

1 **Additively-manufactured PEEK/HA porous scaffolds with highly-**
2 **controllable mechanical properties and excellent osteogenesis for bone**
3 **tissue repairing**

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28 Abstract

29 Polyetheretherketone (PEEK) was widely applied into fabricating of orthopedic implants,
30 benefitting its excellent biocompatibility and similar mechanical properties to native bones.
31 However, the inertness of PEEK hinders its integration with the surrounding bone tissue. Here
32 PEEK scaffolds with a series of hydroxyapatite (HA) contents in gradient were manufactured via
33 fused filament fabrication (FFF) 3D printing techniques. The influence of the pore size, HA content
34 and printing direction on the mechanical properties of the PEEK/HA scaffolds was systematically
35 evaluated. By adjusting the pore size and HA contents, the elastic modulus of the PEEK/HA
36 scaffolds can be widely tuned in the range of 624.7-50.6 MPa, similar to the variation range of
37 natural cancellous bone. Meanwhile, the scaffolds exhibited higher Young's modulus and lower
38 compressive strength along Z printing direction. The mapping relationship among geometric
39 parameters, HA content, printing direction and mechanical properties was established, which gave
40 more accurate predictions and controllability of the modulus and strength of scaffolds. The
41 PEEK/HA scaffolds with the micro-structured surface could promote cell attachment and
42 mineralization *in vitro*. Therefore, the FFF-printed PEEK/HA composites scaffolds can be a good
43 candidate for bone grafting and tissue engineering.

44 **Keywords:** Polyether-ether-ketone (PEEK); Scaffold; Composites; Fused filament fabrication
45 (FFF); Mechanical properties

46 1. Introduction

47 Porous scaffold has been widely used in tissue engineering to provide appropriate environment
48 and architecture for the regeneration of tissue^[1-3]. Compared with conventional preparation
49 techniques of scaffold, 3D printing techniques exhibit attractive attention for constructing
50 controllable macro/microarchitectures^[4, 5], such as the porosity, the pore size, the pore shape and
51 the interconnected network^[6]. Thus, 3D printing techniques simplifies the fabrication process and
52 provides more effective manufacturing method for the porous scaffolds. To date, the 3D printing
53 metallic porous scaffold made from nickel^[7], cobalt-chromium^[8], titanium alloys^[9, 10] and stainless
54 steel^[11] have been widely developed and applied in medicine fields. But the mechanical properties
55 of the metallic scaffolds are quite different from those of natural bone, which lead to stress shielding
56 and prosthetic loosening in the long term.

57 In recent years, high molecular polymers such as polylactic acid^[12, 13], polycaprolactone^[14] and
58 polyetheretherketone (PEEK) were widely used in the tissue engineering for their excellent
59 mechanical properties and biocompatibility. PEEK has been applied into fabricating porous scaffold
60 structures. Su^[15] et al. fabricated PEEK scaffolds by fused filament fabrication (FFF) process and
61 *in vivo* experiments demonstrated that the newly-regenerated soft tissues could grow into the
62 scaffold quickly. However, the soft tissue cannot adhere closely to the surface of the PEEK scaffolds
63 ascribing to the biological inertness. Ahn^[16] et al. fabricated PEEK scaffold by salt leaching method
64 and the animal experiment results showed that the pure PEEK exhibited lower osteogenesis ability
65 and interfacial strength. Thus, biological inertness and poor osteogenic capability restrict the clinical
66 application of PEEK. Numerous studies were conducted to enhance the bioactivity of PEEK via the
67 incorporation of additives such as hydroxyapatite (HA), calcium phosphate (TCP), bioglass (BGA).

68 Zhao^[17] et al. prepared HA/PEEK composite by mixing hydroxyapatite and PEEK, finding that the
69 composite promote osteoblastic cell adhesion and expression of bone-building protein. Rui Ma^[18]
70 et al. synthesized PEEK/CS composites, of which the hydrophilicity, alkaline phosphatase activity,
71 and the mineralized nodule formation of composite were dramatically improved. But the study in
72 3D printing PEEK-based composite porous scaffolds was rare because of the complicated
73 fabrication procedure.

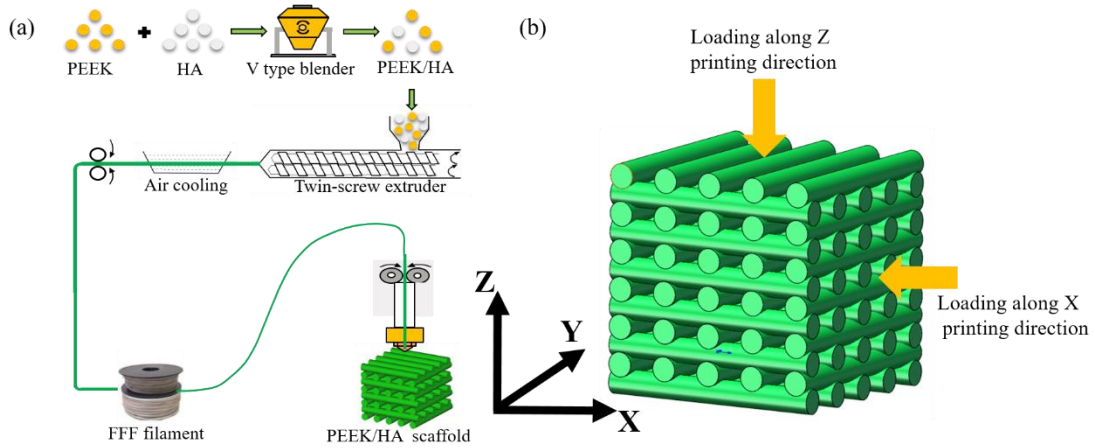
74 In addition to the bioactivity, the mechanical properties of the scaffolds significantly affect the
75 load transmitting and tissue growth^[19-22]. The micro-structure of the scaffolds such as pore
76 architecture^[23, 24], pore size^[25] and porosity^[26, 27], impacted the mechanical performances of the
77 scaffold greatly. Liang^[28] et al. fabricated square, hexagonal and wheel-shaped porous scaffolds by
78 3D printing, finding that the square scaffolds exhibited better mechanical strength and fatigue
79 properties. Melchels^[29] et al. fabricated PLA scaffolds and found that effective modulus of the
80 scaffolds with cube architecture was twice larger than those of gyroid architecture (324 ± 39 MPa vs
81 169 ± 21 MPa). Bagheri^[30] et al. controlled the Young's modulus of the PLA scaffolds in the range
82 of 250-700 MPa by changing the width of the struts. Thus the mechanical properties of scaffold
83 could be controlled to a certain extent by changing the micro-architectures. However, the influences
84 of the printing direction and the material components on the mechanical properties of the PEEK-
85 based scaffold is still unclear.

86 In this study, PEEK/HA composites scaffolds with different pore size and HA content were
87 fabricated via FFF 3D printing technology. The surface microstructure, surface roughness and
88 internal architectures of PEEK/HA porous scaffolds were measured. The thermal analysis
89 measurements of the PEEK composites were carried out to evaluate the crystalline and thermal
90 behaviors. The influences of the micro-structure, HA content and printing direction on the
91 mechanical properties of the PEEK/HA scaffolds was systematically investigated. Meanwhile, high-
92 speed camera was used to record the compressive deformation process of the composite scaffolds
93 to analyze the deformation mechanism. Finally, Cell culture experiments were performed to
94 investigate the biocompatibility and mineralization *in vitro*.

95 **2. Materials and methods**

96 **2.1 Fabrication of PEEK/HA composite scaffolds**

97 The fabrication schematic of the scaffolds via FFF 3D printing process was shown as Figure 1.
98 PEEK (50 μm) and HA (15 μm) powder were mixed with mass ratio of 8:2 and 6:4. Then the mixing
99 composites was fed into a twin-screw extruder, from which they were extruded into filaments with
100 a diameter of 1.75 mm. Tetragonal scaffold samples (length, width and thickness of 10 mm) were
101 fabricated by based FFF process. The basic printing parameters were set as follows: nozzle
102 temperature of 420 °C, layer thickness of 0.2mm, bead width of 0.4 mm and printing speed of 30
103 mm/s.



104
 105 Figure 1. Schematic illustration of the 3D printing process and mechanical properties testing method:
 106 (a) preparation of PEEK/HA composites and FFF printing (b) scaffold compressed along Z or X
 107 printing direction.

108 2.2 Characteristics of the composite scaffolds

109 The structures of the composites scaffolds were measured using a micro computed tomography
 110 (Micro-CT, Y.Cheetah, YXLON, Germany) and reconstructed by the software of VG Studio Max
 111 3.0. The surface morphologies of the scaffolds were characterized via scanning electron microscopy
 112 (SEM, su-8010, Hitachi, Japan) at an acceleration voltage of 5 kV after coated with a thin layer of
 113 Au. PEEK/HA scaffolds were observed on a laser scanning confocal microscope (LSCM, OLS4000,
 114 Olympus, Japan) to evaluate the surface roughness (R_a).

115 2.3 Thermal behaviors of the composites

116 Changes in chemical bonds was measured by Fourier Transform Infrared (FTIR, Nicolet iS10,
 117 Thermo Fisher, U.S.A.) spectrometry. The thermal analysis measurements were carried out by
 118 differential scanning calorimetry (DSC, DSC1, Mettler Toledo, CH). Samples were heated to 400 °C
 119 at a heating rate of 10 °C/min. The samples were all cut from the core of the scaffolds for avoiding
 120 any additional variation due to differences in surface and core crystalline structure. The crystallinity
 121 of the composites could be calculated using the function expressed in Equation (1):

$$122 \quad X_c = \frac{\Delta H_m - \Delta H_c}{\Delta H_f \times W_{PEEK}} \quad (1)$$

123 Where X_c , ΔH_m and ΔH_c refer to the crystallinity, melting enthalpy and crystal enthalpy of
 124 PEEK-based composites, respectively. ΔH_f is the enthalpy of fusion of a 100% crystalline PEEK
 125 sample (130 J/g), W_{PEEK} is the mass fraction of PEEK in the composites. Relationship between
 126 mass and temperature was determined by thermal gravimetric analysis(TGA). PEEK/HA samples
 127 were heated to 800 °C at a ramp rate of 10 °C/min by TGA (DSC3, Mettler Toledo, CH).

