the development and resolution of inflammation, but here too, the question of
- active resolution in the tissue and the mechanisms by which it occurs remains controversial (Weavers
- and Martin, 2020). While this review focuses studies in the eye, it is evident that other diseases and
- 118 disease models, such as arthritis, can be fitted into a similar framework (Jones et al., 2016).
- 119 One essential tool for advancing understanding of these different tissue states is a rigorous method of
- 120 clinical assessment that separates healthy tissue from the vulnerable and that also distinguishes
- between different states of the vulnerable tissue. Such a scheme could then complement studies
- describing gene expression in different forms of ocular inflammation (Heng et al., 2019; Bell et al.,
- 123 2020). Recent advances in the range and quality of techniques that can be applied to quantify ocular
- 124 inflammatory disease make such objective and transferrable assessments increasingly feasible.

125 **5** Assessment of ocular inflammation

126 The measurement of inflammatory activity is a core objective for clinical studies of uveitis and has

- 127 inspired work that seeks to improve its ability to discriminate between lower levels of disease as well
- 128 as improving its sensitivity (Montesano et al., 2018). Progress in this area can also inform animal
- studies.

130 **5.1 Clinical Scoring**

In human eye disease, improvements in imaging have driven diagnostic sensitivity and specificity 131 (Ravin, 1999; Marchese et al., 2020). Scoring systems serve as tools for categorizing disease activity 132 into ordinal groups and as a convenient measure of clinical outcome and directional change. The first 133 134 aqueous and vitreous inflammation scoring systems based on ophthalmic observation of cell counts 135 in patients were published in 1959 (Hogan et al., 1959; Kimura et al., 1959), but consensus recommendations did not emerge until 2005, under the umbrella of the Standardization of Uveitis 136 137 Nomenclature (SUN) workshop (Trusko et al., 2013). For some diseases, for example Behçet's 138 disease, specific scoring systems have proven useful is assessing treatment response (Kaburaki et al., 2014). It is a recognized concern with scoring systems that there is a tension between precision and 139 140 simplicity. Levels of interobserver agreement remain modest and non-linearity in the scaling can lead 141 to poor resolution of differences in disease especially at lower levels of inflammation (Davis et al., 2010; Hornbeak et al., 2014; Denniston et al., 2017). The use of digital images, where biological data 142 is quantified as pixel values, expands the possibilities for analysis by computer imaging (Abramoff et 143 144 al., 2010) for example for automated grading of vitreous haze (Passaglia et al., 2018). Scoring of 145 clinical disease in EAU has evolved from early approaches using slit-lamp aided visualization and semi-quantitative histological scoring to more sophisticated scoring approaches based on blinded 146 147 assessment of fundal photographs (Agarwal and Caspi, 2004; Copland et al., 2008; Xu et al., 2008; Agarwal et al., 2012; Chen and Caspi, 2019a) and most recently using machine learning. Scoring can 148 149 be on a simple ordinal scale (0-4) or can categorize disease into three indicators of inflammation and 150 one of structural damage with inflammation and structural damage reported independently or as a summary score (0-5) calculated as the total or average score for the eye (Xu et al., 2008; Copland et 151 152 al., 2012; Boldison et al., 2014) (Table 1). When applied as a summary score, this approach can be 153 insensitive to differences in aspects of the underlying pathology, for example in Fig. 2, the two 154 images, although clearly different, received the same summary clinical score.

155 ____> Table 1 here

- 156 Complementing photography is optical coherence tomography (OCT). Developed in the 1990s
- 157 (Huang et al., 1991; Drexler et al., 2014) it has rapidly become the state of the art for non-invasive
- 158 retinal imaging. OCT is an interferometric technique providing depth resolved cross sectional images
- 159 of the retina, known as B-scans. In normal eyes the vitreous is optically transparent, retinal layers
- 160 show different degrees of backscatter, and in humans the RPE is one of the most hyper-reflective
- 161 layers. Modern OCT in humans can also go some way to visualizing the choroid beneath the RPE
- 162 (Mrejen and Spaide, 2013). OCT can resolve retinal substructure and its vasculature, can be
- 163 important in the diagnosis and image guided management of human uveitis and can capture changes 164
- in the state of the tissue through time in EAU (Chen et al., 2013; Chu et al., 2013; Yu et al., 2013;
- 165 Chu et al., 2016a).
- 166 ____> Fig 2 here

167 5.2 Ocular tissue analysis

168 In contrast to the wealth of sophisticated imaging that can be directed at the human eye in uveitis, 169 access to human tissue is severely limited. Enucleation of the globe in uveitis is rare and is usually 170 from individuals with long-standing disease (Epps et al., 2020). But in the EAU model, histology was 171 the first accepted standard for disease assessment (Nussenblatt et al., 1980; Kozak et al., 1981; 172 Mochizuki et al., 1985). Immunohistochemistry and immunofluorescence of retinal tissue revealed 173 the profound structural disruption that accompanies acute inflammation, and was used, for example, 174 to show how macrophages reciprocally alter their expression of CD68 and arginase-1 during the persistent (post-peak) phase of uveitis (Chen et al., 2012). For higher dimensional analysis of cell 175 176 infiltrate, investigators have used multiparameter flow cytometry which can quantify many different 177 cell populations (Thurau et al., 2004; Kerr et al., 2008b; Luger et al., 2008). Sampling the cell infiltrate at different time points has been instrumental in demonstrating important changes in the 178 179 relative frequencies of CD4 T regulatory cells (Silver et al., 2015) and CD8 cells (Boldison et al., 180 2014). In EAU this is strong evidence that at the cellular level as well as in serial imaging studies, the 181 tissue and the immune infiltrate change and adapt through time. Developing improved quantitative 182 methods to assess tissue health in EAU offers more sensitive and specific approaches to analyze the 183 impact of therapies for autoimmunity and inflammation.

184 5.3 Quantitative assessment of EAU

185 Using formal criteria, EAU can be assessed semi-quantitatively, but interobserver disagreement and subjectivity limits the usefulness of direct comparison between results from different labs and even 186 187 individual researchers (Xu et al., 2008). As with human clinical graders, experience is required to 188 achieve the highest levels of interobserver agreement (Li et al., 2017b). Employing contemporary 189 technology has the capacity to improve on these limitations. In addition, in EAU as in other medical 190 images, these can be annotated, with the results of end point tissue analysis added to the meta-data 191 associated with the image. This enriches their interpretation and provides a resource that can be 192 applied to other studies. Pooling data from animal cohorts at selected timepoints runs the risk of 193 obscuring subtle patterns, and overweighting the importance of the certain trends. This can be 194 countered by the use of analysis that exploits modern image processing, with its scope for a higher 195 degree of quantitation (Dysli et al., 2015; Li et al., 2017b; Choi et al., 2018b). A critical element of 196 complementary analysis is therefore the use of non-invasive techniques and computational means to 197 maximize information retrieved from the data.

198 Fundus photography, for example obtained by TEFI, correlates well with disease scores from

199 histopathological analysis (Copland et al., 2008) but the images produce a 2D projection of 3D semi-

- 200 transparent biological tissue. Spatial information is only available in two dimensions and artifacts are
- 201 introduced by flattening depth information onto a plane. More accurate measures of infiltrate,
- 202 oedema and structural changes, that are important manifestations of disease, can be obtained with
- 203 OCT (Chen et al., 2013; Chu et al., 2013). Because OCT produces a depth profile of different
- 204 features, it can be more sensitive than 2D fundus imaging in monitoring the appearance and
- 205 development of pathological changes. In particular, cross sectional images are more sensitive to early
- disease because they can visualize small amounts of infiltrate around the optic nerve, and measure
- 207 changes in optic nerve diameter and retinal thickness due to inflammatory oedema (Chen et al., 2013;
- 208 Dysli et al., 2015; Li et al., 2017b; Chen and Caspi, 2019a).

