Sequence ontology terminology for gene regulation



David W. Sant, Michael Sinclair, Christopher J. Mungall, Stefan Schulz, Daniel Zerbino, Ruth C. Lovering, Colin Logie, Karen Eilbeck

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Authors: David W Sant, Michael Sinclair, Christopher J Mungall, Stefan Schulz, Daniel Zerbino, Ruth C Lovering, Colin Logie, Karen Eilbeck

Addresses

DWS: Department of biomedical informatics, University of Utah, Salt Lake City, Utah, USA; Department of Biomedical Sciences, Noorda College of Osteopathic Medicine, Provo, Utah, USA. ORCID: 0000-0001-7372-9896. Email: <u>david.sant@utah.edu</u> MS: Department of biomedical informatics, University of Utr.h, Salt Lake City, Utah, USA. ORCID: 0000-0002-3636-5920. Email: <u>single wornail.com</u> CJM: Environmental Genomics and Systems Biolegy, Lawrence Berkeley National Laboratory: Berkeley, CA, US. ORCID: 0000-100.:-6601-2165. Email <u>cimungall@lbl.gov</u> SS: Institute for Medical Informatics, Str.uetice and Documentation, Medical University of Graz, Austria. ORCID: 0000-0001-722.:-3287 Email: <u>stefan.schulz@medunigraz.at</u> DZ: European Molecular Biology Laboratory, European Bioinformatics Institute, Hinxton, Cambridge, UK, ORCID: 0000-0001-5350-3056. Email: <u>zerbino@ebi.ac.uk</u> RCL: Functional Gene Annotation, Preclinical and Fundamental Science, UCL Institute of Cardiovascular Science, conversity College London, London, UK. ORCID: 0000-0002-9791-0064, email: <u>informatica.uk</u>

CL: Radboud Institute for Molecular Life Sciences, Geert Grooteplein Zuid 28, 6525 GA Nijmegen, Netherlands. OrcID 0000-0002-8534-6582. Email <u>C.Logie@ncmls.ru.nl</u> KE: Department of biomedical informatics, University of Utah, Salt Lake City, Utah, USA. ORCID:0000-0002-0831-6427, email <u>keilbeck@genetics.utah.edu</u>

Abstract

The Sequence Ontology (SO) is a structured, controlled vocabulary that provides terms and definitions for genomic annotation. The Gene Regulation Ensemble Effort for the Knowledge Commons (GREEKC) initiative has gathered input from many groups of researchers, including the SO, the Gene Ontology (GO), and gene regulation experts,

with the goal of curating information about how gene expression is regulated at the molecular level. Here we discuss recent updates to the SO reflecting current knowledge. We have developed more accurate human-readable terms (also known as classes), including new definitions, and relationships related to the expression of genes. New findings continue to give us insight into the biology of gene regulation, including the order of events, and participants in those events. These updates to the SO support logical reasoning with the current understanding of gene expression regulation at the molecular level.

Introduction

With the rapid increase in genomic sequencing across a multitude of species came the need to automate the annotation of genetically encoded sequences. Defining the parts of a genome was key to the unification of the description of genomes across species. To address this issue, the Sequence Ontology was created by the Gene Ontology Consortium to be a structure controlled vocabulary for the definition of biological sequence features^{1,2}. The CO is one of the original members of the OBO Foundry³ (<u>http://www.obofoundry.org/</u> and is interoperable with other ontologies such as the Gene Ontology⁴ (GO) and Colemical Entities of Biological Interest (CHEBI)⁵. Terms (also known as classes in OWL) and relationships between them are added or updated as the team members become aware of new understanding or findings in the field.

The terminology and definitions in the field of gene regulation are intricate. The Sequence Ontology covers the technical language elements necessary to denote the genomic regions involved in regulatory processes. Other aspects of gene regulation are covered by other ontologies such as the GO. The Gene Regulation Ensemble Effort for the Knowledge Commons (GREEKC) initiative was established in 2016 as the European branch of the Gene Regulation Consortium (GRECO) to enable multiple groups of researchers to determine how to accurately represent knowledge of the regulation of gene expression at the molecular level using several ontologies (http://www.greekc.org/, http://thegreco.org). The collaborative nature of this project has

allowed for terms across databases to be updated concurrently, ensuring the interoperability of the ontologies. In this manuscript, we report updates related to gene regulation that have been made to SO as part of GREEKC. When discussing ontology terms in this manuscript, we will italicize and space with underscores to differentiate from the discussion of biological entities.