128 2.4 Mechanical testing

129 To assess the influence of HA content, pore size and printing direction on the mechanical

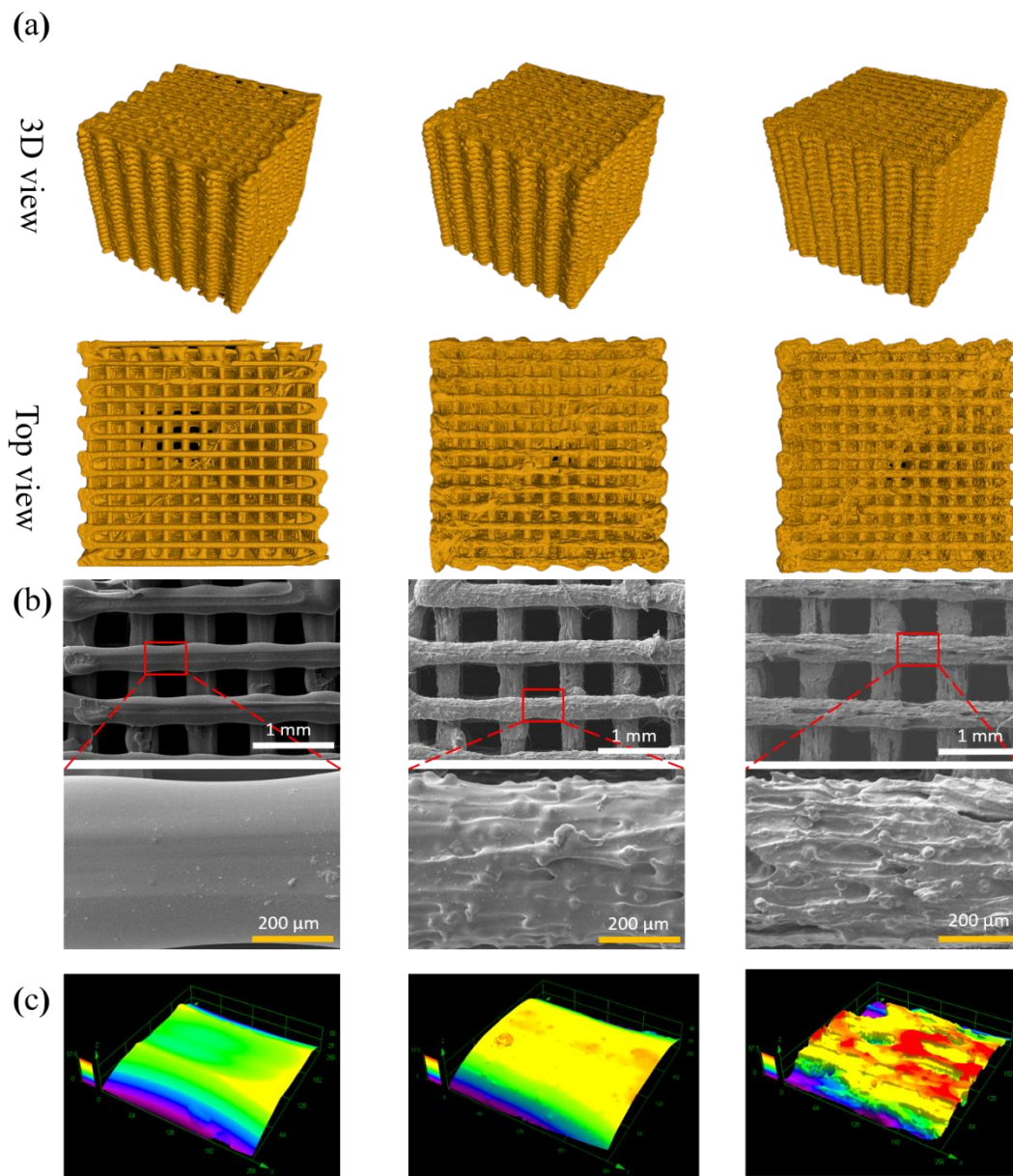
130 properties of the composite scaffolds, PEEK/HA scaffolds were compressed parallel (X printing
131 direction) and perpendicular (Z printing direction) to the printing substrate direction (Figure.1-b) at
132 a compression speed of 3 mm/min. The fabricated PEEK based scaffolds were named as HA
133 content-compressive direction-pore size. For instance, 40%HA-Z-08 means that the HA content was
134 40 wt.% and the compressive direction was Z printing direction and a pore size of 0.8 mm. The
135 stress-strain curves were obtained from the load-displacement data to calculate the Young's modulus
136 and the strength. Meanwhile, a high-speed camera was used to record the compressive responses of
137 the composite scaffolds to analyze the deformation mechanism.

138 **2.5 Cell compatibility of 3D-printed PEEK/HA scaffolds**

139 The scaffolds with a diameter of 14 mm were put into 24-well plates. Then, preosteoblast cells
140 (MC3T3-E1, ATCC, Manassas, VA, USA) were seeded onto the scaffolds at a density of 2×10^4 cells
141 per well and cultured in α -MEM (HyClone, USA). The MC3T3-E1 cells were proliferated for 1, 3,
142 and 5 days, at which time incubated in the CCK-8 solution. The optical density was measured at λ
143 = 450 nm using a microplate reader (RT-6000) to qualitatively evaluate the proliferation. The
144 scaffolds were taken into live/dead staining solution and observed by fluorescence microscope
145 (Olympus Co. Ltd.) to evaluate the cell viability. The cells were fixed with 4% paraformaldehyde
146 solution after 5 days culturing and the cell morphologies were observed using SEM. Alizarin Red S
147 (ARS) staining was used to visualize mineralization ability. MC3T3-E1 were cultured on the disc
148 with the diameter of 14mm at a density of 1×10^5 cells. After incubation with the specimens for 24
149 hours, the culture medium was changed to the osteogenic inductive medium for 21days induction.
150 The stained samples were immersed in DMSO to measure at $\lambda = 550$ nm using a plate reader (RT-
151 6000).

152 **3. Results**

153 **3.1 Micro-structure characteristics of the PEEK/HA scaffold**



154

155 Figure 2. Micro-structure characteristics of the PEEK/HA scaffold samples: (a)Geometry
156 observation by Micro-CT (b) surface morphology by SEM and (c) three-dimensional topography
157 by LSCM. The left column, the middle column and right column represent the pure PEEK, 20%HA
158 and 40%HA scaffolds, respectively. The pore size of three scaffolds is 400μm.

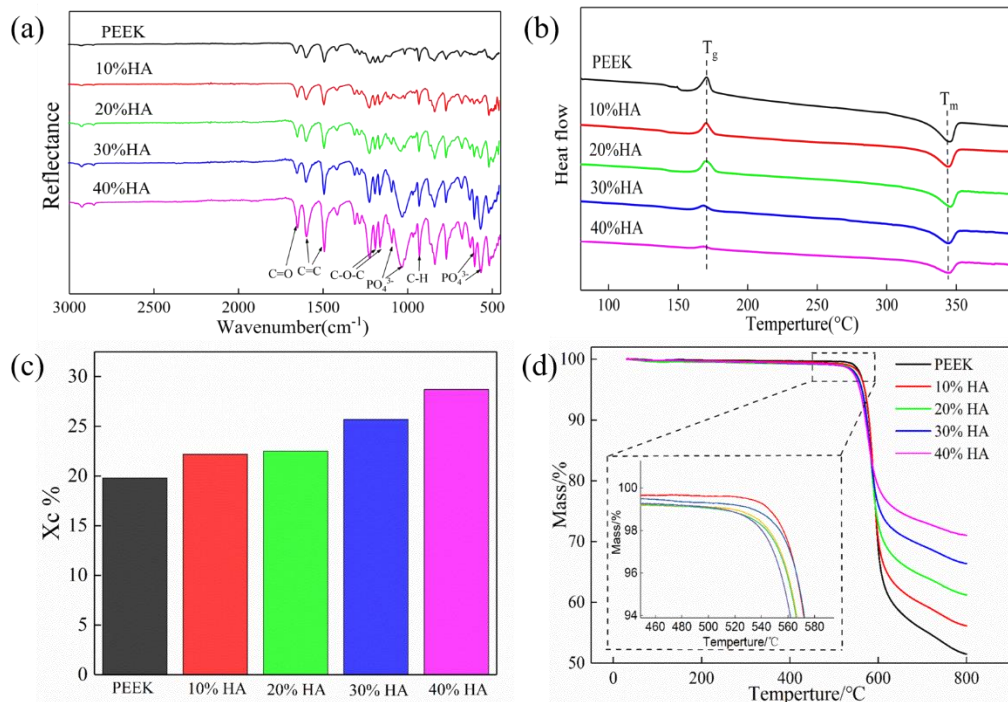
159 Micro-CT was employed to observe the interconnectivity of the scaffolds as shown in Figure
160 2-a. It could be seen that no obvious defect on the 3D and top views and the PEEK/HA composite
161 scaffolds exhibited a well interconnectivity. The Figure 2-b showed that the pure PEEK scaffolds
162 exhibited smooth surface, while numerous HA particles and a coarser morphology were observed
163 on the surface of PEEK/HA composites scaffolds. The LSCM results showed that a rougher

164 morphology consisting of peaks and valleys for PEEK/HA composites scaffolds (Figure 2-c),
 165 confirming the SEM findings. The pure PEEK scaffold featured a Ra of $0.154 \pm 0.05 \mu\text{m}$, while the
 166 Ra of the 20%HA and 40%HA scaffolds increased to 0.2486 ± 0.0216 and $2.6 \pm 0.318 \mu\text{m}$,
 167 respectively. Porosity and pore size are positively correlated with the pore size ranging from 0.2
 168 to 2.0 mm as shown in Table1. When the small pore size (0.2~0.8 mm), the porosity of the PEEK/HA
 169 scaffolds were higher than that of pure PEEK scaffolds under the same diameter, while the deviation
 170 of porosity for the three scaffolds was reduced with the increase of pore size.

171 Table 1. The porosity of three scaffolds under the different pore size

pore size(mm)	0.2	0.4	0.8	1.2	1.6	2.0
Pure PEEK	47.3±1.20%	59.6±0.88%	72.2±0.79%	74.7±0.37%	77.4±0.35%	84.2±0.73%
20% HA	60.4±0.62%	68.7±1.40%	78.2±0.65%	79.8±0.47%	81.2±1.70%	85.9±1.15%
40% HA	60.9±1.10%	70.2±0.92%	78.4±1.49%	82.3±0.45%	83.7±1.15%	87.8±0.66%

172 3.2 Thermal behavior of composites



173
 174 Figure 3. (a) FTIR spectra, (b)DSC curves, (c) crystallinity results and (d)TGA curves of PEEK and
 175 PEEK/HA composites.

176 Figure 3-a showed the FTIR pattern of the pure PEEK and PEEK/HA composites. For the pure
 177 PEEK, the peak at 1650 cm^{-1} represented the C=O carbonyl stretching vibration, and the peaks at
 178 1598 cm^{-1} and 1501 cm^{-1} was contributed to the C=C benzene in-plane vibration. The characteristic
 179 peaks of PEEK could also be detected in PEEK/HA composites. The peaks of PEEK/HA composites
 180 at 1047 cm^{-1} and 570 cm^{-1} are ascribed to the vibration of the PO_4^{3-} , and the increase of HA content
 181 resulted the enhancement of PO_4^{3-} peak intensity. The DSC curves showed that a phase transition
 182 around $170 \text{ }^\circ\text{C}$ during the heating process referred to glass transition temperature (T_c) of PEEK
 183 (Figure 3b). A sharp endothermic peak around $345 \text{ }^\circ\text{C}$ corresponded to melting temperature (T_m) of
 184 PEEK. The degree of crystallinity of PEEK/HA composites increased with the increase of HA