209 5.4 Aqueous and Vitreous assessment

- 210 A defining characteristic of uveitis is cellular infiltrate, and grading is an important quantitative
- 211 metric in preclinical animal model research. In human disease, anterior uveitis produces 'flare' which
- 212 can be categorized by laser flare photometry and which correlates well with conventional clinical
- 213 grading (Holland, 2007; Agrawal et al., 2016) while in the vitreous, 'haze' is an accepted and
- clinically validated proxy for inflammatory status in patients (Passaglia et al., 2018). Moreover, these
- 215 changes have a marked impact on visual acuity in humans and so are biologically and clinically
- 216 relevant outcome measures (Davis et al., 2010).
- 217 In OCT, cells in either chamber appear as hyperreflective dots, whose profile is a function of many
- 218 variables (Ruggeri et al., 2007; Keane et al., 2015; Zarranz-Ventura et al., 2016). Cells and exudate
- 219 incrementally reduce the optical transparency of the ocular media leading to the aqueous and vitreous
- 220 becoming inhomogeneous as disease severity increases. These changes reduce the contrast of object
- boundaries and the results of qualitative or quantitative image analysis lose precision.
- 222 Because of difficulty in imaging the anterior chamber of small eyes, literature for OCT based cell
- counting in these models is relatively sparse (Pepple et al., 2016). However, automated counts of
- absolute cell numbers have been obtained with excellent correspondence to manual image counts.
- This approach has been developed into a fully automated pipeline for cell counting in volumetric
- OCT images, achieving 98% congruence to manual slit lamp counts. Importantly, the subjective
- 227 manual element of the segmentation step was eliminated. The automated segmentation step involved 228 removal of anatomical structures connected to image boundaries (Choi et al., 2018a). Compared with
- removal of anatomical structures connected to image boundaries (Choi et al., 2018a). Compared with counts from histological sections, OCT tended to undercount, which was attributed to insensitivity to
- counts from histological sections, OCT tended to undercount, which was attributed to insensitivity to cell clumps, sediments and exclusion of the extremities of the iris interface (Pepple et al., 2016). It
- may also be contributory that histology is unaffected by overlying opacities, whereas OCT is
- vulnerable to signal degradation. However, histology introduces artifacts and postmortem changes
- that themselves affect tissue measurement (Depple et al. 2016)
- that themselves affect tissue measurement (Pepple et al., 2016).
- Loss of precision becomes more evident when imaging the vitreous, where the optical pathway
- traverses deeper through affected media. Further complicating the analysis of the rodent vitreous, is
- the anatomical vestige of the hyaloid artery (Smith et al., 2002; Ruggeri et al., 2007), protruding
- 237 upwards from the optic disc towards the lens. It confuses the vitreoretinal boundary and can appear
- 238 somewhat discontinuous, with hyperreflective regions that are subjectively indistinguishable from
- cell clusters.
- 240 Automated counting algorithms usually require a preceding segmentation step, that defines a
- boundary for the area or volume of interest. Variations in signal quality and the ambiguity of
- discontinuous image features frustrate the development of accurate, fully automated methods of

- 243 rodent image segmentation and analysis. Quantification of changes in the vitreous has largely been
- restricted to human images, and global signal parameters, as opposed to absolute cell counts.
- 245 To account for signal strength variations in human OCT images, the average intensity of the
- segmented vitreous compartment can be indexed relative to a hyperreflective reference layer such as
- the RPE, providing a relative intensity ratio. These ratios correlate moderately with clinical vitreous
- haze scores, along with other surrogates of disease such as retinal thickness (Keane et al., 2014;
 Zarranz-Ventura et al., 2016). This process has been fully automated using rule-based algorithms f
- Zarranz-Ventura et al., 2016). This process has been fully automated using rule-based algorithms for
 segmentation, reducing subjectivity. The same operation was also performed using a textural
- 250 segmentation, reducing subjectivity. The same operation was also performed using a textural 251 descriptor of the vitreous, which was marginally better correlated to clinical scores than vitreous
- intensity (Keane et al., 2015). These operations were performed on 2D datasets, obtaining an
- averaged intensity ratio based on several B-scans and data analyzed in 3D may potentially offer
- 254 further improvements.
- 255 Since the scan region is much smaller than the ocular globe, one consideration is the selection of a
- representative and informative region of interest (ROI) that must be equivalent between scans and
- subjects. Within human images, landmarks such as the macula can be located automatically and used
- as a central anchor point for region boundary positioning (Keane et al., 2015). In rodents, the optic
- disc is an obvious landmark choice, but the presence of the hyaloid remnant, particularly in severely diseased eyes warrants additional steps to remove its influence. Recently, an automated method of
- 261 quantifying vitreous inflammation in clinical fundus photographs has been suggested (Davis et al.,
- 262 2010; Passaglia et al., 2018)

263 5.5 Retinal layers

OCT of the healthy retina produces good definition of the different layers of light sensitive tissue. In uveitis it can resolve and localize lesions and pathologies, and identify vasodilation and perivascular exudate (Chen et al., 2013; Chu et al., 2013). Standard clinical OCT has an axial resolution of less than 4 microns, which can produce images with near histological detail. Thickness is ascertained from OCT images by measuring the distance between two boundaries of choice (Fig 3). Before measurements can be taken, the layers must be defined.

270 ____> Fig 3 here

271 Techniques for segmentation to define different retinal layers have progressed through manual, semi-272 automated and fully automated protocols, with work on human data leading rodent OCT imaging. 273 Both rule-based algorithms and learner-based approaches have been applied to the problem and new 274 approaches are under active investigation. Retinal thickness can be measured by OCT absolutely. 275 using assumptions such as an average tissue refractive index (Gadjanski et al., 2011), or by fold 276 change compared to pre-disease measurements (Li et al., 2017b). Both are in high agreement with 277 histological measurements (Gadjanski et al., 2011; Chen et al., 2013; Chu et al., 2013; Berger et al., 278 2014; Li et al., 2017b). Several schemes exist for displaying changes in thickness. One that is 279 commonly used shows thickness at different distances from the optic nerve head (Supplementary Fig. 280 1).

281 Rule-based methods execute a pre-programmed set of instructions, designed with the expected

- properties of the image and the desired features in mind. Many image properties can be analyzed,
- 283 including intensity variation, geometric contours and texture (Ishikawa et al., 2005; Mujat et al.,
- 284 2005; Mishra et al., 2009; Kajic et al., 2010; Gonzalez-Lopez et al., 2019). The number of segmented
- 285 layers defined varies between four and nine, and depends on the approach, with the most successful

- techniques to date being learner models (Garvin et al., 2009; Kajic et al., 2010; Kajic et al., 2012;
- Lang et al., 2013; Anantrasirichai et al., 2014; Venhuizen et al., 2017)

288 OCT offers the potential of assessing layer deformation without the artefacts that can be introduced

- by tissue fixation, sectioning and staining (Spaide and Curcio, 2011). Mechanical deformation can
- also introduce ambiguous artifacts, with likeness to retinal detachments (Gadjanski et al., 2011), and
- 291 congenital abnormalities in the retina may also confound the definition of anatomical normality
- (Mattapallil et al., 2012). The literature pertaining to automated quantitation of retinal structure ismore extensive than that related to infiltrate, because retinal layer changes are associated with a wide
- more extensive than that related to infiltrate, because retinal layer changes are associated with a wide variety of ocular diseases (Srinivasan et al., 2006; Fischer et al., 2009). The laminated reflectance
- profile of the retina's architecture also lends itself to image segmentation and the measurement of
- 296 quantitative indices such as layer thickness and geometric descriptors. Protocols for automatic layer
- segmentation developed for human studies have been tested in different mouse strains. These
- 298 performed well when assessing the inner retinal layers, but were less successful in defining the
- 299 murine RPE, whose location displaced distally into the sclera (Dysli et al., 2015).
- 300 Longitudinal studies of retinal thickness have revealed details about the kinetics of disease
- 301 progression, with respect to other important manifestations of pathology (Chen et al., 2013; Li et al.,
- 302 2017b). In the pre-peak to peak phase of disease, retinal thickness increases rapidly due to
- 303 inflammatory oedema, correlating with inflammatory infiltrate, measured longitudinally by OCT and
- 304 confirmed by histology (Gadjanski et al., 2011; Li et al., 2017b). In the post-peak resolution phase,
- the clearance of exudate reveals features on OCT with greater clarity, such as infiltrate, photoreceptor
- 306 atrophy, retinal folds and choroiditis. (Chen et al., 2013). Photoreceptor damage persists beyond the 307 peak phase of disease as retinal oedema is slower to resolve than inflammatory infiltrate. When the
- 308 swelling does subside, the retina thins to below pre-disease levels because of photoreceptor loss.
- 309 OCT confirms that neither infiltrate or retinal thickness returns to baseline in late disease or even
- after resolution is complete (Copland et al., 2008; Gadjanski et al., 2011; Chen et al., 2013).
- 311 Therefore, quantitative directional changes and relative rates of change between retinal thickness and
- 312 inflammatory infiltrate can provide an additional metric for disease activity.
- 313 In severe uveitis, retinal layers are obscured by opacification of the vitreous and aqueous due to
- 314 infiltrate and proteinaceous exudate (Chen et al., 2013) which presents a challenge for scoring
- 315 systems, that must be robust to substantial signal variation and may need to incorporate metrics of
- 316 opacity into the model as proxies of inflammation.

