

Docking and Bioinformatics Tools to Guide Enzyme Engineering

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One general law, leading to the advancement of all organic beings, namely, multiply, vary, let the strongest live and the weakest die.

-Charles Darwin, *The Origin of Species* (1859)

Declaration

I, John Strafford confirm that the work presented in this thesis is my own. Where information has been derived from other sources, I confirm that this has been indicated in the thesis.

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I wish to thank my supervisors, Paul Dalby and David Jones, for their continued support throughout my PhD. I would also like to extend my thanks to all those in the Biochemical Engineering Department of UCL, in particular Nigel Titchener-Hooker for his words of encouragement at difficult times.

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Last, but not least, I would like to thank the Biotechnology and Biological Sciences Research Council for funding my PhD.

Abstract

The carbon-carbon bond forming ability of transketolase (TK), along with its broad substrate specificity, makes it very attractive as a biocatalyst in industrial organic synthesis. Through the production of saturation mutagenesis libraries focused on individual active site residues, several variants of TK have been discovered with enhanced activities on non-natural substrates. We have used computational and bioinformatics tools to increase our understanding of TK and to guide engineering of the enzyme for further improvements in activity.

Computational automated docking is a powerful technique with the potential to identify transient structures along an enzyme reaction pathway that are difficult to obtain by experimental structure determination. We have used the AutoDock algorithm to dock a series of known ketol donor and aldehyde acceptor substrates into the active site of *E. coli* TK, both in the presence and the absence of reactive intermediates. Comparison of docked conformations with available crystal structure complexes allows us to propose a more complete mechanism at a level of detail not currently possible by experimental structure determination alone.

Statistical coupling analysis (SCA) utilises evolutionary sequence data present within multiple sequence alignments to identify energetically coupled networks of residues within protein structures. Using this technique we have identified several coupled networks within the TK enzyme which we have targeted for mutagenesis in multiple mutant variant libraries. Screening of these libraries for increased activity on the non-natural substrate propionaldehyde (PA) has identified combinations of mutations that act synergistically on enzyme activity. Notably, a double variant has

been discovered with a 20-fold improvement in k_{cat} relative to wild type on the PA reaction, this is higher than any other TK variant discovered to date.

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Abbreviations

2OXO:	2-oxoisovalerate dehydrogenase
AA:	Acetaldehyde
ALS:	Acetolactone synthase
BAL:	Benzaldehyde lyase
BFDC:	Benzoylformate decarboxylase
DE:	D-erythrose
DE4P:	D-erythrose-4-phosphate
DG:	D-glyceraldehyde
DG3P:	D-glyceraldehyde-3-phosphate
DHAS:	Dihydroxyacetone synthase
DHP:	1,3-dihydroxypentan-2-one
DR:	D-ribose
DR5P:	D-ribose-5-phosphate
DX5P:	D-xylulose-5-phosphate
DXPS:	D-xylulose-5-phosphate synthase
E4P:	Erythrulose-4-phosphate
epPCR:	Error-prone polymerase chain reaction
GA:	Glycolaldehyde
GXC:	Glyoxylate carboligase
HPA:	Hydroxypyruvic acid
HPLC:	High performance liquid chromatography
IEMR:	Immobilised enzyme microreactor
IPDC:	Indolepyruvate decarboxylase
ISPR:	<i>In situ</i> product removal
MSA:	Multiple sequence alignment
NMR:	Nuclear magnetic resonance
PA:	Propionaldehyde
PCR:	Polymerase chain reaction
PDC:	Pyruvate decarboxylase
PFRD:	Pyruvate ferredoxin reductase
PhPDC:	Phenylpyruvate decarboxylase
PKL:	Phosphoketolase
PO:	Pyruvate oxidase
PPDC:	Phosphopyruvate decarboxylase
R5P:	Ribose-5-phosphate
SCA:	Statistical coupling analysis
SPDC:	Sulfoxyruvate decarboxylase
ThDP:	Thiamine diphosphate
TK:	Transketolase

1 Introduction

Transketolase (TK) is a key constitutive enzyme in metabolic regulation, providing a link between the pentose phosphate pathway and glycolysis through the production of 3 and 6 carbon sugars (glyceraldehyde-3-phosphate and fructose-6-phosphate respectively) (Figure 1.2 a and b) [1]. Found in the non-oxidative branch of the pentose phosphate pathway, TK catalyses the reversible transfer of two carbon ketol groups between several donor and acceptor substrates. In addition to supplying substrates for glycolysis, TK controls the supply of ribose-5-phosphate (R5P), essential for biosynthesis of nucleotides and nucleic acids, and catalyses the production of erythrulose-4-phosphate (E4P) which is utilised by microorganisms in the shikimate pathway for the biosynthesis of aromatic amino acids.

Transketolase was first purified from *Saccharomyces Cerevisiae* [2] and requires divalent cations and thiamine diphosphate (ThDP) for its activity [3]. There is a high level of sequence identity between the TK proteins of different organisms with many residues displaying complete invariance [4]. A second TK encoding gene was identified in *Escherichia Coli* in 1993 [5], this gene was named tktB to distinguish it from tktA. tktA and tktB share high sequence identity (74%) but tktA encodes the major TK activity in *E. coli*. All future references to *E. coli* TK refer to tktA encoded transketolase. In all structures solved to date, TK exists as a homodimer with two identical active sites positioned at the interface between the subunits.

The reaction catalysed by TK proceeds via a Ping Pong Bi Bi mechanism: two substrates are converted into two products as the ThDP cofactor within the enzyme active site shuttles between a free and a substrate modified intermediate state [6].

The Ping Pong Bi Bi model describes a specific type of Bi Bi mechanism in which substrates and products are bound and released sequentially and the enzyme shuttles between a free and a substrate modified intermediate state (Figure 1.1).

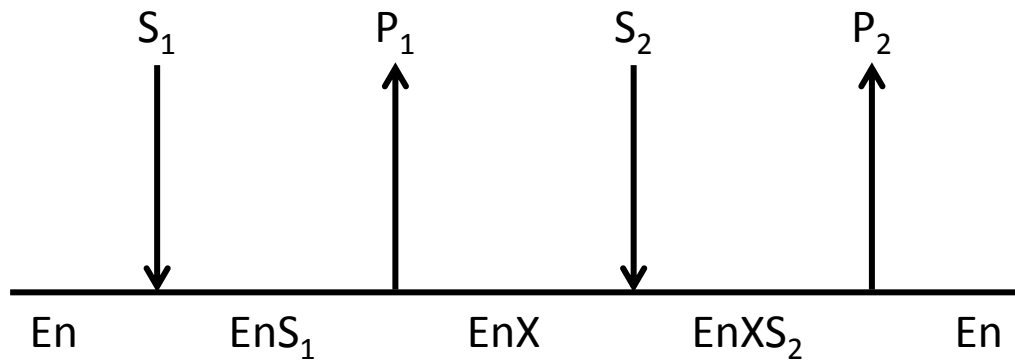


Figure 1.1 Kinetic Scheme of a Bi Bi Ping Pong Mechanism. In the Bi Bi Ping Pong model the enzyme shuttles between a free and a substrate modified intermediate state. In contrast to the standard Bi Bi model, substrates and products are bound and released sequentially in the Ping Pong model

Enzymatic thiamine catalysis is recognised as proceeding through two intermediate states: the ylide of ThDP in which the C2 proton of the thiazolium ring is abstracted, and the 2- α carbanion which is formed following nucleophilic attack by the ylide C2 on the donor substrate [6]. The α -carbanion is stabilised by the thiazolium ring which acts as an electron sink. Further stabilisation is provided by interconversion of the α -carbanion into a neutral enamine, creating a resonance hybrid. Following formation of the intermediate, the two carbon unit is transferred from the carbanion to the acceptor substrate forming a ketose with an extended carbon skeleton through nucleophilic attack. The ThDP in TK is bound in a V conformation which brings the 4-amino group of the pyrimidine ring into close proximity with the C2 carbon atom of the thiazolium ring, this conformation is essential for catalysis [7].

Broad substrate specificity, stereospecificity and stereoselectivity have made TK an attractive target for applications in organic synthesis [8]. The reversible reaction catalysed by TK *in vivo* has been utilised in a synthetic manner, but if the natural ketol donor is replaced with hydroxypyruvic acid (HPA), carbon dioxide is released as a by-product rendering the reaction irreversible and far more industrially useful (Figure 1.2 c). The industrial applicability of TK has been further adapted by engineering the protein sequence of the enzyme to improve attributes such as substrate specificity and enantioselectivity [9-11]. This thesis explores the potential of computational applications to direct and optimise the engineering of this enzyme towards even greater improvements.

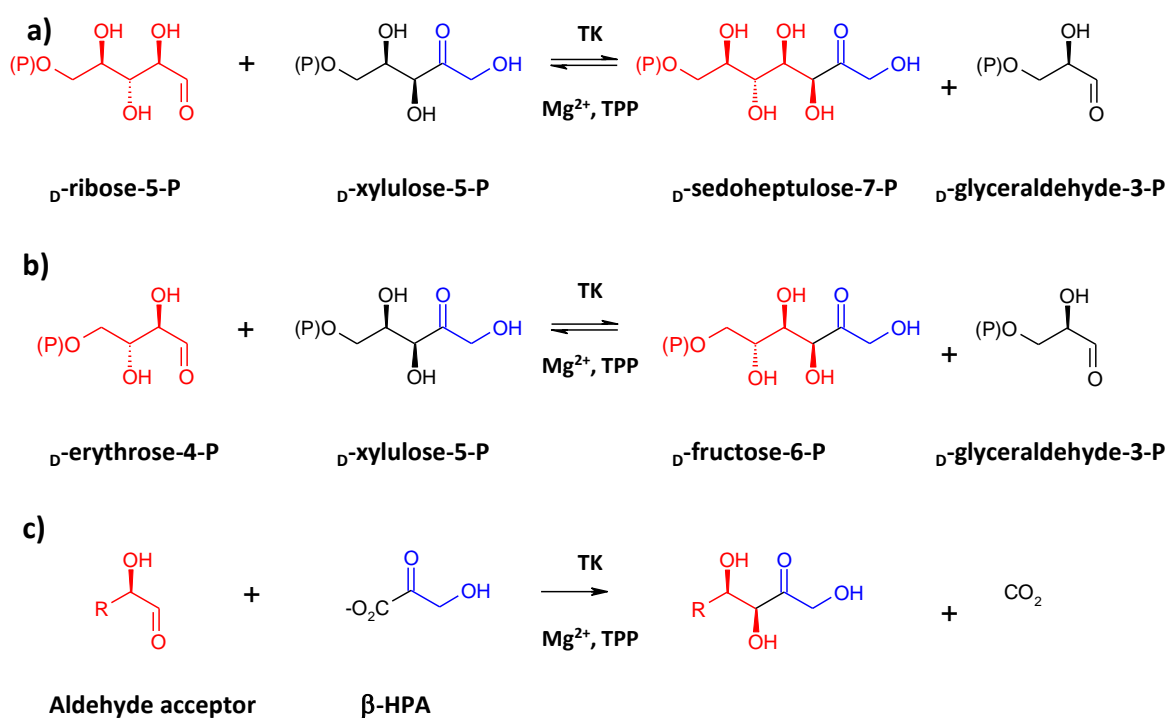


Figure 1.2 *In vivo* and *In vitro* reactions catalysed by transketolase. Reactions (a) and (b) occur *in vivo* in the non-oxidative branch of the pentose phosphate pathway and are reversible. (c) *In vitro* the ketol donor is generally replaced with β -HPA, rendering the reaction irreversible through elimination of CO_2 . Various aldehyde acceptors are accepted by TK but TK preferentially accepts α -hydroxylated aldehydes with the (R)-configuration.

1.1 *Transketolase structure and mechanism*

1.1.1 Transketolase structure

The first transketolase structure was solved for yeast TK in 1992 [12] this was refined to 2Å in 1994 [13] (1TRK). Since then, several other transketolase structures have been solved for TK. The *E. coli* TK structure was solved in 1994 [14] (1QGD), Maize TK in 2003 [15] (1ITZ) and *Leishmania Mexicana* TK in 2004 [16] (1R9J). Several further yeast TK structures have also been solved in the quest to refine our functional understanding of this catalyst. These include Apo TK [17], D-ethrythrose-mutants of TK [19] (1AYO) and several complexes of TK with ThDP analogues [20] (1TKA, 1TKB, 1TKC). More recently, *E. coli* TK structures were determined in covalent complexes with DX5P (2R8O) and DF6P (2R8P), and in non-covalent complex with DR5P (2R5N) [21].

The majority of detailed structural analysis and functional studies have been carried out on the yeast TK protein. Yeast and *E. coli* TK share a very high level of sequence identity and all homologous TK structures defined show near identical conformations of functional residue side chains. The vast majority of functional residues identified in yeast TK are 100% conserved in all TK proteins sequenced so far. We can therefore utilise the data and information gathered for yeast TK and apply it in our study of *E. coli* TK. Throughout this thesis, except where scientific evidence is based solely on yeast TK, numbering refers to *E. coli* TK and is based on the PDB structure 1QGD. Where yeast TK numbering is used, residues are underlined and numbering is based on the PDB structure 1TRK (in these situations *E. coli* numbering is also reported in brackets). Key functional residues are listed in

Table 1.1 together with the corresponding *E. coli* and yeast numbering for reference.

In all TK structures solved to date, TK is a homodimer consisting of two subunits each of 70-74 kDa (Figure 1.3 a). ThDP binds along with the metal ion cofactor at the interface between these two subunits. Each subunit of TK is made up of three domains, the N-terminal domain or the PP domain, the middle domain or Pyr domain, and the C-terminal domain (Figure 1.3 b). Each of the domains is of α/β type and the PP and Pyr domains are structurally similar when superimposed upon each other. The following structural analysis of transketolase is based on the structure for *E. coli* TK [14], but could equally be applied to the structure of yeast TK due to the high degree of structural equivalence.

The PP domain of *E. coli* includes residues 2-317 and comprises of a five stranded parallel β sheet with several helices on either side and some on top of the sheet. The α/β connection after the third strand contains a hairpin loop 187-191, this loop is involved in binding the cofactor and has been shown to be mobile in the apo-transketolase of yeast [17]. In the holo-enzyme of yeast TK, Asp 192 (190) and Ile 191 (189) are in contact with the metal ion and the cofactor. These interactions keep the hairpin in a closed conformation, enclosing the cofactor and shielding it from solvent [14].

The Pyr domain includes residues 318-527. This domain is made up of a parallel β -sheet of six strands. As mentioned above, the Pyr domain is structurally similar to the PP domain. The similarity between these two structural motifs is most pronounced in the last four α/β units of the two domains. Like the PP domain, the

Pyr domain forms interactions with the THDP cofactor through the loops at the carboxy ends of the β -sheet [14].

The C-terminal domain of TK consists of a mixed β -sheet with one antiparallel strand followed by four parallel strands. This domain is not involved in binding the ThDP cofactor and contributes less to the dimer interface interactions than do the other domains. Recent results [22] have demonstrated that the C-terminal domain is not essential for catalysis. The function of this domain remains unknown but it has been suggested that it may have a regulatory or a cellular localisation role.

The interface between the subunits of TK consists of a buried region representing approximately 18% of the accessible surface area of one free monomer [13]. Interactions between the two equivalent PP domains consist of tight packing interactions between the equivalent helices that link β -strands two and three and the equivalent helices that link strands three and four. The main interactions between the Pyr domains of each subunit are limited to the equivalent helices that link strands four and five of the Pyr β -sheet. This dimeric configuration positions the loops at the carboxy ends of the PP β -sheet facing the loops at the carboxy ends of the Pyr β -sheet of the other subunit. The region where these loops come together constructs the ThDP binding site and the active site of TK.

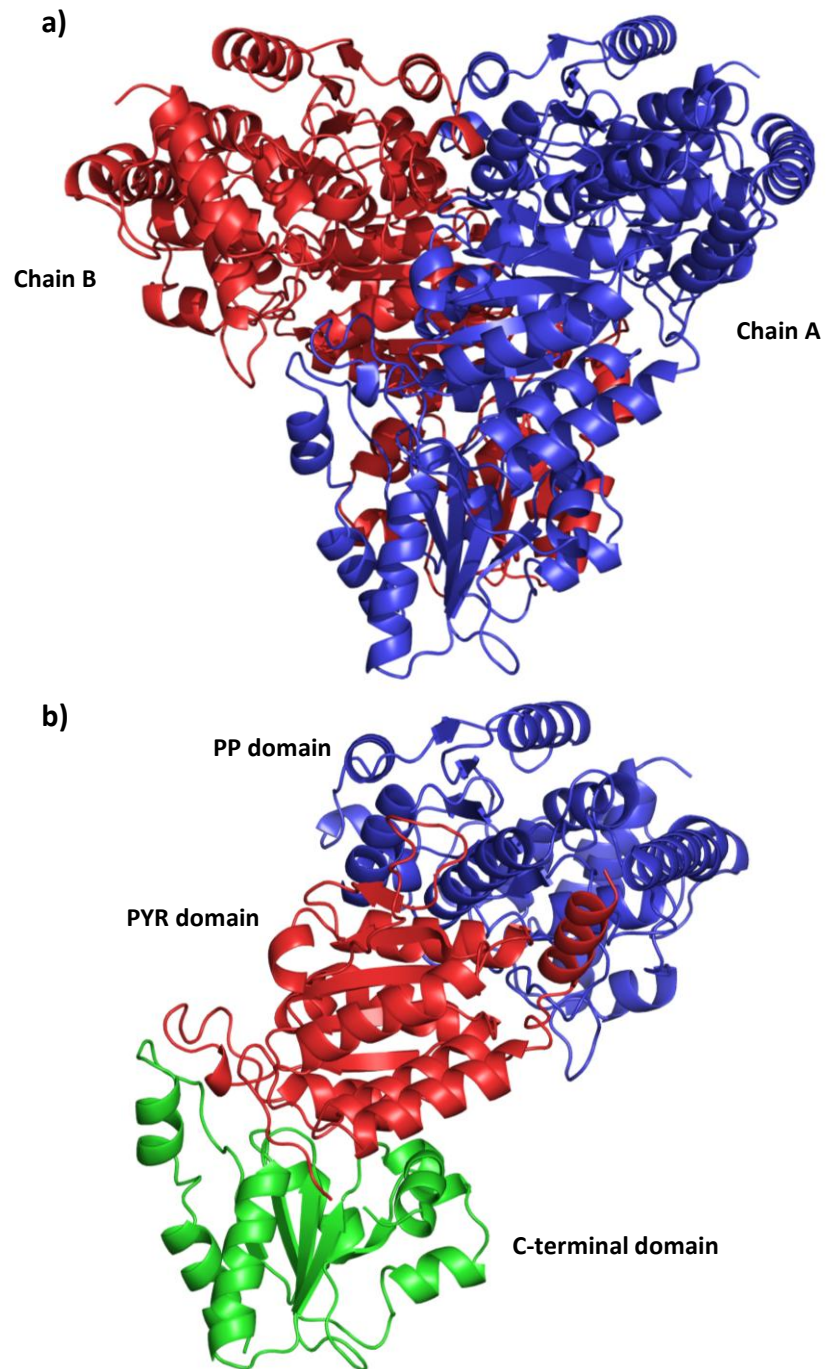


Figure 1.3 (a) *E. coli* transketolase homodimeric structure (1QGD) coloured by chain. (b) Chain A of *E. coli* transketolase coloured by domain.

Residue	TK Numbering		Proposed function
	<i>E. coli</i>	<i>S.cerevisiae</i>	
His	26	30	Interacts with substrates
His	66	69	Interacts with cofactor and substrates
His	100	103	Interacts with substrates
Gly	114	116	Interacts with pyrimidine ring (4'-NH ₂)
Leu	116	118	Interacts with pyrimidine ring (N3')
Asp	155	157	Metal ion ligand
Asn	185	187	Metal ion ligand
Ile	187	189	Metal ion ligand (main chain oxygen)
Ile	189	191	Contact with metal ion and cofactor
Asp	190	192	Contact with metal ion and cofactor
His	261	263	Interacts with cofactor and substrates
Arg	358	359	Interacts with phosphate of substrates
Leu	382	383	Interacts with cofactor thiazolium ring
Ser	385	386	Interacts with phosphate of substrates
Val	409	Ile 416	Interacts with cofactor thiazolium ring
Glu	411	418	Protonates N1' nitrogen pyrimidine ring
Phe	434	442	Interacts with pyrimidine ring
Phe	437	445	Interacts with pyrimidine ring
Tyr	440	448	Interacts with pyrimidine ring
His	461	469	Interacts with phosphate of substrates
Asp	469	477	Interacts with substrates
His	473	481	Interacts with substrates
Arg	520	528	Interacts with phosphate of substrates

Table 1.1 Key functional residues identified in yeast and *E. coli* transketolase with corresponding numbering. All residues are conserved apart from Val 109 in *E. coli* TK, the equivalent of which is Ile 416 in yeast TK.

1.1.2 Cofactor binding

ThDP binds in a deep cleft at the interface of the two TK subunits. Bound ThDP is totally isolated from the surrounding solvent apart from the reactive C2 carbon atom of the thiazolium ring. Unlike the structure of free ThDP, the bound cofactor is strained into a V-conformation. This brings the pyrimidine ring 4'-NH₂ group into close proximity with the reactive C2 carbon and contributes to the catalytic mechanism of all ThDP dependent enzymes (Figure 1.4).

Conserved *E. coli* TK residues His 66 and His 261 form hydrogen bonds with the diphosphate group of ThDP. Two oxygen atoms of diphosphate together with Asp

155, Asn 185 and the main chain oxygen of Ile 187 are ligands of Ca^{2+} and create further indirect interactions [14].

Whilst the diphosphate of ThDP forms interactions with one subunit of TK, the thiazolium and pyrimidine rings of the cofactor are bound in a cleft between the subunits. The thiazolium ring forms hydrophobic interactions with conserved residues Leu 116 and Ile 189. The C4 methyl group of the thiazolium ring interacts with the side chains of Leu 382 and Val 409 [14].

The pyrimidine ring is stacked with the ring system of Phe 437 and forms further interactions with conserved residues Phe 434 and Tyr 440. Main chain atoms of Gly 114 and Leu 116 form H-bonds with the pyrimidine ring 4'-NH₂ group and the N3' nitrogen atom respectively. The N1' nitrogen atom of the pyrimidine ring forms a H-bond with Glu 411 [14]. This interaction is very important in the molecular mechanism of enzymatic thiamine catalysis [23].

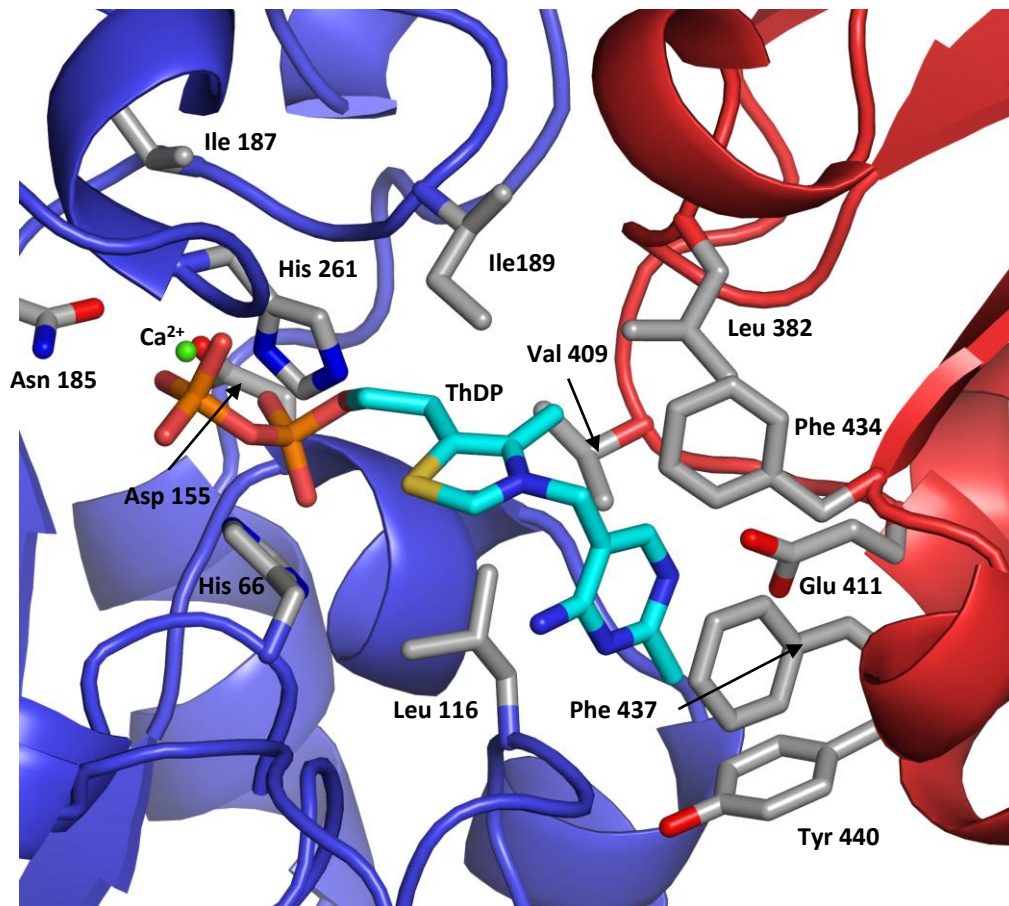


Figure 1.4 ThDP bound in one active site of *E. coli* TK with the Ca^{2+} metal ion. The active site is formed between the PP domain of chain A (blue) and the PYR domain of chain B (red). Interacting residues are labelled.

1.1.3 Substrate binding and recognition

The active site binding cleft of TK is a deep funnel leading towards the exposed reactive C2 of ThDP. Conserved loops make up the walls of this binding funnel. Two conserved arginine residues are positioned at the entrance to the binding funnel. The middle of the binding channel contains several conserved residues including Asp 469, Ser 385 and His 461. Towards the base of the binding cleft, near the thiazolium ring of ThDP, there is a cluster of histidine residues on one side and there are also several conserved hydrophobic residues [14].

The crystal structure of the acceptor substrate, DE4P, bound in yeast TK gives insight into the residues involved in substrate binding and recognition [18]. The phosphate group of the acceptor substrate forms interactions with several conserved residues near the entrance of the binding funnel. Arg 359 (358), Arg 528 (520), Ser 386 (385) and His 469 (461) form interactions with the substrate phosphate group. These interactions position the substrate in the binding channel in the correct orientation, the long side chains of the arginine residues may also provide some flexibility to allow the substrate to move towards the reactive C2 of ThDP [18]. Asp 477 (469) forms polar interactions with the C2-hydroxyl group of the substrate and the aldehyde oxygen atom is within H-bonding distance of His 30 (26) and His 263 (261) (Figure 1.5) [18].

In order to further elucidate the role played by the arginine and histidine residues at the entrance to the binding funnel, these residues have been subjected to site directed mutagenesis in yeast TK [18]. Substitution of residues Arg359 (358), Arg528 (520) and His469 (461) for alanine did not have a great effect on catalytic activity (residual catalytic activities were 31%, 17% and 77% respectively) but did increase K_m values for donor substrate, and in particular acceptor substrates, significantly. Consistent with the crystal structure of DE4P bound in the TK active site, these results support a role for these residues in binding the phosphate group of substrates.

The pattern of H-bonds formed by Asp 477 (469), His 30 (26) and His 263 (261) with the acceptor substrate is consistent with the enantiosensitivity TK displays towards D-threo configured donor substrates. Inversion of the stereocentres in the favoured

configuration would disrupt this H-bond network and reduce enzyme affinity for substrate. The potential of forming a H-bond with Asp 477 (469) also explains the preference for α -hydroxylated acceptor substrates. Replacement of Asp 477 (469) with alanine in yeast TK resulted in an enzyme with severely impaired catalytic activity [24]. K_{cat}/K_m for this variant is reduced relative to wild type TK for D- α -hydroxyaldehydes (DE4P, DR5P) and this reduction is equivalent to the reduction in K_{cat}/K_m for the wild type enzyme with 2-deoxyaldoses or L- α -hydroxyaldehydes [24].

Yeast TK residues His 30 (26) and His 263 (261) have also been mutated to alanine [19]. These residues are within H-bonding distance to the carbonyl oxygen of the acceptor substrate and their mutation to alanine has a large effect on k_{cat} [19].