The Scope of Sequence Ontology

SO was initially developed by the Gene Ontology Consortium, although the scope of SO differed significantly from the Gene Ontology (GC). While GO describes the outward face of gene products - what they do and where they do it, the SO defines the internal parts of genes and genomes. Genomic ann stations and gene models are the parts of genomes demarcated in coordinate space (e.g.; chromosome: start-end). They define where the exons, introns, and Transcription Start Sites (TSS) etc. begin and end. Representing the coordinates of fealures themselves is outside of SO's scope, but has been accomplished by other groups such as FALDO and Biolink^{6,7}. Development of SO terminology is a result of curators' needs to adequately define the parts of their genomic annotations. The updates that have occurred concurrently with the GREEKC initiative are related to nucleotide sequences that are important for different aspects of gene regulation, not the proteins that produce actions at a molecular level. Here we describe the resultant terminology for describing the genomic features involved in gene regulation.

The Understanding *Cis*-Regulatory Modules (CRM)

The correct spatial and temporal expression of genes is required for multicellular organisms to develop properly and maintain the different necessary cell types⁸. Many DNA elements act in tandem to regulate the expression of a specific gene or a set of genes, and these DNA elements are typically clustered into regions commonly referred to as *cis*-regulatory modules (CRMs). The SO definition for *CRM* (SO:0000727) is "A regulatory region where transcription factor binding sites are clustered to regulate various aspects of transcription activities. (CRMs can be located a few kilobases (kb) to hundreds of kb upstream of the basal promoter, in the coding sequence, within introns,

or in the untranslated regions (UTR) sequences, and even on a different chromosome). A single gene can be regulated by multiple CRMs to give precise control of its spatial and temporal expression. CRMs function as nodes in a large, intertwined regulatory network." In short, a CRM is a region of DNA that contains multiple elements that regulate the expression of genes.

While CRMs contain multiple regulatory elements such as transcription factor binding sites, different CRMs have different functions for the regulation of transcription. These different CRMs include enhancers, silencers, locus control regions, and insulators (see Figure). Enhancers are CRMs that activate the expression of their target regardless of orientation and may be distant from the promoter region. Silencers are essentially the opposite of enhancers and function to suppress transcription. Insulators are CRMs that function to prevent another CRM from interacting with the promoter of a nearby gene when the insulator is located between two CRMs. Locus control regions are open chromatin (DNAse hypersensitive) regions of DNA that confer high-level, copy number dependent expression of a gene⁹ A CRM term that has recently been added to SO is *DNA_loop_anchor*, representing the ends of a DNA looping region. This DNA looping allows for areas of DNA that are very distant to remain in close proximity within the cell, allowing for CRMs to interact with distant genes¹⁰.

As noted in the *CRM* definition, a single gene may be regulated by multiple CRMs and the regulation of ex_{μ} ression of a single gene can be very complex. For example, a single gene may be active only when an enhancer region is active, which in turn inactivates a silencer and activates the promoter region of the gene^{11,12}.

While databasch of genes and proteins have been around for decades, the annotation of CRMs in most species aside from yeast and some particular bacteria has lagged, largely due to the difficulty of detecting them¹³⁻¹⁶. Just as gene expression is variable across cell types and conditions, CRMs may be active only in certain cell types and conditions. This would indicate that detecting all CRMs would require analysis using all cell types. Advancements in sequencing technologies have aided greatly in the detection of CRMs, especially the use of chromatin immunoprecipitation sequencing (ChIP-seq) to detect specific chromatin marks or the binding of specific proteins to DNA. For example, active enhancers are detected by the presence of histone 3 lysine 4