317 **5.6 Vasculature**

- Important changes in the vasculature occur in uveitis, including ischemia, neovascularization and 318 319 retinal/choroidal vasculitis (Dingerkus et al., 2019). In disease models these are assessed less 320 commonly than structural changes, but as in humans they are often interrogated by angiography. 321 Confocal scanning laser ophthalmoscopy (SLO) can be coupled to fundus fluorescein angiography 322 (FFA) to quantify vessel diameter and leakage in EAU. When average vascular dilation was 323 measured immediately prior to sacrifice and histology, major vessel diameter was well correlated 324 with retina-choroid thickness and with clinical and histological scores. This indicated that 325 inflammatory vasodilation of superficial vasculature was a novel measure of EAU severity (Li et al., 326 2017b). Complementary to dye-based angiography are OCT based methodologies. Vascular dilation 327 and perivascular exudate attributed to retinal vasculitis can be localized to specific retinal layers 328 during the course of EAU (Chen et al., 2013; Chen and Caspi, 2019a) and OCT has been used for
- 329 imaging vasculature disturbances, such as choroiditis and retinal vasculitis (Marchese et al., 2020).

- Blood flow can be visualized and depth resolved (Alnawaiseh et al., 2016) using OCT angiography
- 331 (OCTA) and this has been used to assess retinal microvascular changes (Chu et al., 2016b; Kim et al.,
- 332 2016).
- 333 Many methods of segmenting retinal blood vessels from fundus photographs have been published
- 334 (Moccia et al., 2018). A much smaller number of approaches have been successfully devised using
- 335 OCT images, which include the use of multimodal imaging (corresponding fundus photographs) and
- learner models (Hu et al., 2012; Rodrigues et al., 2013). In humans, segmentation of fine capillary
- networks has been achieved in OCTA enface images (Zhu et al., 2019) while in mice segmentation of
- retinal vasculature using OCTA has been reported for longitudinal monitoring of angiogenesis (Li et
- al., 2017a). Current advances applying deep learning to vessel segmentation continue to improve the
- performance of these methods and this has been helped by the public access to data sets (Ma et al.,2021).
- - -

342 5.7 Functional

343 As EAU progresses, electroretinogram (ERG) amplitudes change. There is a dramatic reduction in

- function (a and b wave), that accompanies early disease (Chen and Caspi, 2019b), presenting before
- 345 morphologic changes. These findings indicate that functional loss could be mediated by
- 346 inflammation rather than just physical damage, and that retinal function is potentially a sensitive
- early indicator (Chen et al., 2013; Li et al., 2017b). However, photoreceptor damage continues while
- 348 inflammation is receding and in the post-peak phase, ERG amplitudes are correlated with OCT
- 349 measures of retinal thickness. As swelling diminishes, photoreceptor atrophy becomes apparent and
- 350 results in an overall retinal thinning compared to baseline. Neither retinal thickness nor functionality
- ever fully recover (Chen et al., 2013; Chen and Caspi, 2019b).
- 352 Taken together, multimodal quantitative measures can provide information on perceptually subtle,
- 353 but biologically significant changes whose quantification would aid clinical grading and pre-clinical
- 354 research.

355 6 Examples of multimodal measurement

356 A multimodal approach to assessing uveitis is outlined in Fig. 4. EAU was induced by the transfer of 357 pathogenic autoantigen reactive T cells. Sequential imaging of all eyes was carried out by fundal 358 photography and OCT. B-scans were segmented manually and measured by an observer blinded to 359 treatment conditions. Measurements of retinal thickness were made at baseline from all eves (n=11) 360 and these were compared as a Z-score expressing the magnitude of change in thickness on day 13 361 color coded as the number of standard deviations from baseline (Fig. 4D). Fig. 4A-C shows images 362 from a representative single eye at baseline and day 13. The retinal photographs (Fig. 4A) show that 363 at day 13 there is an enlarged optic nerve, sheathing of the vessels due to cell infiltration (white 364 arrow) and infiltrates in the tissue (black arrow). B-scans (Fig. 4B) through the optic nerve, were 365 assembled from multiple averaged frames and are displayed with the accompanying 100 micron scale 366 bars that were used to generate measurements of the retinal thickness following manual segmentation 367 using ImageJ (Schneider et al., 2012). At day 13 it is easy to see objects in the vitreous around the 368 optic nerve. The 3D image (Fig. 4C) is prepared from 512 sequential B scans, processed using code 369 in MATLAB (Natick, Massachusetts: The MathWorks Inc) and ImageJ (Anantrasirichai et al., 2014) 370 adapted for use with murine images and rendered using ImageJ (1.53 3D viewer plugin). These 371 pictures give a better appreciation of the spatial distribution of the vitreal infiltrate and can be used to 372 make a semi-quantitative estimate of the degree of vitreal infiltration.

373 ____> Fig 4 here

374 Following changes in disease scores through time, it is useful to display the aggregate data from the 375 multiple images, and this has been used to produce a color-coded map of the retina, with changes 376 normalized to baseline scans (usually on day 0) and scaled by Z-score. Retinal maps are also useful when comparing the pattern of pathological change between different disease models. For example, 377 378 compare Fig. 4D, which shows that at day 13 the major impact of uveitis is found in the vitreous and 379 the optic nerve with Fig. 5 which shows the does dependent effect of intra-vitreal instillation of paraquat, a model of oxidative stress, in C57BL/6 mice. This induces neuronal degeneration which 380 varies with stain (Cingolani et al., 2006) and in this case particularly impacts the inner retina, seen as 381 382 a negative Z-score increasing in magnitude with dose. But quantitative analysis also reveals that at 383 higher concentrations of paraquat, this is accompanied by an expansion of the outer segments, due to 384 inflammation. This finding, using multimodal analysis is in agreement with a previous report 385 showing more pronounced TUNEL-positive cells in the inner retina than in the outer retina of 386 C57BL/6 mice treated intravitreally with paraquat (Cingolani et al., 2006).

387 ____> Fig 5 here

388 **6.1 Opportunities for automation**

389 Machine learning has made an impact in human clinical care in recent years because of its ability to 390 reach expert-level diagnosis. The automated analysis of ocular disease has led the way in carrying

these methodologies into the clinic, but they have been less extensively utilized in disease models

392 (Liu et al., 2019; Faes et al., 2020).

393 Images are inherently data rich because in theory each pixel can be regarded as a separate input parameter (Faes et al., 2020). This offers opportunities for uncovering novel aspects of pathological 394 395 processes but also challenges, especially in assembling well annotated data sets that are large enough 396 to avoid overparameterization when they are used to train classification algorithms in a machine 397 learning framework. Advances in predictive statistical methods may in time alleviate the need for 398 such extensive input data. One helpful approach, applied in OCT, is decoupling the methods for 399 segmentation from artificial intelligence driven disease classification (De Fauw et al., 2018). This 400 moves practice towards device-independent representation of the disease process, which may aid in 401 comparison between studies carried out by different investigators.

402 Recently the field has advanced with the application of a deep learning model to analyze photographs
403 of the retinas of mice with EAU. Using a data set of images that was extended by data augmentation,
404 disease images were divided into three categories and by applying deep learning methods

404 (convolutional neural networks) the overall performance assessed by area under the receiver

405 (convolutional neural networks) the overall performance assessed by area under the receiver 406 operating characteristic curve (AUC) when the model was applied to an external dataset of 33 images

- 407 was approximately 0.90 (Sun et al., 2020).
- 408 Another area of opportunity in multi-modal ocular imaging is the fusion of information from
- 409 different modalities such as fundal photography and OCT (Mitchell, 2010; Dogra et al., 2017). Image
- 410 fusion aims to yield a more complete, accurate and efficient account of an object by combining
- 411 different visualizations together. Integrating this methodology into the assessment of experimental
- 412 clinical disease will inform our ability to distinguish between different states of tissue health (Fig. 1).