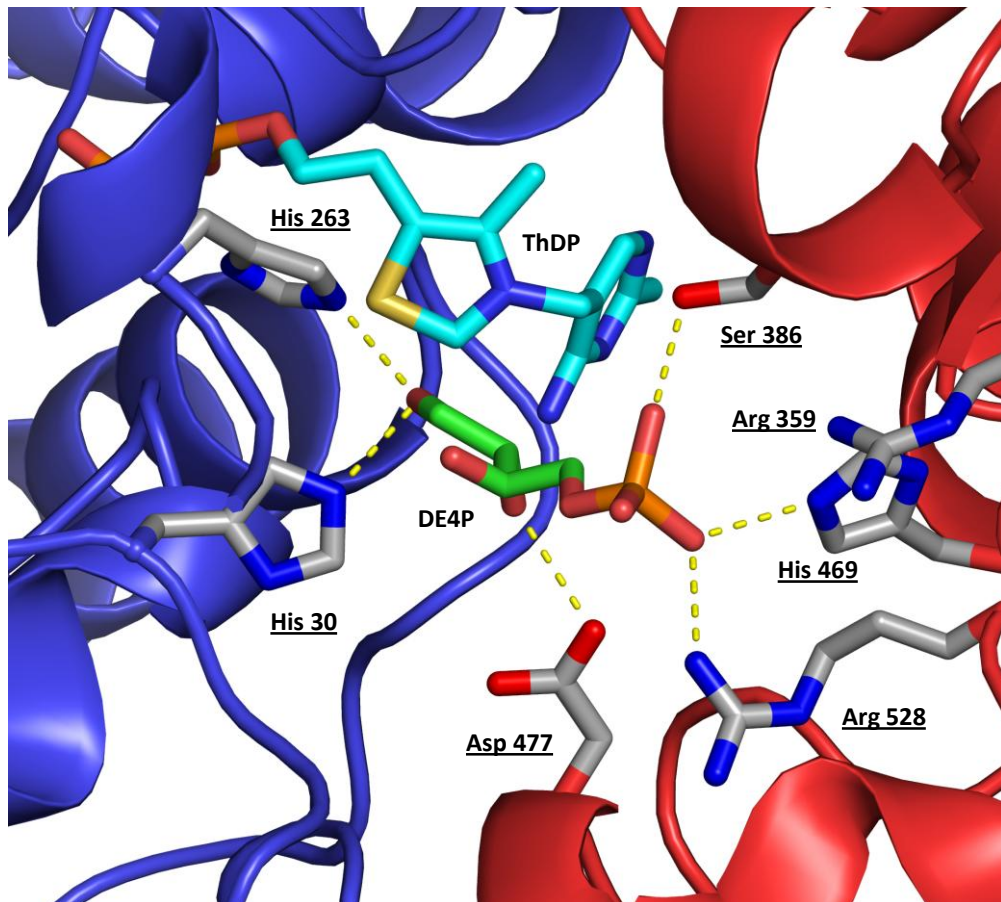


Figure 1.5 DE4P bound by yeast TK. The active site is formed at the interface of the PP domain of chain A (blue) and the PYR domain of chain B (red). Residues from both chains make key interactions with DE4P. Interacting residues are labelled with yeast TK numbering. Hydrogen bonds are displayed by yellow dashed lines.

A cluster of histidine residues is located towards the base of the binding funnel close to the reactive C2 of ThDP. Based on the structures of TK and acceptor substrate (DE4P) bound TK, histidine residues His 69 (66) and His 103 (100) were predicted to form H-bonds with the C1-hydroxyl group of the donor sugar phosphate [18]. In support of this role, replacement of His 69 (66) or His 103 (100) with alanine had little effect on the K_m values for acceptor substrates but significantly increased those of donor substrates [19, 25]. These mutants also displayed a significant decrease in catalytic activity. Although hydroxypyruvate is a donor substrate for TK, pyruvate is not. The recognition of the C1-hydroxyl group of the donor by these two histidine residues might explain this molecular selectivity. In

a crystal structure of the covalent enamine intermediate bound in yeast TK [7], His 103 (100) formed a H-bond with the β -hydroxyl oxygen of the intermediate. The β -hydroxyl oxygen also interacted with His 69 (66), indirectly through a water molecule. These interactions further support a role for His 69 (66) and His 103 (100) in discrimination between hydroxypyruvate and pyruvate.

Recent structures determined of covalent intermediates in *E. coli* TK support the structural studies above and the conserved function of residues between yeast and *E. coli* TK [21]. DX5P and DF6P covalent intermediates adopted very similar extended conformations in the active site of *E. coli* TK, forming at least 11 well defined hydrogen bonds with the side chains of active site residues. The C1-hydroxyl and C2-hydroxyl groups of both substrates formed interactions with His 473 and the 4'-amino group of the ThDP pyrimidine ring. The C1-hydroxyl group also formed a hydrogen bond with His 100. The C3-hydroxyl groups interact with the two histidine residues His 261 and His 26 and the C4-hydroxyl group interacts with Asp 469 and His 26 (Figure 1.6). Phosphate interactions were mediated by residues His 461, Ser 385 and Arg 358 with the phosphate group of DF6P slightly closer to these residues due to the longer carbon chain. The additional C5-hydroxyl group of DF6P interacts with Ser 385.

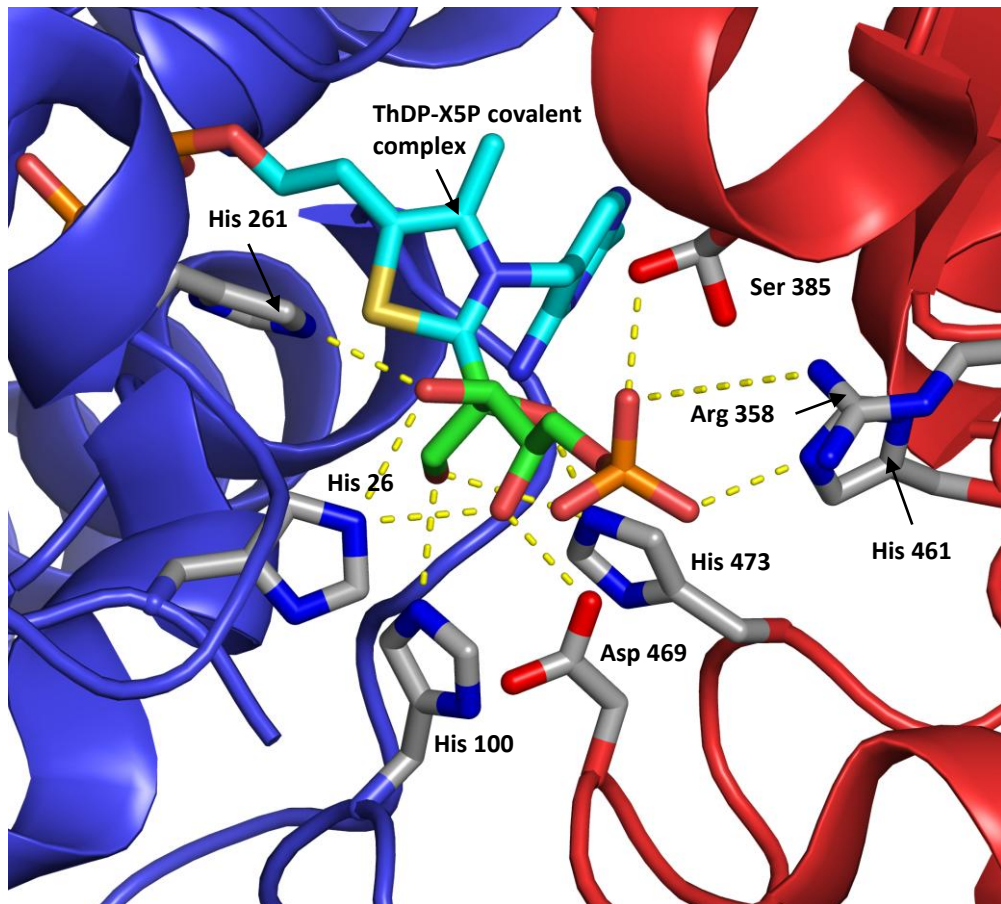


Figure 1.6 Structure of the covalent complex formed between X5P and ThDP in the active site of *E. coli* TK. The PP domain of chain A is coloured blue and the PYR domain of chain B is coloured red. Residues forming interactions with the complex are labelled. Hydrogen bonds are displayed by yellow dashed lines.

1.1.4 Molecular mechanism of Transketolase

Transketolase catalyses the transfer of a two carbon unit from a ketose donor to an aldose acceptor, the reaction proceeds through two major steps. In the first step, the donor substrate is cleaved to produce an aldose and a covalent intermediate, ThDP α -carbanion. The second step is initiated by nucleophilic attack by the α -carbanion on the acceptor substrate, resulting in a ketose product with an extended carbon skeleton. This reaction mechanism, described in further detail below, was proposed by Schneider and Lindqvist (1993) [6] (Figure 1.1 Figure 1.7).

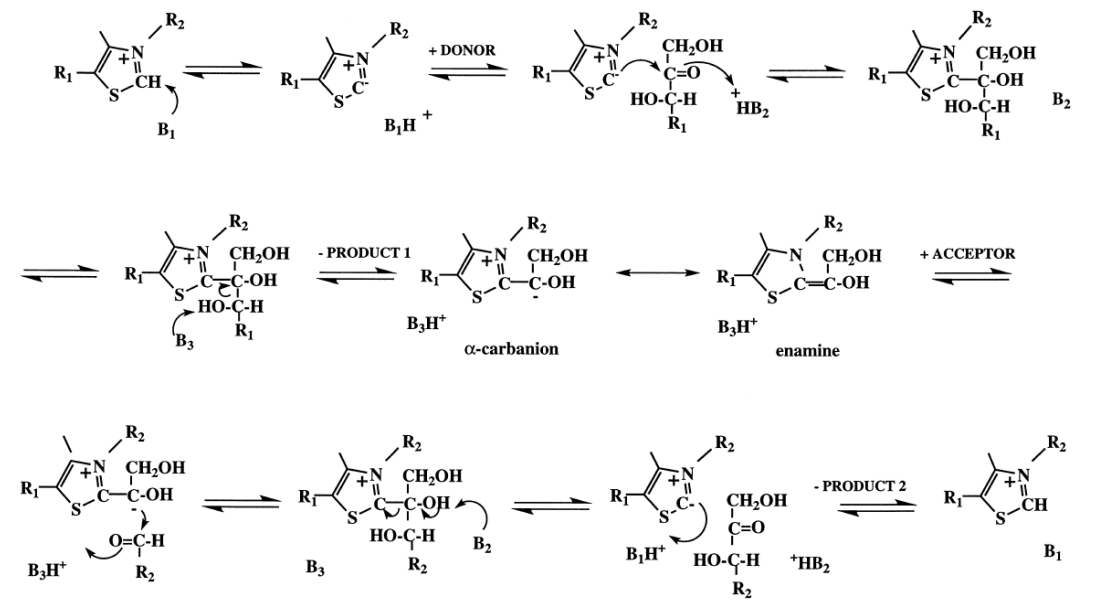


Figure 1.7 Reaction mechanism of transketolase, reproduced from *Biochemica et Biophysica Acta* (1385 387-398) with permission from author and publisher. B₁ represents the 4'-imino group of the pyrimidine ring; B₂ represents either His 473 or the 4'-imino group of ThDP; B₃ represents either His 26 or His 261.

Prior to the first step in catalysis, the C2 carbon of the thiazolium ring must be deprotonated in order to create an ylide that can attack the donor substrate. Evidence suggests that the deprotonation of C2 is catalysed by the cofactor itself [23]. The transketolase molecule contributes to cofactor deprotonation by maintaining the V-conformation of ThDP, which brings the 4'-NH₂ group into close proximity with the C2 carbon, and through protonation of the N1' nitrogen of the pyrimidine ring. The N1' nitrogen is protonated by a H-bond with Glu 411, this interaction alters the pK_a of the 4'-NH₂ group and leads to the production of a 4'-imino group which is sufficiently basic to deprotonate the C2 carbon of the thiazolium ring [6].

Once the C2 carbon has been deprotonated, the carbanion formed attacks the carbonyl oxygen of the donor substrate to create a high energy intermediate. During covalent bond formation between ThDP and the donor substrate, a proton

donor is required to stabilise the negative charge forming at the carbonyl oxygen. His 473 and the charged 4'-imino group of ThDP are possible proton donors at this step in the reaction. While site directed mutagenesis of the His 481 (His 473) in yeast TK suggested a role in transition state stabilisation, the lack of conservation of this histidine residue across other TK enzymes suggests that the 4'-imino group may be responsible for the majority of transition state stabilisation.

The final steps in TK catalysis require an acid/base catalyst that can deprotonate the hydroxyl group of the substrate at C3, catalysing the cleavage that produces the α -carbanion intermediate, and act as a proton donor to the carbonyl oxygen of the acceptor substrate as it is attacked by the α -carbanion. Both His 26 and His 261 are within H-bonding distance from the C3-hydroxyl group of the reaction intermediate and the carbonyl oxygen of the acceptor substrate. Replacement in yeast of either of these residues by alanine severely impairs catalytic activity and they may act together as the acid/base catalyst in the reaction [19].

Crystal structures of the covalent high energy intermediates formed with DX5P and DF6P in *E. coli* TK reveal a conformation in which the newly formed C2-C α bond is out-of-plane with the thiazolium ring by 25-30° (Figure 1.8) [21]. This strained conformation will be relieved upon product elimination conceivably providing the driving force for the reaction. Density functional theory (DFT) calculations supported the above out-of-plane conformation, demonstrating that this conformation is energetically favourable relative to a model with a co-planar C2-C α bond [21]. No structural rearrangements were seen in the active site following intermediate formation, suggesting that the active site is poised for catalysis such

that the substrate binding energy and the enthalpic energy gain following covalent bond formation between ThDP and the donor substrate can be channelled directly into the formation of the high energy, strained intermediate.

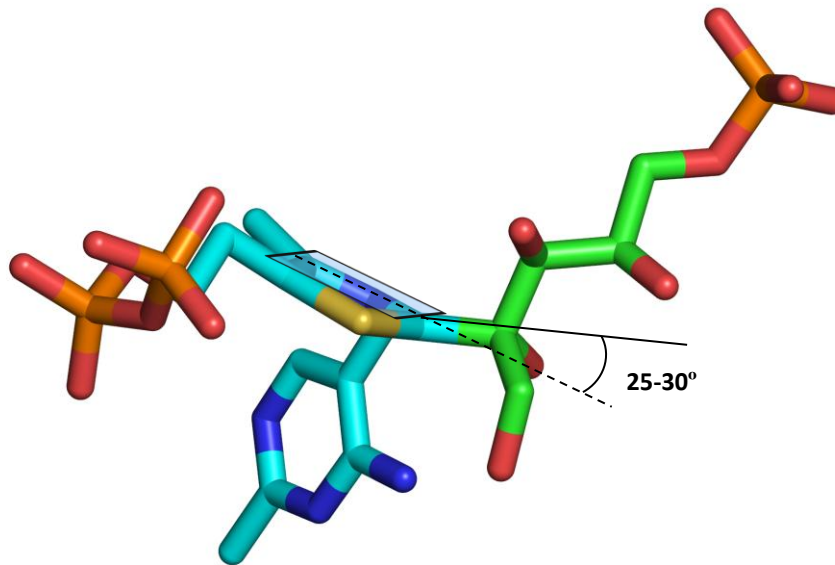


Figure 1.8 Covalent complex of X5P and ThDP isolated in the active site of *E. coli* TK. The strained angle of the out-of-plane C2-C α bond is displayed relative to the thiazolium ring of ThDP.

1.2 *Transketolase as a biocatalyst*

In vivo, transketolase catalyses the transfer of a 2-carbon ketol unit from D-xylulose-5-phosphate to D-ribose-5-phosphate, generating D-sedulose-7-phosphate and D-glyceraldehyde-3-phosphate. The carbon-carbon bond formation catalysed by TK is both stereospecific and stereoselective, making this a very attractive enzyme for the industrial production of complex organic structures. Unlike traditional chemical methods for organic synthesis, enzymatic catalysis does not require complex protection and de-protection steps, additional advantages include greater chiral control and milder reaction conditions.

The reversible reaction catalysed by TK *in vivo* has been utilised for industrial applications but the usefulness of the biocatalyst can be enhanced by replacing the D-xylulose-5-phosphate (X5P) ketol donor with β -hydroxypyruvate (HPA). Use of HPA results in the elimination of CO₂ thereby rendering the reaction irreversible [26].

In addition to varying the ketol donor, much work has been carried out exploring the substrate specificity of TK for aldehyde acceptors. Although TK favours aldehydes containing an α -hydroxylated group in an (R)-configuration [27], specificity with respect to the aldehyde is relatively broad with both non-phosphorylated and phosphorylated aldehydes of varying sizes being accepted [28]. Steric hindrance appears to impact on relative activity with cyclic aldehydes and aldehydes containing bulky groups displaying lower activities.

The broad nature of aldehyde substrate specificity is advantageous for the application of TK in organic synthesis as it provides flexibility in the nature of structures that can be produced. The broad specificity also provides a good starting point for engineering TK to improve activity on non-natural substrates.

Early work in the development of TK as a biocatalyst utilised commercially available *S. cerevisiae* TK, and spinach TK extracted from spinach leaves. These sources are readily available but do not offer the scale required for an industrial process. In 1993, Hobbs *et al* developed an efficient and reliable source of TK by introducing the previously cloned *E. coli* TK gene fragment into a high copy number plasmid and transforming this into *E. coli* [29]. The resulting transformants overexpressed TK with superior specific activity to that obtained previously from *E. coli* TK. Evaluation

of the substrate specificity of *E. coli* TK demonstrated a similar profile to yeast TK and the same preference for α -hydroxylated aldehydes with an (R)-configuration [29]. An additional benefit of *E. coli* TK was its increased activity with HPA (60 U/mg protein) relative to yeast (8.6 U/mg protein) or spinach (2 U/mg protein) [1, 28, 30].

1.2.1 Use of Transketolase in enzymatic syntheses

Transketolase has been used successfully in various organic syntheses of both natural and unnatural complex, chiral compounds. The compounds produced using transketolase are expensive and would require complex multi-step synthesis if produced using traditional chemical methods. Some of the compounds synthesised by transketolase cannot be produced chemically.

In an early example, D-[1,2- $^{13}\text{C}_2$]-xylulose was produced from [2,3- $^{13}\text{C}_2$]-hydroxypyruvate and D-glyceraldehyde using spinach transketolase [31]. The isotopic labelling of sugars is useful for the study of metabolism and structure. Introduction of two adjacent ^{13}C labels is particularly useful due to ^{13}C - ^{13}C coupling. Transketolase has also been utilised to produce glycosidase inhibitors fagomine [32] and 1,4-dideoxy-1,4-imino-D-arabinitol [33], which have applications as agrochemicals and therapeutic agents.

Commercially available TK was used together with chemical steps to produce α -exo brevicamycin, a beetle pheromone with applications in pest control. In this example TK was used to convert a racemic mixture of 2-hydroxy butyraldehyde and HPA into the key tri-hydroxy ketone intermediate in the process [34].

Production of expensive food additives has also utilised TK. Spinach TK was used to produce 6-deoxy-L-sorbose, a precursor for synthetic furaneol, an aromatic product with a caramel like flavour. In the above synthesis, serine glyoxylate amino transferase was used to prepare HPA and 4-deoxy-L-threose was obtained by microbial isomerisation of 4-deoxy-L-erythrulose [35].

Multi-enzyme approaches have been successfully applied to the production of sugars 4-deoxy-D-fructose 6-phosphate and D-xylulose 5-phosphate. In the first example, 4-deoxy-D-fructose 6-phosphate was synthesised in a process including four steps and two enzymatic reactions. Epoxide hydrolase was first used to obtain (3S)-1,1-Diethoxy-3,4-epoxybutane which was subsequently opened by inorganic phosphate to produce 2-deoxy-D-erythrose 4-phosphate. This aldehyde was then reacted with L-erythrulose in the presence of yeast TK to introduce the second chiral centre [36].

In the second example, D-xylulose-5-phosphate, a valuable substrate required for enzymatic assays, was prepared in gram quantities through a one-pot procedure incorporating fructose 1,6 bisphosphate aldolase followed by *E. coli* TK [37]. The aldolase was initially used to produce D-glyceraldehyde-3-phosphate and dihydroxyacetone phosphate (DHAP) through retro-aldolization of D-fructose-1,6-bisphosphate. The D-glyceraldehyde-3-phosphate produced was then coupled with HPA by *E. coli* TK to produce X5P. Zimmermann *et al* also incorporated triosephosphate isomerase (TPI) to equilibrate the two products of the retro-aldolization thereby increasing the overall yield. Thanks to recent advances in

synthesis [38], DHAP is now available as a starting point, opening up a new route to the synthesis of X5P using just TPI and TK [39].

Finally, transketolase was utilised to catalyse the key asymmetric step in a process to produce N-hydroxypyrrolidine, a glucosidase inhibitor. In this synthesis TK was used to couple (+)-3-O-benzylglyceraldehyde with HPA to yield 5-O-benzyl-D-xylulose in multigram quantities [40].

1.2.2 Optimisation of process for industrial application

To utilise TK in large scale industrial enzymatic syntheses, limitations in the process must be identified and overcome to ensure product purity and maximise yield and efficiency. Limitations in TK biocatalysis include stability of the enzyme (reactive α -hydroxy aldehydes inactivate TK [41]), and product inhibition. More general process options also need to be optimised during process development to ensure maximum yield, minimum cost and scalability.

One strategy to overcome the problem of substrate deactivation of TK is to minimise substrate concentration by carrying out the reaction in an enzyme membrane reactor. Applied to the production of L-erythrulose from GA and HPA this strategy was able to increase the half-life of TK from 5.6 hours (repetitive batch reactor) to 106 hours (enzyme membrane reactor). This improvement in stability resulted in an increase in space time yield from 28 g/L/d to 45 g/L/d [41].

An alternative approach to increasing enzyme stability is to immobilise the enzyme on a support. GA is believed to deactivate TK by forming Schiff bases with amino acid side chains on the surface of the enzyme. Formation of Schiff bases can alter

the three dimensional structure of proteins. Stabilisation of TK by immobilisation can be explained by the prevention of the formation of Schiff bases, or by rigidifying the three dimensional structure of TK. Applied to the production of L-erythrulose from GA and HPA, immobilisation of TK on commercially available supports (Amberlite XAD-7 and Eupergit C) increased the stability of TK by 80- and 100-fold respectively. Immobilisation was unable to prevent inactivation by oxidation but this could be reduced by the inclusion of a stabilising solute such as mercaptoethanol [42]. Immobilisation confers additional benefits on process productivity: allowing the enzyme to be retained in the bioreactor, extracted from the product stream, and/or reused.

In situ product removal (ISPR) has been explored as a means to overcome limitations introduced by product inhibition. L-erythrulose was successfully removed using an immobilised phenylboronate resin, however there were also considerable levels of nonspecific substrate binding to the resin which reduced the actual yield. To overcome the problem of nonspecific binding, a fed batch system was utilised. The fed batch mode gave the added benefit of reducing the deactivation of TK by GA. The rate of deactivation of TK by substrate is much higher than by the synthesised product [41], therefore in practice the reduction in aldehyde toxicity overcame the benefits of product removal and negated the need for ISPR [43].

Work has also been carried out to explore more general process development strategies to optimise yield, cost and scalability. The synthesis of X5P using TK and TPI has been utilised as a model reaction to investigate the potential of

semiquantitative process screening, to speed up process development for multienzyme biocatalytic processes. This approach has the potential to reduce the number of process options that need to be screened, applied to the synthesis of X5P the strategy successfully identified new biocatalytic routes and processes for further investigation [44].

As an alternative to reducing the number of process options to screen, high-throughput microwell based methods can be used to screen multiple process options in parallel. For TK process characterisation a more efficient alternative to microwells has recently been developed in the form of an immobilised enzyme microreactor (IEMR). The microreactor developed is composed of a 25 cm long fused silica capillary with a 200 μm internal diameter. His-tagged TK is reversibly immobilised inside the capillary via Ni-NTA linkages. His-tagged TK is expected to be kinetically identical to un-tagged protein based on the location of the His tag and on previous kinetic characterisation. The reactor can be operated in stop flow or continuous flow mode and product is analysed by HPLC. For high throughput screening of different process options or different enzyme variants the IEMR has several advantages over traditional microwell approaches, these include reduction in reactant volume, enhanced productivity, reduced reaction time and increased reusability [45].

1.2.3 Transketolase enzyme engineering

Although TK displays broad substrate specificity, non-natural aldehyde substrates are converted at far lower rates than the phosphorylated, α -hydroxylated natural substrates. The production of non-phosphorylated chiral products is one of the

major advantages of TK over other enzymes such as aldolases, it would therefore be advantageous to increase the activity of TK on these non-natural substrates.

In addition to process engineering, enzyme engineering can be applied to TK to further optimise the biocatalyst for industrial applications. The good structural and mechanistic understanding of TK and its broad substrate specificity make the wild type enzyme a good candidate for enzyme engineering. However, the two-substrate mechanism of TK complicates engineering as any change can result in positive or detrimental effects to the binding of either substrate. In recent years TK has been engineered to improve its activity on non-phosphorylated and non-hydroxylated substrates. The enzyme has also been engineered to improve or reverse its stereoselectivity with these non-natural substrates.

Early work on TK engineering focussed on improving activity in the model reaction of GA and HPA to produce L-erythrulose. Variants with up to 5-fold improvements in activity against GA were identified by screening a library of single point mutants generated by saturation mutagenesis of nineteen positions. Residues were selected for mutagenesis based on structural and phylogenetic criteria. Two sets were included: residues within 4 Å of bound substrates, and phylogenetically varied residues within 10 Å of TPP. Following the screen, twelve variants were identified with enhanced specific activity on GA relative to wild type TK. Six of the nineteen residue libraries yielded variants with improved activity.

The greatest improvements in activity against GA were associated with variations at residues His 461, Arg 520 and Ala 29. His 461 and Arg 520 interact with the phosphate group of natural substrates, mutation of these residues could improve

GA accessibility to the active site by removing a charged group or by reducing the steric bulk around the entrance to the active site. The third position identified, Ala 29, is harder to rationalise as it is in the active site second shell and is in direct contact with the terminal phosphate group of TPP. Overall, residues with high sequence entropy were more likely to confer enhanced activity on GA following mutagenesis. The important exceptions to this rule were the three residues known to interact with the phosphate group of natural substrates. Saturation libraries of His 461, Arg 358 and Arg 520 all yielded at least one mutant with increased specific activity on GA despite the low sequence entropy at these positions. Interestingly the most successful mutations were non-natural variants [9].

In addition to increasing activity on non-phosphorylated substrates it would be advantageous to extend the applicability of TK by enhancing activity on aliphatic, non-hydroxylated aldehydes. Typically, activity of wild type TK on these substrates is only 5%-35% of that for α -hydroxylated aldehydes such as GA. The active site variant libraries described above were screened for TK catalysed production of 1,3-dihydroxypentan-2-one (DHP) using the aldehyde substrate propionaldehyde (PA).

Twenty-six distinct mutants were identified with increased specific activity on PA relative to wild type. These variants represented eight of the nineteen residue libraries. Five of the eight libraries yielding enhanced activity on PA had been previously identified through screening for enhanced activity on GA. These included the group of residues which interacts with the phosphate group of natural substrates, Arg 358, His 461, and Arg 520. The phylogenetically variant residues in the active site second shell, Ala 29 and Asp 259, were also identified. A third group

of residues not previously identified yielded variants with the greatest increase in activity on PA. This group was made up of conserved residues His 26, Asp 469 and His 100. These residues form a pocket in the active site and directly interact with the hydroxyl group at C2 of erythrose-4-phosphate. The greatest increase in activity on PA was observed with D469T which demonstrated a 5-fold improvement in specific activity over wild type [10].

Variation of phylogenetically variant residues or those interacting with the phosphate group of natural substrates led to enhanced activity on both GA and PA [10, 46]. In contrast, mutation of hydroxyl interacting residues produced variants with enhanced substrate specificity for PA over GA. The D469Y mutant displayed the greatest substrate specificity, with a 64-fold higher activity on PA relative to GA [10].