mono-methylation (H3K4me1) and acetylated lysine 27 of histone 3 (H3K27ac), while poised enhancers are repressed by trimethylated lysine at position 9 of histone 3 (H3K9me3) and/or trimethylated lysine at position 27 of histone 3 (H3K27me3)¹⁷. Silencers are typically marked by the binding of polycomb repressive complex 1 or 2 (PRC1/2) and H3K27me3¹⁸. It should be noted that many high-throughput experiments like ChIP-seq provide a starting point for understanding gene regulation, in this case with elucidation of transcription factor binding sites, additional experiments are required to prove the role in the regulation of a gene. The annotation of such genomic features must take into account the level of evidence that supports the role, such as predicted versus validated. This level of belief is not currently articulated in the ontology and therefore should be expressed in the annotation.

A promoter, like a CRM, is a *transcriptional_cic_regulatory_region* (see Figure). Some promoters are characterized by their expression pattern. Constitutive promoters are promoters that have continual transcription. In ducible promoters are those that can be induced for transcription by the presence of a factor. Cryptic promoters are promoters to a cryptic gene, which is a gene that is not transcribed under normal conditions and is not critical to normal cellular function. Bidirectional promoters are promoters that can initiate transcription in either direction¹⁹.

Promoters for DNA template-dependent RNA polymerases have somewhat different structures within different types of organisms. Eukaryotic promoters include a TSS and serve as a region ic: the assembly of a pre-initiation complex (PIC), which is necessary for transcription of the gene. Prokaryotic promoters are regions of binding of a specific RNA polymerase (RNA pol) holoenzyme, which may lead to the transcription of multiple genes²⁰. Prokaryotic promoters include bacterial RNA promoters. Viral promoters include Phage RNA polymerase promoters and they contain the TSS of the gene and will be bound by host machinery that varies with the host species. Eukaryotic promoters include RNA pol I, II, and III promoters, and in plants RNA pol IV and V promoters.

We recently introduced a new term *core_promoter_element* (SO:0002309), defined as "An element that exists within the promoter region of a gene. When multiple transcripts exist for a gene, the separate transcripts may have separate

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core_promoter_elements." The term *core_promoter_element* is further subdivided into subclasses *core_eukaryotic_promoter_element*, *core_prokaryotic_promoter_element* and *core_viral_promoter_element*. In SO the components of a *core_eukaryotic_promoter* are elements that are found within the promoter region of a eukaryotic gene, which indicates that they may be present in RNA pol I, II, or III promoters. These elements include well-known elements such as the *TATA_box* (SO:0000174) and *discontinuous_core_element* (DCE, SO:0001664).

While the parts of bacterial promoters have been described as child terms to bacterial_RNApol_promoter_sigma54_element (minus_12_signal and minus_24_signal) and bacterial_RNApol_promoter_sigma_70_element (minus_35_signal and minus_10_signal), recent work now suggests that these mc ifs are not considered an essential component of bacterial promoters²⁰. This is nue to recent changes in understanding about bacterial gene regulation where'ry these motifs are not necessary in some instances and not sufficient in other instances to promote transcription. Therefore, the term core_prokarytotic_promuter_element currently does not contain motif sequence parts.

In summary, the restructuring of terms under the CRM branch in SO has allowed for a more accurate structuring of terms related to CRM. In particular, the term *core_promoter_element* has been created and the component parts of this region can now be annotated with specific core promoter element SO terms that include general transcription initiation factor binding sites that are distinct from sequence-specific DNA binding transcription inclure binding sites (Gaudet *et al.* in preparation)²¹.

Topologically Defined Regions (TDRs) and Topologically Associated Domains (TADs)

In order for DNA elements to be active and contribute toward gene transcription, the elements must be in regions of open euchromatin²². Some CRMs, such as enhancers, regulate genes located over 100 kb away. An open stretch of DNA 100 kb in length would account for more than 30 (M of distance¹⁰. If all euchromatin existed as free-flowing DNA, it would be highly unlikely that the enhancer region would ever interact with the promoter region of a gene to increase transcription. This is why DNA

within the cell remains in chromatin loops. The ends of the loops are held in close proximity, promoting physical interaction of the elements on either end of the loop. Furthermore, all the DNA within such chromatin loops appears to self-associate efficiently²³. The entire region between these interacting ends has therefore been called a *topologically_associated_domain* (TADs, SO:0002304). Several technologies have emerged over recent years to allow for the identification of TADs, including chromatin conformation capture (3C) and related technologies 4-C, 5-C, GCC, Hi-C, ChIA-PET and GAM²⁴. An area where self-interaction occurs more frequently than expected by chance is known as a *topologically_defined_region* (TDRs, SC:0001412).