413 **7** Conclusion

- 414 Persistent ocular inflammation is a significant and challenging clinical entity that is associated with
- 415 long term changes in the retina and serious sight threatening complications (Dick et al., 2016).
- 416 Experimental models of non-infectious and infectious ocular inflammation have been widely and
- 417 successfully deployed. But fundamental insights regarding how tissue homeostasis is perturbed and
- 418 how it might be restored are still needed (Epps et al., 2018). Such concerns are important in a much 419 broader context than uveitis. Restoring complex tissues, damaged by persistent inflammation, to
- 420 normal physiological function will have wide application. Multimodal and quantitative imaging of
- 421 the eye, in an experimental context, has potential to advance our understanding of the kinetics, cell
- 422 biology, transcriptomic and proteomic architecture of how this multifactorial process is regulated. By
- 423 providing non-invasive techniques to probe the underlaying nature of the tissue, there is an
- 424 opportunity for a more precise and comprehensive discrimination between different states that can be
- 425 used to stratify information gleaned from detailed examination of the transcriptome and microbiome,
- 426 multiparameter flow cytometry and proteomics.
- 427

428 **8 Table**

- Score **Optic disc Retinal vessels** Retinal tissue **Structural damage** infiltration 1 Minimal Cuffing: 1-4 mild 1-4 small lesions or Retinal lesions or retinal inflammation 1 linear lesion atrophy involving 1/4 to 3/4 of retinal area 2 Mild inflammation Cuffing: >4 mild or 5-10 small lesions Panretinal atrophy with 1-3 moderate or 2–3 linear multiple small lesions lesions (scars) or ≤ 3 linear lesions (scars) 3 Moderate Cuffing: >3 >10 small lesions or Pan-retinal atrophy with moderate inflammation >3 linear lesions >3 linear lesions or confluent lesions (scars) 4 Severe Cuffing: >1 severe Linear lesion **Retinal detachment with** inflammation confluent folding
- 429 Table 1. Scheme for scoring clinical ocular inflammation.

5	Not visible (white-	Not visible (white-	Not visible (white-	Not visible (white-out or
	out or extreme	out or extreme	out or extreme	extreme detachment)
	detachment)	detachment)	detachment)	

430 A blinded observer assigns scores to retinal photographs for changes that relate to inflammation of

the optic disc, retinal vessels and retinal tissue and a score for structural damage. These scores can 431

432 then be summed independently (score of 0-20) or given as a summary score of the average of all

features (score of 0-5). (Xu et al., 2008; Copland et al., 2012; Boldison et al., 2014). 433

434

435 9 **Captions for Figures**

- 436 Figure 1 Tissue states in ocular inflammation.
- Healthy ocular tissue is 'immune-privileged' and under low-level immunosurveillance. Specific 437
- 438 (ocular antigen driven) and non-specific (extra-ocular inflammation) stimuli disturb this homeostasis
- 439 and increase interactions across the blood retinal barrier making the tissue more vulnerable to the
- development of disease. In uveitis following active immunization, this starts with the prodrome (Kerr 440
- et al., 2008), which can resolve back to the healthy state. When the prodrome progresses to clinical 441
- 442 EAU in immunocompetent animals, there is an influx of cells to a maximum (peak) followed by a
- reduction in immune cell content, which does not return to base line. The post-peak (in EAU 443
- 444 described as secondary regulation) is distinguished from the pre-peak by changes in the relative
- proportion of different lymphocyte populations (CD4 T regulatory cells, CD8 T resident memory 445 446
- cells). There is currently no evidence that disease proceeds directly from pre-peak to post-peak, nor
- 447 that eves that have reached peak disease ever return to the normal healthy state.
- 448 Figure 2. Clinical score can be insensitive to underlying pathology.
- Mouse eyes imaged using Micron IV with OCT (Phoenix technology group, CA). Left (A&C) and 449
- right (B&D) eyes assessed by fundal photography (A&B) and OCT (C&D). Retinal photographs 450
- scored in a set of images by an observer blinded to the treatment groups, both received the same 451
- summary clinical score. Scale bar 100 µm. 452
- 453 Figure 3: OCT of the normal mouse retina delineates layers and allows retinal dimensions to be
- 454 quantified. Scale bars are 100 microns and illustrate differences in axial and lateral resolution. GCL
- 455 ganglion cell layer; IPL inner plexiform layer; INL inner nuclear layer; OPL outer plexiform layer;
- ONL outer nuclear layer; ELM external limiting membrane; IS/OS inner and outer segments; RPE 456
- 457 retinal pigment epithelium (Dysli et al., 2015).
- 458 Figure 4: Multimodal analysis of EAU.
- 459 Mouse eyes were imaged at day 0 and day 13 after the induction of EAU and one representative
- image of the same eye is shown (A-C). Clinical disease can be assessed by photography (A), 460
- measurements of retinal thickness and optic nerve diameter at three points from the temporal, nasal 461
- and optic nerve regions of the OCT B-scans (B), 3D-reconstuction of retinal infiltrate (C) and 462
- summary data of retinal scores from all groups (D). Summary scores are assembled from 463
- 464 unsupervised quantitative assessment of vitreal involvement, manual segmentation and measurement
- 465 of inner and outer layer thickness and optic nerve diameter transformed and represented as Z-scores.

466 Figure 5: Changes in retinal thickness in mouse eyes following intra-vitreal paraquat instillation were

467 measured on day 10. Images were visualized by OCT, manually segmented, and measured at three

- 468 points in the temporal, nasal, and optic nerve regions. Measurements are expressed as positive and
- 469 negative Z-scores relative to a PBS injected control group. Changes in the inner and outer layers are
- 470 decoupled.
- 471

472 **10** Conflict of Interest

473 *The authors declare that the research was conducted in the absence of any commercial or financial* 474 *relationships that could be construed as a potential conflict of interest.*

475 **11** Author Contributions

Literature survey and drafting of original manuscript and figures (LJB, AW, LBN); Preparation and
 provision of data (AW, MCYH, JL, DAC); Discussion, editing and critical revision of manuscript (all
 authors)

478 authors).

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482 13 References

- Abramoff, M.D., Garvin, M.K., and Sonka, M. (2010). Retinal imaging and image analysis. *IEEE reviews in biomedical engineering* 3, 169-208. doi: 10.1109/rbme.2010.2084567.
- 485 Agarwal, R.K., and Caspi, R.R. (2004). Rodent models of experimental autoimmune uveitis.
 486 *Methods Mol Med* 102, 395-419. doi: 10.1385/1-59259-805-6:395.
- 487 Agarwal, R.K., Silver, P.B., and Caspi, R.R. (2012). Rodent models of experimental autoimmune
 488 uveitis. *Methods Mol Biol* 900, 443-469. doi: 10.1007/978-1-60761-720-4_22.
- Agrawal, R., Keane, P.A., Singh, J., Saihan, Z., Kontos, A., and Pavesio, C.E. (2016). Classification
 of semi-automated flare readings using the Kowa FM 700 laser cell flare meter in patients
 with uveitis. *Acta Ophthalmologica* 94(2), e135-e141. doi: 10.1111/aos.12833.
- Alnawaiseh, M., Rosentreter, A., Hillmann, A., Alex, A.F., Niekamper, D., Heiduschka, P., et al.
 (2016). OCT angiography in the mouse: A novel evaluation method for vascular pathologies
 of the mouse retina. *Exp Eye Res* 145, 417-423. doi: 10.1016/j.exer.2016.02.012.
- Anantrasirichai, N., Nicholson, L.B., Morgan, J.E., Erchova, I., Mortlock, K., North, R.V., et al.
 (2014). Adaptive-weighted bilateral filtering and other pre-processing techniques for optical
 coherence tomography. *Computerized Medical Imaging and Graphics* 38(6), 526-539. doi:
 10.1016/j.compmedimag.2014.06.012.
- Bell, O.H., Copland, D.A., Ward, A., Nicholson, L.B., Lange, C.A.K., Chu, C.J., et al. (2020). Single
 Eye mRNA-Seq Reveals Normalisation of the Retinal Microglial Transcriptome Following
 Acute Inflammation. *Frontiers in Immunology* 10(3033). doi: 10.3389/fimmu.2019.03033.
- Berger, A., Cavallero, S., Dominguez, E., Barbe, P., Simonutti, M., Sahel, J.A., et al. (2014).
 Spectral-domain optical coherence tomography of the rodent eye: highlighting layers of the