Wild type TK catalyses the production of L-erythrulose from GA and HPA with 95% ee, but 3S-DHP is only produced in 58% ee using PA and HPA. To identify variants with increased stereoselectivity for the production of 3S-DHP, Smith *et al* screened the three variant libraries of residues that interact with the hydroxyl group of natural substrates, His 26, Asp 469 and His 100. Significant increases in stereoselectivity were obtained with the D469E variant which produced 3S-DHP in 90% ee. Interestingly the majority of His 26 variants lead to the formation of 3R-DHP and H26Y produced 3R-DHP in 88% ee [11].

This work was extended to establish the enantioselectivity of wild type TK and selected variants on linear aliphatic aldehydes of increasing length and cyclic aldehydes. Compared to wild type TK, D469E TK produced products in greater yields

for the longer chain and cyclic aliphatic aldehydes although yield did decrease as chain length increased. Although yields were lower, D469E displayed enhanced stereoselectivity with longer chain aldehydes and over 99% ee for cyclic aldehydes cyclopropanecarbaldehyde and cyclopentanecarbaldehyde. H26Y gave product with lower yields but the reversal of ee was maintained across all the aldehydes tested, with the highest ee noted for butanal at 92% [47].

Although work to date has been limited to single point variants and residues in close proximity to the active site, considerable success has been achieved in the engineering of TK. Activity on both non-phosphorylated and non-hydroxylated aldehydes has been improved by 5-fold relative to wild type, and enantioselectivity has also been improved for the non-natural substrates. Surprisingly, a single point mutation was also able to reverse the enantioselectivity of the enzyme. The variants identified further expand the potential applications of TK for industrial synthesis applications. However, there is significant potential to further engineer TK to the point where non-natural substrates convert at the same rate and with the same exquisite stereoselectivity as the natural substrates.

1.3 Current methods in enzyme engineering

Directed evolution can be used to engineer the catalytic properties of enzymes and has been used successfully to modify properties such as specificity, selectivity and enantioselectivity. The general process of directed evolution can be broken down into three steps; generation of a variant library, screening for a desired property and selection of positive variants for the next cycle. Although most directed evolution experiments follow this overall process, there are many different ways to carry out each step. The main limitation in directed evolution is the size of the library that can be screened. Typically, libraries of 10^3 to 10^6 variants can be screened using high-throughput techniques and robotic equipment. In some cases a desired attribute can be linked to a growth advantage in bacteria, allowing significantly larger libraries (up to 10^{9-13}) to be screened. Unfortunately for most directed evolution experiments this cannot be applied and we are limited to libraries containing thousands rather than millions of variants. In some situations we are limited to even smaller libraries, for example where a suitable colorimetric or fluorogenic assay is not available.

The limitation in the size of libraries means we can only sample a tiny fraction of the possible sequence space. A protein with 300 amino acids has 20^{300} distinct possible sequences so even with a large screening effort of 10^6 variants we can only sample a minute fraction of the potential sequence space. The strategy chosen to produce the variant library must therefore be selected very carefully in order to target the best section of sequence space for the desired property.

1.3.1 Error-prone polymerase chain reaction

The most commonly used method to produce a library is error-prone polymerase chain reaction (epPCR). Error-prone PCR involves the introduction of random copying errors through imperfect reaction conditions (e.g. by adding Mn^{2+} or Mg^{2+} to the PCR reaction mixture) usually with the aim of introducing approximately one mutation each time the gene is copied [48]. Such a technique applied to a 300 amino acid protein will produce a library of 5,700 potential variants, easily screenable even accounting for the oversampling required. In such examples it is usually possible to find a variant with improved properties although several cycles may be required as multiple variations are usually needed to generate the required level of improvement in a particular property. Previous experiments have demonstrated that on average 30% - 50% of random mutations are deleterious, 50% - 70% are neutral, and just 0.01% - 0.5% are beneficial [49]. In our example of a 300 amino acid protein we could expect to find 1 – 30 beneficial mutations in the 5,700 variant library.

Using epPCR the full length of the protein sequence can be probed; but the cycling nature of directed evolution, and the introduction of one change at a time, means that an evolutionary trajectory is entered once the first variant has been selected. This trajectory theoretically limits the potential optimal sequence that can be achieved. If all the individual variations that constitute an improved activity are independently beneficial and additive in nature then this is not a problem because all trajectories should arrive at the same optimal solution. Unfortunately, variation

at one site in a protein sequence often requires the simultaneous variation at another site to be beneficial.

Following several rounds of selection directed evolution experiments often plateau with further rounds failing to generate improvements. This has been suggested to represent protein sequences becoming stuck on fitness peaks in sequence space. Usually this can be overcome by one or two rounds of selection for stabilising mutations, it is hypothesised that the accumulation of beneficial mutations gradually reduces stability until no further mutations can be tolerated [49]; introduction of stabilising mutations allows the protein to tolerate further mutations. We can envisage a situation where an incredibly beneficial variant is so destabilising that it cannot be introduced without the simultaneous introduction of a specific stabilising variation, which by itself may have no impact on the stability of the wild type sequence. In such a situation, it would be impossible to identify the beneficial variant even following pre-stabilisation of the protein fold.

1.3.2 Saturation mutagenesis

Another common method of library generation is saturation mutagenesis. This approach requires the selection of one, or a small number of sites in the protein sequence; randomised codons are then used in PCR primers to generate all the possible variants at the individual sites. Single site libraries created in this manner are small enough to allow the use of conventional GC or HPLC methods for screening and therefore allow selection for properties that are intractable with high throughput techniques. However, as more sites are added to the library, the size quickly becomes intractable to screening. Complete saturation at three sites would

create 7,999 potential variant sequences (including 57 single point mutants and 1,083 double mutants), four sites results in approximately 1.6×10^5 possible sequences, representing the approximate upper limit even for high throughput screening techniques. Such a mutagenesis strategy generally requires structural information and a good understanding of the functionally important residues. Sites are normally selected based on their proximity to the active site and on their hypothesised role in specific elements of the reaction being catalysed.

Saturation mutagenesis overcomes some of the issues which limit epPCR, allowing multiple amino acids to be varied simultaneously means we can identify combinations of variations that may be deleterious or neutral in isolation. Reetz *et al* identified multiple beneficial variants by saturated mutagenesis of three positions in the epoxide hydrolase enzyme from *Aspergillus niger* (ANEH), variants were screened for activity on a new substrate. 5000 variants were screened resulting in 26 unique hits. Twenty two (85%) of the resulting hits were triple mutants and four (15%) were double mutants; none were single mutants [50]. The authors did not create double and triple mutant cycles to investigate whether the multiple beneficial variants were synergistic or additive in nature but it is interesting that no single variants were identified by the screen.

1.3.3 Limitations in enzyme engineering

Many variations and combinations of the above methods have been used in directed evolution experiments. A further method, DNA shuffling, represents another technique used for library generation but this is not covered in detail here. For each enzyme engineering experiment a choice of technique is made in an effort

to maximise the proportion of beneficial variants in the limited library size. However, no one method has proved universally superior to the others and each has its own benefits and limitations.

In examples where mutagenesis is focused on residues likely to confer beneficial properties, positions are usually selected based on their proximity to the active site. Where an attempt is made to introduce multiple mutations, such sites are often selected based on their proximity to each other [51]. Such a simple selection strategy illustrates the limitations of current enzyme engineering approaches. Enzyme properties such as substrate specificity are sometimes determined by sites distant from the active site. Hedstrom *et al* successfully engineered Trypsin serine protease to accept Chymotrypsin substrates, but to do so required mutation of both the binding pocket and distributed surface loops which don't interact directly with the substrate. A particular residue (172) was identified as a determinant of substrate specificity through interaction with both the binding pocket residues and surface loops [52, 53]. Using proximity to the active site to select residues for mutagenesis, it would not be possible to engineer a Trypsin enzyme to accept Chymotrypsin substrates.

Modern computational techniques such as structural modelling and statistical coupling analysis provide a new resource to refine our choice of enzyme residues to target in enzyme engineering experiments. In addition to supporting library design, structural modelling can be used to rationalise positive hits identified in a successful enzyme engineering campaign. This new information can be cycled around for further potential benefit in later rounds of design. Statistical coupling analysis has

the potential to identify relationships between residues that are not apparent from examination of the structure. Such information could lead to the production of multiple variant libraries that are not limited to a proximal shell around the active site. In the following sections these techniques are discussed in further detail.

1.4 Computational methods for enhanced of enzyme engineering

1.4.1 Computational structural modelling

Crystal structures provide a great deal of information that can be utilised in the generation of hypotheses on enzyme mechanism and to guide the design of variant libraries for enzyme engineering. Protein crystallisation does however have various limitations- the time required to produce crystals is often limiting; the nature of the crystallisation process renders it impossible to derive reactive structures that only exist transiently; this process also limits crystal structures to static structure solutions, crystal structures often fail to indicate the dynamic nature of a particular protein structure. Computational docking of ligands in protein active sites allows us to address some of the issues that limit crystal structures of proteins and extends the use of structure data. With a protein structure as a starting point, computational docking allows us to generate multiple structures representing the likely conformations of different ligands bound in the active site of the protein in a fraction of the time required to achieve this experimentally. We can also model the structures of intermediate, transient, structures within the protein active site. This would not be possible experimentally. Computational modelling has even been extended to design *de novo* functional enzymes, Baker *et al* have utilised computational enzyme design to produce both Kemp elimination catalysts [54] and a Diels-Alderase [55].

Computational automated docking involves searching for a conformation of a ligand in an active site that has minimal energy. The energy of the ligand in the context of

the active site is calculated using a molecular mechanics forcefield with parameters for all the different types of interaction that contribute to the bound energy of the ligand. There are two main categories of automated docking methods: matching methods and docking simulation methods. In matching methods, a model of the active site is created and rigid ligands are docked into this model. Dock is a good example of an automated docking algorithm using a matching method. In contrast to matching methods, docking simulation methods involve exploration of flexible ligand translations and orientations until an ideal conformation is found within the protein active site. Docking simulation methods are more computationally intensive than matching methods but this is not a problem unless a large chemical database of lead compounds needs to be screened against an active site. Docking simulation methods allow the docking of a flexible ligand and the use of a more detailed molecular mechanics forcefield which can more accurately calculate the binding energy of the ligand.

Although other programs such as DOCK are available for ligand-protein docking, Autodock is the best known example of a docking simulation method. This program couples a well optimised empirical molecular mechanics forcefield with an efficient search algorithm. These attributes, together with the fact that Autodock is freely available, have led to a good support network for this program and many publications utilising it. Docking simulation aims to identify the minimum energy binding conformation in a huge energy landscape, resulting in a very computationally intensive problem requiring sophisticated search algorithms to reduce the search space to a tractable size. Autodock (Version 3.0.5) uses a

Lamarckian genetic algorithm and an empirical free energy function to find the minimal energy binding conformation

A genetic algorithm is a search technique that utilises the principles of biological evolution to find a solution which exhibits maximum fitness. In the genetic algorithm used by Autodock, the state of the ligand in the context of the protein is defined by a set of state variables which describe the translation, orientation and conformation of the ligand. Each state variable corresponds to a gene in the genetic algorithm. The state of the whole ligand corresponds to the genotype, and the atomic coordinates of the ligand correspond to the phenotype. Finally, the fitness of the ligand state is defined by the total interaction energy of the ligand with the protein.

The genetic algorithm initiates with a random set of genotypes which makes up the population. Individuals with better fitness in the initial population are allowed to reproduce whereas others die. Reproduction involves the mating of random pairs of individuals, during this process crossover takes place with new individuals inheriting genes from either parent. Some of the offspring also undergo random mutation where one gene changes by a random amount.

Autodock 3.0.5 combines a genetic algorithm with a local search method which performs energy minimisation. The genetic algorithm stage is a global search of the energy landscape and allows transitions over energy barriers which may separate energy valleys. The local search method uses the same forcefield as the genetic algorithm to make fine adjustments and find the energetic minimum within the energy trough. The step size of the local search method is adaptive, becoming

smaller in response to a series of consecutive successes. Following a local search the individual can be replaced by the result of the local search. As the local search is carried out at the phenotypic level, this is an example of an inverse mapping function, a genotype can be derived from a given phenotype. The term “Lamarckian genetic algorithm” refers to Jean Baptiste de Lamarck’s belief that phenotypic characteristics acquired during an individual’s lifetime can become inheritable traits [56].

The scoring function used by Autodock to represent fitness is based on an empirical free energy function that can reproduce experimentally derived binding constants of ligands. The energy function consists of five entropic terms which represent Van der Waals forces, H-bonding, electrostatic forces, entropy of the ligand and solvation. Coefficients for each of these terms have been determined using linear regression from a set of protein ligand complexes with known binding constants. Autodock utilises a fast grid based method for energy evaluation in which ligand-protein pairwise interaction energies are precalculated and used as a look up table during the simulation. Summations are performed for all ligand (i) and protein (j) atom pairs as well as all ligand atom pairs three or more bonds apart (Equation 1.1).

$$\Delta G = \Delta G_{vdw} + \Delta G_{Hbond} + \Delta G_{elec} + \Delta G_{tor} + \Delta G_{sol}$$

$$\Delta G_{vdw} = \sum_{i,j} \left(\frac{A_{ij}}{r_{ij}^{12}} \frac{B_{ij}}{r_{ij}^6} \right)$$

$$\Delta G_{Hbond} = \sum_{i,j} E(t) \left(\frac{C_{ij}}{r_{ij}^{12}} \frac{D_{ij}}{r_{ij}^{10}} E_{Hbond} \right)$$

$$\Delta G_{elec} = \sum_{i,j} \frac{q_i q_j}{\epsilon(r_{ij}) r_{ij}}$$

$$\Delta G_{tor} = N_{tor}$$

Equation 1.1 Energy function utilised by Autodock 3.0.5 calculations of binding energy.

The three terms that describe the interaction energies of atom pairs include a Lennard Jones 12-6 van der waals term, a directional 12-10 H-bonding term in which $E(t)$ is a directional weight based on the angle between the H-bond donator and the H-bond acceptor atom, and a coulombic electrostatic potential with a distance cut off. Ligand binding is accompanied by unfavourable entropy as the ligands conformational degrees of freedom are reduced. This contribution to the total binding energy is proportional to the number of SP3 bonds in the ligand and is represented by N_{tor} . The grid based method for energy evaluation used by Autodock limits the choice of solvation terms to use as most of these methods are based on surface area calculations. Autodock uses the pairwise volume based method of Stouten *et al*, in which the percentage of volume around a ligand atom that is occupied by protein atoms is weighted against the atomic solvation parameter of the ligand atom [57]. This gives the desolvation energy contribution from the ligand atom upon binding.

1.4.2 Statistical coupling analysis

There are many examples of epistatic coupling within proteins. Signalling proteins such as GPCRs rely on information transfer between distant residues [58, 59], the exquisite specificity of antibodies generated through B-cell maturation is often determined by residues distant from the antigen binding site [60], co-operative binding of oxygen in haemoglobin is mediated by networks of interacting residues [61-64]. In all of these examples, energy transduction mechanisms have evolved which make possible the highly adapted functions of these varied proteins.

Identification of these interacting networks of residues could enable enhanced engineering of new properties into protein scaffolds.

A new method termed Statistical Coupling Analysis (SCA) has been developed to identify epistatically coupled networks of residues [65]. SCA utilises evolutionary data contained within multiple sequence alignments (MSA's) to identify co-evolved positions within a protein sequence. The method is based on two hypotheses, firstly that without evolutionary constraint, the amino acids at a specific position in an MSA will approach their mean distribution in all proteins. Secondly, that functional coupling of two positions in a protein should mutually constrain their evolution. If two positions are functionally coupled, alteration of the distribution of amino acids at one site (by the removal of sequences from the MSA) should result in a change in the distribution of amino acids at the other site. Importantly, this does not require that the level of conservation change at the second site, just that the distribution of amino acids be altered [65].

In the first application of the SCA technique, Lockless *et al* applied the method to the identification of coupled sites within the PDZ domain, a small protein binding motif [65]. PDZ domains can be divided into two classes based on their target sequence specificity. The identity of the residue at position 76 in the PDZ domain is known to be an important determinant of this property. Lockless *et al* constructed a multiple sequence alignment consisting of 274 eukaryotic PDZ domains, including 4 PDZ domains with known structures. This MSA was then perturbed by removing all the sequences apart from those with a histidine at position 76. In response to this perturbation, the distributions of amino acids at several other positions in the MSA

were found to be altered. These sites are statistically coupled to position 76. Positions identified included both sites in close proximity to position 76 and other surface residues which are involved in sequence recognition. Coupling to proximal cooperative surface sites can be explained by energy propagation through the bound substrate. A third, unexpected, class of residues were also identified by SCA; these sites were a long distance from position 76 and were found in the core or on the opposite surface of the PDZ domain. Although the mechanism and function of this coupling is unknown, pathways of sterically connected, coupled residues were identified that connect position 76 to these distant residues. These pathways may represent routes of signal transduction through the tertiary structure of the protein. Lockless *et al* went on to verify the coupling interactions identified through thermodynamic mutant cycle analysis. This verification demonstrated good correlation between the statistically coupled sites and the thermodynamically coupled sites including those both proximal and distant from position 76.

Since this early demonstration of statistical coupling analysis, the technique has been applied to several different protein folds including G-protein coupled receptors, haemoglobin and serine proteases [66]. In these examples, Suel *et al* hypothesised that if networks of coupled residues exist and are conserved, perturbations at positions within the network should redundantly identify each other. In each of the examples above, Suel *et al* carried out global perturbation analysis and displayed the resulting statistical coupling energies on a matrix with perturbations represented by columns and positions represented by rows. Using

two-dimensional cluster analysis, global patterns of statistical coupling could be identified in the protein folds.

Of particular interest is the analysis of statistical coupling in serine proteases. A multiple sequence alignment was constructed consisting of 616 chymotrypsin serine proteases. Global SCA was carried out involving 69 site specific perturbations. Iterative two-dimensional clustering of the resulting matrix identified two distinct clusters, each containing positions that demonstrate similar patterns of coupling. One of the clusters was found to encompass both the S1 binding pocket and the surface loops known to determine substrate specificity. Residue 172 was also present in this cluster. Although distant from the active site and the binding pocket, SCA was able to identify positions known to determine substrate specificity.

Substrate specificity of the transketolase enzyme has previously been modified using saturation mutagenesis. Improved specific activity towards non-natural substrates such as glycoaldehyde [9] and propionaldehyde [10] has been engineered into transketolase by targeting residues in close proximity to the TPP cofactor. Phylogenetic information was also used to select residues but in both cases no residues more than 10Å from the ThDP cofactor were modified. In the examples above, single point variant libraries were constructed and screened using a colorimetric assay or conventional HPLC. Although hits were identified for different non-natural substrates within these libraries, the screening process limited the number of residues that could be probed and multiple simultaneous mutations could not be assessed. Statistical coupling analysis was able to identify

sites in chymotrypsin that appear to have co-evolved to determine substrate specificity. Identification of such networks in the transketolase enzyme may allow screening to be directed towards areas of sequence space unidentifiable by traditional structure and phylogeny directed selection methods.

1.5 Conclusions

The natural activity of transketolase can be applied to many applications where regio- and stereo-specific carbon-carbon bonds need to be created. This activity is hugely sort after in the synthesis of fine chemicals and pharmaceuticals. In addition to the added specificity afforded by transketolase over more traditional chemical processes, the use of a biocatalyst reduces the need for harsh reaction conditions, organic solvents, and multistep processes. To date, all biotransformations using transketolase have used the wild type molecule, an enzyme which has evolved over millions of years to catalyse two specific reactions *in vivo*. Through protein engineering, the tools exist to tailor-make transketolase variants with improved properties desirable to the fine chemical and pharmaceutical industries.

Arguably the most powerful method in enzyme engineering is directed evolution. However, limitations in library size impose limits on the sequence space that can be searched using this technique. Effort is being made to reduce these size limitations through advancements in high throughput screening and improved ligation steps. But, as long as there is any limit at all in library size, it will be necessary to target the sequence space to be searched to areas most likely to lead to functional improvements. Various strategies can be adopted to select residues to target; from simple spatial constraints to more complex phylogenetic strategies, such as common ancestor rebuilding. We can also take a rational approach to choose changes that are likely to improve characteristics. Most of these selection methods utilise one branch of knowledge and data. Here we attempt to utilise sophisticated computational techniques to merge information sources, creating a more

sophisticated knowledge base that can be used to direct the creation of more intelligent variant libraries.

Crystal structures of proteins represent one of the richest data sources available to us as enzyme engineers. But, these structures are limited to long-lived, static targets through the nature of crystallisation. Our understanding of the nature of non-covalent interatomic interactions and data from ligand binding experiments allow us to computationally model the energy landscape of substrate-protein interactions, and predict conformations in which substrates are likely to bind. This extends our understanding of function and structure beyond what is possible with X-ray crystallography alone. Computational docking of substrates can identify residues directly involved in substrate interactions as well as rationalising the results from previous library screens.

The second great data source available to protein engineers is the vast, ever growing, collection of sequence data. Using statistical coupling analysis we can delve into this data and discover energetic coupling between sites within proteins. The true power of this technique becomes apparent when the networks of energetic coupling are superimposed onto the three-dimensional structure of the protein. Using the results from computational docking of substrates together with the knowledge of coupled networks of sites in the protein structure we can start to target our variant library very efficiently.

Structural and sequence data represents the culmination of a 3.5 billion year experiment in evolution. It is evolution which ties each of these data sets inextricably together. Random mutation of sequence affects the structure of

proteins, altering the chemical properties of active sites, leading to changes in the function of enzymes. These changes in enzyme function drive evolution. Only when each class of data is viewed in the context of the others does its true potential become apparent. Using the most modern computational methods, we hope to develop rich, combinatorial, information which can help direct the production of variant libraries to those regions which hold the most potential for improved activities.

2 Mechanistic analysis of *Escherichia coli* transketolase by *in silico* docking of substrates in the active site

2.1 Introduction

The carbon-carbon bond forming ability of TK, along with its broad substrate specificity, makes it very attractive as a biocatalyst in industrial organic synthesis [8, 39, 67]. If the ketol donor in the reaction is replaced by hydroxypyruvic acid (HPA) the reaction is rendered irreversible by the release of carbon dioxide. The use of α -hydroxyaldehydes as acceptors together with HPA as the ketol donor allows the creation of enantiomerically pure chiral triols. The potential for producing non-phosphorylated products simplifies their isolation and avoids the requirement to remove phosphate from the product [8, 67]. The TK enzyme from *E. coli* is a preferable biocatalyst to that from yeast due to the higher specific activity of *E. coli* TK towards HPA [68]. Therefore, it is useful to establish that the structural and mechanistic information gained for yeast TK is equally applicable to TK from *E. coli*, for which there is also a crystal structure available [14]. Considerable mechanistic detail has been obtained for *S. cerevisiae* and *E. coli* TK from crystal structures and NMR experiments [7, 18, 21]. Crystal structures have been obtained for the DE4P acceptor substrate bound to the yeast-TK active-site [18], and also for the enamine intermediate formed upon reaction of the yeast enzyme with the DX5P donor substrate [7]. More recently, *E. coli* TK structures were obtained in covalent complexes with DF6P (2R8P.pdb) and DX5P (2R8O.pdb) prior to enamine formation, as well as a non-covalent complex with the cyclic form of DR5P (2R5N) [21].

Despite these impressive structural studies, there is still little information regarding the mode of binding for the donor substrate before it reacts to form the covalent complex and subsequent enamine intermediate, or for the aldehyde acceptor binding in the presence of the enamine intermediate. While crystal structures offer valuable insights into how ligands interact with protein binding-pockets, the binding of enzyme substrates is more challenging as the substrate will usually only bind transiently in the correct conformation before reaction occurs. Notably, for the crystal structure of DE4P bound to the TK active-site [18], the original aim was to solve the structure with DF6P, but the electron density of the resulting structure revealed that the donor substrate had been cleaved into DE4P by TK during the crystal formation process. This exemplifies the difficulties in obtaining structures of substrates bound in active sites.

There is considerable interest in the further development of TK as an efficient biocatalyst, with rational mutagenesis and directed evolution approaches previously having resulted in mutants with altered or improved activity [9], substrate specificity [10, 69] and enantioselectivity [11]. However, further protein engineering to accept an even broader range of substrates would benefit from methods to rationalise the behaviour of existing mutants in structural terms, and to understand how non-natural substrates bind to the active site of this enzyme. Unlike DE4P, many of the non-natural aldehyde acceptor substrates so far examined for biocatalysis with TK, do not contain a phosphate group or an α -hydroxyl group, which are both known to have an important role in substrate recognition [18]. Furthermore, the engineering of TK variants that are less

susceptible to substrate or product inhibition will require a better understanding of the roles played by various residues within the enzyme active-site.

Automated computational docking presents an alternative and complementary means to x-ray crystallography for probing the binding of reactive substrates in short-lived conformations, and also for studying the many non-natural substrates and products, for which crystallography would be time-consuming. Computational automated docking involves searching for the conformation of a ligand bound within in an enzyme active-site that has minimal energy. AutoDock is the best known example of a docking simulation method in which the active site is created and ligands are docked into an enzyme active-site model with an accurate calculation of the binding energy. Flexible ligand translations and orientations are explored until an ideal conformation is found within the protein active site [70]. The observation that very little structural change occurs in the TK active site upon formation of covalent complexes with substrates [21], suggests that it would be an excellent system for docking different substrate complexes without requiring the modelling of amino-acid sidechains for induced fit.

Here we show that automated docking can produce accurate models of substrates bound in the active site of TK. The accuracy of our results is demonstrated by comparison of a computationally derived structure with the crystal structure of DE4P in yeast TK. Further validation is provided by a correlation of experimentally derived K_m values for yeast TK, with those calculated from computationally derived docking energies in AutoDock. Having demonstrated the accuracy of the approach we explored the differences and similarities between the binding of DE4P in the

active-sites of *E. coli* TK and yeast TK, and the impact this may have on function. We then examined the binding of natural and non-natural substrates in *E. coli* TK in non-covalently associated complexes that are too reactive to be obtained by crystallography. We also discuss the implications on the potential nucleophilic attack of the deprotonated ThDP cofactor upon the ketol substrate at an unusual Bergi-Dunitz angle, and also the mechanism for ring opening of the cyclic form of D-ribose-5-phosphate. These results will have a significant bearing on attempts to further engineer TK as a biocatalyst for organic synthesis, as well as generating useful hypotheses for future experimental studies to understand the enzyme mechanism of TK.

2.2 Materials and methods

2.2.1 AutoDock 3.0.5

The open source AutoDock software version 3.0.5 was used for all the automated docking reported. AutoDock combines a Lamarckian Genetic algorithm with an empirical free energy function to obtain ligand docked conformations [70]. Substrate docking models were obtained using the *E. coli* TK structure 1qgd.pdb with a cubic grid in the active site of sides 80 Å. Defaults were used for docking each substrate except for the following: the maximum number of energy evaluations was increased to 1 million, the number of genetic algorithm runs was increased from 10 to 200, and the grid spacing used was 0.375 Å. AutoDock performed a cluster analysis to each final conformation obtained from the 200 GA runs such that two conformations with an RMSD less than 0.5 Å are stored in the same cluster. Clusters are output in ranked order of increasing energy following completion of analysis. Manual visual analysis of docked conformations and further analysis of the docked conformations was carried out with Pymol and Ligplot.

2.2.2 Docking of D-erythrose 4-phosphate in yeast TK

D-erythrose 4-phosphate (DE4P) was removed from the yeast TK PDB file 1NGS. AutoDock was used to re-dock the substrate back into the binding site. Grid centre and size used for AutoDock run: (-12.645, 56.02, 19.419) 80 Åx80 Åx80 Å.

2.2.3 Docking of D-erythrose 4-phosphate in *E. coli* TK

DE4P was docked into the binding site of *E. coli* TK (1QGD). Grid centre and size used for AutoDock run: (-10.6, 27.6, 36.4) 80 Åx80 Åx80 Å.

2.2.4 Creation of a model of the ThDP-enamine intermediate in *E. coli* TK

The ThDP-enamine intermediate was docked into *E. coli* TK (1QGD). Grid centre and size for AutoDock run: (-10.0, 28.1, 36.0) 80 Åx80 Åx80 Å.