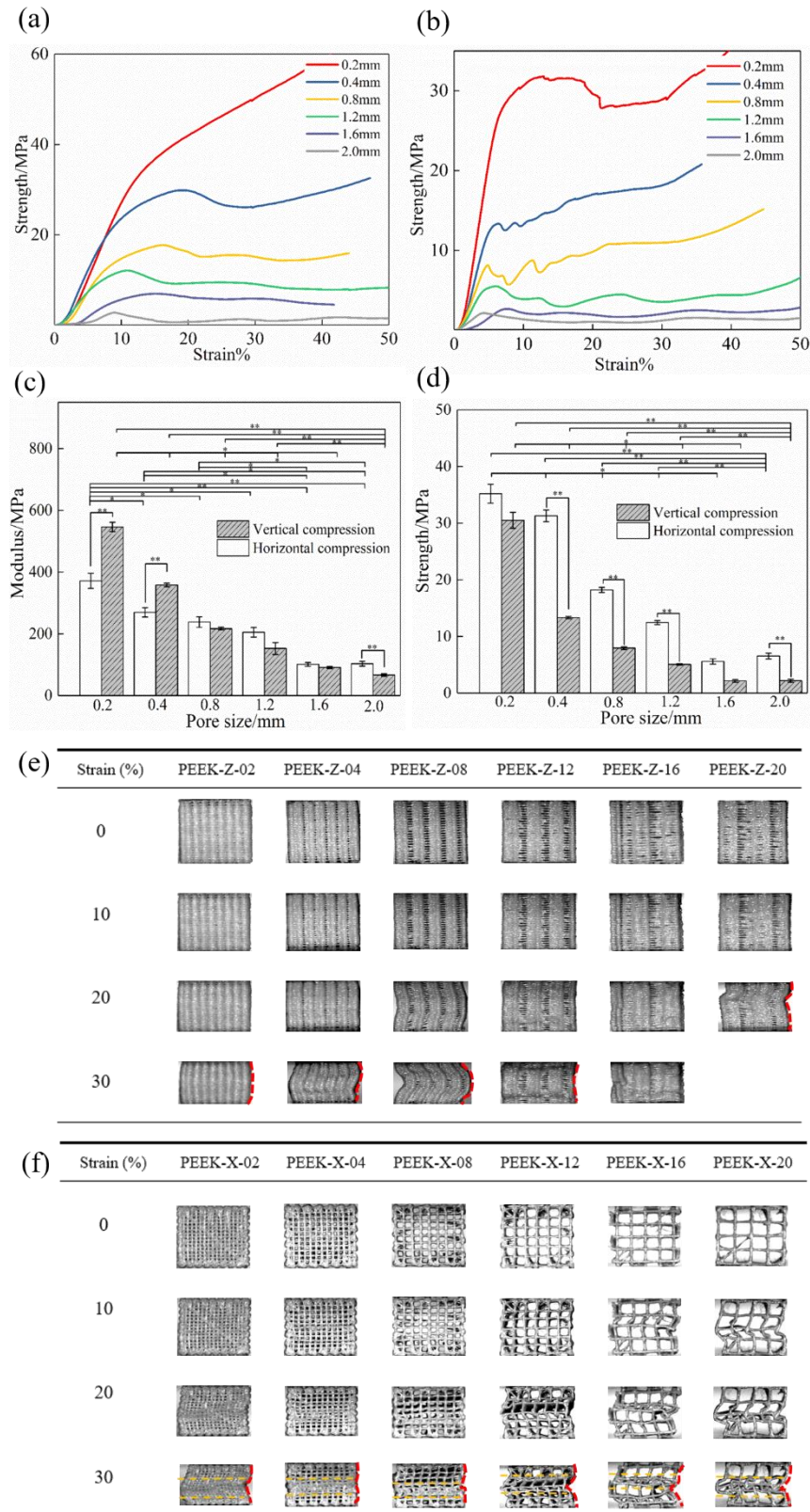
185 content (Figure 3-c). When the HA content increased to 40 wt.%, the degree of crystallinity of the
186 composite reached 28.7%. The TGA results show that both PEEK and PEEK/HA composites
187 undergo initial degradation approximately 550 °C (Figure 3d). A sharp drop in mass was observed
188 when the temperature exceeded 560 °C. The initial decomposition temperature shifted to slightly
189 lower temperatures with the increase of the HA content. Meanwhile the residual mass of PEEK/HA
190 composites at 800 °C increased with the increase of HA content.

191 **3.3 Mechanical tests**

192 Figure 4-a and Figure 4-b showed the stress-strain curves of scaffolds under compressive
193 process. The stress-strain curves of the scaffolds could be divided into three stages: elastic, plateau
194 and densification, which was consistent with previous investigations^[31]. For the PEEK scaffolds
195 compressed along Z printing direction, smooth curves were observed at the plateau stage. However,
196 negligible stress fluctuations were observed at the plateau stage for the scaffolds compressed along
197 X printing direction.

198 The compressive modulus and the strength were shown as in the Figure 4-c and Figure 4-d.
199 The Young's modulus of the pure PEEK scaffolds compressed along the Z and X printing direction
200 decreased from 371MPa to 102.9 MPa and from the 545.8MPa to 66.3MPa as the pore size increased
201 from 0.2 mm to 2 mm. Compressive strength along the Z compressive direction was higher than
202 that of scaffold along X direction for the scaffolds with the same pore size. The compressive strength
203 of scaffolds can be tuned in the range of 35.2 MPa to 2.2 MPa.

204 The compressive responses of the 3D-printed scaffolds were shown in Figure 4-e and Figure
205 4-f. When the scaffolds were compressed along the Z printing direction, the struts of square units
206 were horizontal to loading plate. The gap between layers of scaffolds gradually decreases and then
207 buckling deformation occurred. With the increase of the pore size, the strain corresponding the
208 beginning of the buckling deformation decreases. When the scaffolds were compressed along the X
209 printing direction, the buckling deformation and collapse occurred layer-by-layer, resulting the
210 crush bands (shown by the yellow dotted lines). The crush bands gradually accumulated and the
211 scaffolds were finally compressed into nearly compacted solids.



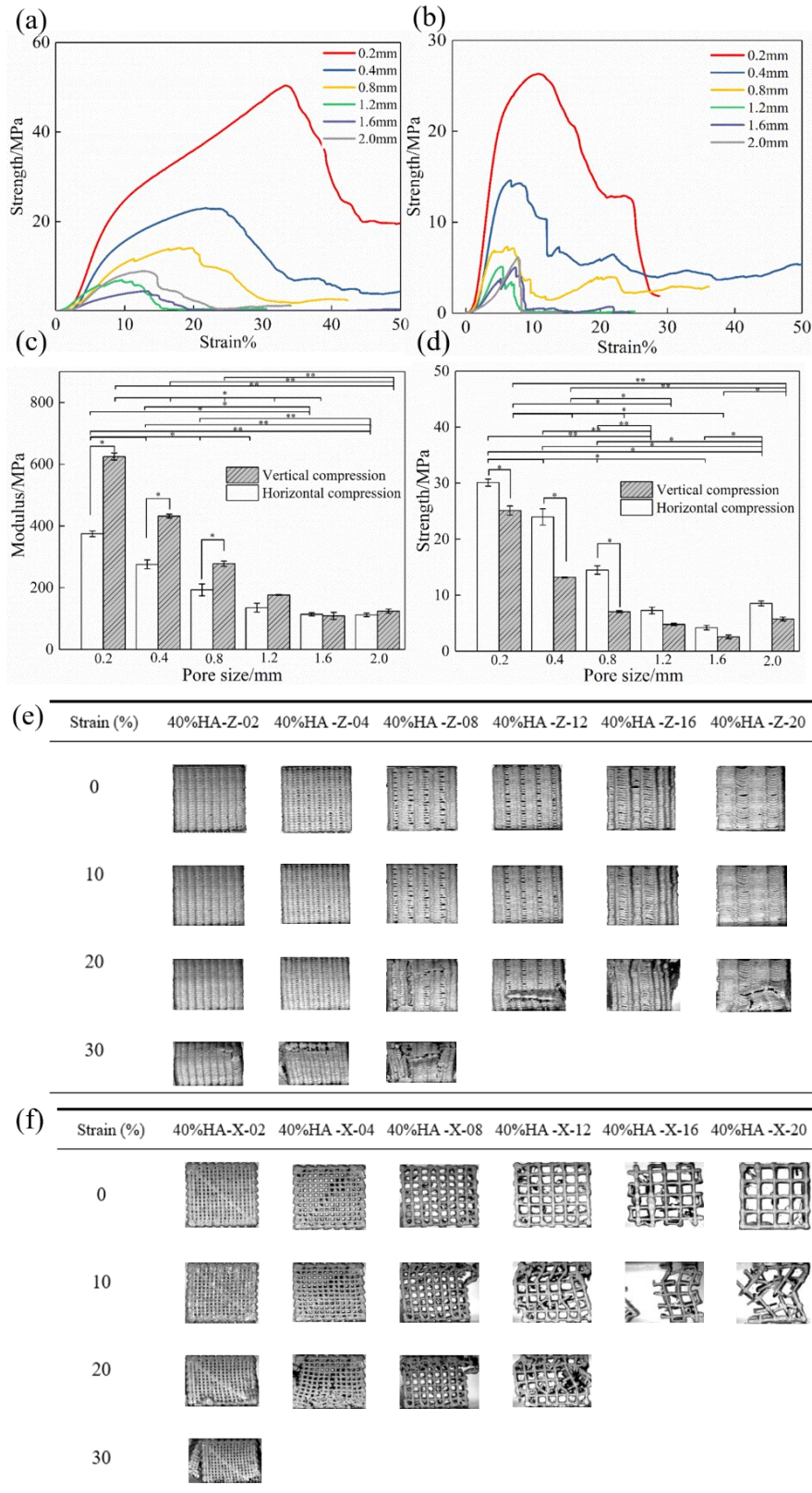
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213 Figure 4. The stress -strain curves of pure PEEK scaffolds compressed along Z (a) and X (b) printing
 214 direction. The modulus (c) and strength (d) results of the pure PEEK scaffolds with different pore
 215 sizes. The compressive responses of the pure PEEK composite scaffolds compressed along Z (e)
 216 and X (f) printing direction.

217 The mechanical properties of the composite scaffolds with 20 wt.% and 40 wt.% HA content
218 were evaluated. Taking composites scaffold with 40 wt.% HA content as an illustration, the stress-
219 strain curves of PEEK/HA composite scaffolds were shown in Figure 5-a and Figure 5-b. The
220 additive of HA particle reduced toughness of the scaffolds and the stress-strain curves of the
221 composite scaffolds showed a breakage stage instead of plateau and densification after the elastic
222 stage.

223 The compressive modulus and the strength were shown as in the Figure 5-c and Figure 5-d.
224 Compared with mechanical properties along the X printing direction, the composites scaffold
225 exhibits a higher Young's modulus and a lower compressive strength along the Z printing direction.
226 As the pore size increased from 0.2 mm to 2 mm, the value of Young's modulus along the Z and X
227 printing direction decreased from 374.7 MPa to 112.0 MPa and from 624.7 MPa to 124.2 MPa
228 respectively.

229 Figure5-e and Figure5-f showed the compressive responses of the PEEK/HA composite
230 scaffolds. The 40%HA-Z-02 and 40%HA-Z-04 scaffolds exhibited a brittle fracture as the strain
231 was > 30%. The larger pore size and porosity reduced the strain corresponding the beginning of
232 brittle fracture. The brittle fracture of 40%HA-Z-12, 40%HA-Z-16 and 40%HA-Z-20 occurred
233 when the strain reached 20%. As the scaffolds were compressed along X printing direction, the
234 struts of the scaffolds were collapsed layer by layer when pore size was <1.6 mm, complied with
235 crush bands accumulation and numerous brittle cracks. 40%HA-X-16 and 40%HA-X-20 scaffold
236 was crushed almost without any deformation when the strain was >10%.



237

238 Figure 5. The stress-strain curves of PEEK/HA scaffolds with 40 wt.% HA compressed along Z (a)

239 and X (b) printing direction. The modulus (c) and strength (d) results of the scaffolds with different

240 pore sizes. The compressive responses of the PEEK/HA composite scaffolds compressed along Z

241 (e) and X (f) printing direction.