TADs are flanked by a *topologically_associated_do.maii_boundary* (TAD boundary) (SO:0002305) on both sides. The DNA inside a `AD can form a *DNA_loop* (SO:0002307). The term *DNA_loop* refers to the phonomenon of loop formation of DNA. Importantly, loops are molecular *conformations*, and as such they are continuants, i.e. static entities, opposed to intrinsically related or ping *processes* displayed by a chromosome in a cell at one specific time are occurrents, since processes are always occurrents (https://en.wikipedia.org/wiki/Pasic_Formal_Ontology)²⁵. The TAD and the TAD boundary are continuants that respectively refer to the regions of DNA that self-associate frequently and to the region across which chromatin loops occur infrequently. A DNA loop ancher will usually occur at the TAD boundary where the ends of the loop are held in close previmity, but a majority of loop anchors actually reside inside TADs¹⁰. During interplase, the DNA loop anchors are CCCTC-binding factor (CTCF) binding sites. Surgeral studies have investigated the binding of CTCF in different tissues to determine the endpoints of DNA loops and help decipher TADs¹⁰.

While the concept of regions of self-interaction of DNA for gene regulation has been established for some time^{23,26-29}, these new updates to SO allow for an accurate representation of the current understanding of TADs and the related concepts of TAD boundary and insulator elements. This new terminology enables these regions to be annotated in databases and knowledgebases whereby the precise biological condition and cell type can be captured. The hierarchical structure of TAD-related SO terms and their relationship to CRM is shown in the Figure.

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Discussion

The GREEKC initiative has provided scientists and creators of biological ontologies an opportunity to collectively discuss how to accurately represent terms and relationships pertaining to gene regulation. Many terms have been either added to SO or updated in SO to allow it to better represent the current understanding of gene regulation. Specifically, several terms have been updated under the branch of *cis*-regulatory module (*CRM*). A new term, *core_promoter_element*, has been created to annotate elements that exist within the promoter region of a gene. The term *topologically_associated_domain* (TAD) has been added along with terms describing parts of TADs. Since insulators harbor CTCF sites and since C TCF sites form loop anchors, the addition of the TAD boundary term should allo v different data types and analysis approaches that focus on either chromosome is poping, enhancer insulation or topological segregation to be accurately annotated in an experimental entity-oriented fashion so as to permit objective discovery of engenetic patterns and mechanisms of gene regulation in humans and other eut crystes for biomedical research in particular.

These updates to SO have been conducted in parallel with updates to other biological ontologies, including the Gone Ontology (Gaudet *et al.* in preparation). The concurrent updates have allowed the untologies to be interoperable, which will allow for the most accurate representation or complex concepts. For example, these updates to SO can already and will soon on used by reference annotations such as the Ensembl Regulatory Build³⁰, which accurates CRMs across genomes based on available public epigenomic data. Altrough already using SO, this new annotation will thus express more precisely the nature of the elements. These CRMs can be associated with transcription factor binding events (e.g. through motif analysis or ChIP-Seq), and therefore to upstream genes. In future, using cis-regulatory evidence (e.g. eQTLs or Hi-C), these CRMs will further be attached to their downstream target genes. Therefore, these sequence elements will constitute links between GO annotations.