504 505	outer retina using signal averaging and comparison with histology. <i>PLoS One</i> 9(5), e96494. doi: 10.1371/journal.pone.0096494.
506	Boldison, J., Chu, C.J., Copland, D.A., Lait, P.J.P., Khera, T.K., Dick, A.D., et al. (2014). Tissue-
507	Resident Exhausted Effector Memory CD8+ T Cells Accumulate in the Retina during
508	Chronic Experimental Autoimmune Uveoretinitis. <i>The Journal of Immunology</i> 192, 4541-
509	4550. doi: 10.4049/jimmunol.1301390.
510 511	Caspi, R.R. (2010). A look at autoimmunity and inflammation in the eye. <i>The Journal of Clinical Investigation</i> 120(9), 3073-3083.
512 513 514	Caspi, R.R., Roberge, F.G., Chan, C.C., Wiggert, B., Chader, G.J., Rozenszajn, L.A., et al. (1988). A new model of autoimmune disease. Experimental autoimmune uveoretinitis induced in mice with two different retinal antigens. <i>Journal of Immunology</i> 140(5), 1490-1495.
515	Chen, J., and Caspi, R.R. (2019a). "Clinical and Functional Evaluation of Ocular Inflammatory
516	Disease Using the Model of Experimental Autoimmune Uveitis," in <i>Immunological</i>
517	<i>Tolerance: Methods and Protocols</i> , ed. A.S. Boyd. (New York, NY: Springer New York),
518	211-227.
519	Chen, J., and Caspi, R.R. (2019b). Clinical and Functional Evaluation of Ocular Inflammatory
520	Disease Using the Model of Experimental Autoimmune Uveitis. <i>Methods Mol Biol</i> 1899,
521	211-227. doi: 10.1007/978-1-4939-8938-6_15.
522 523 524	Chen, J., Qian, H., Horai, R., Chan, C.C., and Caspi, R.R. (2013). Use of optical coherence tomography and electroretinography to evaluate retinal pathology in a mouse model of autoimmune uveitis. <i>PLoS One</i> 8(5), e63904. doi: 10.1371/journal.pone.0063904.
525	Chen, M., Copland, D.A., Zhao, J., Liu, J., Forrester, J.V., Dick, A.D., et al. (2012). Persistent
526	Inflammation Subverts Thrombospondin-1–Induced Regulation of Retinal Angiogenesis and
527	Is Driven by CCR2 Ligation. <i>American Journal of Pathology</i> 180(1), 235-245.
528	Choi, W.J., Pepple, K.L., and Wang, R.K. (2018a). Automated three-dimensional cell counting
529	method for grading uveitis of rodent eye in vivo with optical coherence tomography. J
530	Biophotonics 11(9), e201800140. doi: 10.1002/jbio.201800140.
531 532 533	Choi, W.J., Pepple, K.L., and Wang, R.K. (2018b). Automated three-dimensional cell counting method for grading uveitis of rodent eye in vivo with optical coherence tomography. <i>Journal of Biophotonics</i> 11(9), e201800140. doi: 10.1002/jbio.201800140.
534	Chu, C.J., Gardner, P.J., Copland, D.A., Liyanage, S.E., Gonzalez-Cordero, A., kleine Holthaus, S
535	M., et al. (2016a). Multimodal analysis of ocular inflammation using the endotoxin-induced
536	uveitis mouse model. <i>Disease Models & Mechanisms</i> 9(4), 473-481. doi:
537	10.1242/dmm.022475.
538	 Chu, C.J., Herrmann, P., Carvalho, L.S., Liyanage, S.E., Bainbridge, J.W.B., Ali, R.R., et al. (2013).
539	Assessment and <i>In Vivo</i> Scoring of Murine Experimental Autoimmune Uveoretinitis Using
540	Optical Coherence Tomography. <i>PLoS ONE</i> 8(5), e63002. doi:
541	10.1371/journal.pone.0063002.
542	Chu, Z., Lin, J., Gao, C., Xin, C., Zhang, Q., Chen, C.L., et al. (2016b). Quantitative assessment of
543	the retinal microvasculature using optical coherence tomography angiography. <i>J Biomed Opt</i>
544	21(6), 66008. doi: 10.1117/1.JBO.21.6.066008.

545 Cingolani, C., Rogers, B., Lu, L., Kachi, S., Shen, J., and Campochiaro, P.A. (2006). Retinal 546 degeneration from oxidative damage. Free Radical Biology and Medicine 40(4), 660-669. 547 doi: 10.1016/j.freeradbiomed.2005.09.032. 548 Copland, D.A., Liu, J., Schewitz-Bowers, L.P., Brinkmann, V., Anderson, K., Nicholson, L.B., et al. 549 (2012). Therapeutic Dosing of Fingolimod (FTY720) Prevents Cell Infiltration, Rapidly 550 Suppresses Ocular Inflammation, and Maintains the Blood-Ocular Barrier. American Journal of Pathology 180, 672-681. 551 552 Copland, D.A., Wertheim, M.S., Raveney, B.J.E., Armitage, W.J., Nicholson, L.B., and Dick, A.D. 553 (2008). The clinical time-course of experimental autoimmune uveoretinitis using topical 554 endoscopic fundal imaging with histological and cellular infiltrate correlation. *Investigative* 555 Ophthalmology & Visual Science 49, 5458-5465. 556 Davis, J.L., Madow, B., Cornett, J., Stratton, R., Hess, D., Porciatti, V., et al. (2010). Scale for 557 Photographic Grading of Vitreous Haze in Uveitis. American Journal of Ophthalmology 558 150(5), 637-641.e631. doi: 10.1016/j.ajo.2010.05.036. 559 De Fauw, J., Ledsam, J.R., Romera-Paredes, B., Nikolov, S., Tomasev, N., Blackwell, S., et al. 560 (2018). Clinically applicable deep learning for diagnosis and referral in retinal disease. Nature 561 Medicine 24(9), 1342-1350. doi: 10.1038/s41591-018-0107-6. 562 Denniston, A.K., Keane, P.A., and Srivastava, S.K. (2017). Biomarkers and Surrogate Endpoints in 563 Uveitis: The Impact of Quantitative Imaging. Investigative Ophthalmology & Visual Science 564 58(6), BIO131-BIO140. doi: 10.1167/iovs.17-21788. 565 Dick, A.D., Tundia, N., Sorg, R., Zhao, C., Chao, J.D., Joshi, A., et al. (2016). Risk of Ocular Complications in Patients with Noninfectious Intermediate Uveitis, Posterior Uveitis, or 566 567 Panuveitis. Ophthalmology 123(3), 655-662. doi: 10.1016/j.ophtha.2015.10.028. 568 Diedrichs-Möhring, M., Kaufmann, U., and Wildner, G. (2018). The immunopathogenesis of chronic 569 and relapsing autoimmune uveitis – Lessons from experimental rat models. Progress in 570 Retinal and Eye Research. doi: 10.1016/j.preteyeres.2018.02.003. 571 Dingerkus, V.L.S., Munk, M.R., Brinkmann, M.P., Freiberg, F.J., Heussen, F.M.A., Kinzl, S., et al. 572 (2019). Optical coherence tomography angiography (OCTA) as a new diagnostic tool in 573 uveitis. J Ophthalmic Inflamm Infect 9(1), 10. doi: 10.1186/s12348-019-0176-9. 574 Dogra, A., Goyal, B., Agrawal, S., and Ahuja, C.K. (2017). Efficient fusion of osseous and vascular 575 details in wavelet domain. Pattern Recognition Letters 94, 189-193. doi: 576 10.1016/j.patrec.2017.03.002. 577 Drexler, W., Liu, M.Y., Kumar, A., Kamali, T., Unterhuber, A., and Leitgeb, R.A. (2014). Optical 578 coherence tomography today: speed, contrast, and multimodality. Journal of Biomedical 579 Optics 19(7). doi: 10.1117/1.jbo.19.7.071412. 580 Dysli, C., Enzmann, V., Sznitman, R., and Zinkernagel, M.S. (2015). Quantitative Analysis of Mouse 581 Retinal Layers Using Automated Segmentation of Spectral Domain Optical Coherence 582 Tomography Images. Translational Vision Science & Technology 4(4), 9-9. doi: 583 10.1167/tvst.4.4.9. 584 Epps, S.J., Boldison, J., Stimpson, M.L., Khera, T.K., Lait, P.J.P., Copland, D.A., et al. (2018). Re-585 programming immunosurveillance in persistent non-infectious ocular inflammation. Progress 586 in Retinal and Eye Research 65, 93-106. doi: 10.1016/j.preteveres.2018.03.001.