2.2.5 Docking DE4P and glycolaldehyde in ThDP-enamine complexed forms of yeast and *E. coli* TK

DE4P was docked into the yeast ThDP-enamine-TK complex (1GPU) and in the modelled *E. coli* ThDP-enamine-TK complex. Glycolaldehyde (GA) was docked into the modelled *E. coli* ThDP-enamine-TK complex. Grid centres and sizes were (-6.6, 56.7, 18.4) 60 Åx60 Åx60 Å for DE4P in yeast ThDP-enamine-TK, and (-11.4, 26.3, 36.4) 60 Åx60 Åx60 Å for DE4P and GA in *E. coli* ThDP-enamine-TK.

2.2.6 Docking of natural and non-natural aldehyde substrates into *E. coli* TK

PDB files for the ten TK substrates for which there are published K_m values, and also fluoropyruvate, found to be a potential inhibitor (unpublished data), were generated using the Dundee PRODRG server [71]. Each substrate was docked into the active site of *E. coli* TK. Preliminary docking identified two docking regions within the binding funnel of *E. coli* TK for some of these substrates. Grid sizes and positions were altered to obtain docked conformations for each substrate in the binding region closest to the ThDP cofactor. For some substrates the grid centres were adapted to avoid inaccessible pocket “traps” within the protein. Grid centres and sizes were as follows (grid centres in brackets):

Hydroxypyruvate:	(-15.991 21.945 37.096)	60 Å x 60 Å x 60 Å
Acetaldehyde:	(-18.344 24.016 40.547)	60 Å x 60 Å x 60 Å
D-erythrose 4-phosphate:	(-18.344 24.016 40.547)	40 Å x 40 Å x 40 Å
D-erythrose:	(-18.344 24.016 40.547)	40 Å x 40 Å x 40 Å
D-glyceraldehyde 3-phosphate:	(-10.586 27.153 35.586)	80 Å x 80 Å x 80 Å
D-glyceraldehyde:	(-18.344 24.016 40.547)	40 Å x 40 Å x 40 Å
D-ribose 5-phosphate:	(-10.586 27.153 35.586)	80 Å x 80 Å x 80 Å
D-ribose:	(-10.586 27.153 35.586)	80 Å x 80 Å x 80 Å
Glycolaldehyde:	(-15.991 21.945 37.096)	60 Å x 60 Å x 60 Å
Xylulose 5-phosphate:	(-10.586 27.153 35.586)	80 Å x 80 Å x 80 Å
Fluoropyruvate:	(-15.991 21.945 37.096)	60 Å x 60 Å x 60 Å

Docking energies (ΔG) were converted to a K_m values using $\Delta G = -RT \ln(K_m)$, where R is the gas constant and T is the temperature in Kelvin.

2.2.7 PyMol Molecular Graphics System

All visualisations of docked conformations were produced using PyMol, available from <http://www.pymol.org> [72].

2.3 Results and discussion

2.3.1 Automated docking of D-erythrose-4-phosphate in the active site of yeast TK

Nilsson *et al* (1997) previously solved the crystal structure for a substrate protein complex of DE4P bound in the active site of yeast TK [18]. The original aim of their crystallisation was to solve the structure with DF6P but the electron density of the resulting structure would not fit a six-carbon chain and the conclusion was drawn that the donor substrate had been cleaved into DE4P by TK during the long period necessary for crystal formation. This cleavage would also yield the α,β -dihydroxyethyl thiamine diphosphate intermediate but this would degrade in a few hours into ThDP and glycolaldehyde explaining the lack of an intermediate in the electron density map.

To assess the accuracy and potential of automated computational docking on the substrates of TK we initially docked DE4P into the empty active site of yeast holo-TK, to recreate the substrate-holoenzyme complex. AutoDock accurately predicts the binding conformation of DE4P producing a docked structure within 1.65 Å RMSD of the crystal structure (Figure 2.1). The hydrogen bonding network of the docked substrate is accurately predicted by AutoDock, supporting the role and evolution of this network in determining the stereospecificity of TK. In the crystal structure the C1 aldo carbon atom of the acceptor substrate is positioned 4.16 Å away from the reactive C2 carbon of the thiazolium ring of ThDP. This distance is sufficient to allow the presence of an enamine intermediate in a reactive conformation with the acceptor substrate. The computationally docked

conformation positioned the C1 aldo carbon 4.81 Å away from the thiazolium ring of ThDP. Although AutoDock docked DE4P slightly further away from the ThDP reactive centre, it was still within an acceptable distance to react in the presence of the intermediate.

The main discrepancy between the experimentally determined structure and that created by computational docking was the position of the phosphate group. In the computational docking the phosphate group of DE4P was pulled closer to the side chains of the Arg 358, Arg 520, Ser 385 and His 461 residues (*E. coli* numbering) at the entrance to the binding funnel (by just over 1 Å). This has the effect of pulling DE4P slightly out of the funnel away from the ThDP cofactor (Figure 2.1). The interaction between the phosphate group and the positively charged arginine residues of yeast TK is strongly affected by the electrostatic interactions between these groups. The force field of AutoDock includes a term to model electrostatic interactions but the discrepancy between the modelled and the crystal structures may be explained by errors in the electrostatic term of the forcefield. While all of the thirty ligand-protein complexes used to calibrate the AutoDock forcefield included H-bond interactions, only a small proportion involved electrostatic interactions of explicitly charged groups [70]. Electrostatic interactions between these groups would also be strongly influenced by pH which may further explain the slight discrepancy. However, the TK structure itself was resolved to 2.4 Å and so a 1 Å shift is acceptable within error. Despite the possible error in the modelling of phosphate binding, the results show that AutoDock is capable of reproducing accurate docked conformations of substrates in the active site of TK.

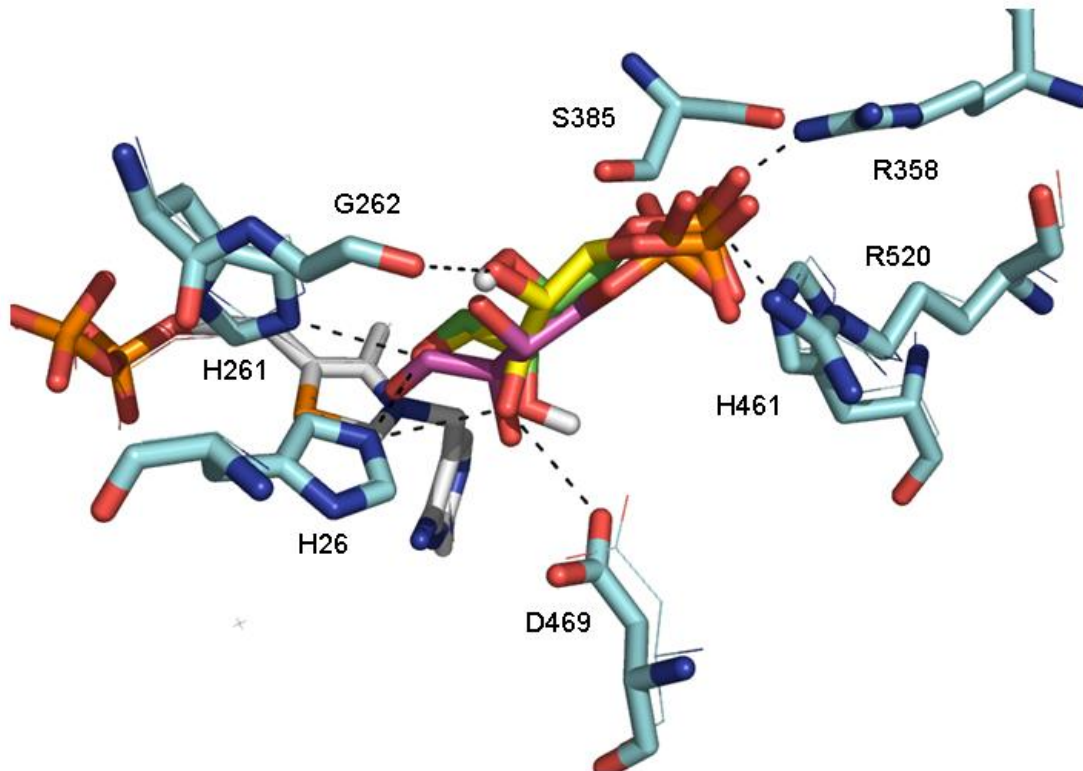


Figure 2.1 Comparison of D-erythrose-4-phosphate (DE4P) binding in yeast and *E. coli* transketolases. *E. coli* TK residues are shown as blue sticks and the aligned yeast TK residues are shown as lines. Three DE4P structures are compared: (magenta) crystal structure in yeast (1NGS); (yellow) docked into the uncomplexed *E. coli* TK structure (1QGD); (green) re-docked into the yeast TK structure (1NGS).

2.3.2 Automated docking of D-erythrose-4-phosphate in the active site of *E. coli* TK

The active sites of yeast and *E. coli* TK, including the orientation of conserved residues, are nearly identical. For *E. coli* TK, structures are available in covalent complexes with DF6P (2R8P.pdb) and DX5P (2R8O.pdb), as well as a non-covalent complex with the cyclic form of DR5P (2R5N) [21]. However, no crystal structure exists for the non-covalent complex of *E. coli* TK with DE4P. If *E. coli* TK binds DE4P in the same conformation as for yeast TK then mechanistic insights derived from studies of yeast TK could be used with confidence to infer equivalent mechanistic details in the *E. coli* protein. This would be of great value as much of the work on the function of TK to date has been carried out on the yeast enzyme.

DE4P was docked into the active site of *E. coli* TK (1QGD) using AutoDock. Many of the docked conformations predicted by AutoDock involved an inversion of the DE4P, with the phosphate group oriented towards the ThDP and the carbonyl active centre of DE4P pointing out of the active site in an un-reactive conformation. This could be explained by the error in the handling of phosphate interactions. Like the arginine residues at the entrance to the active site normally involved in phosphate binding, the ThDP molecule carries an explicit positive charge. Hypothetically, the DE4P may actually be able to bind in this orientation and form an un-reactive inhibitory complex. If this is not an artefact of computational docking it may therefore have implications for substrate inhibition of TK at high concentrations.

AutoDock also docked DE4P in the reactive orientation in *E. coli* TK (Figure 2.1). This conformation is very similar to that of DE4P bound in yeast TK (compared in Figure 2.1). The phosphate group is positioned close to the entrance of the active site and interacts with residues Arg 358, Arg 520, Ser 385 and His 461. These interactions are equivalent to those maintaining the position of the DE4P phosphate group in yeast TK. The carbon chain of DE4P extends down the active site of *E. coli* TK forming a hydrogen-bonding network with the side chains of several conserved residues. The C3 hydroxyl group forms an interaction with the backbone oxygen of Gly 262 that is not seen in the yeast TK bound structure. Like yeast TK, the conserved Asp 469 residue of the *E. coli* enzyme interacts with the C2 hydroxyl group of DE4P but an additional interaction is formed through His 26. The C1 aldo oxygen atom interacts with residues His 261 and His 26 in *E. coli* as

observed in yeast TK, to position the C1 aldo carbon at 4.89 Å away from the C2 atom of the ThDP thiazolium ring.

The general binding conformation for DE4P in *E. coli* TK is the same as that for yeast TK. The minor differences in the hydrogen bonding network do not change the favoured stereospecificity of the recognition and these interactions could be transiently present in the yeast structure with only a small degree of dynamic movement. The conformation of DE4P docked in *E. coli* TK supports the hypothesis that the conserved residues of the TK active sites of *E. coli* and yeast TK have the same roles in substrate binding.

2.3.3 Modelling the enamine intermediate in *E. coli* TK and docking of D-erythrose-4-phosphate into the yeast and *E. coli* TK-ThDP-enamine complexes

We have demonstrated the ability of AutoDock to accurately model the binding conformation of DE4P in yeast TK and shown that the same binding conformation is formed in *E. coli* TK. However, in the TK catalysed reaction, DE4P cannot bind productively until an enamine intermediate has first been formed between the ThDP cofactor and the ketol donor substrate. It is possible that the binding conformation of DE4P in the TK active site is different in the presence of this intermediate. Due to the reactive nature of the intermediate, DE4P would not bind and exist in the presence of the enamine intermediate for long enough to obtain crystals for structural analysis. Currently, the only way to solve the structure for DE4P bound in the presence of ThDP-enamine intermediate is to model this structure computationally. In the yeast-TK crystal structure of the ThDP-enamine

intermediate there were no significant variations in the orientations or positions of active site residue side chains relative to the ThDP bound structure. The ThDP-enamine intermediate could therefore be confidently docked into the known *E. coli* holo-TK structure using AutoDock to obtain a model of the *E. coli* TK-ThDP-enamine complex.

As seen in Figure 2.2, the complex obtained for *E. coli* TK-ThDP-enamine was nearly identical to the solved structure of the yeast TK-ThDP-enamine complex (1GPU) [7]. All the major functional interactions were present in the modelled complex, including the Glu 411 (Glu 418 in yeast TK) interaction with N1' of the pyrimidine ring [73]. The ketol donor derived enamine intermediate is co-ordinated by hydrogen bonds to the conserved *E. coli* histidine residues His 100 and His 473. The only difference between the TK-enamine interactions of yeast and TK is that in the yeast-TK complex His 481 (His 473 in *E. coli*) interacts with both hydroxyl groups of the enamine whereas in the *E. coli* complex His 473 only interacts with the α -hydroxyl group. Some other minor differences are present in the hydrogen-bonding network of the di-phosphate group but none of these differences significantly alter the position of ThDP-enamine relative to the TK molecule (Figure 2.2).

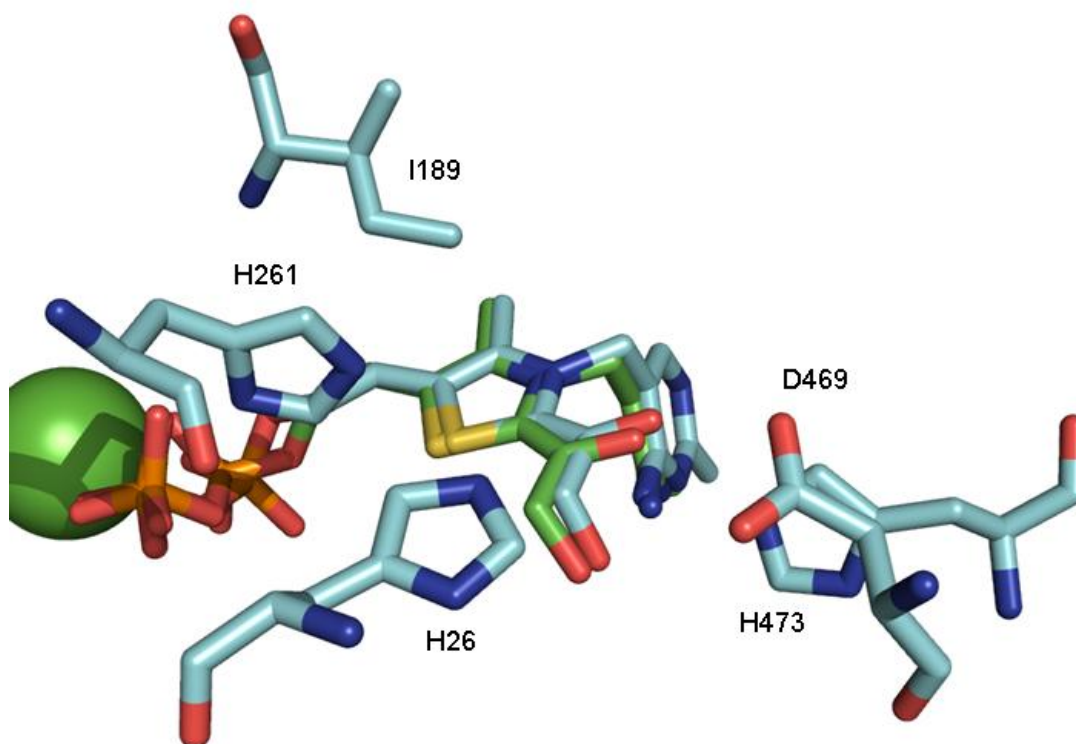


Figure 2.2 Comparison of the ThDP-enamine intermediate in yeast and *E. coli* transketolases. The ThDP-enamine intermediate in yeast TK (blue sticks) is from an available structure (1GPU), whereas that for *E. coli* TK was obtained by docking (green sticks) in 1QGD. The bound calcium ion is shown as a green sphere.

Following the creation of the *E. coli* TK-ThDP-enamine complex model, AutoDock was used to obtain docked conformations of DE4P in both the yeast (crystal structure) and the *E. coli* (modelled) TK-ThDP-enamine complexes. In each case the DE4P molecule docked in the same conformations as previously observed in Figure 2.1, in the absence of the enamine intermediate. The hydrogen bonding interactions between the TK and DE4P observed in the absence of the enamine are also preserved along with the few differences between yeast and *E. coli* TK described above. The DE4P carbonyl C1-atom is placed within 3.44 Å and 4.28 Å of the enamine α -carbon in the yeast TK and *E. coli* TK models respectively. As well as positioning the carbonyl group of DE4P in close proximity to the α -carbon of the enamine, the hydrogen-bonding network described orientates the carbonyl group

for the formation of the S-enantiomer product upon carbon-carbon bond formation as expected for TK (see Figure 2.4 a) [74].

2.3.4 Docking of other substrates in *E. coli* holo-TK

AutoDock has proven accuracy in predicting the binding affinities of ligand-protein complexes. Published kinetic data including K_m values are available for several substrates in the reaction catalysed by *E. coli* TK [1]. Ten substrates, namely glycolaldehyde (GA), β -hydroxypyruvate (HPA), D-xylulose-5-phosphate (DX5P), D-ribose-5-phosphate (DR5P), D-glyceraldehyde-3-phosphate (DG3P), D-erythrose-4-phosphate (DE4P), acetaldehyde (AA), D-ribose (DR), D-glyceraldehyde (DG), D-erythrose (DE), were docked in the binding site of *E. coli* TK and their K_m values were calculated from the binding affinities reported by AutoDock.

Initial docking using a grid that encompassed the entire binding funnel of *E. coli* TK revealed two regions in which substrates could bind in the active site of the enzyme. Following the identification of these separate binding sites, grid sizes and positions were altered to remove the non-productive and higher energy binding region, which was in the active-site funnel but further out from the ThDP molecule and the active centre of the enzyme. Such a site which involved mainly protein backbone interactions may prove to be physiologically relevant, for example resulting in substrate or product inhibition, but was not studied further here.

The calculated K_m values for the docked substrates were compared against the experimentally determined values previously published [1] as shown in Figure 2.3. The calculated K_m values we obtain from the binding energies of substrates in the

active site of TK were obtained only using the acyclic and monomeric forms of substrates. However, the values of K_m can be affected in some cases by the equilibrium between dimeric, monomeric, cyclic and linear forms which decreases the availability of the reactive substrate. DR5P, DR and DE can all form ring structures and the proportion of acyclic monosaccharides in aqueous solution have been determined respectively as 0.6% [75], 0.05% [76] and 12.1% [77]. These equilibria were used to adjust the experimental K_m data obtained by Sprenger and co-workers as presented in Table 2.1 and Figure 2.3. A recent crystal structure of *E. coli* TK bound to DR5P has shown that the substrate can bind directly in the cyclic form. Therefore, the adjustment of K_m values above assumed that the ring opening of the sugars is not rate limiting, and therefore does not affect the observed K_m . Other substrates such as glycolaldehyde and D-glyceraldehyde form dimers at high concentrations but in dilute aqueous solutions they become essentially monomeric and so the experimental data for these substrates were not adjusted [77]. The calculated and experimental $\log(K_m)$ values correlate well with a Pearson R^2 value of 0.82 indicating that the AutoDock algorithm is capable of predicting the conformations and binding energies of a broad range of mostly natural substrates. In all cases except DR5P, the calculated K_m values are lower than the experimentally determined values indicating a systematic underestimation of K_m values by AutoDock.

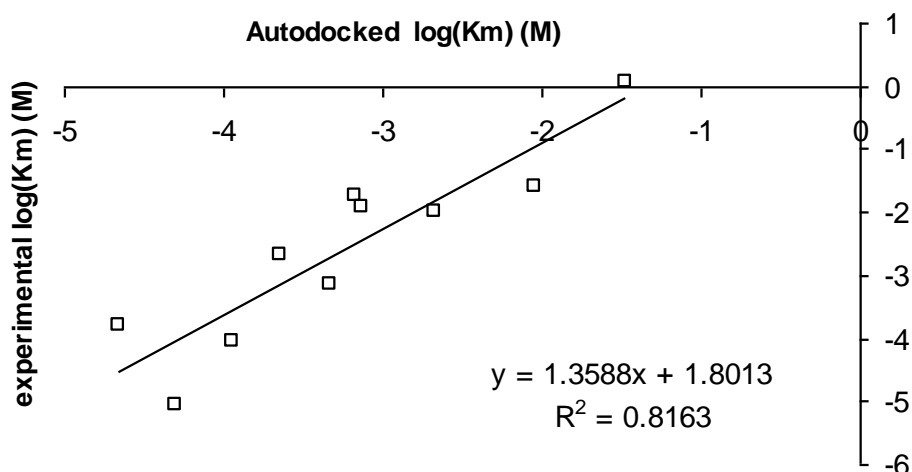


Figure 2.3 Comparison of $\log(K_m)$ values obtained experimentally and calculated from the docking energies reported by AutoDock for ten substrates of TK. All values were taken from Sprenger *et al* [1], except the values for HPA and GA which were averaged with those from Hibbert *et al* [9]. Values for D-ribose, D-ribose-5-P and erythrose, were adjusted by experimentally determined equilibrium constants for the fraction of the acyclic forms [75-77].

Substrate	K_m (calc) (mM)	K_m (exp) ^a (mM)	Distance to nucleophile ^g (Å)
D-Ribose	0.47	0.7 ^b	2.64
D-Ribose-5-phosphate	0.05	0.0084 ^c	3.13
D-Erythrose	0.67	18.15 ^d	3.93
D-Erythrose-4-phosphate	0.12	0.09	4.28
D-Glyceraldehyde	2.10	10 ^e	2.33
D-Glyceraldehyde-3-phosphate	0.23	2.1 ^e	5.01
Glycolaldehyde	9.14	24.5 ^f	2.66
Acetaldehyde	33.6	1200	2.69
β -Hydroxy pyruvate	3.61	11.65 ^f	3.14
D-Xylulose-5-phosphate	0.022	0.16	4.73

Table 2.1 Comparison of *E. coli* TK K_m -values obtained by docking with those from experiment. ^a Experimental data obtained by Sprenger *et al* [1]. ^b adjusted by equilibrium for 0.05% acyclic form [76]. ^c adjusted by equilibrium for 0.6% acyclic form [75]. ^d adjusted by equilibrium for 12.1% acyclic form [77]. ^e Experimental data obtained with D,L racemate. ^f average of independent values from Sprenger *et al* [1] (glycolaldehyde 14 mM, β -Hydroxy pyruvate 18 mM) and Hibbert *et al* [9] (glycolaldehyde 35 mM, β -Hydroxy pyruvate 5.3 mM). ^g distances measured between either a) aldehyde carbon atom of aldol acceptor and the ThDP-enamine nucleophile, or b) carbonyl carbon atom of ketol donor and the ThDP thiazolium C2 nucleophile.

Examination of the structures of each substrate bound in the active site of *E. coli* TK reveals the same general binding conformation as seen for DE4P described above (Figure 2.4 a-b and Figure 2.5 a). Notably, all phosphorylated substrates docked with the phosphate group interacting with the conserved Arg 358, Arg 520, Ser 385 and His 461 residues at the entrance of the binding funnel as expected. This

conformation orientates the substrates so that their active centres are positioned within the binding funnel in an active conformation, but also creates small differences in their proximity to the ThDP/enamine depending on chain length which plays a role in substrate preference. The distance from the aldehyde carbon atom to the enamine nucleophile for phosphorylated substrates decreases progressively as the chain length increases from the three-carbon DG3P, and four-carbon DE4P, to the preferred five-carbon DR5P which has the shortest distance of 3.13 Å (Table 2.1). The phosphorylated substrates had generally lower experimental and predicted K_m values (Table 2.1) which can be explained by this strong interaction at the entrance to the binding cleft. When overlaid with the docked enamine structure (Figure 2.4 a) the phosphorylated aldol acceptors are all positioned and oriented with the Re-face of their aldehydic carbonyls ready for nucleophilic attack by the nearby α -carbon of the enamine to give products of the S-enantiomer.

Docking of the non-phosphorylated aldol acceptors was found to be dominated by hydrogen-bonding to His 26, His 261 and, with the exception of acetaldehyde, Asp 469 (Figure 2.4 b). Interestingly, D-ribose forms a hydrogen bond to Arg 520 via the C5-hydroxyl group in the absence of the phosphate, though this does not fully compensate for the much tighter binding achieved with the phosphate in DR5P. As for the phosphorylated aldehydes, an overlaid structure of the docked enamine (Figure 2.4 b) shows that the non-phosphorylated aldol acceptors are also all positioned and oriented with the Re-face of the aldehyde carbonyl prone to

nucleophilic attack by the nearby α -carbon of the enamine to give the *S*-enantiomer products.

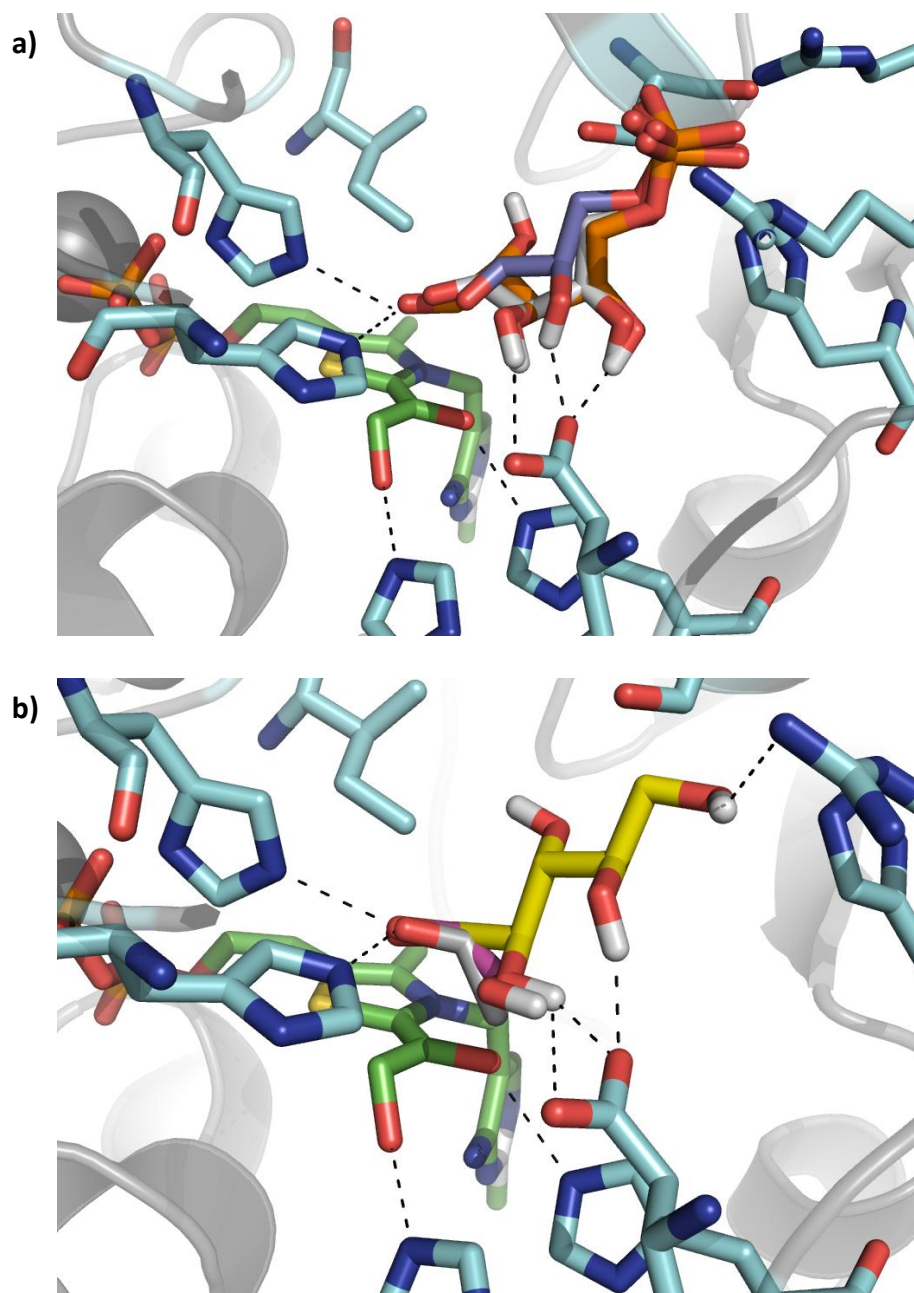


Figure 2.4 Comparison of the docking of phosphorylated and non-phosphorylated aldol acceptor and ketol donor substrates in *E. coli* TK. The enamine intermediate (not present during docking) is overlaid for reference in each panel (green sticks). a) Phosphorylated aldol acceptor substrates. DE4P (grey), G3P (dark blue) and DR5P (orange) are each oriented to present the *Re* face of their *aldo* carbonyls to give the expected *S*-enantiomer products. The phosphate binding location is highly defined in the R520 and R358 pocket, and creates differences in the proximity of the *aldo* carbonyl to the $C\alpha$ enamine carbon depending on substrate chain length. b) Non-phosphorylated aldol acceptor substrates. Acetaldehyde (grey), Glycolaldehyde (magenta), and Ribose (yellow) are also oriented to present the *Re* face of their *aldo* carbonyls to give the expected *S*-enantiomer products. Non-phosphorylated substrate binding is dominated by hydrogen-bonding to H26, H261, and D469 (except acetaldehyde and ribose which hydrogen-bonds to R520 even without the phosphate).