242 Figure 6. showed the influence of the HA content and pore size on the mechanical properties
 243 of the scaffolds. The relations between the increase in pore size and decrease in modulus and
 244 strength can be intuitively understood by the response surface. Changes of HA content have different
 245 effects on the modulus anisotropy of the scaffold. When scaffolds were compressed along Z printing
 246 direction, no significant changes in modulus was found for the scaffolds with the same pore size,
 247 while the modulus of scaffolds remarkably increased with the increase of HA content along the X
 248 printing direction. The mathematic relationships between mechanical properties and parameter was
 249 obtained by polynomial fitting:

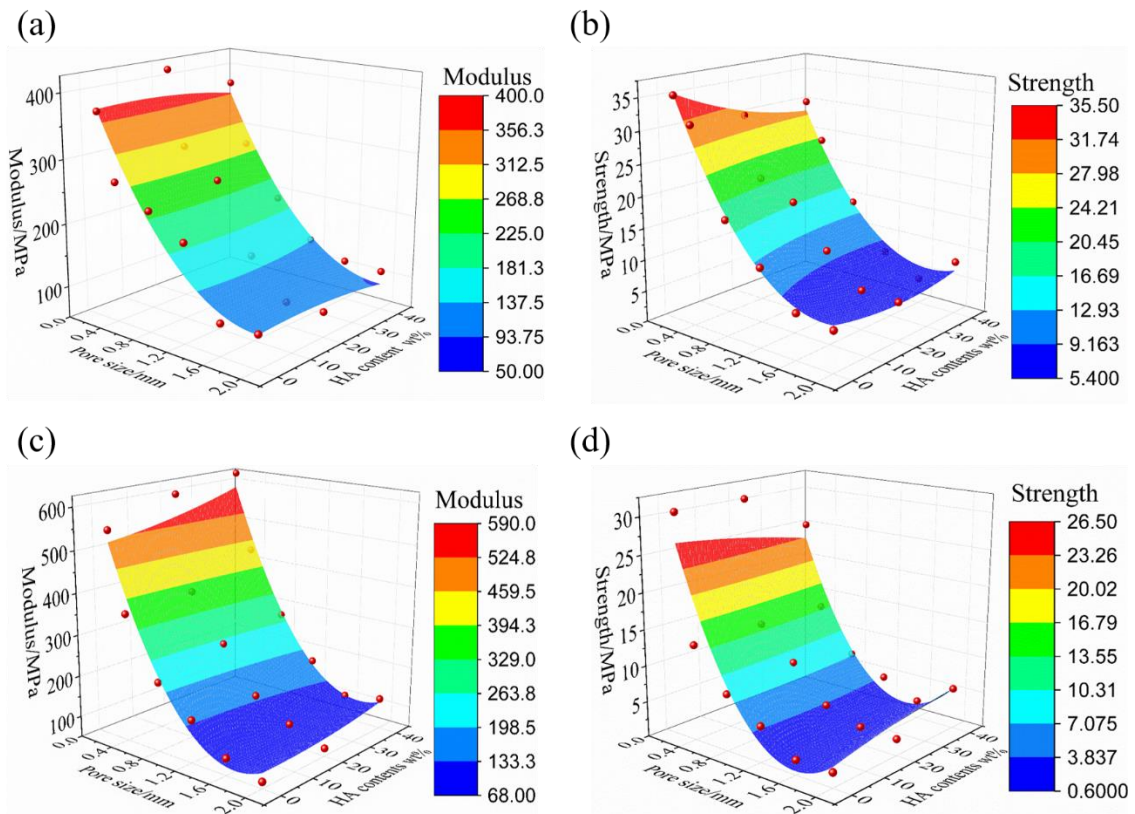
250
$$E_z = 483.7 - 337d + 0.313r + 84.41d^2 - 0.0187r^2 + 0.0865dr \quad (R^2 = 0.9431) \quad (2)$$

251
$$\sigma_z = 42.5 - 37.2d - 0.32r + 9.8d^2 + 0.003r^2 + 0.0936dr \quad (R^2 = 0.9712) \quad (3)$$

252
$$E_x = 644.7 - 676.7d + 1.05r + 198.8d^2 - 0.0214r^2 - 0.583dr \quad (R^2 = 0.9712) \quad (5)$$

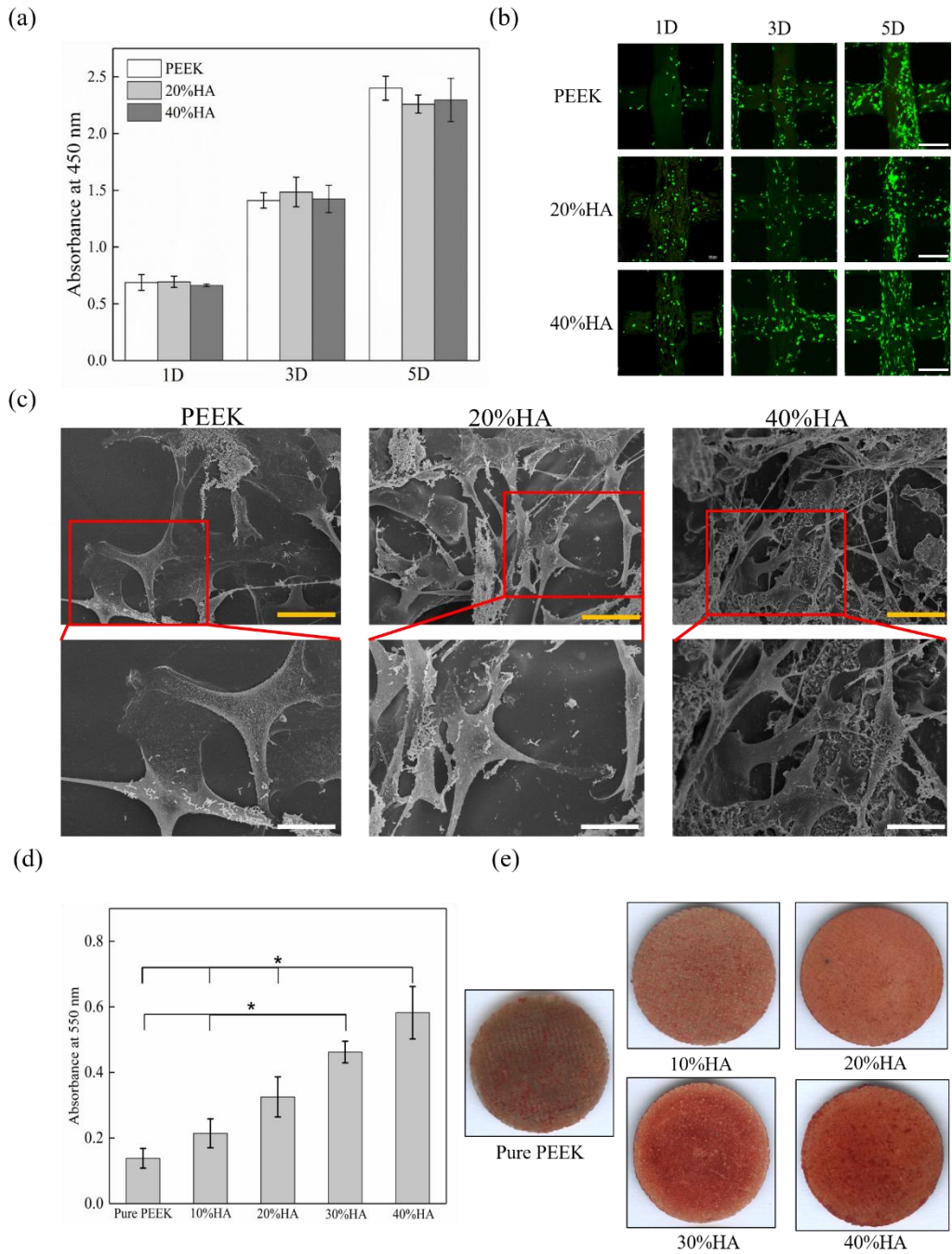
253
$$\sigma_x = 34.5 - 42.6d + 0.064r + 13.38d^2 - 0.001r^2 + 0.0875dr \quad (R^2 = 0.8996) \quad (5)$$

254 Where E_z and σ_z are the Young's modulus and strength compressed along Z printing direction.
 255 The E_x and σ_x are the Young's modulus and strength compressed along X printing direction. d
 256 is the pore size and r is the HA content.



257
 258 Figure 6. The modulus and strength varied with pore size and HA content: (a) modulus and (b)
 259 strength compressed along Z printing direction, (c) modulus and (d) strength compressed along X
 260 printing direction.
 261

262 **3.4 Cell compatibility of 3D-printed PEEK/HA scaffolds**



263

264 Figure 7. Proliferation, viability, morphology and mineralization of MC3T3-E1 on PEEK/HA

265 scaffolds. (a) Proliferation of MC3T3-E1 cell cultured on the scaffolds. (b) viability of MC3T3-E1

266 cultured for 1, 3, 5 days on the scaffolds. The white bar represented 400 μm. (c) SEM results of

267 adhering MC3T3-E1 cultured on the scaffolds for 5 days. The yellow bar represents 50 μm, and the

268 white bar represents 20 μm. (d) quantitative analysis of the alizarin red staining. (e) alizarin red

269 staining of samples with different HA content.

270 The proliferation of MC3T3-E1 cells on the scaffolds was reflected by OD values as shown in

271 Figure7-a. The OD values of cells on the scaffolds exhibited a persistent increasing proliferation

272 tendency from day 1 to day 7, indicating that both PEEK and PEEK/HA scaffolds exhibited good
273 cytocompatibility. There is no significant difference in the OD values between PEEK and PEEK/HA
274 under the same culture time. Figure7-b displayed that live cells were bright green and elongated
275 filopodia on the scaffolds. A higher number of cells were found on the scaffold with the increase of
276 culture time. Figure7-c showed the morphology of MC3T3-E1 cells on the scaffolds after 5 days of
277 culture and a morphology with an expanded polygonal shape was observed. It could be observed that
278 cells on scaffolds with 40 wt.% HA content showed more filopodia than that of cells on pure PEEK
279 scaffold. The mineralization ability of cells was visualized after 21 days of induction as shown in
280 Figure7-d and Figure7-e. Limited calcium nodule was observed on the pure PEEK sample, while
281 obvious calcium deposition was formed on the PEEK/HA scaffolds. The OD values were enhanced
282 with the increase of HA content.

283 4. Discussion

284 PEEK, as a polymeric material, was widely used in orthopedic clinics owing to its good
285 biocompatibility and satisfactory mechanical properties. But the biological inertness of PEEK
286 limited its applications for bone repairing. In this study, to enhance the bioactivity and
287 osseointegration ability of PEEK, PEEK/HA composites were prepared by incorporating HA
288 particles into PEEK and the PEEK/HA scaffolds were manufactured via FFF-3D printing technology.
289 Afterwards, the Micro-structure characteristics, thermal behavior, mechanical properties and the
290 biological compatibility of the scaffolds were systematically evaluated.

291 The printed scaffolds exhibited interconnected structure and the HA content reached up to 40
292 wt. %. The SEM results for the scaffolds surfaces indicated that the HA particles were not
293 completely embedded in the PEEK matrix but evenly distributed on the surfaces of the composites.
294 The surface roughness and morphology play a critical role in the cell responses for tissue
295 engineering material^[32]. Han^[33] et al. fabricated the PEEK and PEEK/CF composites samples and
296 found that special printing structures caused by FFF-3D printing process exhibited higher cell
297 densities than the samples being polished and sandblasted process. In this study, the LSCM results
298 showed that the Ra of composite scaffolds with 40 wt. % HA increased to $2.6 \pm 0.318 \mu\text{m}$. Although
299 PEEK/HA scaffolds had no significant effect on cell proliferation (Figure7-a), more filopodia of
300 cells on PEEK/HA scaffolds indicate that addition of HA particles and micro-structured surface
301 could promote cell attachment and spread, which was approved by previous study^[34]. The thermal
302 properties of the PEEK scaffold material were measured. For the PEEK/HA composites, because
303 HA particles provided heterogeneous crystallization nucleation points and reduced the nucleation
304 barrier of PEEK in the crystallization process, the degree of crystallinity of composites increased
305 with the increase of HA content. Previous studies^[35, 36] had demonstrated that the crystallinity of
306 PEEK could be adjusted by controlling the ambient temperature of FFF 3D printing process.
307 Meanwhile, the increase of crystallinity would reduce the breaking elongation and toughness.