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References

- K. Eilbeck, S.E. Lewis, C.J. Mungall, M. Yandell, L. Stein, R. Durbin, M. Ashburner. The Sequence Ontology: a tool for the unification of genome annotations. *Genome Biol.* 2005;6(5):R44.
- 2. C.J. Mungall, C. Batchelor, K. Eilbeck. Evolution of the Sequence Ontology terms and relationships. *J Biomed Inform* 2011;44(1):87-93.
- B. Smith, M. Asnburner, C. Rosso J. Bard, W. Bug. *et al.* The OBO Foundry: coordinated evolution of ontologies to support biomedical data integration. *Nat Biotechnol.* 2007;25(11):125¹-¹ 255.
- 4. Gene Ontology Consort: m. Creating the gene ontology resource: design and implementation. *Ger. and Res.* 2001;11(8):1425-1433.
- K. Degtyarenko K, P. de Matos, M. Ennis, J. Hastings, M. Zbinden, A. McNaught, R. Alcantara, M. Dorsow, M. Guedj, M. Ashburner. ChEBI: a database and ontology for charaical entities of biological interest. *Nucleic Acids Res.* 2008;36:D344-350.
- J.T. Bolleman, C.J. Mungall, F. Strozzi, J. Baran, M. Dumontier, R.J.P. Bonnal, R. Buels, R. Hoehndorf, T. Fujisawa, T. Katayama, P.J.A. Cook. FALDO: a semantic standard for describing the location of nucleotide and protein feature annotation. *J Biomed Semantics*. 2016;7:39.
- K. Verspoor, H. Shatkay, L. Hirschman, C. Blanschke, A. Valencia. Summary of the BioLINK SIG 2013 meeting at ISMB/ECCB 2013. *Bioinformatics*. 2013;31(2):297-298.

- D.M. Jeziorska, K.W. Jordan, K.W. Vance. A systems biology approach to understanding cis-regulatory module function. *Semin Cell Dev Biol.* 2009;20(7):856-862.
- Q. Li, K.R. Peterson, X. Fang, G. Stamatoyannopoulos. Locus control regions. Blood. 2002;100(9):3077-3086.
- 10. L. Nanni, S. Ceri, C. Logie. Spatial patterns of CTCF sites define the anatomy of TADs and their boundaries. *Genome Biol.* 2020;21(1):197.
- 11. L.T. Huong, M. Kobayashi, M. Nakata, G. Shioi, H. Miyachi, T. Honjo, H. Nagaoka. In vivo analysis of Aicda gene regulations: a critical balance between upstream enhancers and intronic silencers governs op opriate expression. *PLoS One*. 2013;8(4):e61433.
- 12. P. Kolovos, T.A. Knoch, F.G. Grosveld, P.R. Could, A. Papantonis. Enhancers and silencers: an integrated and simple model for their function. *Epigenetics Chromatin*. 2012;5:1.
- 13.S. Lisser and H. Margalit. Compilation of E. coli mRNA promoter sequences. Nucleic Acids Res. 1993;21(7):1307-1516.
- 14. R. Hershberg, G, Bejerano, A. Santos-Zavaleta, H. Margalit. PromEC: An updated database of Escherich a coli mRNA promoters with experimentally identified transcriptional start sites. *Nucleic Acids Res.* 2001;29(1):277.
- 15. J. Zhu and M.Q. Zhang. SCPD: a promoter database of the yeast Saccharomyces corectisiae. *Bioinformatics*. 1999;15(7-8):607-611.
- 16. H. Li, J. Hou, L Eci, C. Hu, P. Tong, Y. Kang, X. Zhao, Z. Shao. Genmoe-wide analysis of corc promoter structures in Schizosaccharomyuces pombe with DeepCAGE. RNA Biol. 2015;12(5):525-537.
- G.E. Zenter, P.J. Tesar, P.C. Scacheri. Epigenetic signatures distinguish multiple classes of enhancers with distinct cellular function. *Genome Res.* 2011;21(8):1273-1283.
- A. Laugesen, J.W. Hojfeldt, K. Helin. Role of the Polycomb Repressive Complex
 2 (PRC2) in Transcriptional Regulation and Cancer. *Cold Spring Harb Perspect. Med.* 2016;6(9):a026575.