2.3.5 Comparison of docked and crystallized DR5P conformations

Asztalos and co-workers previously obtained a structure of *E. coli* TK with a mixture of both the cyclic and acyclic forms of DR5P bound in the active site [21]. However, the apparently ambiguous electron density due to low occupancy of the acyclic form, positioned it such that it would clash sterically with our modelled enamine intermediate, indicating that this might not be the exact productive binding conformation. While modelling of the yeast TK enamine structure coordinates by Asztalos *et al* found the acyclic DR5P to approach the enamine C α at a distance of 1.6 Å with a favourable angle of 112° for the C1 aldo carbon relative to the C2-C α of the enamine, inspection of their structure indicates that the enamine would be poised to attack the incorrect Se face of the C1 aldo carbonyl. Our docked acyclic DR5P gives a more plausible (or earlier) conformation for binding prior to a productive reaction in which the reactive enamine is 3.13 Å from the Re face of the aldehyde carbon atom of DR5P. The aldehyde O-atom is coordinated by His 26 (3.0 Å) and His 261 (2.8 Å), the C2 hydroxyl hydrogen bonds with Ser 385 (3.0 Å), the C3 (2.6 Å) and C4 (2.8 Å) hydroxyl groups both hydrogen bond with Asp 469, and the phosphate is coordinated to Arg 358, Arg 520, Ser 385 and His 461 as usual. The contacts observed in the binding of the cyclic form by Asztalos *et al* are different to those in our modelled acyclic form with the C1 to C3 hydroxyl groups being rotated around the protein side-chains by one position (Figure 2.5 b-c). Comparison of the two suggests that TK binds the cyclic DR5P, catalyses the ring opening, and then re-arranges the position of the aldehyde closer to the enamine prior to reaction.

His 261 is highly polarised by the phosphate in ThDP (2.7 Å) making it a good base for catalysis of the ring opening via deprotonation of the C1-hydroxyl group in DR5P which is 3.33 Å away. Ser 385 was also found in two occupancies with the first coordinating the DR5P phosphate [21], and the second in position to stabilise the protonation of the furanose O-atom of DR5P during ring opening. As the aldehyde forms at C1 upon ring opening, the hydrogen bond to His 261 is retained via the O-atom which moves by 3.2 Å into the position previously occupied by the C2 hydroxyl group, and forms an additional hydrogen bond with His 26. This movement is facilitated by a 180° rotation of the C2-C3 bond, as suggested by Asztalos *et al*, but forms a new hydrogen bond between the C2 hydroxyl group and Ser 385. Meanwhile the hydrogen bond between Asp 469 and the C3 hydroxyl group becomes shorter by 0.15 Å and the C3-C4 bond rotates by 180° to form a new hydrogen bond between the C4 hydroxyl group and Asp 469. Collectively these hydrogen bond exchanges serve to unwind the now acyclic DR5P backbone and free up the space near His 26 and the enamine-ThDP intermediate for the newly formed C1 aldehyde.

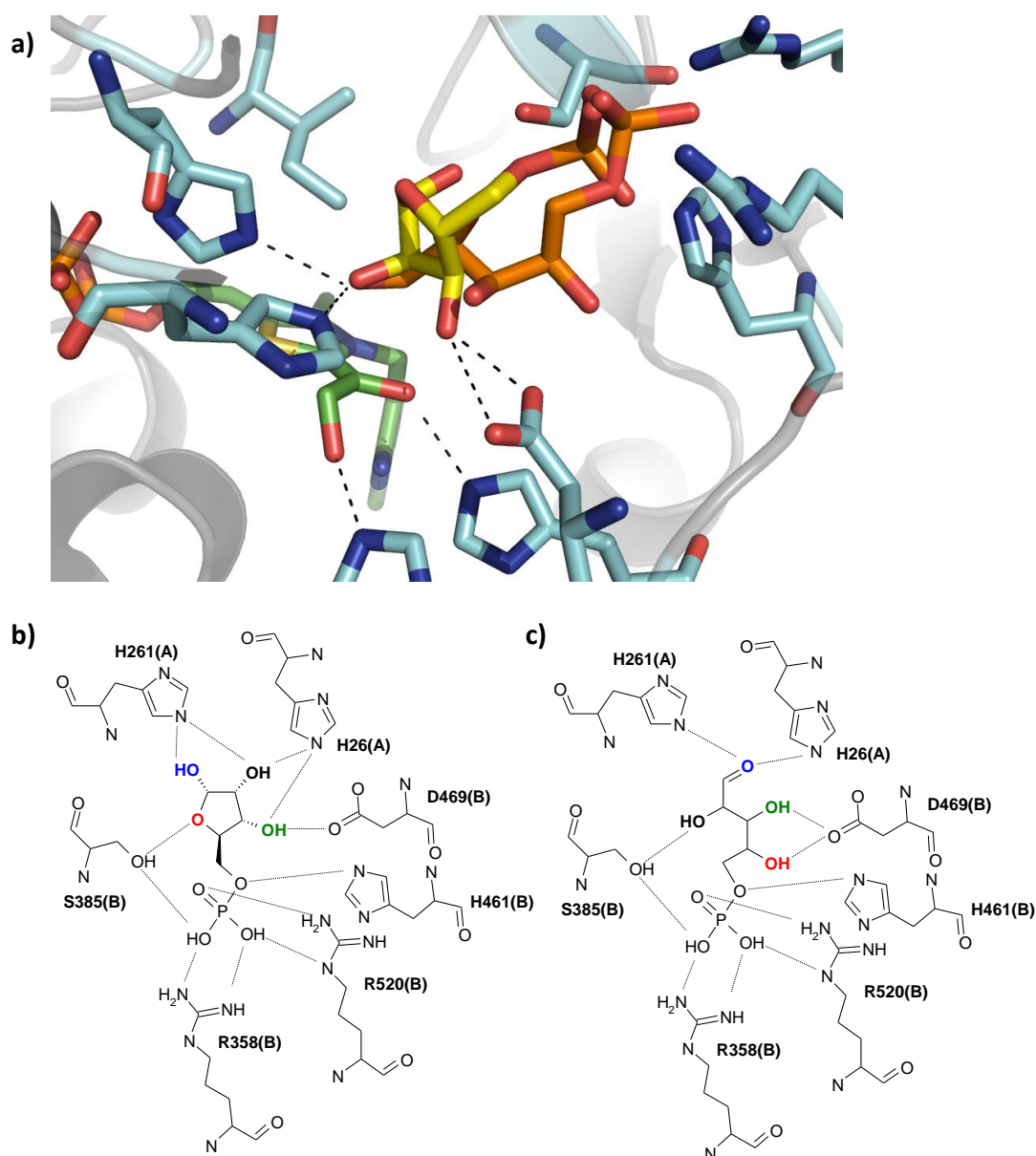


Figure 2.5 a) Cyclic and acyclic forms of DR5P. Cyclic DR5P (yellow) from a TK crystal structure [21] and the modeled acyclic DR5P (orange). b) and c) Hydrogen bonding network changes upon ring opening of DR5P. Cyclic (left) and acyclic (right) forms of DR5P with hydrogen bond interactions with transketolase shown schematically as dashed lines.

2.3.6 Glycolaldehyde docking

Glycolaldehyde (GA) is one of the smallest aldehydes accepted by TK and the reaction of GA with TK has been studied extensively [1, 26, 41, 74, 78]. As for DE4P, the docking of GA was performed both in the presence and absence of the modelled enamine. Comparison of the two docks indicated that they differed by an RMSD of only 0.53 Å and that their hydrogen bonding interactions were identical.

As with the other aldehydes, GA binding presented the Re face of the aldehyde to the nucleophilic enamine such that it would give the expected L-erythrulose product (Figure 2.6 a). Overall the very small differences, for both GA and DE4P docked with and without the enamine, indicate that the productive positioning of the aldehyde acceptors relative to the enamine is sufficiently optimised through specific interactions other than those to the enamine in the active site. This observation is analogous to that by Asztalos *et al* where the active site is catalytically poised to maximally translate substrate binding interactions directly into the strained tetrahedral intermediate with the ketol donor. It is possible then that the reaction between aldol acceptor and the enamine may introduce a similarly strained conformation, driven by optimised substrate interactions without the need for induced fit or the unfavourable flexibility this introduces.

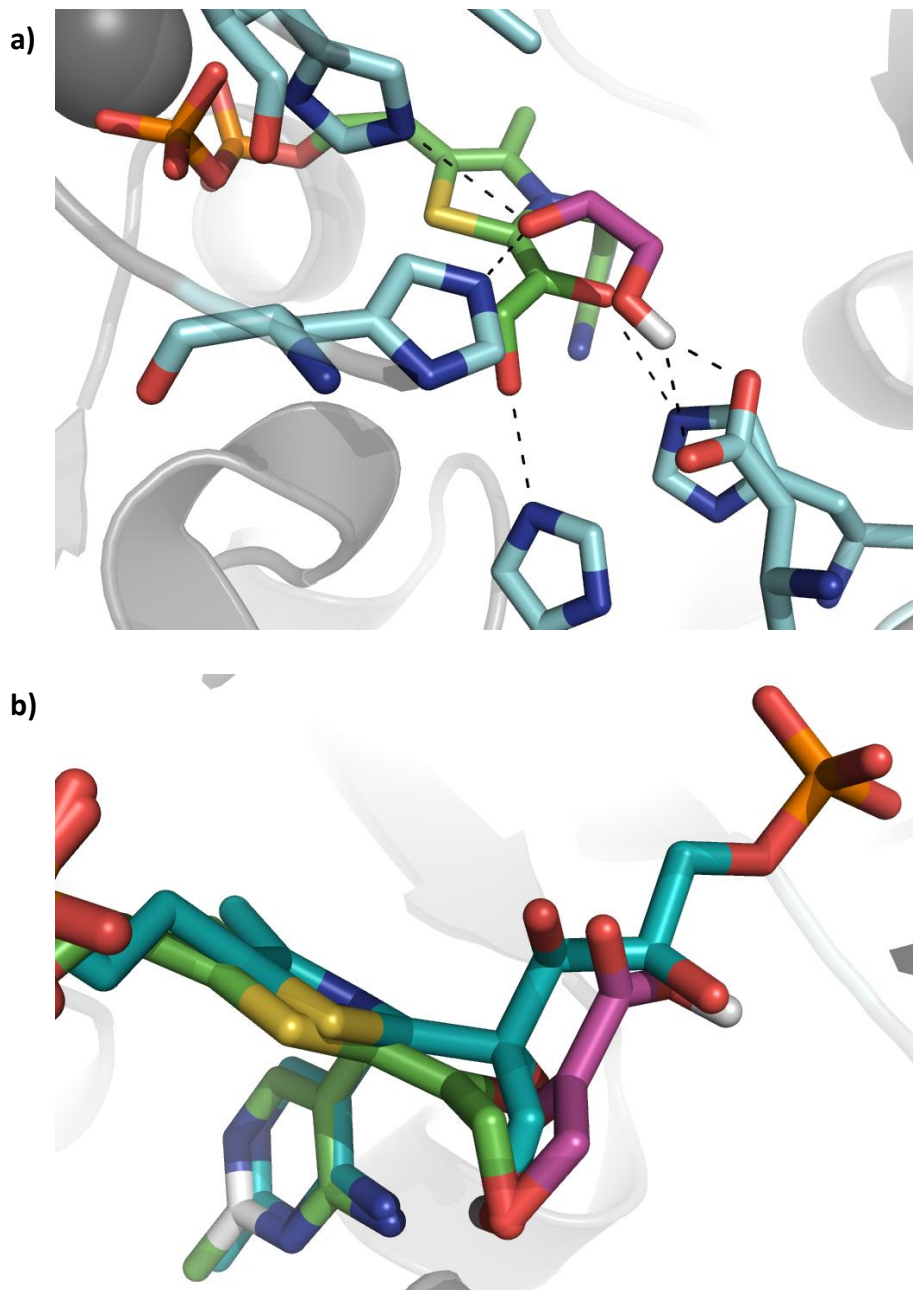


Figure 2.6 Docking of substrates into *E. coli* TK for the reaction of hydroxypyruvate and glycolaldehyde. a) Glycolaldehyde docked into *E. coli* TK in the presence of the modeled enamine. b) Transition from docked non-covalently bound HPA (magenta), via the covalently bound and strained tetrahedral intermediate with DX5P (blue - from crystal structure of Asztalos *et al* [21]), to the docked enamine (green).

2.3.7 Comparison of docking for the ketol donors

Hydroxypyruvate (HPA) and D-xylulose-5-phosphate (DX5P) are both donor substrates for the TK catalysed reaction for which no structures of the non-covalent complexes have been obtained experimentally. Fluoropyruvate (FPA) is an analogue of HPA for which no measurable reaction is observed with TK. FPA and

HPA are both known competitive inhibitors of the pyruvate decarboxylase from *Zymomonas mobilis* [79] and preliminary data from our lab suggest that FPA can also inhibit the TK reaction when using HPA as the ketol donor (unpublished). Both of the ketol donor substrates and also FPA were found to dock in conformations that position the carbonyl within a reactive distance from the C2 carbon of the ThDP thiazolium ring (Figure 2.6 b). However, the DX5P docked conformation and position was found to be unusually different to all other substrates and the carbonyl was at an unfavourable orientation for reaction with the ThDP thiazolium ring (not shown). This dock was deemed to be suspect upon visual inspection, and indeed removal of DX5P from the plot of calculated and experimental $\log(K_m)$ values in Figure 2.3 gave a slightly improved Pearson R² correlation of 0.84 (not shown).

To examine the mechanism of binding for ketol donors, and their reaction with ThDP, the docking of HPA was compared to the covalently bound DX5P intermediate from an available crystal structure with *E. coli* TK [21], and also to the modelled ThDP-enamine intermediate (Figure 2.6 b). The most striking observation is that the reaction mechanism for HPA binding and subsequent covalent bond formation with ThDP (as observed by analogy in the DX5P complex), followed by the final enamine intermediate formation, involves a stepwise shortening of the emergent covalent bond between the C2 carbanion of ThDP and the substrate carbonyl, accompanied by an almost 90° rotation of the molecule. Throughout this reaction, the carbonyl O-atom and α -hydroxyl group in HPA remain in the same position, both maintaining hydrogen-bond interactions to His 473 and the amino

group on the pyrimidine ring of ThDP, and additionally between the carbonyl and His 100.

Hydrogen bonds from His 26 and His 261 to just one O-atom of the carboxyl group in HPA mirror those observed previously by crystallography in the covalently associated DX5P complex [21], where the two histidines interact with the C3 hydroxyl group (Figure 1.6). Asp 469 hydrogen bonds to the other carboxyl group O-atom in HPA which also mirrors an interaction with the C4 hydroxyl group in DX5P. However these interactions are all lost upon enamine formation by cleavage of the two-carbon ketol unit from HPA and DX5P.

Surprisingly, the Bergi-Dunitz angle for nucleophilic attack of the carbonyl of HPA by the ThDP carbanion is only 68° , which is a long way from the traditionally preferred 107° [80]. Such an unusual acute angle of attack is potentially made possible through a number of factors. First, the substrate is tightly anchored by interactions to the amino group on the pyrimidine ring of ThDP, His 100 and His 473, whereas Bergi-Dunitz angles of 107° are typically preferred in reactions between two unconstrained molecules. Second, the ThDP may attack in the carbene form, especially in the proximity of the negatively charged carboxyl group in HPA, which can take place from above a pi-system such as in a carbonyl. Finally, the conserved Asp 469 residue appears to be hydrogen bonded to one of the carboxyl group O-atoms in HPA (equivalent to the C1 hydroxyl group of DX5P). This may serve to lengthen the carbonyl bond in HPA giving it less double bond character, along with protonation of the carbonyl by the amino group on the pyrimidine ring of ThDP to

remove electron density from the O-atom, thus making an acute angle of attack more favourable.

FPA was found to dock in an identical manner to HPA with an RMSD of 0.75 Å suggesting that the fluorine atom can hydrogen bond with equivalent contacts to those found with HPA. FPA is less reactive than HPA as the high electronegativity of the fluorine atom decreases the polarity of the neighbouring carbonyl making it less reactive to the ThDP carbanion in TK. The identical binding of docked FPA and HPA indicates that FPA should provide a great opportunity for crystallographers to obtain a non-covalent complex of TK with an analogue of the ketol donor HPA.

The initial formation of a non-covalent ketol donor complex at an unusual angle of attack, through maximised substrate interactions, is consistent with the previous observation of strain in the subsequently formed tetrahedral intermediate [21]. This builds up a picture of the enzyme active site being poised catalytically such that the binding energetics allow the unusually constrained attack angle. The strain in the complex is released partially to form the out-of-plane covalent intermediate, and then released further upon formation of the enamine.

2.4 Conclusions

Although computational docking is not completely accurate, the most obvious errors can be eliminated by visual inspection as was the case for DX5P in this work. Furthermore, the lack of significant side-chain movements in the TK active site upon substrate binding eliminates errors that might otherwise have arisen by not modelling induced fit mechanisms. Computational docking in TK provided supportive evidence for understanding the enzyme mechanism where experimental structural studies have been difficult, and more importantly it has generated interesting hypotheses that can be tested in future experimental studies. The AutoDock algorithm used was able to determine binding conformations of several ketol donor and aldehyde acceptor substrates in the active site of *E. coli* TK with a good correlation between experimental and computationally derived K_m values. We were also able to model aldol acceptors into TK with the ThDP-enamine intermediate present as would be formed after reaction with the ketol donor substrate. The binding of aldol acceptors was found to be identical both in the presence and absence of the enamine intermediate indicating that the interactions made independently of the enamine are sufficient for binding in a reactive conformation. This has an important consequence for protein engineering attempts to alter the substrate specificity of a two-substrate reaction in which the interactions of the second substrate with the enzyme are partially overlapping with those made by the first.

An interesting potential mechanism for the ring opening of cyclic DR5P is suggested by comparison of a previous crystal structure to our docked acyclic DR5P. We have

identified potential general acid and general base residues for the ring opening, and also suggest how the molecule moved around a network of hydrogen bonding interactions to bring the linearised DR5P into place for reaction with the enamine intermediate.

Finally, a comparison of HPA docking to the modelled enamine and to a structurally determined covalent DX5P intermediate indicates a nucleophilic attack by the deprotonated ThDP cofactor with an acute Bergi-Dunitz angle of just 68°, rather than the typically preferred obtuse angle of 107°. A repeat of the HPA docking conformation but with that of the non-reactive analogue FPA suggests a possible method for testing this unusual hypothesis by crystallography.

3 Statistical Coupling Analysis of *Escherichia coli* transketolase

3.1 Introduction

Elucidation of the structure and mechanism of transketolase has led to the identification of several key residues involved in substrate recognition [18, 21], protonation of cofactor [23], transition state stabilisation [7], and deprotonation of substrate. Efforts to engineer the substrate specificity and enantioselectivity of TK have both utilised and built upon this knowledge of structure and mechanism.

Despite the detailed understanding of TK, work to date has focussed on the function of individual residues in isolation. Early experimental mechanistic work on yeast TK involved the site directed mutagenesis of individual residues [18, 19, 24], and engineering of *E. coli* TK has focussed on saturation mutagenesis targeted to individual residues [9-11, 47]. In addition to identifying individual residues with key functions, it is also desirable to identify key synergistic networks of residues. Identification of such networks will lead to a better understanding of function and aid in the engineering of TK.

Statistical coupling analysis (SCA) is a powerful tool for identifying co-evolved residues in protein multiple sequence alignments (MSA) [65, 66]. The co-evolution of residues indicates potential synergy between them that is driven by overall protein fitness or function. Various properties can impact on protein function; these include expression, folding, solubility, stability and allostery; synergistic relationships could be linked to any one of these properties. Here we have used the

SCA method to identify potentially synergistic networks of residues in *E. coli* TK. Other methods to measure co-evolution of residues include Explicit Likelihood of Subset Variation (ELSC), mutual information and correlation-based methods.

Statistical coupling analysis is based on two hypotheses, firstly that without evolutionary constraint, all amino acids at a specific position in a multiple sequence alignment will approach their mean distribution in all proteins. Secondly, that the epistatic coupling of two positions in a protein should mutually constrain their evolution [65]. In order to identify such coupled positions in a multiple sequence alignment, the distribution of amino acids at one site is altered (by removing a subset of sequences from the MSA) and other sites are monitored for concurrent changes in distribution of amino acids. Notably, identification of such relationships does not require that the level of conservation change at the second site, just that the distribution of amino acids be altered.

TK exists as a homodimer of two 680-residue chains, each consisting of three domains, the PP (pyrophosphate binding), the Pyr (pyrimidine binding), and the C-terminal domain. All TPP-dependent enzymes contain catalytic PP and Pyr domains which bind the ThDP cofactor, although different enzyme types have different domain architecture [81, 82]. Although the TPP-binding sites of these enzymes are very similar, each enzyme type has different substrate specificity. As the different enzyme types have diverged from a common ancestor, any synergistic networks of residues could be expected to create evolutionary constraints on the residues involved. Identification of such co-evolved networks could provide important

insights into substrate specificity and aid in the engineering of substrate specificity of *E. coli* TK.

The wild type *E. coli* TK enzyme has been engineered for improved substrate specificity towards the non-hydroxylated aldehyde acceptor substrate, propionaldehyde (PA) [10]. The D469T mutant was identified as giving the greatest improvement in activity on PA with a 5-fold increase in specific activity relative to wild type. A further mutant at site 469, D469Y, displayed the greatest substrate specificity with a 64-fold higher activity on PA relative to GA. Finally the D469E mutant displayed the greatest improvement in enantioselectivity with PA [10]. The repeated identification of Asp 469 variants among seventeen active site residues probed for three different selection criteria illustrates the importance of this residue. Identification of networks of synergistic residues involving Asp 469 could lead to further improvements in the engineering of *E. coli* TK.

Using SCA, we have identified residues that are statistically coupled to Asp 469. These include both a proximal connected network and a more distal and network distributed throughout the protein fold. Some of the residues had been previously identified as improving activity on non-natural substrates [9, 10] but the vast majority have not been investigated by mutagenesis before and represent new potential targets for rational design of expanded variant libraries.

In addition to measuring coupling between individual residues in an alignment, SCA allows global coupling analysis across all positions in the alignment that meet certain criteria. Two-dimensional hierarchical clustering, originally developed for microarray data analysis, can be used to identify networks of similarly coupled

residues throughout the protein sequence. For robust and conserved networks of residues, perturbations at network positions are expected to redundantly identify each other, clustering the coupling matrix allows the identification of such networks. We applied this method to identify inter- and intra-domain networks of coupled residues throughout the PP and Pyr domains of *E. coli* TK. Many of these networks were located in the interface between the two domains but we also identified interesting connected networks of residues in and around the active site.

3.2 Materials and Methods

3.2.1 Multiple sequence alignments

Costelloe *et al* carried out a phylogenetic analysis of seventeen different TPP dependant enzymes using sequence alignments of the conserved PP and PYR domains [82]. Transketolase (TK), D-xylulose-5-phosphate synthase (DXPS), dihydroxyacetone synthase (DHAS), phosphoketolase (PKL), 2-oxoisovalerate dehydrogenase (2OXO), pyruvate ferredoxin reductase (PFRD), pyruvate decarboxylase (PDC), indolepyruvate decarboxylase (IPDC), phenylpyruvate decarboxylase (PhPDC), pyruvate oxidase (PO), acetolactone synthase (ALS), glyoxylate carboligase (GXC), benzoylformate decarboxylase (BFDC), benzaldehyde lyase (BAL), oxalyl CoA decarboxylase (OCADC), sulfopyruvate decarboxylase (SPDC), and phosphopyruvate decarboxylase (PPDC) enzymes were included.

Costelloe *et al* identified TPP dependent enzyme sequences using BLASTP searches of the Swissprot and nr databases (Blosum62 matrix and default settings). Query sequences for each enzyme were selected from those with known structure or well defined biochemistry and homologous sequences were identified with >30% sequence identity. In total 382 sequences representing the 17 different enzymes were identified. Following removal of putative sequences these hits were aligned using ClustalW.

Alignments of the PP and PYR domains were generated separately. The PP domain was defined as residues 1-350 in *E. coli* TK and the PYR domain was defined as residues 323-528 in *EcoTK*. Crystal structures of *EcoTK* (1QGD.pdb), *P. putida* 2OXO

(2BP7.pdb), *D. africanus* PFRD (1B0P.pdb), *S. cerevisiae* PDC (1PVD.pdb), and *L. plantarum* PO (1POX.pdb) were used to refine the alignment of functionally important residues and secondary structural elements. Finally alignments were de-gapped to leave only residues found in *E. coli* TK.

3.2.2 Statistical coupling energy calculation

SCA Version 1.5 was used to calculate statistical coupling energies between sites in multiple sequence alignments (MSA's) of the PP and PYR domains. SCA Version 1.5 is a MATLAB implementation of the calculation described by Lockless et al, 1999 [65]. The calculation gives a quantitative measure of the change in amino acid distribution at site j given a perturbation at another position i . This is calculated as a statistical coupling energy ($\Delta\Delta G_{j,i}^{\text{stat}}$).

Briefly, each site in the MSA is described by a 20 element vector of binomial probabilities of individual amino acid frequencies given their frequencies in all proteins ($P_j = [P_j^{\text{ala}}, P_j^{\text{cys}}, P_j^{\text{asp}}, \dots, P_j^{\text{tyr}}]$). P_j^x gives the probability of the observed number of x amino acids at position j , given its mean frequency in all proteins. The 20 element vector of P_j^x can then be converted into a vector of statistical energies that represents the evolutionary constraint at site j ($\Delta G_j^{\text{stat}} = [\Delta G_j^{\text{ala}}, \Delta G_j^{\text{cys}}, \Delta G_j^{\text{asp}}, \dots, \Delta G_j^{\text{tyr}}]$). Each term in the vector is the value for amino acid x at site j and is given by $\Delta G_j^x = kT^* \ln(P_j^x/P_{\text{MSA}}^x)$, where kT^* is an arbitrary energy unit and P_{MSA}^x represents a hypothetical site where all amino acids are observed at their mean frequencies in the MSA as a whole. This hypothetical site serves as a reference state for all sites and ΔG_j^x represents the statistical free energy separating site j from the

hypothetical site for amino acid x by the Boltzmann distribution. The magnitude of the ΔG^{stat}_j vector represents an evolutionary conservation parameter for site j.

To measure functional coupling between two positions j and i in the MSA, two statistical energy vectors are calculated, one from the full MSA (ΔG^{stat}_j) and one from a subalignment representing a perturbation of the amino acid frequencies at a second site i ($\Delta G^{\text{stat}}_{i|j}$). The magnitude of the difference in these two statistical energy vectors represents a quantitative measure of the degree to which the probability of individual amino acids at site j is dependent on the perturbation at i, $\Delta\Delta G^{\text{stat}}_{j,i} = \Delta G^{\text{stat}}_j - \Delta G^{\text{stat}}_{i|j}$. $\Delta\Delta G^{\text{stat}}_{j,i}$ is calculated for all sites j given a perturbation at position i.