- 587 Epps, S.J., Coplin, N., Luthert, P.J., Dick, A.D., Coupland, S.E., and Nicholson, L.B. (2020).
 588 Features of ectopic lymphoid-like structures in human uveitis. *Experimental Eye Research* 589 191. doi: 10.1016/j.exer.2019.107901.
- Faes, L., Liu, X., Wagner, S.K., Fu, D.J., Balaskas, K., Sim, D.A., et al. (2020). A Clinician's Guide
 to Artificial Intelligence: How to Critically Appraise Machine Learning Studies.
 Translational Vision Science & Technology 9(2), 7-7. doi: 10.1167/tvst.9.2.7.
- Fischer, M.D., Huber, G., Beck, S.C., Tanimoto, N., Muehlfriedel, R., Fahl, E., et al. (2009).
 Noninvasive, In Vivo Assessment of Mouse Retinal Structure Using Optical Coherence
 Tomography. *Plos One* 4(10). doi: 10.1371/journal.pone.0007507.
- Forrester, J.V., Kuffova, L., and Dick, A.D. (2018). Autoimmunity, Autoinflammation, and Infection
 in Uveitis. *American Journal of Ophthalmology* 189, 77-85. doi: 10.1016/j.ajo.2018.02.019.
- Forrester, J.V., Worgul, B.V., and Merriam, G.R., Jr. (1980). Endotoxin-induced uveitis in the rat.
 Albrecht Von Graefes Archiv fur Klinische und Experimentelle Ophthalmologie 213(4), 221 233.
- Gadjanski, I., Williams, S.K., Hein, K., Sattler, M.B., Bahr, M., and Diem, R. (2011). Correlation of
 optical coherence tomography with clinical and histopathological findings in experimental
 autoimmune uveoretinitis. *Exp Eye Res* 93(1), 82-90. doi: 10.1016/j.exer.2011.04.012.
- Garvin, M.K., Abramoff, M.D., Wu, X., Russell, S.R., Burns, T.L., and Sonka, M. (2009).
 Automated 3-D intraretinal layer segmentation of macular spectral-domain optical coherence tomography images. *IEEE Trans Med Imaging* 28(9), 1436-1447. doi: 10.1109/TMI.2009.2016958.
- Gonzalez-Lopez, A., de Moura, J., Novo, J., Ortega, M., and Penedo, M.G. (2019). Robust
 segmentation of retinal layers in optical coherence tomography images based on a multistage
 active contour model. *Heliyon* 5(2), e01271. doi: 10.1016/j.heliyon.2019.e01271.
- Gutowski, M.B., Wilson, L., Van Gelder, R.N., and Pepple, K.L. (2017). In Vivo Bioluminescence
 Imaging for Longitudinal Monitoring of Inflammation in Animal Models of Uveitis. *Invest Ophthalmol Vis Sci* 58(3), 1521-1528. doi: 10.1167/iovs.16-20824.
- Heng, J.S., Hackett, S.F., Stein-O'Brien, G.L., Winer, B.L., Williams, J., Goff, L.A., et al. (2019).
 Comprehensive analysis of a mouse model of spontaneous uveoretinitis using single-cell
 RNA sequencing. *Proceedings of the National Academy of Sciences*, 201915571. doi:
 10.1073/pnas.1915571116.
- Hogan, M.J., Kimura, S.J., and Thygeson, P. (1959). Signs and symptoms of uveitis. I. Anterior
 uveitis. *Am J Ophthalmol* 47(5 Pt 2), 155-170. doi: 10.1016/s0002-9394(14)78239-x.
- Holland, G.N. (2007). A reconsideration of anterior chamber flare and its clinical relevance for
 children with chronic anterior uveitis (an American Ophthalmological Society thesis).
 Transactions of the American Ophthalmological Society 105, 344-364.
- Horai, R., Zárate-Bladés, Carlos R., Dillenburg-Pilla, P., Chen, J., Kielczewski, Jennifer L., Silver,
 Phyllis B., et al. (2015). Microbiota-Dependent Activation of an Autoreactive T Cell Receptor
 Provokes Autoimmunity in an Immunologically Privileged Site. *Immunity* 43(2), 343-353.
 doi: 10.1016/j.immuni.2015.07.014.
- Hornbeak, D.M., Payal, A., Pistilli, M., Biswas, J., Ganesh, S.K., Gupta, V., et al. (2014).
 Interobserver agreement in clinical grading of vitreous haze using alternative grading scales. *Ophthalmology* 121(8), 1643-1648. doi: 10.1016/j.ophtha.2014.02.018.

630 Hu, Z., Niemeijer, M., Abramoff, M.D., and Garvin, M.K. (2012). Multimodal retinal vessel 631 segmentation from spectral-domain optical coherence tomography and fundus photography. 632 IEEE Trans Med Imaging 31(10), 1900-1911. doi: 10.1109/TMI.2012.2206822. 633 Huang, D., Swanson, E., Lin, C., Schuman, J., Stinson, W., Chang, W., et al. (1991). Optical 634 coherence tomography. Science 254(5035), 1178-1181. doi: 10.1126/science.1957169. 635 Ishikawa, H., Stein, D.M., Wollstein, G., Beaton, S., Fujimoto, J.G., and Schuman, J.S. (2005). Macular segmentation with optical coherence tomography. Invest Ophthalmol Vis Sci 46(6). 636 637 2012-2017. doi: 10.1167/iovs.04-0335. 638 John, S., Rolnick, K., Wilson, L., Wong, S., Van Gelder, R.N., and Pepple, K.L. (2020). 639 Bioluminescence for in vivo detection of cell-type-specific inflammation in a mouse model of 640 uveitis. Scientific Reports 10(1), 11377. doi: 10.1038/s41598-020-68227-4. 641 Jones, G.W., Hill, D.G., and Jones, S.A. (2016). Understanding Immune Cells in Tertiary Lymphoid Organ Development: It Is All Starting to Come Together. Front Immunol 7, 401. doi: 642 10.3389/fimmu.2016.00401. 643 644 Kaburaki, T., Namba, K., Sonoda, K.-h., Kezuka, T., Keino, H., Fukuhara, T., et al. (2014). Behcet's 645 disease ocular attack score 24: evaluation of ocular disease activity before and after initiation 646 of infliximab. Japanese Journal of Ophthalmology 58(2), 120-130. doi: 10.1007/s10384-013-647 0294-0. 648 Kajic, V., Esmaeelpour, M., Povazay, B., Marshall, D., Rosin, P.L., and Drexler, W. (2012). 649 Automated choroidal segmentation of 1060 nm OCT in healthy and pathologic eyes using a 650 statistical model. Biomed Opt Express 3(1), 86-103. doi: 10.1364/BOE.3.000086. 651 Kajic, V., Povazay, B., Hermann, B., Hofer, B., Marshall, D., Rosin, P.L., et al. (2010). Robust 652 segmentation of intraretinal layers in the normal human fovea using a novel statistical model 653 based on texture and shape analysis. Opt Express 18(14), 14730-14744. doi: 654 10.1364/OE.18.014730. 655 Keane, P.A., Balaskas, K., Sim, D.A., Aman, K., Denniston, A.K., Aslam, T., et al. (2015). 656 Automated Analysis of Vitreous Inflammation Using Spectral-Domain Optical Coherence 657 Tomography. Translational Vision Science & Technology 4(5). doi: 10.1167/tvst.4.5.4. 658 Keane, P.A., Karampelas, M., Sim, D.A., Sadda, S.R., Tufail, A., Sen, H.N., et al. (2014). Objective 659 measurement of vitreous inflammation using optical coherence tomography. *Ophthalmology* 121(9), 1706-1714. doi: 10.1016/j.ophtha.2014.03.006. 660 661 Kerr, E.C., Copland, D.A., Dick, A.D., and Nicholson, L.B. (2008a). The Dynamics of Leukocyte Infiltration in Experimental Autoimmune Uveoretinitis. Progress in Retinal and Eye 662 Research 27, 527-535. 663 664 Kerr, E.C., Raveney, B.J.E., Copland, D.A., Dick, A.D., and Nicholson, L.B. (2008b). Analysis of 665 Retinal Cellular Infiltrate in Experimental Autoimmune Uveoretinitis Reveals Multiple 666 Regulatory Cell Populations. Journal of Autoimmunity 31, 354-361. 667 Kielczewski, J.L., Horai, R., Jittayasothorn, Y., Chan, C.-C., and Caspi, R.R. (2016). Tertiary 668 Lymphoid Tissue Forms in Retinas of Mice with Spontaneous Autoimmune Uveitis and Has 669 Consequences on Visual Function. The Journal of Immunology 196(3), 1013-1025. doi: 670 10.4049/jimmunol.1501570. 671 Kim, A.Y., Rodger, D.C., Shahidzadeh, A., Chu, Z., Koulisis, N., Burkemper, B., et al. (2016). 672 Quantifying Retinal Microvascular Changes in Uveitis Using Spectral-Domain Optical