- 19. W. Wei, V. Pelechano, A.I. Jarvelin, L.M. Steinmetz. Functional consequences of bidirectional promoters. *Trends Genet.* 2011;27(7):267-276.
- 20.C. Mejia-Almonte, S.J.W. Busby, J.T. Wade, J. van Helden, A.P. Arkin, G.D. Stormo, K. Eilbeck, B.O. Palsson, J.E. Galagan, J. Collado-Vides. Redefining fundamental concepts of transcription initiation in bacteria. *Nat Rev Genet.* 2020.
- 21. R. Lovering, P. Gaudet, M.L. Acencio, A. Ignatchenko, A. Jolma, O. Fornes, M. Kuiper, I.V. Kulakovskiy, A. Lægrid, M.J. Martin, C. Logie. A GO catalogue of human DNA-binding transcription factors. *bioRxiv* 2020; doi:10.1101/2020.10.28.359232.
- 22. M. Falk, Y. Feodorova, N. Naumova, M. Imakaev, B.R. Lajoie, H. Leonhardt, B. Joffe, J. Dekker, G. Fudenberg, I. Solovei, L.A. Nirm. Heterochromatin drives compartmentalization of inverted and conventional nuclei. *Nature*. 2019;570(7761):395-399.
- 23. J.R. Dixon, S Selvaraj, F. Yue, A. Kim, Y Li, Y. Shen, *et al.* Topological domains in mammalian genomes identified by analysis of chromatin interactions. *Nature*. 2012;485:376–80.
- 24. R.A. Beagrie, A. Scialdone, M. Schueler, D.C.A. Kraemer, M. Chotalia, *et al.* Complex multi-enhancer ccn².acus captured by Genome Architecture Mapping (GAM). *Nature*. 2017;54 3(7646):519-524.
- 25. A. Galton. On generically dependent entities. *Applied Ontology*. 2014; 9(2):129-153.
- 26. E.P. Nora, B.K. Lejole, E.G. Schulz, L. Giorgetti, I Okamoto, *et al.* Spatial partitioning of the regulatory landscape of the X-inactivation centre. *Nature*. 2012;485(7398):381-385.
- 27. E. Alipour E, J.F. Marko. Self-organization of domain structures by DNA-loopextruding enzymes. *Nucleic Acids Res.* 2012;40:11202–12.
- 28.A.L. Sanborn, S.S.P. Rao, S.C. Huang, N.C. Durand, M.H. Huntley, A.I. Jewett, et al. Chromatin extrusion explains key features of loop and domain formation in wild-type and engineered genomes. *Proc Natl Acad Sci USA*. 2015;112:E6456– 65.

- 29. K. Nasmyth. Disseminating the Genome: Joining, Resolving, and Separating Sister Chromatids During Mitosis and Meiosis. *Annu Rev Genet.* 2001;35:673– 745.
- 30. D.R. Zerbino, N. Johnson, T. Juetteman, D. Sheppard, S.P Wilder, *et al.* Ensembl regulation resources. *Database (Oxford)* 2016;bav119.

Figure.

Dendrogram showing the relationships between SO terms related to gene regulation discussed in this manuscript. Black arrows represent 'is_a' elationships, red arrows represent 'part_of' relationships and green arrows represent 'overlaps' relationships.

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Author Statement

David W Sant: Writing – original draft, data curation, visualization, software. Michael Sinclair: Data curation Christopher J Mungall: Writing – review and editing, data curation, software. Stefan Schulz: Writing – review and editing, data curation. Daniel Zerbino: Writing – review and editing, data curation. Ruth C Lovering: Writing – review and editing, data curation. Colin Logie: Writing – original draft, data curation, visualization. Karen Eilbeck: Writing – original draft, supervision, resources, funding acquisition.

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Conflict of Interest

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The authors of this manuscript declare that they have no competing interests related to this manuscript.

Sincerely,

David W. Sant, Ph.D. Postdoctoral Fellow University of Utah Department of Biomedical Informatics Salt Lake City, UT 84108

Highlights

Sequence Ontology has updated terminology related gene expression for GREEKC Project.

Cis-regulatory modules contain multiple elements that regulate gene expression. Locus control regions confer high-level, copy number dependent expression of a gene. Topologically associated domains promote physical proximity to regulate transcription.

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