3.2.3 Matrix assembly and cluster analysis

All acceptable perturbations, and their associated coupling energies to other positions in the sequence, are displayed as a matrix of statistical coupling energies. Each row in the matrix represents a position in the alignment and each column represents a specific perturbation.

Iterative clustering methods originally developed for microarray analysis are then used to identify co-evolving networks of positions. The process of iterative clustering involves sequential rounds of 2-dimensional clustering, after each round, sub-matrices representing areas of low signal are eliminated. The next round of clustering further refines the clusters focussing around positions and perturbations with significant statistical coupling. This process is repeated until the clusters converge and no further refinement is possible.

3.3 Results and Discussion

3.3.1 TPP-dependent multiple sequence alignment validation and perturbation of position 469

A 382 protein multiple sequence alignment (MSA) of the PP and PYR domains of TPP-dependent enzymes was converted into a matrix of residue frequencies and positions using SCA version 1.5. Calculation of evolutionary constraint at each site in the MSA reveals a diverse and well evolved collection of proteins. Figure 3.1 a illustrates graphically the evolutionary constraint at each position as measured by ΔG^{stat} . High values of ΔG^{stat} indicate a high degree of conservation or evolutionary constraint. Among the residues which display a very high degree of evolutionary constraint are Asp 155, Glu 411, and His 473. These three residues have important functions in the active site of TK. Asp 155 acts as a ligand for the metal ion Ca^{2+} or Mg^{2+} [14], Glu 411 protonates the N_1' Nitrogen of the pyrimidine ring [23], and His 473 has a potential involvement in transition state stabilisation [7].

The MSA must be validated before carrying out SCA to ensure that it meets certain criteria. In order for functional constraints in the MSA to be exposed, the alignment should have diversified to the extent that frequencies of amino acids at un-conserved sites have relaxed near to their mean values in all natural proteins [65]. This is evident from the frequencies of residues at the un-conserved site 38 which are approaching the mean frequencies of residues in all proteins (Figure 3.1 b). This is in contrast to a conserved site such as 469 where the amino acid frequencies differ considerably from the mean values in all proteins (Figure 3.2 a).

Secondly, the MSA should be large enough that random elimination of sequences does not change the amino acid frequencies at un-conserved sites [65]. To validate the MSA for this criteria, the five least conserved positions that still represent at least 85% occupancy were identified (1QGD numbered positions: 38, 211, 363, 508, and 509) and average ΔG^{stat} values were determined across these five sites following elimination of different proportions of the MSA. Random elimination of large proportions of the MSA was possible without increasing the average ΔG^{stat} at these un-conserved positions (Figure 3.1 c). The alignment was therefore judged to have reached statistical equilibrium in sequence space, a necessary condition for applying Boltzmann statistics.

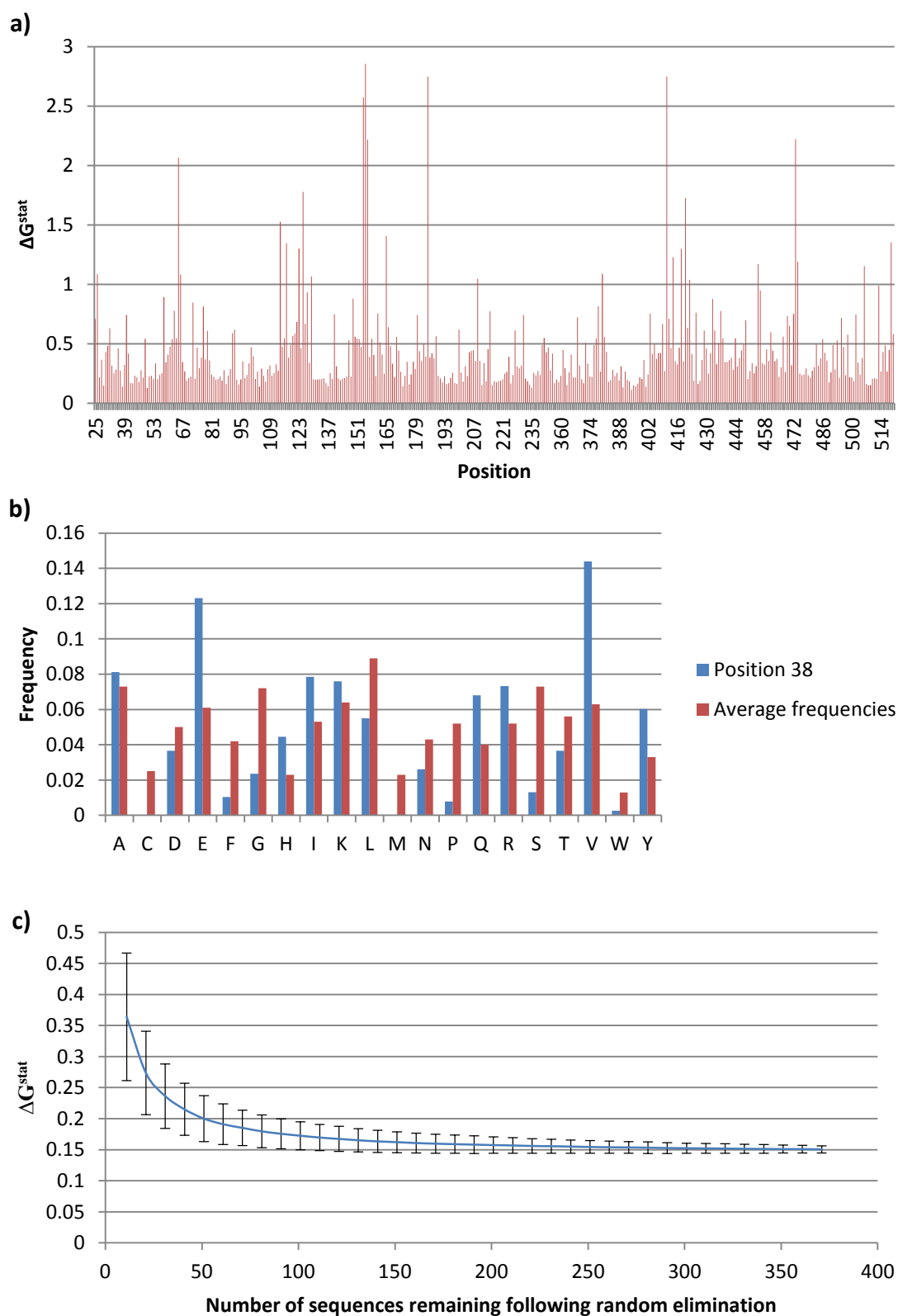


Figure 3.1 a Overall conservation across the MSA as measured by dG_{stat} , an expression of divergence from the frequency of residues expected in all proteins. b Frequency of residues at position 38, the least conserved position in the MSA. c The 5 least conserved sites that retain at least 85% occupancy were selected and average dG_{stat} at these 5 sites was evaluated following random elimination of increasing proportions of the MSA.

3.3.2 Identification of residues statistically coupled to Asp 469

Residue Asp 469 is known to be a functionally important residue in *E. coli* TK. Early studies identified the equivalent residue in yeast TK as having an important function in enantioselectivity [24] and recent work to engineer *E. coli* TK to accept non-natural substrates has identified several variants at this position that confer improved and desirable properties to the enzyme [9, 10]. Given the important function of this residue we used SCA to identify residues which display statistical coupling to Asp 469. Identification of such a network has important implications for the understanding of TK evolution and function but could also be applied to aid in the design variant libraries for engineering TK.

The residue Asp 469 is highly conserved in TK and in some of the other TPP-dependent enzymes (Figure 3.2 a). Using the SCA Toolbox we created a perturbation of the MSA to remove all sequences that do not contain an Asp at position 469, this perturbation resulted in a subalignment of 173 sequences. Statistical coupling to position 469 was then determined by calculation of the $\Delta\Delta G^{\text{stat}}$ values at all other positions in the alignment (Figure 3.2 b).

The statistical coupling energies between the 469D perturbation and other positions in the alignment are generally insignificant, however a relatively small number of positions display significant coupling to the 469D perturbation. Focusing on the seven sites displaying the highest levels of coupling to position 469 identifies an interesting cluster of residues surrounding the active site.

Mapping these sites onto the structure of *E. coli* TK reveals a network of connected residues within the protein core that display coupling with position 469.

Interestingly these are not restricted to residues in close proximity to position 469, coupling is also seen in distant residues and across the PP-PYR domain boundary (Figure 3.2 c).

Focusing on the active site of *E. coli* TK and the Asp 469 residue itself, four of the seven coupled residues form a connected cluster in and around the active site tunnel which is centred on Asp 469 (Figure 3.2 c). This cluster spans from residue Phe 437 to Thr 472 and then to Asp 469 of the PP domain. Asp 469 then forms a connecting bridge across the inter-domain barrier to His 26 which in turn interacts with His 66. Arg 520 is not physically in contact with this contiguous cluster but is relatively close to Asp 469 and reaches into the active site forming part of the entrance to the active site tunnel.

The two remaining residues, Tyr 72 and Pro 486, are more distant from the active site. Tyr 72 is located on the α -helix which leads towards His 66 of the active site and may therefore be responsible for positioning the loop containing His 66 to maintain the integrity of the active site. Pro 486 forms a hairpin loop at the opposite end of an α -helix which leads to residues Thr 472 and Asp 469. This residue may therefore have a similar function to Tyr 72 in maintaining tertiary structure and active site integrity.

Of the seven residues identified as coupling with Asp 469, two have been identified previously for their propensity to improve activity on non-natural substrates when mutated [9, 10]. Arg 520 was identified in screens for improved activity on both GA and PA and His 26 was identified in screens for improved activity on PA [10]. As Arg 520 is known to interact with the phosphate group of natural substrates, a rational

explanation for the increase in activity for non-phosphorylated substrates was a removal of the steric bulk around the entrance to the active site. The coupling between Arg 520 and Asp 469 suggests there may be a subtler synergistic explanation for the increased activity. Mutation of His 26 in *E. coli* TK led to variants with reversed enantioselectivity for PA [11]. This residue forms a pocket with Asp 469 which interacts with the α -hydroxyl group of natural substrates, coupling is not surprising for a residue with such a close functional and structural relationship with Asp 469.

Apart from Asp 469, His 26, and Arg 520, none of the remaining three active site residues in this network have been targeted for mutagenesis. The high hit-rate for obtaining improved mutants among these three sites indicates that similar success may be obtained at the other sites in the cluster. Identification of this network provides a good starting opportunity for creating targeted libraries of single and multiple mutants with potentially improved function on non-natural substrates.

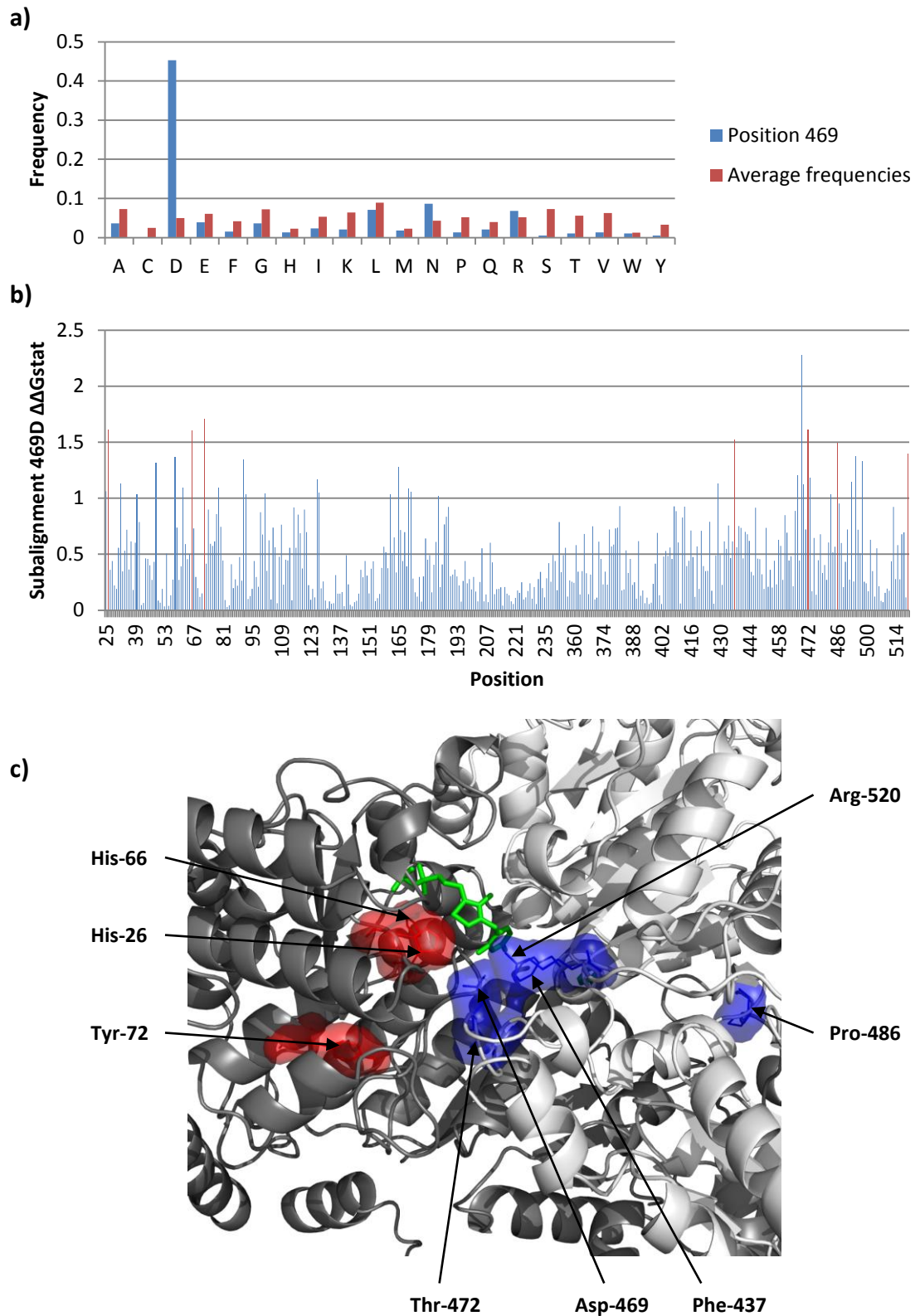


Figure 3.2 a Frequency of residues at position 469 in the MSA versus the expected frequency of residues in all proteins. b $\Delta\Delta G^{\text{stat}}$ values between the 469D perturbation and different positions in the MSA. The seven positions representing the highest coupling energies to position 469 are coloured red. c The seven positions coupled to 469 mapped onto the structure of *E. coli* TK (Asp 469 is also shown). Residues in Chain A are coloured red and in chain B are coloured blue. A structurally contiguous group of coupled residues spans the PYR domain of chain A and the PP domain of chain B around the active site tunnel.

3.3.3 Global SCA analysis of TPP-dependent enzymes

The results above demonstrate the ability of SCA to identify residues displaying evolutionary constraint to perturbations at residue Asp 469. Both proximal connected and distant unconnected residues were identified that express an evolutionary dependence on the identity of the residue at position 469. The residues identified as coupled to Asp 469 may be part of a larger interconnected network of coupled residues, not necessarily directly coupled to Asp 469. To identify such an extended network of interactions and to identify other networks that don't include Asp 469 we need to carry out a global SCA across the whole alignment.

To conduct a global SCA, each site in the MSA is subjected to perturbations to create subalignments. Perturbations are allowed that produce sub-alignments large enough to not represent global changes in conservation relative to the parent alignment [65]. To identify the cut off for sub-alignment size, the statistical coupling energy ($\Delta\Delta G^{\text{stat}}$) at un-conserved sites is monitored following random eliminations of different fractions of the MSA. The fraction of sequence elimination that begins to show coupling at un-conserved sites represents the limit of sequence elimination tolerated for the specific alignment. For the MSA of TPP dependent enzymes, un-conserved residues begin to display coupling where sub-alignments of less than 80 sequences are selected (Figure 3.3). Therefore for global SCA, perturbations were allowed that resulted in sub-alignments greater than 0.21 as a fraction of the total alignment size (382 sequences).

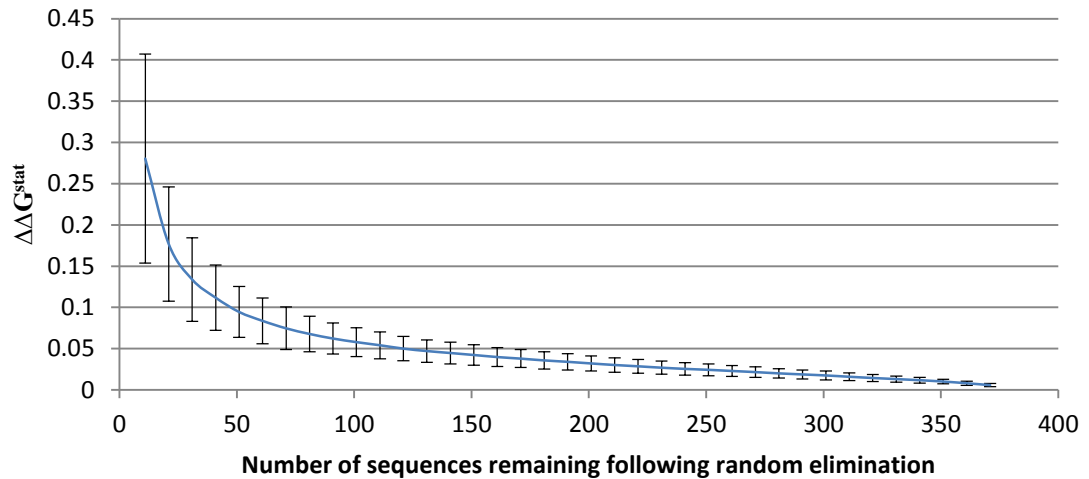


Figure 3.3 The 5 least conserved sites that retain at least 85% occupancy were selected and average $\Delta\Delta G^{\text{stat}}$ values were calculated at the 5 sites following random elimination of different proportions of the MSA.

Global SCA was carried out on the MSA in line with the criteria determined above and $\Delta\Delta G^{\text{stat}}$ values were determined for all positions in the alignment. Figure 3.4 shows the initial matrix with perturbations represented by columns and positions represented by rows. There is not a perturbation for each position in the alignment as some positions could not meet the cut-off for subalignment size. Therefore the matrix is naturally taller than it is wide. $\Delta\Delta G^{\text{stat}}$ values between residues are displayed as a linear colour scale from blue (0) to deep red (4). The highest level of coupling to position 469, identified following the Asp 469 perturbation, was Tyr72 with a $\Delta\Delta G^{\text{stat}}$ of 1.72. In contrast, many of the coupling interactions identified following global SCA are associated with coupling energies ($\Delta\Delta G^{\text{stat}}$) greater than 3 (Figure 3.4).

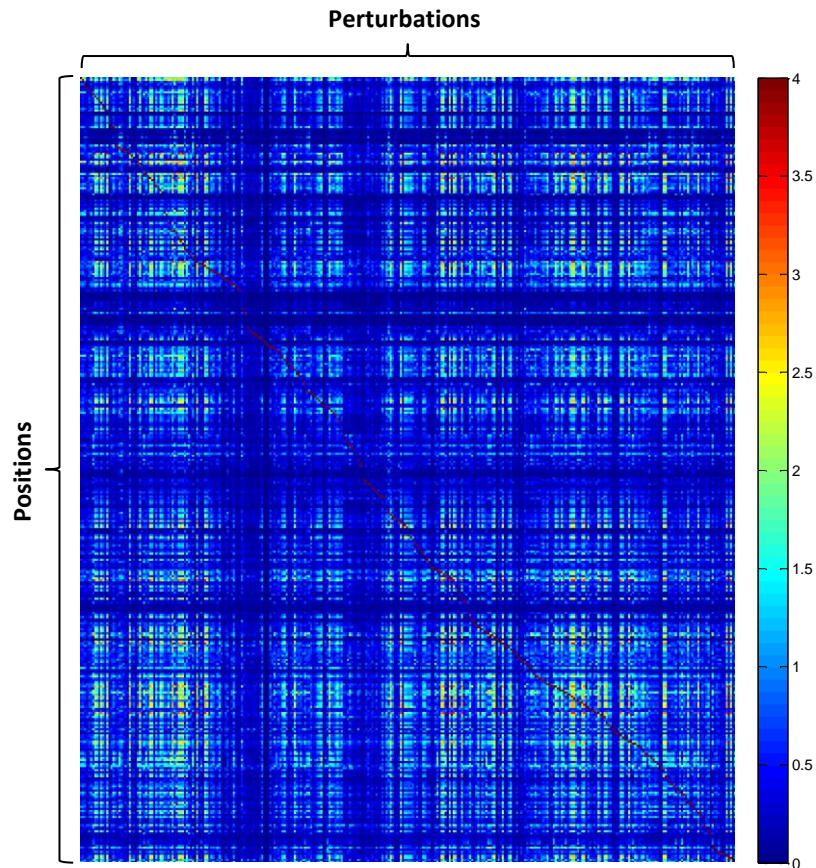


Figure 3.4 Matrix of $\Delta\Delta G^{\text{stat}}$ values following global statistical coupling analysis of TPP dependent enzymes. Columns represent specific perturbations of the MSA. Rows represent positions in the PP and PYR domains.

We carried out 2-dimensional hierarchical clustering to identify networks of coupled residues. Figure 3.5 shows the initial round of clustering. Large areas of the matrix are made up of low level coupling. Following clustering, large groups of perturbations or positions with low coupling energy were sequentially removed and the matrix was re-clustered to iteratively focus on the networks representing the highest statistical coupling.

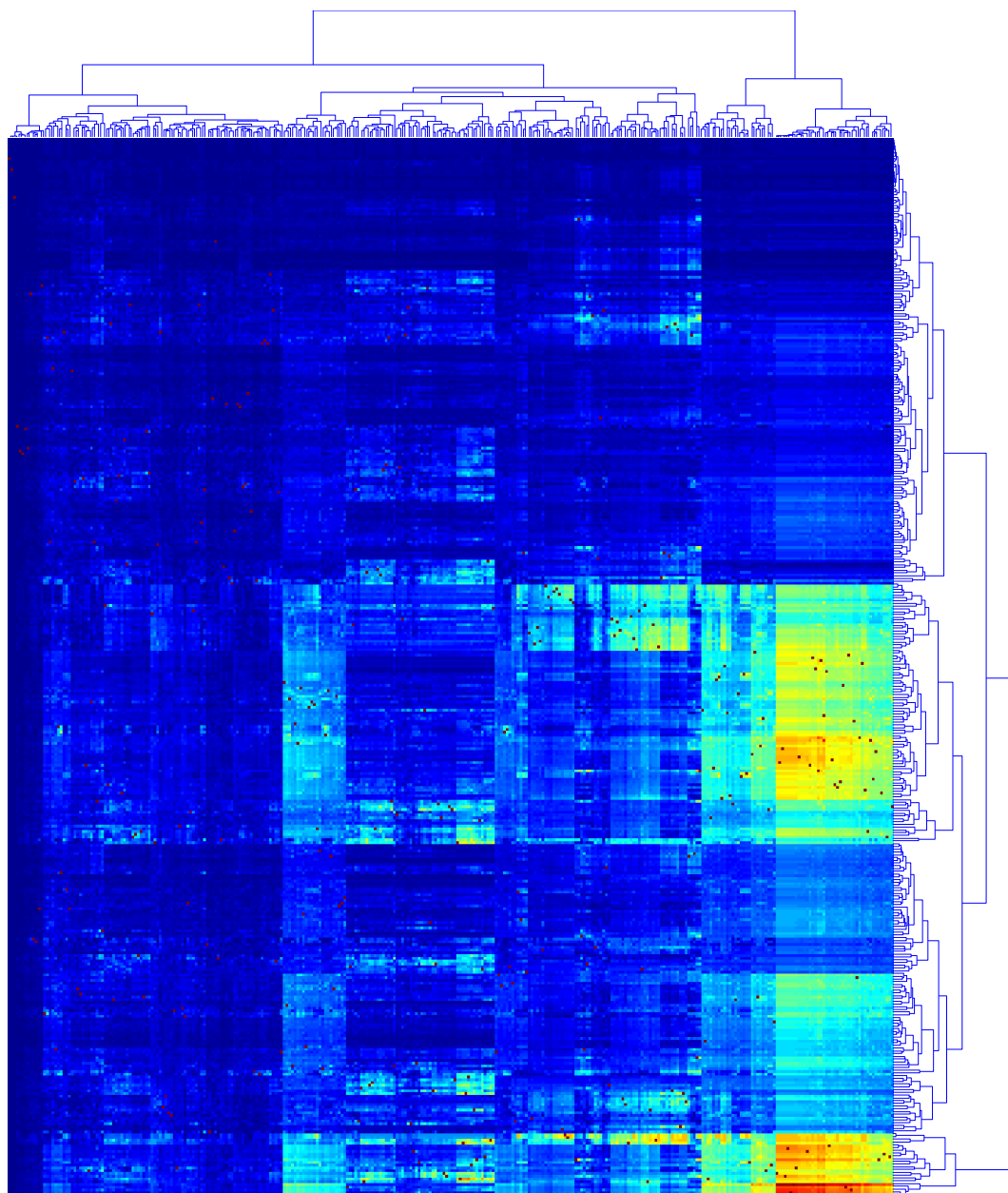


Figure 3.5 Initial clustered matrix of coupling energies following global perturbation analysis of PP and PYR domains.

Following five rounds of iterative clustering we identified a highly coupled network of 30 residues and 45 specific perturbations (Figure 3.6). Based on the clustering dendrogram, the 30 residues can be clustered into six groups of different sizes representing different coupling profiles. Six of the positions identified are known to have important functions in *E. coli* TK (Table 3.1). These include His 26 which forms a hydrogen bond with the α -hydroxyl group of natural acceptor substrates, His 66 which forms a hydrogen bond with the diphosphate group of ThDP and interacts

with the C1 hydroxyl group of donor substrates, Tyr 440 and Phe 434 which interact with the pyrimidine ring of ThDP, His 461 which binds the phosphate group of both donor and acceptor substrates, and Ile 187 which interacts with the divalent metal ion through its main chain oxygen. The identification of functionally defined residues demonstrates the robustness of SCA and indicates a potential application of the method where structure and functional information is lacking. Perhaps more interesting than the known residues are the residues for which we are unaware a function.

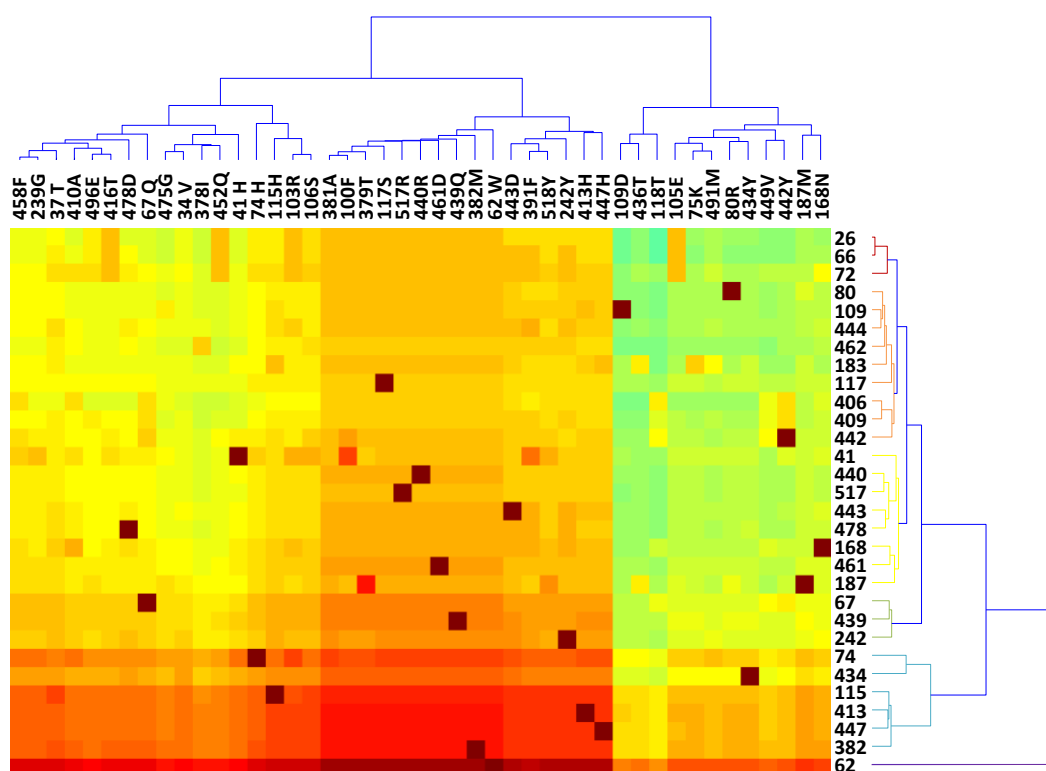


Figure 3.6 Final global SCA matrix following iterative focusing and reclustering around areas of high signal. As in previous figures, perturbations are represented by columns and positions by rows. The dendrogram of positions is coloured according to clusters.