- 673 Coherence Tomography Angiography. *Am J Ophthalmol* 171, 101-112. doi:
 674 10.1016/j.ajo.2016.08.035.
- Kimura, S.J., Thygeson, P., and Hogan, M.J. (1959). Signs and symptoms of uveitis. II.
 Classification of the posterior manifestations of uveitis. *Am J Ophthalmol* 47(5 Pt 2), 171176. doi: 10.1016/s0002-9394(14)78240-6.
- Kozak, Y.d., Sakai, J., Thillaye, B., and Faure, J.P. (1981). S antigen-induced experimental
 autoimmune uveo-retinitis in rats. *Current Eye Research* 1(6), 327-337. doi:
 10.3109/02713688108998359.
- Lang, A., Carass, A., Hauser, M., Sotirchos, E.S., Calabresi, P.A., Ying, H.S., et al. (2013). Retinal
 layer segmentation of macular OCT images using boundary classification. *Biomed Opt Express* 4(7), 1133-1152. doi: 10.1364/BOE.4.001133.
- Lee, D.J., and Taylor, A.W. (2015). Recovery from experimental autoimmune uveitis promotes
 induction of antiuveitic inducible Tregs. *Journal of Leukocyte Biology* 97(6), 1101-1109. doi:
 doi:10.1189/jlb.3A1014-466RR.
- Lee, R.W.J., Nicholson, L.B., Sen, H.N., Chan, C.C., Wei, L., Nussenblatt, R.B., et al. (2014).
 Autoimmune and autoinflammatory mechanisms in uveitis. *Seminars in Immunopathology* 36, 581-594. doi: DOI 10.1007/s00281-014-0433-9.
- Li, A., You, J., Du, C., and Pan, Y. (2017a). Automated segmentation and quantification of OCT
 angiography for tracking angiogenesis progression. *Biomed Opt Express* 8(12), 5604-5616.
 doi: 10.1364/BOE.8.005604.
- Li, J., Ren, J., Yip, Y.W.Y., Zhang, X., Chu, K.O., Ng, T.K., et al. (2017b). Quantitative
 Characterization of Autoimmune Uveoretinitis in an Experimental Mouse Model. *Invest Ophthalmol Vis Sci* 58(10), 4193-4200. doi: 10.1167/iovs.17-22436.

Lin, H.H., Faunce, D.E., Stacey, M., Terajewicz, A., Nakamura, T., Zhang-Hoover, J., et al. (2005).
 The macrophage F4/80 receptor is required for the induction of antigen-specific efferent
 regulatory T cells in peripheral tolerance. *Journal of Experimental Medicine* 201(10), 1615 1625.

- Liu, X.X., Faes, L., Kale, A.U., Wagner, S.K., Fu, D.J., Bruynseels, A., et al. (2019). A comparison of deep learning performance against health-care professionals in detecting diseases from medical imaging: a systematic review and meta-analysis. *Lancet Digital Health* 1(6), E271-E297. doi: 10.1016/s2589-7500(19)30123-2.
- Luger, D., Silver, P.B., Tang, J., Cua, D., Chen, Z., Iwakura, Y., et al. (2008). Either a Th17 or a Th1
 effector response can drive autoimmunity: conditions of disease induction affect dominant
 effector category. *Journal of Experimental Medicine* 205(4), 799-810.
- Ma, Y., Hao, H., Xie, J., Fu, H., Zhang, J., Yang, J., et al. (2021). ROSE: A Retinal OCT Angiography Vessel Segmentation Dataset and New Model. *IEEE Transactions on Medical Imaging* 40(3), 928-939. doi: 10.1109/TMI.2020.3042802.
- Marchese, A., Agarwal, A., Moretti, A.G., Handa, S., Modorati, G., Querques, G., et al. (2020).
 Advances in imaging of uveitis. *Ther Adv Ophthalmol* 12, 2515841420917781. doi:
 10.1177/2515841420917781.

Mattapallil, M.J., Wawrousek, E.F., Chan, C.-C., Zhao, H., Roychoudhury, J., Ferguson, T.A., et al. (2012). The rd8 mutation of the Crb1 gene is present in vendor lines of C57BL/6N mice and

- embryonic stem cells, and confounds ocular induced mutant phenotypes. *Investigative Ophthalmology & Visual Science* 53(6), 2921-2927. doi: 10.1167/iovs.12-9662.
- Mishra, A., Wong, A., Bizheva, K., and Clausi, D.A. (2009). Intra-retinal layer segmentation in
 optical coherence tomography images. *Opt Express* 17(26), 23719-23728. doi:
 10.1364/OE.17.023719.
- Mitchell, H.B. (2010). *Image fusion: Theories, techniques and applications*. Springer Berlin
 Heidelberg.
- Moccia, S., De Momi, E., El Hadji, S., and Mattos, L.S. (2018). Blood vessel segmentation
 algorithms Review of methods, datasets and evaluation metrics. *Comput Methods Programs Biomed* 158, 71-91. doi: 10.1016/j.cmpb.2018.02.001.
- Mochizuki, M., Kuwabara, T., McAllister, C., Nussenblatt, R.B., and Gery, I. (1985). Adoptive
 transfer of experimental autoimmune uveoretinitis in rats. Immunopathogenic mechanisms
 and histologic features. *Investigative Ophthalmology & Visual Science* 26(1), 1-9.
- Montesano, G., Way, C.M., Ometto, G., Ibrahim, H., Jones, P.R., Carmichael, R., et al. (2018).
 Optimizing OCT acquisition parameters for assessments of vitreous haze for application in uveitis. *Scientific Reports* 8. doi: 10.1038/s41598-018-20092-y.
- Mrejen, S., and Spaide, R.F. (2013). Optical coherence tomography: Imaging of the choroid and
 beyond. *Survey of Ophthalmology* 58(5), 387-429. doi: 10.1016/j.survophthal.2012.12.001.
- Mujat, M., Chan, R., Cense, B., Park, B., Joo, C., Akkin, T., et al. (2005). Retinal nerve fiber layer
 thickness map determined from optical coherence tomography images. *Opt Express* 13(23),
 9480-9491. doi: 10.1364/opex.13.009480.
- Nussenblatt, R.B., Gery, I., and Wacker, W.B. (1980). EXPERIMENTAL AUTOIMMUNE
 UVEITIS CELLULAR IMMUNE RESPONSIVENESS. *Investigative Ophthalmology & Visual Science* 19(6), 686-690.
- Oh, H.-M., Yu, C.-R., Lee, Y., Chan, C.-C., Maminishkis, A., and Egwuagu, C.E. (2011).
 Autoreactive Memory CD4+ T Lymphocytes That Mediate Chronic Uveitis Reside in the
 Bone Marrow through STAT3-Dependent Mechanisms. *The Journal of Immunology* 187(6),
 3338-3346. doi: 10.4049/jimmunol.1004019.
- Paques, M., Guyomard, J.L., Simonutti, M., Roux, M.J., Picaud, S., LeGargasson, J.F., et al. (2007).
 Panretinal, High-Resolution Color Photography of the Mouse Fundus. *Investigative Ophthalmology Visual Science* 48(6), 2769-2774.
- Passaglia, C.L., Arvaneh, T., Greenberg, E., Richards, D., and Madow, B. (2018). Automated
 Method of Grading Vitreous Haze in Patients With Uveitis for Clinical Trials. *Transl Vis Sci Technol* 7(2), 10. doi: 10.1167/tvst.7.2.10.
- Pepple, K.L., Choi, W.J., Wilson, L., Van Gelder, R.N., and Wang, R.K. (2016). Quantitative
 Assessment of Anterior Segment Inflammation in a Rat Model of Uveitis Using SpectralDomain Optical Coherence Tomography. *Invest Ophthalmol Vis Sci* 57(8), 3567-3575. doi:
 10.1167/iovs.16-19276.
- Pepple, K.L., Rotkis, L., Van Grol, J., Wilson, L., Sandt, A., Lam, D.L., et al. (2015). Primed
 Mycobacterial Uveitis (PMU): Histologic and Cytokine Characterization of a Model of
 Uveitis in Rats. *Investigative Ophthalmology & Visual Science* 56(13), 8438-8448. doi:
 10.1167/iovs.15-17523.