308 The appropriate mechanical performance could remain the geometric characteristics of the
309 porous scaffold and provide a stress stimulation for the ingrowth of the bone tissue under
310 physiological loading conditions^[28]. Meanwhile, the mechanical anisotropy of the porous structure
311 fabricated by 3D printing process directly determines the applications site and stress transmission
312 *in vivo*. However, there were no systematic studies on the mechanical properties and the anisotropy
313 of the FFF printed PEEK/HA scaffolds reported. Results showed that the increase of pore size could

314 reduce the compressive modulus attributed to the increase of the porosity^[12]. When the pore size
315 increased to 2.0 mm, the modulus and the strength of the scaffolds dropped below 120 MPa and 9
316 MPa, respectively. The scaffolds compressed along Z printing direction exhibited higher
317 compressive strength and lower modulus compared with those of the scaffolds compressed along X
318 printing direction. The different loading state of the micro-rods inside the scaffolds was the main
319 reason to the differences of mechanical properties. For the scaffolds compressed along X printing
320 direction, the micro-rods are horizontal to the loading direction and directly bear the compressive
321 loading until bucking deformation. For the scaffolds compressed along Z printing direction, the
322 micro-rods are perpendicular to the loading direction and the interlayer gaps provides space for the
323 bending deformation of the micro-rod, which leads to a lower modulus compared to that of X
324 printing direction. Meanwhile, the higher effective bearing area along Z printing direction
325 contributed to a better compressive strength. The compressive response of pure PEEK scaffolds was
326 mainly plastic deformation, indicating its excellent ductility. The cracks and brittle fractures in the
327 compressive response of PEEK/HA composites demonstrated that the addition of HA particles can
328 significantly reduce the toughness. When the HA content range from 0 to 40 wt. %, the crystallinity
329 increased from 19.8% to 28.7%. The increase of crystallinity and poor interfacial strength between
330 the PEEK matrix and HA particles finally caused toughness decrease of the PEEK/HA composites^{[35,}
331 ^{37, 38]}.

332 The 3D surface representation of the mechanical properties changing with pore size and HA
333 content was investigated and the mathematical relationship among mechanical properties, geometric
334 structure and material components was further established by polynomial fitting. With the porosity
335 of scaffold decreased from about 55% to 85% (Table 1), the Young's modulus and the strength
336 decreased gradually ranging from 624.7-50.6 MPa and 35.2-2.2 MPa, respectively. The adjustable
337 range of the strength and modulus similar to the variation range of the natural trabecular bone
338 (strength: 2–17 MPa, modulus: 344–3230 MPa) ^[39, 40]. However, the scaffolds exhibited higher
339 Young's modulus and lower compressive strength along Z printing direction. Meanwhile, the
340 addition of HA particle led the decrease of the toughness. In clinical application, the appropriate
341 porosity, printing direction and HA content of the scaffolds should be selected and designed
342 according to the actual biomechanical environment for mechanical stability. The developed
343 mapping relationship could be used in adjusting the mechanical properties of the porous structure
344 to match the properties of the surrounding native bone. The printed scaffolds could be used in low-
345 load-bearing sites to enhance the integration with surrounding bone tissues. The mechanical bionics
346 of the scaffolds would be realized by balancing the mechanical mismatch between the PEEK-based
347 implant with natural bone.

348 Furthermore, the proliferation, attachment and mineralization ability of MC3T3-E1 cells on
349 the printed scaffolds were fully examined to study the cytocompatibility of the scaffolds. In this
350 study, a pore size of 400 μm was chosen for the scaffold, which has been reported in the literature
351 to be associated with excellent bone ingrowth, vascularization and permeability^[41, 42]. The CCK-8
352 assay results indicated that both the pure PEEK and PEEK/HA scaffolds showed good
353 cytocompatibility and the addition of HA particles had no significant effect on the proliferation the
354 MC3T3-E1 cells. However, Deng et al.^[43] and Swaminathan et al.^[44] reported that PEEK/HA
355 composites with nanoscale HA particles had a significantly positive influence on cells proliferation.
356 The difference of cells proliferation could be explained that bigger superficial area and nanoscale
357 roughness caused by nanoscale HA particles may exhibit better bioactivity. The obvious spreading

358 of cells on the struts of scaffolds demonstrated that both pure PEEK and PEEK/HA scaffolds
359 provided a suitable biological microenvironment for cell adhesion^[15, 45]. Specially, the cells on
360 PEEK/HA scaffolds with 40 wt. % HA content showed more filopodia than that of the cells on pure
361 PEEK scaffolds, demonstrating the better cell adhesion on the PEEK/HA composites. The results
362 of alizarin red staining showed that the addition of HA can significantly increase the mineralization
363 ability of cells on PEEK/HA scaffolds surface, similar to previous studies^[46, 47]. But the excessive
364 HA content would reduce the strength and toughness, thus the HA content should be selected to
365 balance the bioactivity and mechanical properties of PEEK/HA scaffolds.

366 Some limitations of this work must also be recognized. Only the mechanical behaviors of
367 scaffolds with tetragonal structures were evaluated, more bionic architectures, such as dodecahedron,
368 spiral structure, need to be addressed and evaluated for more thorough investigation in the future.
369 Additionally, the mechanical behaviors of the scaffolds were studied under static loading condition.
370 However, the fatigue behaviors of the 3D-printed scaffolds should also be evaluated. Cell
371 experiments *in vitro* was employed to assess the biocompatibility of the scaffolds, but the results
372 cannot fully represent the growth situation of native bone tissue *in vivo*. Therefore, tests under
373 realistic physiological environments such as the animal experiments should be carried out in the
374 future.

375 **5. Conclusion**

376 In this study, the 3D-printed PEEK/HA composites scaffolds with various lever of HA content
377 and pore sizes were fabricated using FFF 3D printing technology. The effects of the pore size,
378 compressive direction and HA content on the mechanical properties were systematically evaluated.
379 The Young's modulus and strength of the PEEK/HA scaffolds could be widely tuned in the range of
380 624.7-50.6MPa and 35.2-2.2 MPa respectively by increasing the pore size from 0.2 mm to 2.0 mm.
381 The participant of HA was found to significantly enhance the modulus along X printing direction,
382 while the modulus of scaffolds along Z printing direction has no obvious change. The mapping
383 relationship among geometric parameters, HA content and mechanical properties was established,
384 giving more accurate predictions and controllability for the mechanical properties of the scaffolds.
385 The PEEK/HA composites with the micro-structured surface could significantly promote cell
386 attachment and mineralization. Presented study will have a clear impact in the design and
387 mechanical properties control of the PEEK/HA 3D-printed scaffolds for bone tissue engineering.

388 **Acknowledgements**

389 The work was financially supported by the Program of the National Natural Science
390 Foundation of China [51835010], the Key R&D Program of Guangdong Province
391 [2018B090906001], the Project funded by China Postdoctoral Science Foundation [2020M683458],
392 the National Key R&D Program of China [2018YFE0207900], the Fundamental Research Funds
393 for the Central Universities, and "The Youth Innovation Team of Shaanxi Universities."

394 CL Would like to acknowledge the support of European Union via H2020-MSCA-RISE program(BAMOS project, grant no:
734156) and Engineering and Physical Science Research Council (EPSRC) via DTP CASE programme (Grant No: EP/
T517793/1).

395 **Reference**

- 396 [1] C. Wang, Q.L. Zhao, M. Wang, Cryogenic 3D printing for producing hierarchical porous
397 and rhBMP-2loaded Ca-P/PLLA nanocomposite scaffolds for bone tissue engineering,
398 *Biofabrication* 9(2) (2017) 12.
- 399 [2] L. Wang, J.F. Kang, C.N. Sun, D.C. Li, Y. Cao, Z.M. Jin, Mapping porous microstructures
400 to yield desired mechanical properties for application in 3D printed bone scaffolds and
401 orthopaedic implants, *Mater. Des.* 133 (2017) 62-68.
- 402 [3] N. Sears, P. Dhavalikar, M. Whitely, E. Cosgriff-Hernandez, Fabrication of biomimetic bone
403 grafts with multi-material 3D printing, *Biofabrication* 9(2) (2017) 11.
- 404 [4] S. Ghouse, N. Reznikov, O.R. Boughton, S. Babu, K.C.G. Ng, G. Blunn, et al., The design
405 and in vivo testing of a locally stiffness-matched porous scaffold, *Appl. Mater. Today* 15
406 (2019) 377-388.
- 407 [5] C.J. Shuai, Y. Cheng, W.J. Yang, P. Feng, Y.W. Yang, C.X. He, et al., Magnetically
408 actuated bone scaffold: Microstructure, cell response and osteogenesis, *Compos. Pt. B-
409 Eng.* 192 (2020) 11.
- 410 [6] M.R. Zhou, J.F. Hou, G. Zhang, C. Luo, Y.Y. Zeng, S. Mou, et al., Tuning the mechanics
411 of 3D-printed scaffolds by crystal lattice-like structural design for breast tissue engineering,
412 *Biofabrication* 12(1) (2020) 17.
- 413 [7] H.M.A. Kolken, S. Janbaz, S.M.A. Leeflang, K. Lietaert, H.H. Weinans, A.A. Zadpoor,
414 Rationally designed meta-implants: a combination of auxetic and conventional meta-
415 biomaterials, *Mater. Horizons* 5(1) (2018) 28-35.
- 416 [8] S. Limmahakhun, A. Oloyede, K. Sitthiseriratip, Y. Xiao, C. Yan, Stiffness and strength

417 tailoring of cobalt chromium graded cellular structures for stress-shielding reduction, Mater.
418 Des. 114 (2017) 633-641.

419 [9] F.S.L. Bobbert, K. Lietaert, A.A. Eftekhari, B. Pouran, S.M. Ahmadi, H. Weinans, et al.,
420 Additively manufactured metallic porous biomaterials based on minimal surfaces: A unique
421 combination of topological, mechanical, and mass transport properties, Acta Biomater. 53
422 (2017) 572-584.

423 [10] S.M. Ahmadi, R. Hedayati, Y. Li, K. Lietaert, N. Tumer, A. Fatemi, et al., Fatigue
424 performance of additively manufactured meta-biomaterials: The effects of topology and
425 material type, Acta Biomater. 65 (2018) 292-304.

426 [11] C. Yan, L. Hao, A. Hussein, P. Young, D. Raymont, Advanced lightweight 316L
427 stainless steel cellular lattice structures fabricated via selective laser melting, Mater. Des.
428 55 (2014) 533-541.

429 [12] R. Baptista, M. Guedes, Morphological and mechanical characterization of 3D printed
430 PLA scaffolds with controlled porosity for trabecular bone tissue replacement, Materials
431 science & engineering. C, Materials for biological applications 118 (2021) 111528-111528.

432 [13] S. Chen, W. Chen, Y. Chen, X. Mo, C. Fan, Chondroitin sulfate modified 3D porous
433 electrospun nanofiber scaffolds promote cartilage regeneration, Materials science &
434 engineering. C, Materials for biological applications 118 (2021) 111312-111312.

435 [14] D. Sridharan, A. Palaniappan, B.N. Blackstone, J.A. Dougherty, N. Kumar, P.B.
436 Seshagiri, et al., In situ differentiation of human-induced pluripotent stem cells into
437 functional cardiomyocytes on a coaxial PCL-gelatin nanofibrous scaffold, Materials science
438 & engineering. C, Materials for biological applications 118 (2021) 111354-111354.