The thirty residues identified with strong coupling interactions are not the most conserved residues in the alignment. This can be expected as highly conserved positions in an alignment are less likely to display changes in amino acid distribution

upon perturbation. Comparison of the thirty residues identified by SCA and the thirty most conserved residues in the alignment, as measured by evolutionary constraint (ΔG^{stat}), identifies only three residues present in both lists. These are His 26, His 66, and Gly 117. ΔG^{stat} , a measure of conservation, for the thirty highly coupled residues ranges from 0.27 to 1.35 with a mean of 0.51; for the thirty most conserved residues, ΔG^{stat} ranges from 0.88 to 2.86 with a mean of 1.50. Conservation is often used as the first tool to indicate potentially functional residues where structural and functional information is lacking, here we demonstrate an alternative approach to the identification of potentially functional residues which can complement the simple test of conservation.

Group	EcoTK Residue	Position	Known function
1	His 26	Active site surface	Catalysis/stereospecificity
	His 66	Active site surface	Substrate recognition
	Tyr 72	Internal	
2	Tyr 80	Internal	
	Val 109	Internal	
	Ala 444	Internal	
	Asp 462	Internal	
	Asp 183	Internal	
	Gly 117	Internal	
	His 406	Interdomain surface	
	Val 409	Interdomain surface	
	Arg 442	Interdomain surface	
3	Trp 41	Internal	
	Tyr 440	Interdomain surface	Forms hydrophobic pocket
	Ile 517	Internal	
	Asn 443	Interdomain surface	
	Gln 478	Internal	
	Ser 168	Interdomain surface	
	His 461	Interdomain surface	Phosphate binding
	Ile 187	Internal	Metal binding
4	Gly 67	Internal	
	Glu 439	Interdomain surface	
	Met 242	Internal	
5	Leu 74	Internal	
	Phe 434	Interdomain surface	Forms hydrophobic pocket
	Pro 115	Internal	
	Gly 413	Internal	
	Met 447	Interdomain surface	
	Leu 382	Interdomain surface	
6	Leu 62	Internal	

Table 3.1 Hierarchical clusters of coupled positions identified (Figure 3.6) together with their identities in *E. coli* TK and their function where known.

When mapped onto the surface of the *E. coli* TK enzyme, the thirty residues identified display an interesting pattern. Relatively few are exposed on the surface of the holoenzyme apart from in and around the active site (Figure 3.7 a). However, when the two chains of TK are separated (Figure 3.7 b) we see several of the coupled residues exposed on the subunit surface in a connected network which spans out from the active site itself. Rotating the two chains apart further displays the surface residues identified that make up the interface between the two

subunits (Figure 3.7 c). In total, fourteen of the thirty residues identified are located on the subunit surface in and around the active sites.

Some of the interface residues form the binding pocket for the ThDP cofactor, these include Asp 183, Ile 187, Val 409, Phe 434, and Tyr 440. Four residues that make up the active site entrance tunnel are also identified, Lue 382 and His 461 form one side of the active site tunnel and His 26 and His 66 make up the other side. The other five surface residues are located on the interface between the two subunits and are buried in the holoenzyme. These include His 406, Arg 442, Asn 443, Glu 439 and Met 447. Presence of coupled networks of residues on the interface between the two subunits of transketolase could be expected, as a change in the residue identity on one surface of the inter-subunit interface may require a corresponding change on the other face to avoid deleterious effects on subunit aggregation. This also holds true for residues on opposite sides of the active site or on opposite sides of the cofactor binding site.

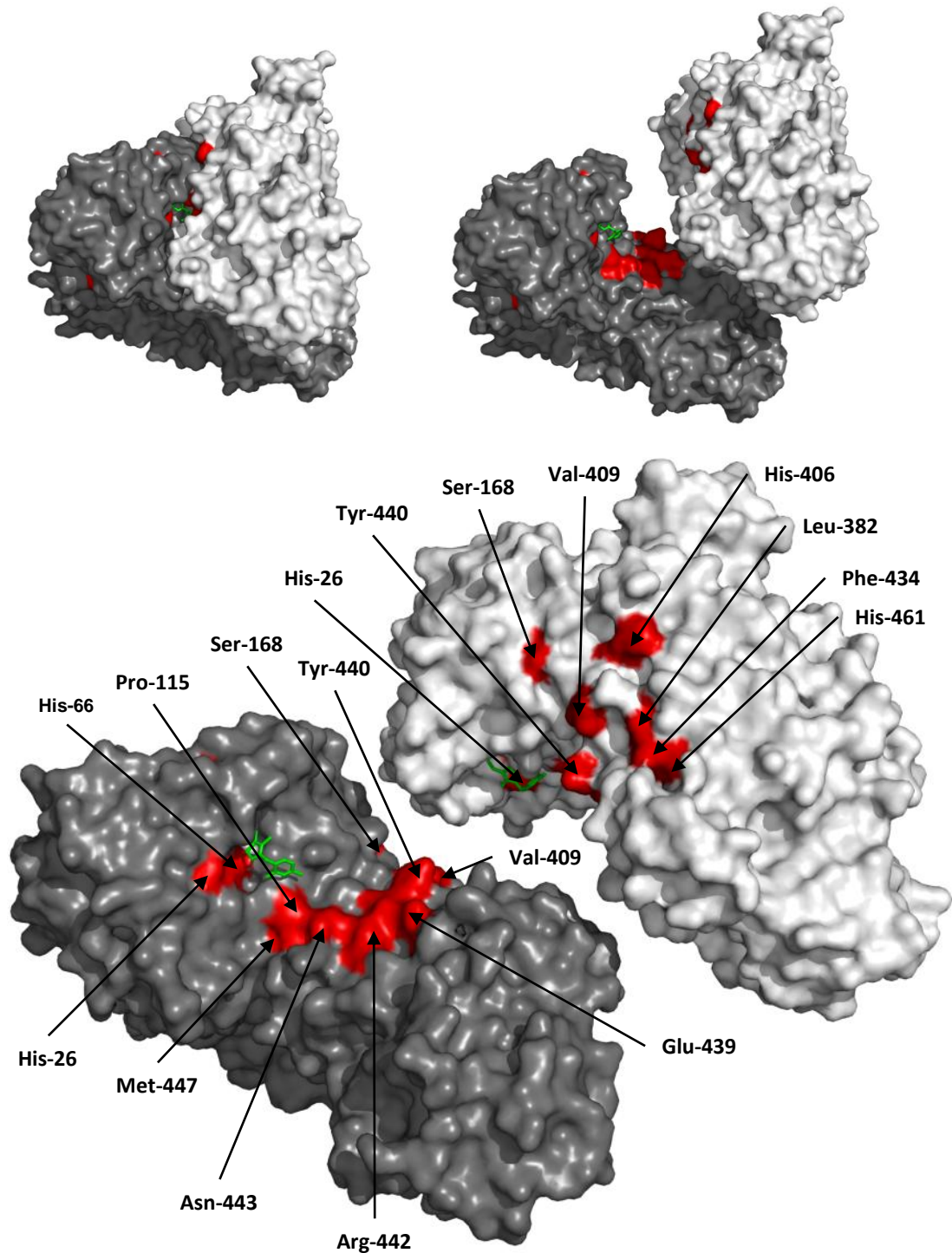


Figure 3.7 Surface rendering of TK showing two chains in different shades of gray and the coupled residues identified in red. a *E. coli* TK showing the active site tunnel with the TPP cofactor in green. b Separation of the two chains to expose the coupled residues that form the interaction face. c Rotation of the A chain by -45 degrees and the B chain by +45 degrees to expose the fourteen coupled residues in the interaction face and the active site.

In order to see the other sixteen coupled residues identified through SCA we need to look into the centre of the protein structure. In Figure 3.8 we see the same view

points but the individual clusters of coupled residues have been surface rendered and they are visualised with the cartoon secondary structure of the PP and PYR domains of chain B and coloured by hierarchical clustering. Although connected networks are apparent among the thirty residues, they are relatively widely distributed throughout the subunit. Examination of the residues in the structure indicates the presence of connected networks, but despite one or two exceptions, grouping of the residues by hierarchical clustering does not resolve all of these networks (Table 3.1). Several connected networks include residues from more than one clustering group. Despite the difficulty in resolving the individual networks, the majority of the residues identified are located on the subunit interface or one layer back from this interface, suggesting that the overriding pressure driving the coevolution of coupled networks in TK and other TPP-dependent enzymes is to support the interactions between the PP and Pyr domains which together form the active site.

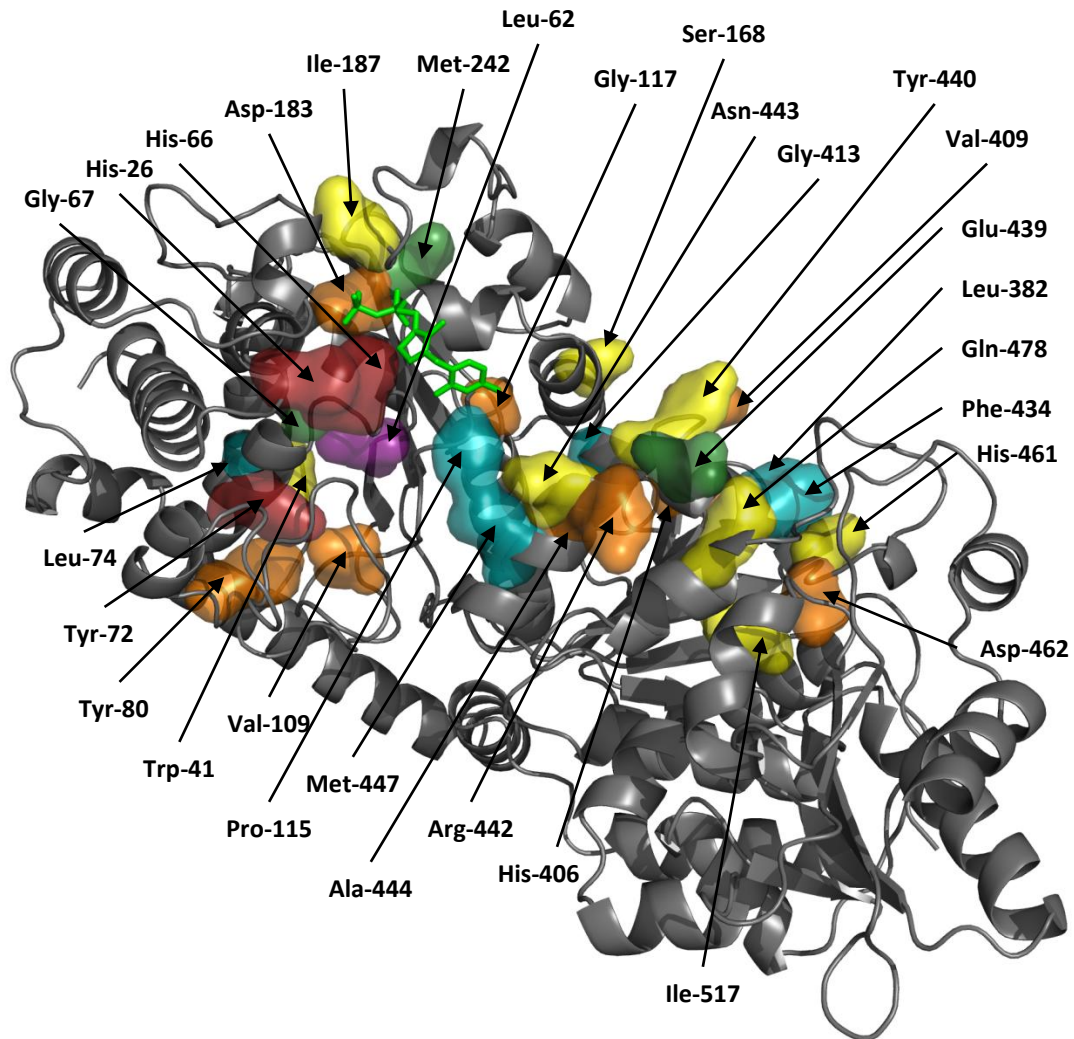


Figure 3.8 All residues identified following global SCA of PP and PYR domains. Residues are coloured by the cluster groupings in Figure 3.6 and mapped onto one subunit of *E. coli* TK.

Although Asp 469 was not identified by global SCA (Asp 469 displays relatively low coupling compared to the highly coupled residues identified), three of the seven residues that are coupled with Asp 469 were identified and these residues were clustered into an independent group by hierarchical clustering. These residues were His 26, His 66 and Tyr 72, the three PP residues identified for their coupling to Asp 469.

This group of residues has a distinct coupling pattern involving good coupling signals with the perturbations 416T, 452Q, 103R, and 105E (Figure 3.6). In contrast

to the vast majority of the coupled residues identified, this group did not co-select their perturbations in the final clustered matrix. This may be due to a distinct group of perturbations specific to the His 26 cluster being removed during the iterative clustering process. This further demonstrates the independence of this cluster from the other residues identified. The location of the three residues, in the active site (His 26 and His 66) and deep within the protein fold (Tyr 72) further suggests a co-evolutionary constraint on catalytic function as opposed to structural dimerization pressures which may explain the coupling of other residues.

The highly coupled but independent residue 62 is also located close to His 26 group. Residue 62 is not highly conserved but it exhibits the highest level of coupling of all the residues in the sequence and falls in a separate, independent cluster. We are unaware of the function of this residue but it displays particularly high coupling energies with other functionally defined residues and its function warrants further investigation.

3.3.4 Single PYR domain global SCA analysis

Position 469 did not display strong enough coupling to be selected in the network of highly coupled residues identified above, where the strongest coupling was seen in the interface between the two subunits of TK. The homodimer of holo-TK is formed through interaction of the PP domain of one subunit interacting with the Pyr domain of the other subunit. In order to identify subtler networks of coupled residues within the Pyr domain of TK we carried out global SCA on just the Pyr domain alignment. By focusing on the Pyr domain we hope to identify networks

containing the 469 position which could aid in the design of multiple mutant libraries with enhanced properties over the individual 469 variant libraries.

The MSA was divided to leave just the Pyr domains and global SCA was carried out as described above. Following initial clustering, a similar set of residues were identified with high coupling energies indicating that the coupling networks even if driven by inter-domain interactions can be identified within one domain in isolation. In addition to the high signal clusters, an interesting cluster of coupled residues was identified in a separate part of the matrix (red sections of dendrograms in Figure 3.9 a below). Upon closer examination this cluster was found to contain Asp 469.

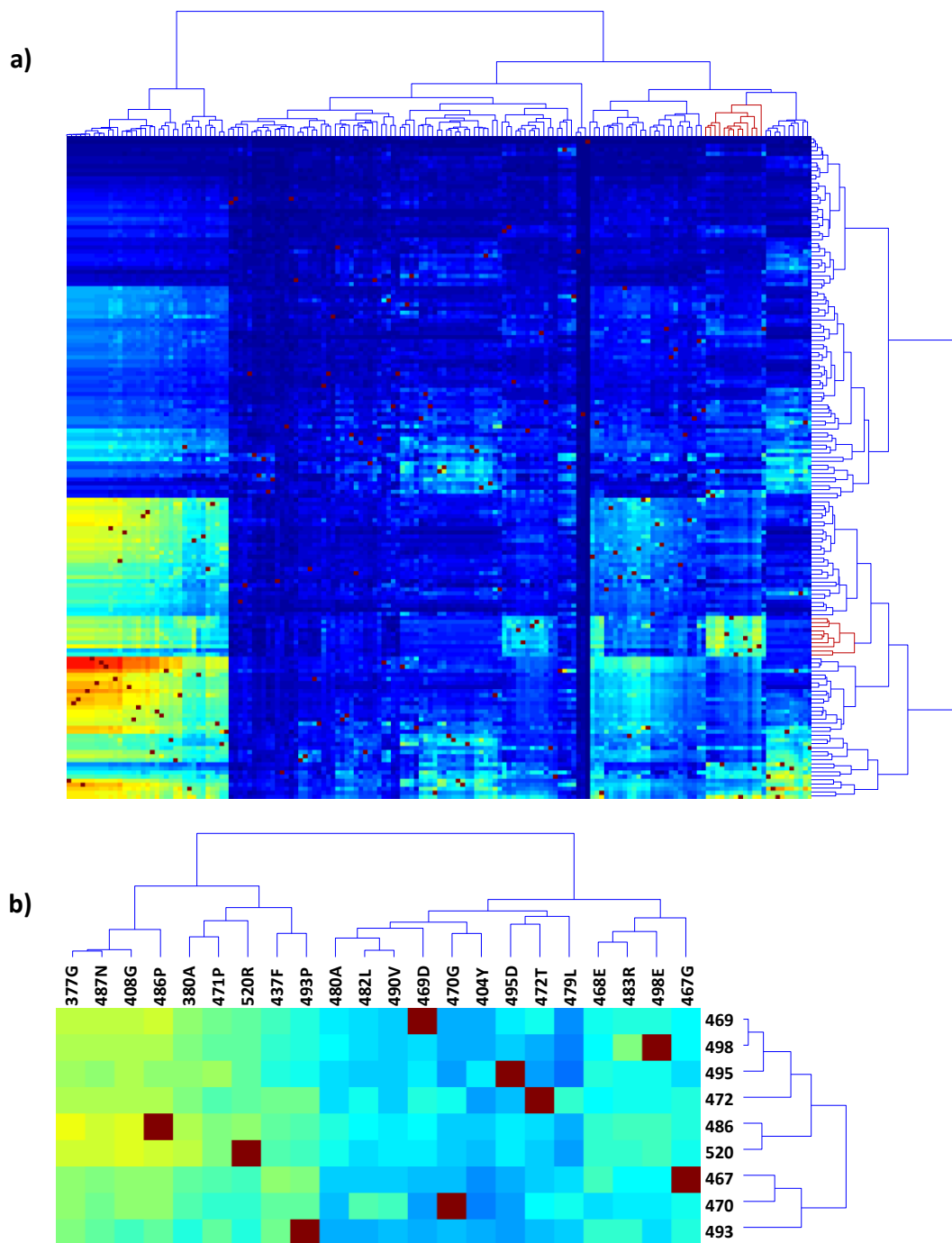


Figure 3.9 a Clustered matrix of $\Delta\Delta G^{\text{stat}}$ values following global SCA analysis of PYR domains. Interesting signal cluster containing position 469 identified by red colouring of dendrogram on perturbations and positions. b Matrix following iterative focusing and reclustering around interesting cluster identified in initial clustering

Iterative clustering to focus down to this cluster results in the final matrix seen in Figure 3.9 b. Nine positions are identified in this group including the 469 position. In contrast to the three position His 26 cluster identified following global SCA of the

PP and Pyr domains, this cluster shows self-consistency: the nine positions are also represented in the perturbations that identify them. Such self-consistency gives confidence in the robustness of the evolutionary coupled network.

The nine positions identified include Gly 467, Asp 469, Gly 470, Thr 472, Pro 486, Pro 493, Asp 495, Glu 498, and Arg 520. Three of these residues, Thr 472, Pro 486 and Arg 520, were previously identified by their coupling energy with Asp 469 but the remaining residues have not been identified before. When these positions are mapped onto the PYR domain, two connected networks are visualised, the first containing residues Asp 469, Gly 467, Gly 470, Thr 472, and Arg 520 and the second containing residues Pro 493, Asp 495, and Glu 498 (Figure 3.10 a). The ninth member of this conserved network, Pro 486, is located at a distant point relative to the other groups. However, in *E. coli* TK, Pro 486 forms a tight hairpin turn in the protein structure between an alpha helix and a beta sheet (Figure 3.10 b). This turn forms at a midway point between residues in the two groups described above. It seems likely that the coupling of Pro 486 to the other two groups is related to its function in maintaining the tertiary structure around these two groups of important residues.

By just selecting residues evolutionarily coupled to Asp 469 we identified part of the cluster above. By carrying out a global SCA of the Pyr domain we have identified other physically connected members of this network together with a second, distant group linked to the first via the Pro 486 hairpin turn.

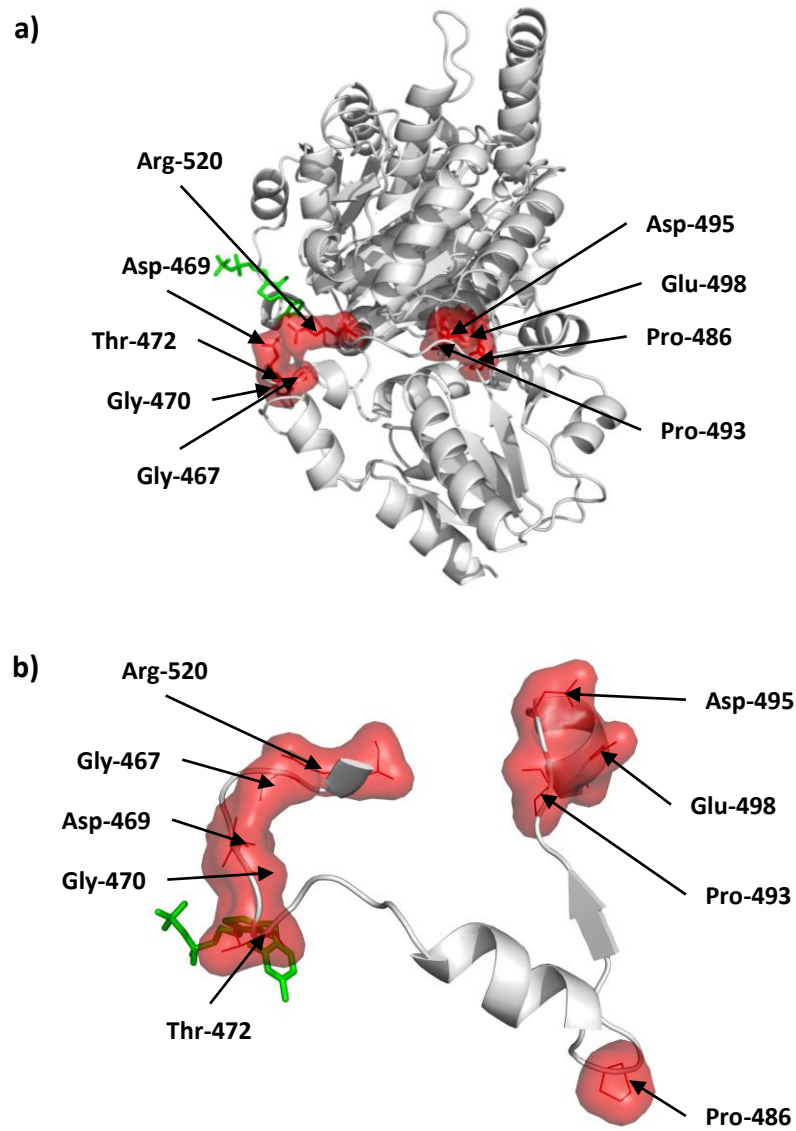


Figure 3.10 a Group of residues coupled to 469 when focusing on PYR domain. PYR domain of chain B displayed. b chain B is rotated away from view by 90 degrees in the X-plane to display relationship between the residues in the cluster. Secondary structure is only displayed for the region spanning residues 465 to 500 to emphasise the relationship between the residues

3.4 Conclusions

We initially investigated the evolutionary coupling between Asp 469 and other positions throughout the MSA of TPP-dependent enzymes. This approach identified seven residues which were either close in proximity to Asp 469 or more distant but with a potential to impact on the tertiary structure around Asp 469. In addition to residues in the same subunit of TK, Asp 469 appears to express evolutionary coupling to sites in the other subunit of the enzyme across the interface barrier.

Having identified residues coupled to Asp 469 we carried out global SCA on the whole MSA to identify further networks within the protein fold. Global SCA identified coupling energies far greater than those associated with Asp 469. Focusing on those high energy positions identified thirty residues dispersed across the two domains. The location of these residues in the interface between the two subunits of TK suggests that co-evolution of the networks identified is driven by structural requirements for dimerization to form the active site. Identification of such interaction surfaces indicates a potential new application for SCA in the identification of protein-protein interaction sites by combining the multiple sequence alignments of proteins known to interact.

Although the clustering of the highly coupled residues did not appear to resolve the detailed sub-networks between the thirty residues, one cluster of three residues did demonstrate structural connectivity and did not appear to be involved in interface interactions. The three residues in this group had previously been identified as the only PP domain residues coupled to Asp 469. The independent identification of this small network by global perturbation analysis suggests that

these residues form a strongly coupled sub-network within the network centred on Asp 469.

In order to identify networks of evolutionary coupling containing the Asp 469 residue we focused the global SCA on the Pyr domain of the MSA. As the subunit interactions are formed between the PP and the Pyr domains of opposite subunits it was hoped that this would filter out the strong coupling signals from the interface between the subunits. The networks of coupled interface residues appear to be so strong that they can be identified by perturbations of the Pyr domain alone. However, an interesting lower signal cluster was investigated and found to contain Asp 469.

The Asp 469 cluster identified some of the residues that had previously shown coupling to Asp 469, however, an additional five residues were newly identified. Eight of the nine residues in the cluster, are arranged in two distinct connected groups in the Pyr domain of TK. In between these two structures, the ninth residue in the group forms a hairpin loop that maintains the tertiary structure around the active site.

SCA is a powerful tool that has previously been applied to several different families of proteins to discover evolutionarily coupled networks of residues [66]. Here we have used the tool to investigate evolutionary coupling within the TPP-dependent enzymes, and specifically the networks of coupled residues in the *E. coli* TK enzyme. Focusing on a key active site residue that has been mutated to engineer improved TK activity on non-natural substrates, we have identified a further eleven residues that could be targeted either singly or in combination to engineer the properties of

E. coli TK. Nine of these residues have not been targeted for directed mutagenesis before.

4 Statistical coupling guided library design for enzyme engineering of Transketolase

4.1 Introduction

Through the reversible transfer of a two carbon ketol unit from D-xylulose-5-phosphate to either D-ribose-5-phosphate or D-erythrose-4-phosphate, transketolase bridges the non-oxidative pentose-phosphate pathway and glycolysis [1]. The substrate specificity of transketolase is relatively broad and the reaction can be made irreversible by replacing the ketol donor with β -hydroxypyruvate (HPA) [8, 27-29, 44, 67, 83]. These attributes make transketolase a very attractive biocatalyst for the stereoselective formation of carbon-carbon bonds in industrial synthesis. *Escherichia coli* transketolase has become the favoured enzyme for biocatalysis applications as it has a higher specific activity with HPA compared to yeast or spinach derived enzyme [1, 28, 30]. Bacterial transketolase is also easier and cheaper to produce in the high yields required for industrial applications [29].

Although transketolase has broad substrate specificity, activity is typically lower with non-phosphorylated aldehyde acceptors such as glycolaldehyde. Use of non-phosphorylated substrates is desirable as it removes the requirement of a de-phosphorylation step later on in synthesis. Various strategies have been applied to increase transketolase activity with non-phosphorylated substrates such as glycolaldehyde. In one example, saturation mutagenesis was targeted at residues in the active site identifying variants with up to 5-fold improvement in specific activity with non-phosphorylated substrate relative to wild-type [9]. Transketolase activity is

lower still with non-hydroxylated aliphatic aldehydes [84] (typically 5-35% of those for α -hydroxylated substrates), however saturation mutagenesis targeted at the active site residues identified variants with up to 5-fold improvements in specific activity towards propionaldehyde, an aliphatic non-hydroxylated aldehyde [10].