- Radtke, A.J., Kandov, E., Lowekamp, B., Speranza, E., Chu, C.J., Gola, A., et al. (2020). IBEX: A
 versatile multiplex optical imaging approach for deep phenotyping and spatial analysis of
 cells in complex tissues. *Proceedings of the National Academy of Sciences* 117(52), 3345533465. doi: 10.1073/pnas.2018488117.
- Raveney, B.J.E., Copland, D.A., Dick, A.D., and Nicholson, L.B. (2009). TNFR1-Dependent
 Regulation of Myeloid Cell Function in Experimental Autoimmune Uveoretinitis. *Journal of Immunology* 183, 2321-2329.
- Ravin, J.G. (1999). Sesquicentennial of the ophthalmoscope. *Arch Ophthalmol* 117(12), 1634-1638.
 doi: 10.1001/archopht.117.12.1634.
- Rodrigues, P., Guimaraes, P., Santos, T., Simao, S., Miranda, T., Serranho, P., et al. (2013). Two dimensional segmentation of the retinal vascular network from optical coherence tomography.
 J Biomed Opt 18(12), 126011. doi: 10.1117/1.JBO.18.12.126011.
- Ruggeri, M., Wehbe, H., Jiao, S., Gregori, G., Jockovich, M.E., Hackam, A., et al. (2007). In vivo
 three-dimensional high-resolution imaging of rodent retina with spectral-domain optical
 coherence tomography. *Invest Ophthalmol Vis Sci* 48(4), 1808-1814. doi: 10.1167/iovs.060815.
- Schneider, C.A., Rasband, W.S., and Eliceiri, K.W. (2012). NIH Image to ImageJ: 25 years of image
 analysis. *Nature Methods* 9(7), 671-675. doi: 10.1038/nmeth.2089.
- Shao, H., Liao, T., Ke, Y., Shi, H., Kaplan, H.J., and Sun, D. (2006). Severe chronic experimental autoimmune uveitis (EAU) of the C57BL/6 mouse induced by adoptive transfer of IRBP1–20-specific T cells. *Experimental Eye Research* 82(2), 323-331. doi: 10.1016/j.exer.2005.07.008.
- Silver, P.B., Horai, R., Chen, J., Jittayasothorn, Y., Chan, C.-C., Villasmil, R., et al. (2015). Retina Specific T Regulatory Cells Bring About Resolution and Maintain Remission of Autoimmune
 Uveitis. *The Journal of Immunology* 194(7), 3011-3019. doi: 10.4049/jimmunol.1402650.
- Smith, R.S., John, S.W.M., Nishina, P.M., and Sundberg, J.P. (2002). "Systematic Evaluation of the
 Mouse Eye: Anatomy, Pathology, and Biomethods". (Boca Raton, FL: CRC Press).
- Spaide, R.F., and Curcio, C.A. (2011). Anatomical correlates to the bands seen in the outer retina by
 optical coherence tomography: literature review and model. *Retina* 31(8), 1609-1619. doi:
 10.1097/IAE.0b013e3182247535.
- Srinivasan, V.J., Ko, T.H., Wojtkowski, M., Carvalho, M., Clermont, A., Bursell, S.E., et al. (2006).
 Noninvasive Volumetric Imaging and Morphometry of the Rodent Retina with High-Speed,
 Ultrahigh-Resolution Optical Coherence Tomography. *Investigative Ophthalmology Visual Science* 47(12), 5522-5528.
- Sun, J., Huang, X., Egwuagu, C., Badr, Y., Dryden, S.C., Fowler, B.T., et al. (2020). Identifying
 Mouse Autoimmune Uveitis from Fundus Photographs Using Deep Learning. *Translational Vision Science & Technology* 9(2), 59-59. doi: 10.1167/tvst.9.2.59.
- Thurau, S.R., Mempel, T.R., Flugel, A., edrichs-Mohring, M., Krombach, F., Kawakami, N., et al.
 (2004). The fate of autoreactive, GFP+ T cells in rat models of uveitis analyzed by intravital
 fluorescence microscopy and FACS. *International Immunology* 16(11), 1573-1582.
- Trusko, B., Thorne, J., Jabs, D., Belfort, R., Dick, A., Gangaputra, S., et al. (2013). The
 Standardization of Uveitis Nomenclature (SUN) Project Development of a Clinical Evidence

- Base Utilizing Informatics Tools and Techniques. *Methods of Information in Medicine* 52(3),
 259-265. doi: 10.3414/me12-01-0063.
- Venhuizen, F.G., van Ginneken, B., Liefers, B., van Grinsven, M., Fauser, S., Hoyng, C., et al.
 (2017). Robust total retina thickness segmentation in optical coherence tomography images
 using convolutional neural networks. *Biomed Opt Express* 8(7), 3292-3316. doi:
 10.1364/BOE.8.003292.
- Wang, R.-X., Yu, C.-R., Dambuza, I.M., Mahdi, R.M., Dolinska, M., Sergeey, Y.V., et al. (2014).
 Interleukin-35 induces regulatory B cells that suppress autoimmune disease. *Nat Med* 20(6),
 633-641. doi: 10.1038/nm.3554.
- Weavers, H., and Martin, P. (2020). The cell biology of inflammation: From common traits to
 remarkable immunological adaptations. *Journal of Cell Biology* 219(7). doi:
 10.1083/jcb.202004003.
- Xu, H.P., Koch, P., Chen, M., Lau, A., Reid, D.M., and Forrester, J.V. (2008). A clinical grading
 system for retinal inflammation in the chronic model of experimental autoimmune
 uveoretinitis using digital fundus images. *Experimental Eye Research* 87(4), 319-326. doi:
 10.1016/j.exer.2008.06.012.
- Yu, C.-R., Kim, S.-H., Mahdi, R.M., and Egwuagu, C.E. (2013). SOCS3 Deletion in T Lymphocytes
 Suppresses Development of Chronic Ocular Inflammation via Upregulation of CTLA-4 and
 Expansion of Regulatory T Cells. *Journal of Immunology* 191(10), 5036-5043. doi:
 10.4049/jimmunol.1301132.
- Zarranz-Ventura, J., Keane, P.A., Sim, D.A., Llorens, V., Tufail, A., Sadda, S.R., et al. (2016).
 Evaluation of Objective Vitritis Grading Method Using Optical Coherence Tomography:
 Influence of Phakic Status and Previous Vitrectomy. *Am J Ophthalmol* 161, 172-180 e171174. doi: 10.1016/j.ajo.2015.10.009.
- Zhong, X., Aredo, B., Ding, Y., Zhang, K., Zhao, C.X., and Ufret-Vincenty, R.L. (2016). Fundus
 Camera-Delivered Light-Induced Retinal Degeneration in Mice With the RPE65 Leu450Met
 Variant is Associated With Oxidative Stress and Apoptosis. *Investigative Ophthalmology & Visual Science* 57(13), 5558-5567. doi: 10.1167/iovs.16-19965.
- Zhu, Q., Xing, X., Zhu, M., Xiao, H., Ma, L., Chen, L., et al. (2019). A New Approach for the
 Segmentation of Three Distinct Retinal Capillary Plexuses Using Optical Coherence
 Tomography Angiography. *Transl Vis Sci Technol* 8(3), 57. doi: 10.1167/tvst.8.3.57.
- Zinkernagel, M.S., Bolinger, B., Krebs, P., Onder, L., Miller, S., and Ludewig, B. (2009).
 Immunopathological Basis of Lymphocytic Choriomeningitis Virus-Induced Chorioretinitis and Keratitis. *Journal of Virology* 83(1), 159-166. doi: 10.1128/jvi.01211-08.
- Zinkernagel, M.S., Chinnery, H.R., Ong, M.L., Petitjean, C., Voigt, V., McLenachan, S., et al.
 (2013). Interferon γ–Dependent Migration of Microglial Cells in the Retina after Systemic
 Cytomegalovirus Infection. *The American Journal of Pathology* 182(3), 875-885. doi:
 10.1016/j.ajpath.2012.11.031.
- Zinkernagel, M.S., Petitjean, C., Wikstrom, M.E., and Degli-Esposti, M.A. (2012). Kinetics of ocular
 and systemic antigen-specific T-cell responses elicited during murine cytomegalovirus
 retinitis. *Immunology and Cell Biology* 90(3), 330-336. doi: 10.1038/icb.2011.43.

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Figure 1.









Figure 5.TIF