- 439 [15] Y. Su, J. He, N. Jiang, H. Zhang, L. Wang, X. Liu, et al., Additively-manufactured poly-
440 ether-ether-ketone (PEEK) lattice scaffolds with uniform microporous architectures for
441 enhanced cellular response and soft tissue adhesion, *Mater. Des.* 191 (2020).
- 442 [16] H. Ahn, R.R. Patel, A.J. Hoyt, A.S.P. Lin, F.B. Torstrick, R.E. Guldborg, et al.,
443 Biological evaluation and finite-element modeling of porous poly(para-phenylene) for
444 orthopaedic implants, *Acta Biomater.* 72 (2018) 352-361.
- 445 [17] M.Z. Zhao, H.Y. Li, X.C. Liu, J. Wei, J.G. Ji, S. Yang, et al., Response of Human
446 Osteoblast to n-HA/PEEK-Quantitative Proteomic Study of Bio-effects of Nano-
447 Hydroxyapatite Composite, *Sci Rep* 6 (2016) 13.
- 448 [18] R. Ma, S. Tang, H. Tan, J. Qian, W. Lin, Y. Wang, et al., Preparation, characterization,
449 in vitro bioactivity, and cellular responses to a polyetheretherketone bioactive composite
450 containing nanocalcium silicate for bone repair, *ACS applied materials & interfaces* 6(15)
451 (2014) 12214-12225.
- 452 [19] P. Egan, R. Sinko, P.R. LeDuc, S. Keten, The role of mechanics in biological and bio-
453 inspired systems, *Nat. Commun.* 6 (2015) 12.
- 454 [20] A. Higuchi, Q.D. Ling, Y. Chang, S.T. Hsu, A. Umezawa, Physical Cues of
455 Biomaterials Guide Stem Cell Differentiation Fate, *Chem. Rev.* 113(5) (2013) 3297-3328.
- 456 [21] T.Z. Luo, K. Mohan, P.A. Iglesias, D.N. Robinson, Molecular mechanisms of cellular
457 mechanosensing, *Nat. Mater.* 12(11) (2013) 1063-1070.
- 458 [22] C. Shuai, W. Yang, P. Feng, S. Peng, H. Pan, Accelerated degradation of HAP/PLLA
459 bone scaffold by PGA blending facilitates bioactivity and osteoconductivity, *Bioactive*
460 *Materials* 6(2) (2021) 490-502.

- 461 [23] A. Barba, Y. Maazouz, A. Diez-Escudero, K. Rappe, M. Espanol, E.B. Montufar, et
462 al., Osteogenesis by foamed and 3D-printed nanostructured calcium phosphate scaffolds:
463 Effect of pore architecture, *Acta Biomater.* 79 (2018) 135-147.
- 464 [24] J.F. Kang, E.C. Dong, D.C. Li, S.P. Dong, C. Zhang, L. Wang, Anisotropy
465 characteristics of microstructures for bone substitutes and porous implants with application
466 of additive manufacturing in orthopaedic, *Mater. Des.* 191 (2020) 10.
- 467 [25] K. Hayashi, M.L. Munar, K. Ishikawa, Effects of macropore size in carbonate apatite
468 honeycomb scaffolds on bone regeneration, *Mater. Sci. Eng. C-Mater. Biol. Appl.* 111
469 (2020) 9.
- 470 [26] D. Melancon, Z.S. Bagheri, R.B. Johnston, L. Liu, M. Tanzer, D. Pasini, Mechanical
471 characterization of structurally porous biomaterials built via additive manufacturing:
472 experiments, predictive models, and design maps for load-bearing bone replacement
473 implants, *Acta Biomater.* 63 (2017) 350-368.
- 474 [27] K. Hazeli, B.B. Babamiri, J. Indeck, A. Minor, H. Askari, Microstructure-topology
475 relationship effects on the quasi-static and dynamic behavior of additively manufactured
476 lattice structures, *Mater. Des.* 176 (2019) 21.
- 477 [28] X.Y. Liang, J.M. Gao, W.K. Xu, X.L. Wang, Y. Shen, J.Y. Tang, et al., Structural
478 mechanics of 3D-printed poly(lactic acid) scaffolds with tetragonal, hexagonal and wheel-
479 like designs, *Biofabrication* 11(3) (2019) 22.
- 480 [29] F.P.W. Melchels, K. Bertoldi, R. Gabbriellini, A.H. Velders, J. Feijen, D.W. Grijpma,
481 Mathematically defined tissue engineering scaffold architectures prepared by
482 stereolithography, *Biomaterials* 31(27) (2010) 6909-6916.

- 483 [30] A. Bagheri, I. Buj-Corral, M.F. Ballester, M.M. Pastor, F.R. Fernandez, Determination
484 of the Elasticity Modulus of 3D-Printed Octet-Truss Structures for Use in Porous Prosthesis
485 Implants, *Materials* 11(12) (2018) 16.
- 486 [31] Z. Meng, J. He, Z. Cai, F. Wang, J. Zhang, L. Wang, et al., Design and additive
487 manufacturing of flexible polycaprolactone scaffolds with highly-tunable mechanical
488 properties for soft tissue engineering, *Mater. Des.* 189 (2020).
- 489 [32] S. Metwally, S. Ferraris, S. Spriano, Z.J. Krysiak, L. Kaniuk, M.M. Marzec, et al.,
490 Surface potential and roughness controlled cell adhesion and collagen formation in
491 electrospun PCL fibers for bone regeneration, *Mater. Des.* 194 (2020).
- 492 [33] X.T. Han, D. Yang, C.C. Yang, S. Spintzyk, L. Scheideler, P. Li, et al., Carbon Fiber
493 Reinforced PEEK Composites Based on 3D-Printing Technology for Orthopedic and
494 Dental Applications, *J. Clin. Med.* 8(2) (2019) 17.
- 495 [34] P. Jiang, Y. Zhang, R. Hu, X. Wang, Y. Lai, G. Rui, et al., Hydroxyapatite-modified
496 micro/nanostructured titania surfaces with different crystalline phases for osteoblast
497 regulation, *Bioactive materials* 6(4) (2021) 1118-1129.
- 498 [35] C.C. Yang, X.Y. Tian, D.C. Li, Y. Cao, F. Zhao, C.Q. Shi, Influence of thermal
499 processing conditions in 3D printing on the crystallinity and mechanical properties of PEEK
500 material, *J. Mater. Process. Technol.* 248 (2017) 1-7.
- 501 [36] A. El Magri, K. El Mabrouk, S. Vaudreuil, H. Chibane, M.E. Touhami, Optimization of
502 printing parameters for improvement of mechanical and thermal performances of 3D
503 printed poly(ether ether ketone) parts, *J. Appl. Polym. Sci.* 137(37) (2020) 14.
- 504 [37] M.S. Abu Bakar, M.H.W. Cheng, S.M. Tang, S.C. Yu, K. Liao, C.T. Tan, et al., Tensile

505 properties, tension-tension fatigue and biological response of polyetheretherketone-
506 hydroxyapatite composites for load-bearing orthopedic implants, *Biomaterials* 24(13)
507 (2003) 2245-2250.

508 [38] E.A.B. Hughes, A. Parkes, R.L. Williams, M.J. Jenkins, L.M. Grover, Formulation of a
509 covalently bonded hydroxyapatite and poly(ether ether ketone) composite, *Journal of*
510 *Tissue Engineering* 9 (2018).

511 [39] V. Martin, I.A. Ribeiro, M.M. Alves, L. Goncalves, R.A. Claudio, L. Grenho, et al.,
512 Engineering a multifunctional 3D-printed PLA-collagen-minocycline-nanoHydroxyapatite
513 scaffold with combined antimicrobial and osteogenic effects for bone regeneration, *Mater.*
514 *Sci. Eng. C-Mater. Biol. Appl.* 101 (2019) 15-26.

515 [40] E.F. Morgan, T.M. Keaveny, Dependence of yield strain of human trabecular bone on
516 anatomic site, *J. Biomech.* 34(5) (2001) 569-577.

517 [41] T. Yu, H. Gao, T. Liu, Y.D. Huang, C. Wang, Effects of immediately static loading on
518 osteointegration and osteogenesis around 3D-printed porous implant: A histological and
519 biomechanical study, *Mater. Sci. Eng. C-Mater. Biol. Appl.* 108 (2020) 12.

520 [42] X.P. Tan, Y.J. Tan, C.S.L. Chow, S.B. Tor, W.Y. Yeong, Metallic powder-bed based
521 3D printing of cellular scaffolds for orthopaedic implants: A state-of-the-art review on
522 manufacturing, topological design, mechanical properties and biocompatibility, *Mater. Sci.*
523 *Eng. C-Mater. Biol. Appl.* 76 (2017) 1328-1343.

524 [43] Y. Deng, P. Zhou, X.C. Liu, L.X. Wang, X.L. Xiong, Z.H. Tang, et al., Preparation,
525 characterization, cellular response and in vivo osseointegration of
526 polyetheretherketone/nano-hydroxyapatite/carbon fiber ternary biocomposite, *Colloid Surf.*

527 B-Biointerfaces 136 (2015) 64-73.

528 [44] P.D. Swaminathan, M.N. Uddin, P. Wooley, R. Asmatulu, Fabrication and Biological
529 Analysis of Highly Porous PEEK Bionanocomposites Incorporated with Carbon and
530 Hydroxyapatite Nanoparticles for Biological Applications, *Molecules* 25(16) (2020).

531 [45] H. Spece, T. Yu, A.W. Law, M. Marcolongo, S.M. Kurtz, 3D printed porous PEEK
532 created via fused filament fabrication for osteoconductive orthopaedic surfaces, *J. Mech.
533 Behav. Biomed. Mater.* 109 (2020).

534 [46] R. Ma, D.G. Guo, Evaluating the bioactivity of a hydroxyapatite-incorporated
535 polyetheretherketone biocomposite, *J. Orthop. Surg. Res.* 14 (2019) 13.

536 [47] R. Ma, Z.F. Yu, S.C. Tang, Y.K. Pan, J. Wei, T.T. Tang, Osseointegration of
537 nanohydroxyapatite- or nanocalcium silicate-incorporated polyetheretherketone bioactive
538 composites in vivo, *Int. J. Nanomed.* 11 (2016) 6023-6033.

539

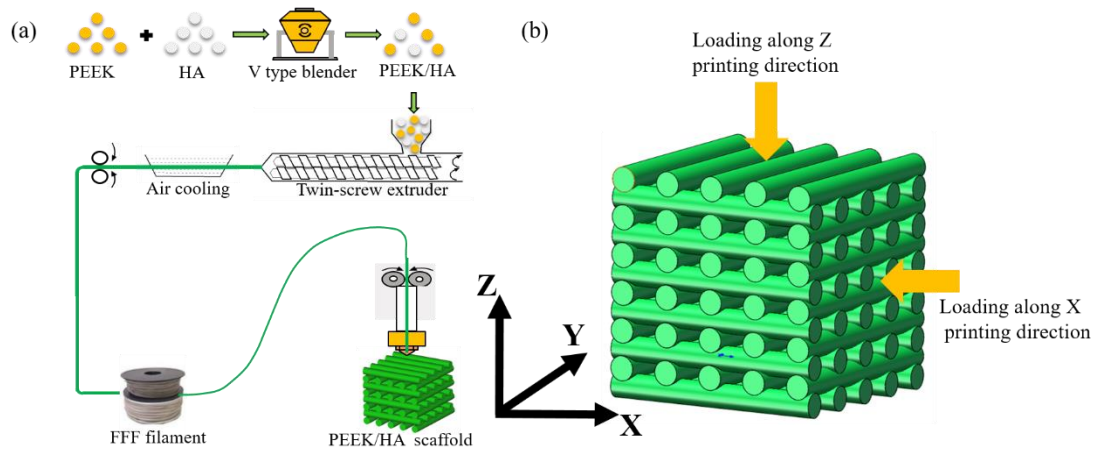


Figure 1. Schematic illustration of the 3D printing process and mechanical properties testing method: (a) preparation of PEEK/HA composites and FFF printing (b) scaffold compressed along Z or X printing direction.

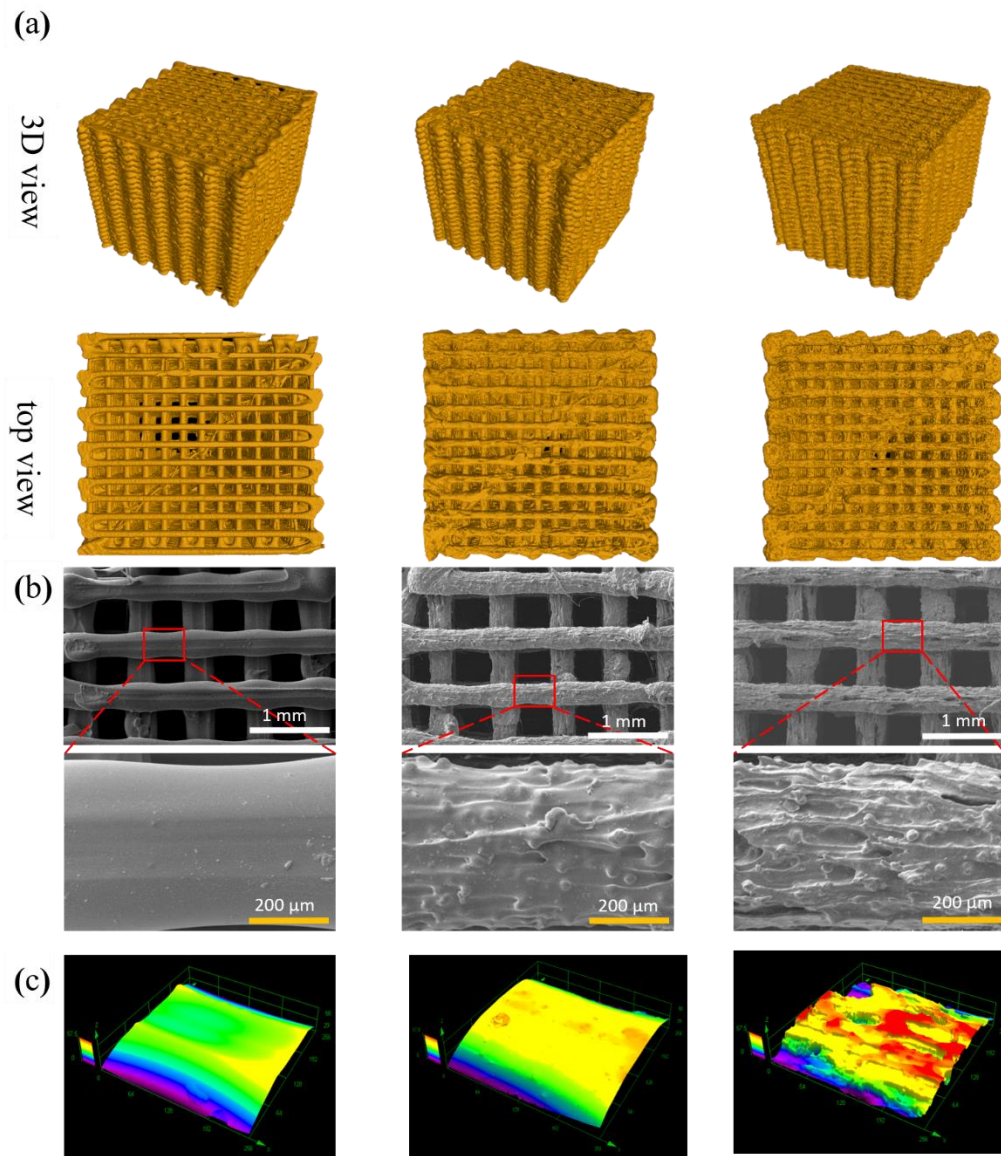


Figure 2. (a) structural characteristics, (b) surface morphology and (c) three-dimensional topography of the scaffolds with pure PEEK, 20 wt.% HA and 40 wt.% HA content.

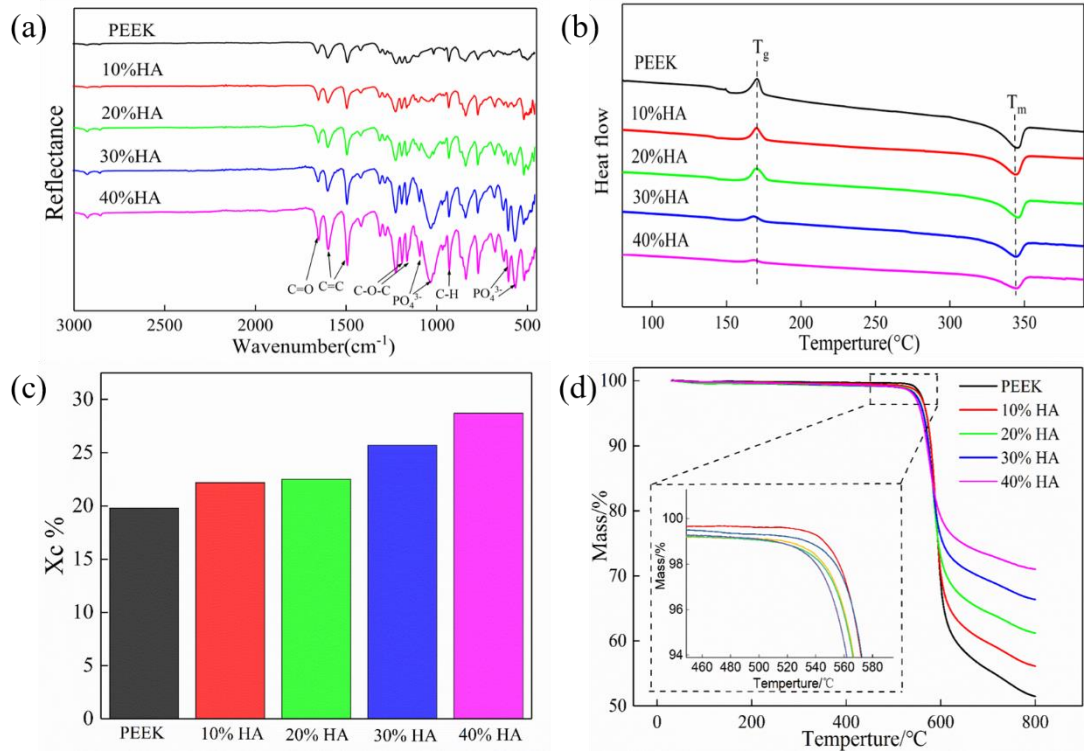


Figure 3. (a) FTIR pattern, (b)DSC results, (c) crystallinity and (d)TGA curves of PEEK and PEEK/HA composites.

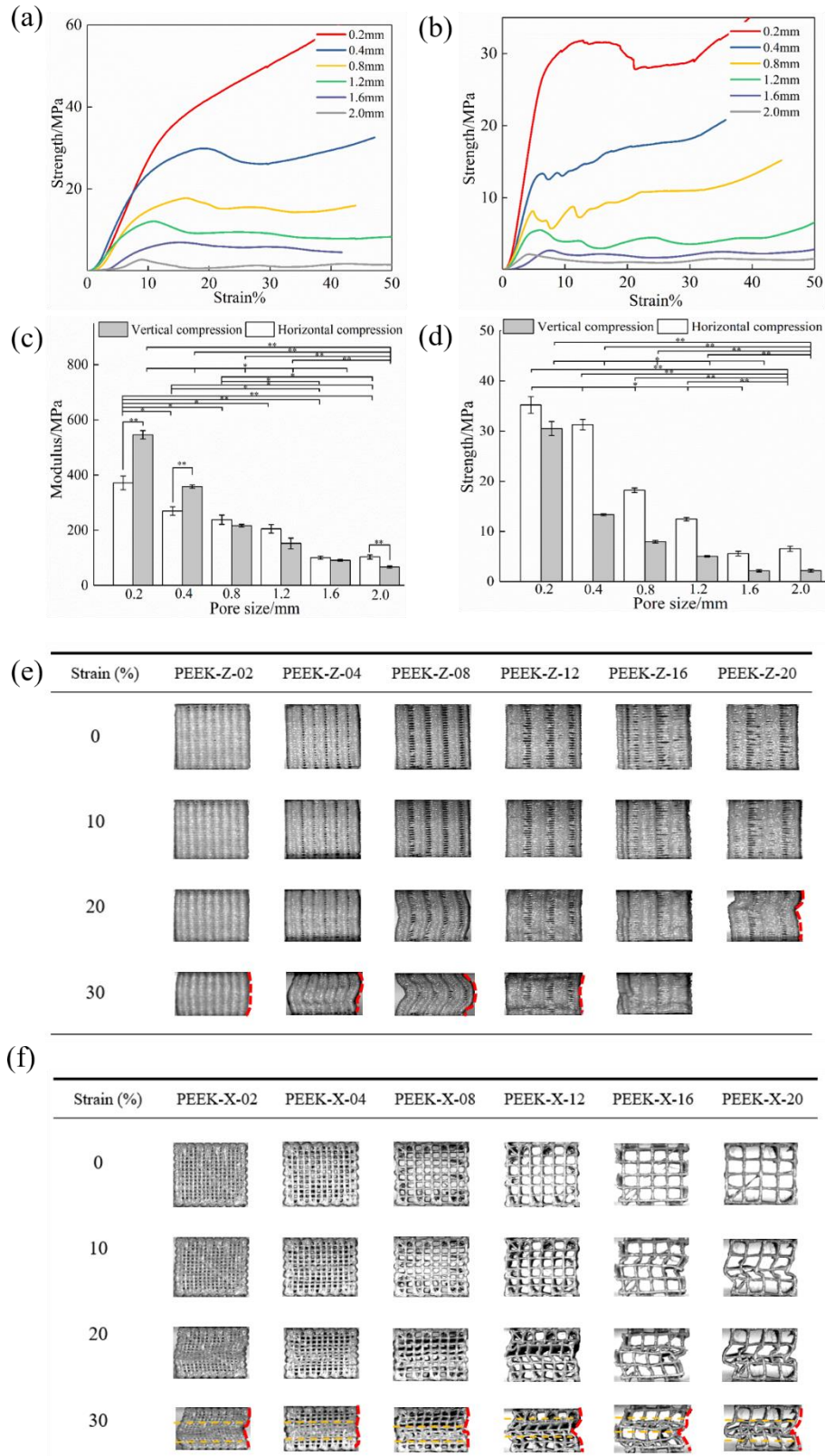


Figure 4. The stress-strain curves of pure PEEK scaffolds compressed along Z (a) and X (b) printing direction. The modulus (c) and strength (d) results of the pure PEEK scaffolds with different pore sizes. The compressive responses of the pure PEEK composite scaffolds compressed along Z (e) and X (f) printing direction.

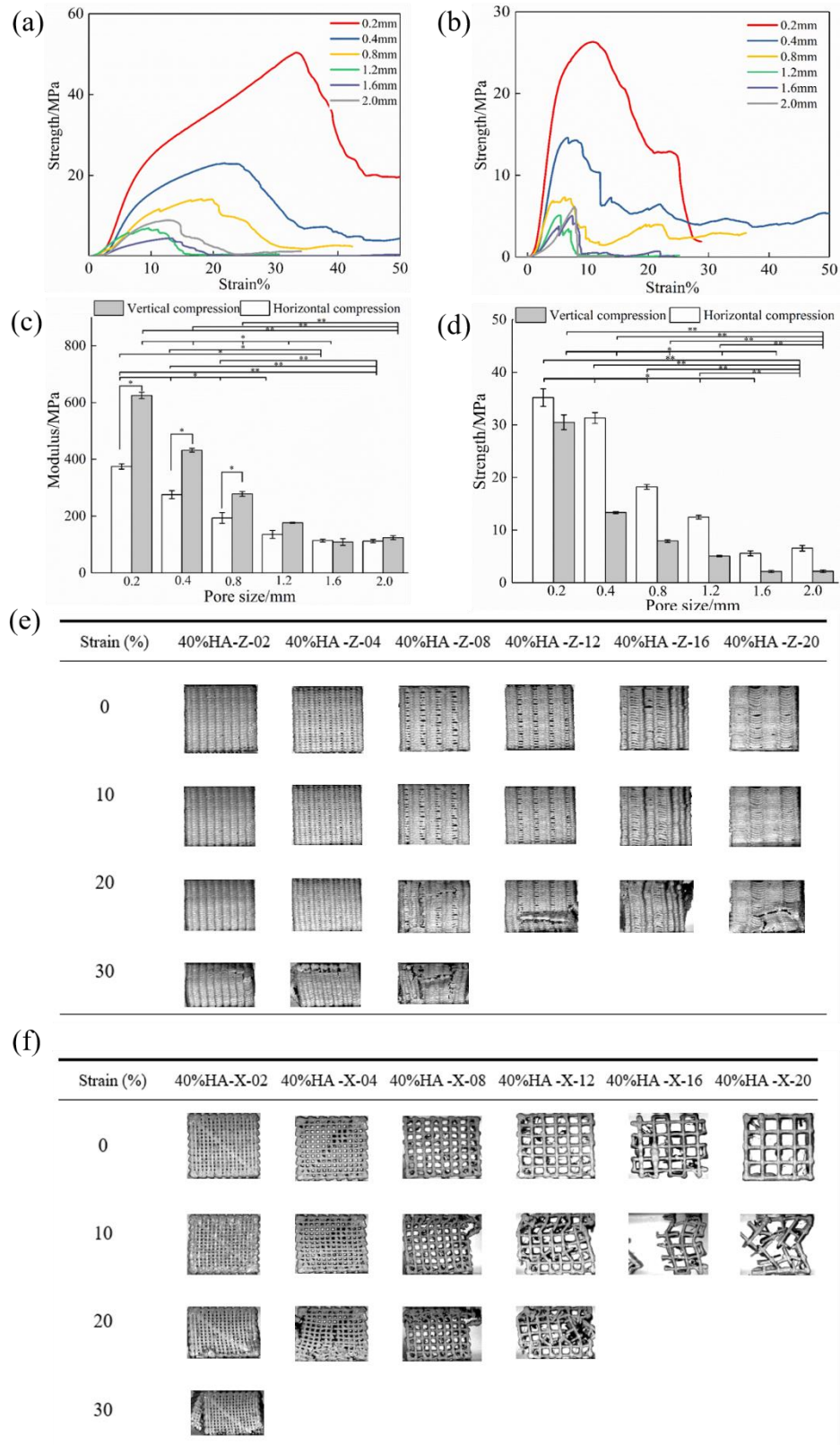


Figure 5. The stress-strain curves of PEEK/HA scaffolds compressed along Z (a) and X (b) printing direction. The modulus (c) and strength (d) results of the scaffolds with different pore sizes. The compressive responses of the PEEK/HA composite scaffolds compressed along Z (e) and X (f) printing direction.

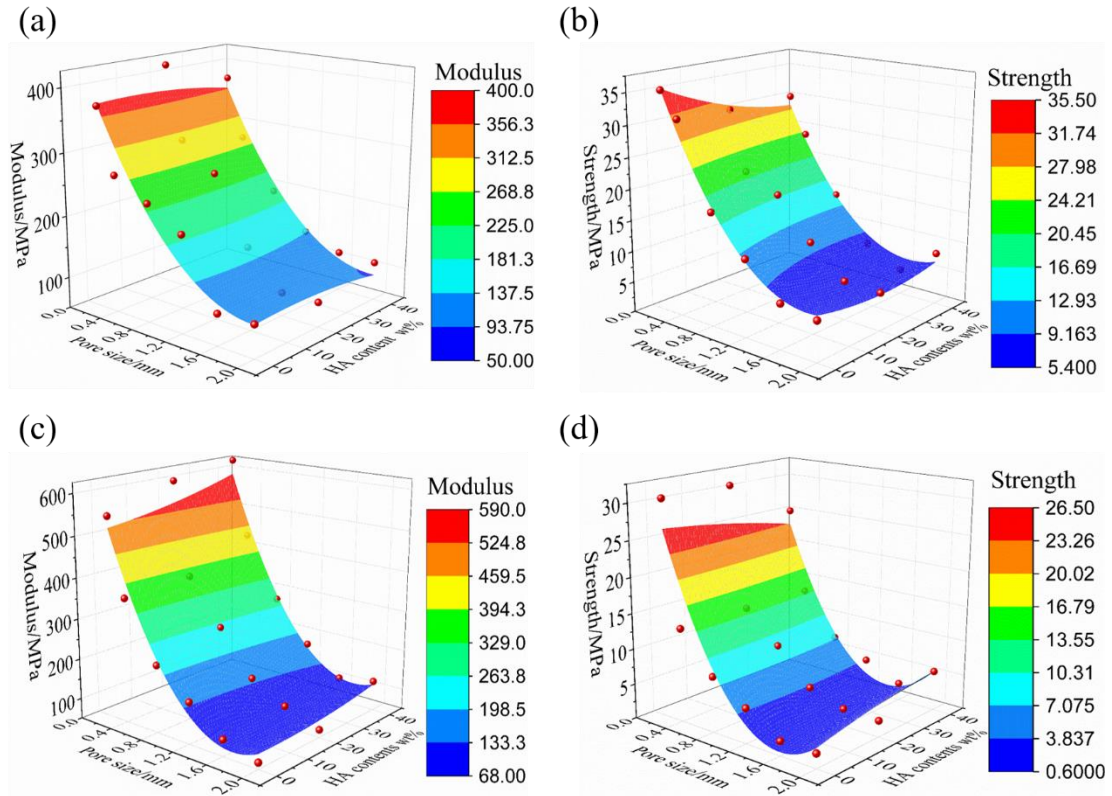


Figure 6. The modulus and strength varied with pore size and HA content: (a) modulus and (b) strength compressed along Z printing direction, (c) modulus and (d) strength compressed along X printing direction.

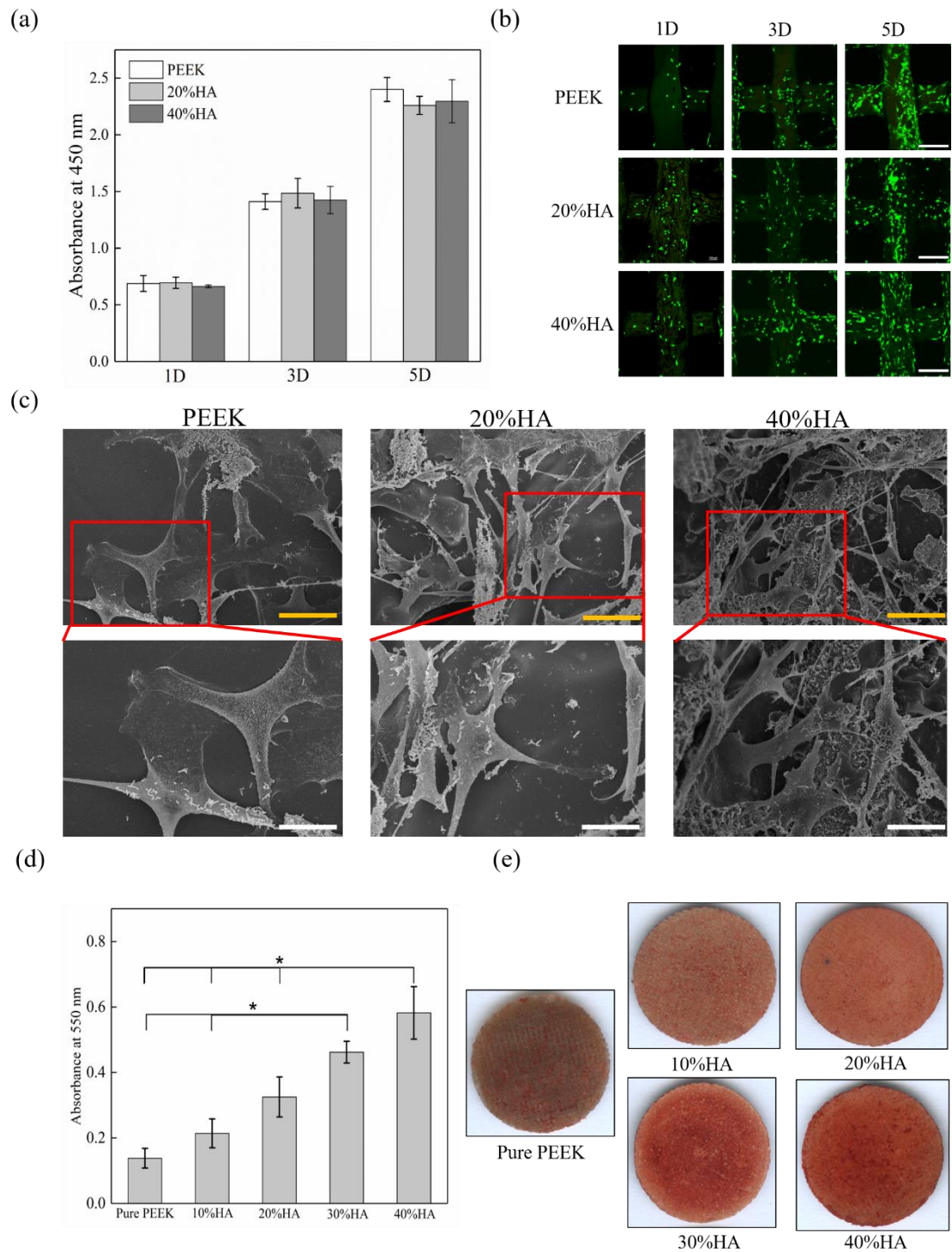


Fig 7. Proliferation, viability, morphology and mineralization of MC3T3-E1 on PEEK/HA scaffolds. (a) Proliferation of MC3T3-E1 cell cultured on the scaffolds. (b) viability of MC3T3-E1 cultured for 1, 3, 5 days on the scaffolds. The white bar represented 400 μm . (c) SEM results of adhering MC3T3-E1 cultured on the scaffolds for 5 days. The yellow bar represents 50 μm , and the white bar represents 20 μm . (d) quantitative analysis of the alizarin red staining. (e) alizarin red staining of samples with different HA content.

Table 1. The porosity of the scaffolds with different pore size and HA content

Scaffold/pore size(mm)	0.2	0.4	0.8	1.2	1.6	2.0
Pure PEEK	47.3±1.20%	59.6±0.88%	72.2±0.79%	74.7±0.37%	77.4±0.35%	84.2±0.73%
20% HA	60.4±0.62%	68.7±1.40%	78.2±0.65%	79.8±0.47%	81.2±1.70%	85.9±1.15%
40% HA	60.9±1.10%	70.2±0.92%	78.4±1.49%	82.3±0.45%	83.7±1.15%	87.8±0.66%