The examples above demonstrate the potential for transketolase to be engineered to accept non-natural substrates, producing variants with beneficial activities relative to the wild type in industrial applications. Although specific activity has been improved up to 5-fold relative to wild-type for two different non-natural substrates, as we move further away from the natural substrates the ability to enhance activity with single changes to residues in the active site is expected to become more limited, with large increases in specific activity requiring multiple simultaneous mutations. The best mutations identified by saturation mutagenesis of *E. coli* transketolase have been recombined with the hope of achieving further improvements in activity on non-natural substrates. Unfortunately such improvements have proven elusive owing to the negative impact of multiple mutations on aspects of protein structure and function (unpublished data). New strategies are therefore required to identify multiple variants with the potential for enhanced activity towards non-natural substrates.

Different methods have been applied to identify residues which interact synergistically upon combined mutation to produce improvements that are greater than the sum of the respective single mutations. Synergistically interacting double mutants have been identified by targeting pairs of residues close in sequence [50, 85]. An alternative approach is to analyse the results of the first rounds of random

mutagenesis and recombine those predicted to be mutually beneficial [86, 87]. However, it is difficult to predict networks of synergistically acting residues not necessarily adjacent in sequence without prior experimental data on a considerable number of variants.

Statistical coupling analysis (SCA) is a powerful bioinformatics method for identifying co-evolved residues in aligned protein sequences. This technique has been applied to the transketolase enzyme and has identified potentially coupled networks of residues in and around the active site. The co-evolution of these residues may represent synergistic interactions for retained catalytic function, such synergy can be driven by various protein properties such as expression, folding, solubility, stability, activity or allostery.

Asp 469 has been identified as having a key role in substrate recognition and enantioselectivity in *E. coli* TK. Variants of Asp 469 were repeatedly identified from a large number of residues around the active site following directed evolution for improved catalytic activity on non-phosphorylated and non- α -hydroxylated substrates. Statistical coupling analysis of the TK enzyme also identified this residue among a cluster of co-evolving, potentially synergistic, residues. Individual or multiple variant libraries focused on these SCA identified residues could lead to further improvements in the activity of TK on non-natural substrates such as glycolaldehyde and propionaldehyde.

Following pre-incubation with HPA as a ketol donor, we have challenged both single and multiple SCA libraries for enhanced activity with the non-natural, non-hydroxylated substrate propionaldehyde. We used Asp 469 mutations known to

improve activity towards PA as a required initial perturbation and explored double and triple mutants using only the most frequently observed natural variant of six evolutionary coupled residues. This mutagenesis strategy successfully identified a triple mutant cluster which behaves synergistically, whereby the combination of all three mutations leads to greater activity than would be expected from the additive effects of each mutation in isolation. Further evaluation of the triple mutant cycle by kinetic analysis identified a double mutant with significantly improved k_{cat} . For the production of (3S) 1,3-dihydroxypentan-2-one (DHP), this is the highest k_{cat} obtained for any TK mutant to date. In industrial biocatalytic transformations the turnover constant k_{cat} is a very valuable property to improve. The identified variant therefore has considerable potential in the production of DHP.

4.2 *Materials and methods*

4.2.1 Materials

All chemicals were obtained from Sigma and used as supplied except β -hydroxypyruvate (HPA) which was prepared as the lithium salt as described in section 4.2.1.5.

4.2.1.1 Luria Bertani medium

Luria Bertani (LB) medium was prepared with 10 g L⁻¹ tryptone, 10 g L⁻¹ NaCl, and 5 g L⁻¹ yeast extract in pure water. pH was adjusted to 7 using 4M sodium hydroxide solution and media was sterilised by autoclave (15 minutes, 2 bar, 124 °C).

4.2.1.2 LB agar plates

LB medium was prepared as described in section 4.2.1.1 above, with the addition of 20 g L⁻¹ select agar. LB agar was sterilised by autoclave (15 minutes, 2 bar, 124 °C). After cooling, ampicillin was added to a final concentration of 150 mg L⁻¹ before the solution was transferred to petri dishes to solidify.

4.2.1.3 NZYM broth

NXYM broth was prepared with 22 g L⁻¹ NZYM broth in pure water. pH was adjusted to 7 using 4 M sodium hydroxide solution and broth was sterilised by autoclave (15 minutes, 2 bar, 124 °C).

4.2.1.4 Ampicillin

Ampicillin was prepared in pure water to a concentration of 150 g L⁻¹. 1ml stocks were sterilised by filtration and stored at -20 °C in 1.5 ml Eppendorf tubes.

4.2.1.5 β-Hydroxypyruvate

The lithium salt of β-hydroxypyruvate was prepared by modification of a previously described procedure [27]. A 0.06 M solution of bromopyruvic acid was prepared. Using a Metrohm autotitrator, 1 M LiOH was added to 100 ml bromopyruvic acid solution at 3 ml.min⁻¹ to bring the pH to 7.0 (approximately 70 ml LiOH was required). LiOH flow rate was then reduced to 1 ml.min⁻¹ and the pH was maintained at 9.0 until a total of 110 ml LiOH had been added. At the end of the reaction, the pH was immediately adjusted to pH 5 using glacial acetic acid and the solution was frozen at -20 °C (unless concentrated on the same day). The mixture was concentrated under low pressure (using a vacuum pump) to approximately 20 ml final volume. Concentrate was stored at 4 °C overnight before the crude product was filtered off and washed with ethanol. Crude product was suspended in 50 ml ethanol and 40 °C for 30 minutes using a rotavap with no vacuum. The white solid was then washed with further ethanol and dried under vacuum to give a white powder. This final product (approximately 2 g) was stored at 4 °C.

4.2.1.6 Stock co-factor solution

A 12x stock solution of co-factors was prepared with 28.8 mM TPP and 108 mM MgCl₂. 12x co-factors were aliquoted into 1.5 ml Eppendorfs and frozen immediately at -20 °C until required.

4.2.2 Standard procedures

4.2.2.1 Streaking agar plates

Cultures were streaked out on LB agar ampicillin plates with a sterilised wire loop. Plates were incubated at 37 °C for 16 hours and then stored at 4 °C.

4.2.2.2 Overnight cultures

Single colonies were picked with a sterilised wire loop from an agar plate into 10 ml LB medium with ampicillin (to a final concentration of 150 mg L⁻¹) in 50 ml Falcon tubes. Tubes were incubated for 16 hours at 37 °C with shaking at 200 rpm. 500 µl glycerol stocks were prepared (4.2.2.4) and the remaining culture was spun down at 4500 rpm for 10 minutes. Supernatant was discarded and the pellets were used immediately or stored at -20 °C.

4.2.2.3 100 ml fermentations

10 ml overnight culture was added to 90 ml LBamp and incubated at 37 °C with shaking at 200 rpm. 1 ml samples were taken each hour to determine biomass by spectrophotometry at OD₆₀₀. Fermentation was grown until stationary phase which was determined by biomass measurements (approximately 8 hours). At stationary phase 1 ml glycerol stocks were prepared (4.2.2.4) and the remaining fermentation was distributed to two 50 ml Falcon tubes and centrifuged at 4000 rpm for 10 minutes. Supernatant was discarded and pellets were frozen at -20 °C.

4.2.2.4 Glycerol stocks

Glycerol stocks of cultures were prepared by adding filter sterilised 40% glycerol to overnight culture in a 1:1 volume ratio to give a final glycerol concentration of 20%. Glycerol stocks were aliquoted into 1.5 ml Eppendorf tubes and stored at -80 °C.

4.2.2.5 Preparation of plasmid DNA

A Qiagen QIAprep Spin Miniprep Kit was used with a microcentrifuge to prepare plasmid DNA from pelleted overnight cultures. Plasmid DNA was eluted in 50 µl 10 mM Tris-Cl, pH 8.5. Concentration and purity were assessed by absorbance at OD₂₆₀ and OD₂₈₀ using a Thermo Scientific NanoDrop Spectropotometer. Final plasmid preparations were stored at -20 °C.

4.2.2.6 Quickchange™ site-directed mutagenesis

Polymerase chain reaction (PCR) mixture was prepared with 5 µl 10x reaction buffer (supplied by Stratagene), 2 µl pQR791 plasmid DNA (diluted to 10 ng/µl), 1.25 µl +ve primer (100 ng/µl), 1.25 µl -ve primer (100 ng/µl), 1 µl dNTP mix, 1 µl DMSO, 39.5 µl ddH₂O (to total 50 µl), and 1 µl Pfu Turbo DNA Polymerase (2.5 U/µl). PCR was run with a 95 °C 30 second initialisation step followed by 16 cycles of PCR. Each cycle of PCR included a 95 °C 30 sec denaturing step, a 55 °C 1 min annealing step and a 68 °C 22 min elongation step (4 min/Kb). Finally, the plasmid mixture was digested by adding 1 µl Dpn1 and incubating at 37 °C for 2 hours.

4.2.2.7 XL10 Gold transformation

25 μ l of XL10 Gold cells (Stratagene Ltd.) were thawed on ice and aliquoted into prechilled falcon tubes. 1 μ l 2-mercaptoethanol was added and the mixture was incubated on ice for 10 minutes with gentle mixing by swirling every 2 minutes. 1 μ l of the *dpn1* digest reaction was added and incubated on ice for 30 minutes. The mixture was heat pulsed at 42 $^{\circ}$ C for 30 seconds then returned to ice for 2 minutes. 225 μ l NZYM broth (4.2.1.3) pre-heated to 42 $^{\circ}$ C was added prior to incubation at 37 $^{\circ}$ C for 1 hour with shaking at 225 rpm. The NZYM broth mixture was then plated out on LB agar plates and incubated overnight at 37 $^{\circ}$ C.

For Multi Site-Directed Mutagenesis 45 μ l cells, 2 μ l 2-mercaptoethanol, 1.5 μ l *Dpn1* treated multi-site DNA, and 0.5 ml NZYM broth were used.

4.2.2.8 Sequencing

LBamp plates were streaked out from glycerol stock and incubated for 16 hours at 37 $^{\circ}$ C. Single colonies were used to inoculate 10 ml overnight cultures. After 16 hour incubation at 37 $^{\circ}$ C, 250 μ l was taken to create 20% glycerol stocks and the remaining culture was spun down at 4000 rpm for 10 minutes. Plasmid DNA was extracted from pellets using the standard Miniprep protocol (4.2.2.5). Plasmid DNA was quantified using a NanoDrop spectrophotometer. DNA was then diluted to 16 fmole/ μ l using ddH₂O. Plasmid DNA (24 μ l) was sent to Wolfson Institute Scientific Services for sequencing together with 12 μ l sequencing primer (TKLibSeqS) per reaction (diluted 1:25).

4.2.2.9 His-tag protein purification

Binding buffer was prepared with 0.5 M NaCl, 20 mM Tris-HCl, and 40 mM Imidazole. Wash buffer was prepared with 0.5 M NaCl, 20 mM Tris-HCl, and 60 mM Imidazole. Elution buffer was prepared with 0.5 M NaCl, 20 mM Tris-HCl, and 0.5 M Imidazole.

Cell pellets from 100 ml fermentations were resuspended in 10 ml binding buffer on ice. Suspended cells were then lysed by 7 cycles of sonication (20 sec on, 20 sec off). The sonicate was centrifuged at 4000 rpm for 10 minutes at 4 °C. The supernatant was recovered and filtered using a Whatman Puradisc 25 AS 1.0 µm Polyethersulfone membrane followed by Minisart Sterile-EO, non-pyrogenic, Hydrophilic 0.20 µm filters. 0.5 ml samples were taken of sonicate, supernatant and filtrate for evaluation by SDS page gel electrophoresis.

Filtrate was loaded onto a Novagen His bind quick 900 cartridge which had been wetted and equilibrated with 6 ml binding buffer. The cartridge was then washed with 20 ml binding buffer followed by 10 ml wash buffer (provided by Novagen). Finally, protein was eluted with 4 ml elution buffer. Purified protein was dialysed overnight against 2 L 0.5 M NaCl 20 mM Tris (pH 7.0).

4.2.2.10 Protein concentration determination

Concentration of purified TK was determined using densitometry. The extinction coefficient of the TK dimer was calculated from the Escherichia coli TK1 sequence (P27302) [88]. The dimer of *E. coli* TK contains 1326 amino acids including 22 tryptophan residues, 46 tyrosine residues, and 10 cysteine residues. The molecular

weight of the TK dimer is 144405.4. The extinction coefficient (ϵ_{280}) calculated as described by Pace *et al* is $190790 \text{ M}^{-1}\text{cm}^{-1}$.

$$\text{Concentration } mg/ml = \frac{Abs_{280} \times \text{Molecular weight}}{\epsilon_{280}}$$

Concentration of TK in total cell lysates was determined by densitometry of TK bands in coomassie stained SDS gels using the ImageJ program available from <http://rsbweb.nih.gov/ij/>. Concentrations were benchmarked against bands of purified TK for which concentration had been determined by spectrophotometry.

4.2.2.11 SDS PAGE gel electrophoresis

An 8% acrylamide:bisacrylamide resolving gel solution was prepared using 13.35 ml ProtoGel (supplied by National Diagnostics), 12.25 ml Resolving buffer, 23.6 ml ddH₂O, 0.5 ml Ammonium persulfate (10% w/v), and 0.05 ml TEMED. The resolving gel was poured into a preassembled electrophoresis gel chamber and overlaid with ethanol. Stacking gel was prepared with 1.3 ml ProtoGel, 2.5 ml Stacking buffer, 6.1 ml ddH₂O, 0.05 ml Ammonium persulfate (10% w/v) and 0.01 ml TEMED. Once the resolving gel had solidified, the ethanol was poured off and the stacking gel solution was poured into the chamber. Finally a Teflon comb was inserted to create the wells.

After polymerisation had completed, the gel was mounted in the SDS chamber. Tris-glycine electrophoresis buffer was used to fill the chamber and flush the wells prior to sample loading. Samples were prepared for SDS by mixing with 2x Laemmli Sample Buffer (Bio-Rad Laboratories) and heating at 100 °C for 2 minutes. 20 μ l of

prepared samples were loaded per well. Gels were run at 80 V for 30 minutes followed by 120 V for an additional 50 minutes. Gels were stained with Coomassie Blue.

4.2.3 Mutant library construction

Single point mutations were introduced to the PQR791 plasmid template using Quickchange™ site directed mutagenesis as described in section 4.2.2.6. Primers for mutagenesis were designed according to the criteria defined by Stratagene and are listed in Table 4.1. Primers were sourced from Operon Biotechnologies. Following transformation of DPN1 digests, colonies were picked for overnight cultures. Pellets from overnight cultures were minipreped and sequenced to verify mutation. Following sequence verification, a 96-well plate (Deep Square Well) containing 900 µl LBamp in each well was inoculated in triplicate with each variant.

Double mutant libraries were generated using pools of variant plasmids as templates and pools of primers to introduce the second mutation. A G467X/D495X library was created by pooling the four G467 variant plasmids (G467X plasmids) and the four D495 variant plasmids (D495X plasmids) separately. Primers were also pooled to give G467X(-), G467X(+), D495X(-), and D495X(+) primer pools. Two Quickchange™ site directed mutagenesis reactions were set up, one with G467X plasmid templates and D495X(-)/D495X(+) primers, and one with D495X plasmid templates and G467X(-)/G467X(+) primers. The *dpn1* digest of both reactions were pooled to create a mixture of plasmids harbouring different combinations of G467X/D495X double mutations. The final pool of plasmid DNA was transformed into XL10 Gold cells and the transformation was plated out on an LBamp agar plate.

The mutagenesis strategy described above was repeated to create three further double mutant libraries, D469X/E498X, D469X/R520X and E498X/R520X. Following incubation at 37 °C overnight, a QPix robot was used to pick 90 colonies from each double mutant library into 96-well plates (Deep Square Well) containing 900 µl LBamp in each well.

A triple mutant library was created using double mutant libraries as templates. Entire library plates of D469X/E498X, D469X/R520X and E498X/R520X double mutants (each containing approximately 200 colonies) were used to inoculate three 10 ml overnight cultures. Pellets from overnight cultures were minipreped to isolate plasmid template pools of the D469X/E498X, D469X/R520X and E498X/R520X libraries. A quickchange reaction was set up using D469X/E498X plasmid template together with R520X(-)/R520X(+) primers. Two further quickchange reactions were set up, one with D469X/R520X template and E498X(-)/E498X(+) primers, and one with E498X/R520X template and D469X(-)/D469X(+) primers. The *dpn1* digestion products from all three reactions were pooled to create a D469X/E498X/R520X triple library. The plasmid library was transformed into XL10 Gold cells and plated out on six LBamp agar plates. Following incubation at 37 °C overnight, a QPix robot was used to pick 540 colonies from the six triple mutant library plates into six 96-well plates (Deep Square Well) containing 900 µl LBamp in each well.

The 96-well plates of single, double and triple libraries were incubated for 16 hours at 37 °C with shaking at 1000 rpm. Following incubation, each 96-well plate was split into six reaction plates (100 µl/well), four glycerol plates (50 µl culture per

well, 50 μ l 40% glycerol per well), and one OD read plate (20 μ l culture, 180 μ l LB per well) using a Tecan high-throughput robot. Reaction and glycerol plates were frozen at -80 °C. Absorbance at 600 nm was measured from the OD read plate using the Tecan spectrophotometer.

Variant	(+) Primer sequence	(-) Primer sequence
G467D	ctccatcggctctggatgaagacgggcccac	gtcggcccgtcttcatccagaccgatggag
G467V	ctccatcggctctggatgaagacgggcccac	gtcggcccgtcttccaccagaccgatggag
G467S	ctccatcggctctgagcgaagacggggcc	ggcccgtcttctcagaccgatggag
G467A	tccatcggctctggcgaagacggggccga	tccggcccgtcttccagaccgatgga
D469A	ggctctggcgaagcggggccgactcacc	ggtagtcggccccgttccggccagacc
D469L	tccatcggctctggcgaactggggccgactcac	gtgagtcggccccagttccggccagaccgatgga
D469T	tccatcggctctggcgaaccggggccgact	agtccggcccgttccggccagaccgatgga
D469S	tccatcggctctggcgaagcggggccgact	agtccggcccgttccggccagaccgatgga
G470T	ggctctggcgaagacaccccactcaccagccg	cggctggtgagtcgggggtcttccggccagacc
G470N	ggctctggcgaagacaaccccactcaccagccg	cggctggtgagtcgggggtcttccggccagacc
G470L	gtctggcgaagacctgccgactcaccagc	gctggtgagtcggcaggtcttccggccagacc
G470I	ggctctggcgaagacattccgactcaccagccg	cggctggtgagtcggaatgcttccggccagacc
T472S	gaagacgggcccagaccagccggttg	caaccggctggtgctcggcccgtcttc
T472D	gcgaagacgggcccgatcaccagccggttg	caaccggctggtgatccggcccgtcttcg
T472A	cgaagacgggcccgcaccagccggttg	caaccggctggtgcgcccgtcttcg
T472L	ggcgaagacgggcccgtcaccagccggttgag	ctcaaccggctggtgcagcggcccgtcttcg
D495E	tggcgtccgtgtgaacaggttgaatccgc	gcggttcaacctgttccacggaccgcca
D495N	acatgtctacatggcgtccgtgaaccaggttgaat	attcaacctggttacacggaccgatgtagacatgt
D495Q	catggcgtccgtgtcagcaggttgaatccgc	gcggttcaacctgtgacacggaccgcatg
D495L	tacatggcgtccgtgtcagcaggttgaatccgcg	ccggttcaacctgcagacacggaccgcatgta
E498I	tggcgtccgtgtgaccaggttatttccggtcgc	gcgaccgcggaaataacctggtcaccggaccgcca
E498D	ccgtgtgaccaggttgattccggtc	gaccgcggaatcaacctggtcaccg
E498A	gtgtgaccaggttgcgtccggtcgcgtg	cacgcgaccgcggacgcaacctggtcacc
E498V	gtgtgaccaggttgcgtccggtcgcgtg	cacgcgaccgcggacacacctggtcacc
R520G	cactgatcctctcggccagaacctggcgca	tgcaccaggttctggccggagaggatcagtg
R520K	cgactgatcctctccaacagaacctggcgagc	gctcgcgaggttctgtttggagaggatcagtg
R520Q	cactgatcctctccagcagaacctggcgag	ctgcgaggttctgctgggagaggatcagtg
R520A	gcactgatcctctcggcgaacctggcgag	ctgcgaggttctgctgggagaggatcagtg

Table 4.1 Primers used to create site directed mutations in *E. coli* TK.

4.2.4 Colorimetric screening of libraries for activity

Biotransformations were prepared using reaction plates containing 100 μ l of cell culture per well, lysed by a double freeze-thaw from -80 °C. Cell lysate was pre-incubated with co-factors (2.4 mM TPP, 9 mM MgCl₂ final concentration) for 20 minutes. After pre-incubation, Lithium Hydroxypyruvate (50 mM final concentration) followed by Propionaldehyde (50 mM final concentration) were

added to a total volume of 300 μ l. All reagents were prepared in 50 mM Tris buffer at pH 7.0. Propionaldehyde was prepared fresh on the day of the biotransformation. The biotransformations were incubated for 48 hours at 21 °C.

Colorimetric screening was carried out on biotransformation reaction plates as described previously [89]. Fresh 96-well plates (Shallow Round Well) were prepared with 50 μ l Tris buffer (50 mM, pH 7.0) and 20 mg MP-Carbonate Scavenger Resin (supplied by Biotage) in each well. Individual biotransformation reactions (50 μ l) were then added to each well and incubated for 3 hours at 21 °C to quench any excess hydroxypyruvate. Following quenching, the reaction-resin mixture was diluted with 100 μ l Tris buffer (50 mM, pH 7.0) and 100 μ l of the reaction mixture was transferred to a new plate containing 20 μ l 2,3,5-triphenyltetrazolium chloride solution (0.2% in methanol), leaving the resin beads in the first plate. A plate reader (Fluostar, BMG-labtech) fitted with an autoinjector was used to add 10 μ l NaOH (3 M) to each well of the new plate. Following addition of NaOH, the plate was shaken for 10 seconds and left to stand for 1 minute before an OD measurement was taken for each of the wells at 485 nm (50 flashes per well).

4.2.4.1 Validation of colorimetric screen

The colorimetric screen was validated for its ability to identify positive variants relative to wild type level activity. Variants D469A, D469E, and D469T, known to demonstrate increased activity relative to wild type, were screened against wild type TK. In total, 93 positive controls and 93 wild type controls were screened (Figure 4.1). Results from the assay indicated twelve potential false positives in the

wild type group and five potential false negatives in the positive group. The colorimetric assay therefore displays good predictability of activity.

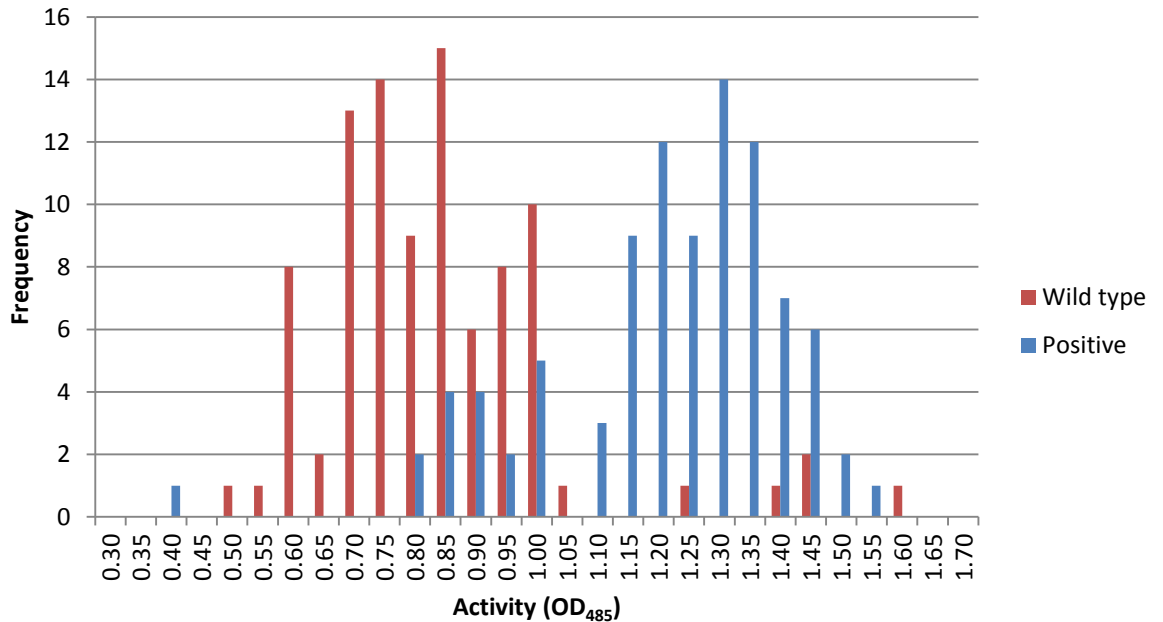


Figure 4.1 Validation of the colorimetric assay to distinguish between wild-type transketolase and improved variants on the PA reaction.

4.2.5 Solubilisation and refolding of insoluble variants

4.2.5.1 Solubilisation of transketolase variants

Pellets from 100 ml fermentations were resuspended in 40 ml binding buffer, sonicated (20 sec on, 20 sec off, 20 cycles), and spun down for 15 min at 5000 rpm. This wash step was repeated once more with 20 ml binding buffer. The final pellet was resuspended in 5 ml binding buffer with 3 M Guanidine HCl (concentration required to remove TK from insoluble fractions), sonicated and incubated overnight at 4 °C to solubilise the TK enzyme. The solubilised TK was spun down at 5000 rpm for 30 minutes and the supernatant was purified on an NiNTA column as described previously (4.2.2.9) but with 3 M Guanidine HCl in each buffer. Imidazole

concentration in the wash buffer was reduced to 20 mM to account for the Guanidine HCl.

4.2.5.2 Refolding by dialysis

Eluted TK was dialysed against decreasing concentrations of Guanidine HCl in 0.5 M NaCl, 20 mM Tris over a course of 20 hours. Dialysed samples in 0.5 M NaCl, 20 mM Tris were spun down at 5000 rpm for 30 minutes at 4 °C and analysed for protein concentration at OD_{280 nm}.

4.2.5.3 Refolding by drip feeding

5 ml of eluted TK was drip fed into 45 ml 50 mM Tris HCl, 5 mM DTT over the course of 8 hours. Dilute solution of refolded protein was concentrated to 1 ml sequentially using 10,000 micron followed by 3,000 micron spin columns. Concentrated protein solution was dialysed against 20 mM Tris, 0.5 M NaCl.

4.2.6 Activity of purified enzymes

Wild-type and mutant transketolases were over-expressed and purified using His-tag affinity chromatography, and enzyme kinetics were determined using HPLC. 300 µl reactions were prepared in sealed glass vials using 150 µl purified protein. Purified protein was pre-incubated with co-factors (2.4 mM TPP, 9 mM MgCl₂ final concentration) for 20 minutes prior to the addition of Lithium Hydroxypyruvate (50 mM final concentration). Propionaldehyde was then added to varying final concentrations. Samples (20 µl) of reaction mixture were taken at twelve separate time points (at least one hour apart) and quenched with 180 µl 0.1% Trifluoroacetic acid (TFA). Quenching was carried out in a microplate which was stored at -20 °C in-

between time points. For each variant, reactions were prepared with seven different final concentrations of propionaldehyde, from 8 mM to 60 mM.

The reaction product was measured by HPLC using a 15 cm C18 column and a 15 min isocratic protocol of 0.1% TFA in 5% acetonitrile with a flow rate of 0.6 ml min⁻¹. Product was detected by UV at 200 nm. Standard curves were produced with 10-50 mM 1,3-dihydroxypentan-2-one and these were used to determine the product concentrations. New standard curves were produced for each microplate analysed.

4.2.7 Enantioselectivity of purified enzymes

The stereoselectivity of purified TK variants was established by gas chromatography as described previously [11]. Reactions were carried out to completion using purified TK with 50 mM LiHPA, 50 mM PA, 50 mM Tris-HCl, 2.4 mM TPP, 9 mM MgCl₂, 250 mM NaCl, pH 7.0 at 300 µl scale in sealed glass vials. Reaction mixture (100 µl) was transferred to vials containing 300 µl EtOAc. Vials were shaken and allowed to partition. The organic phase (100 µl) was transferred to fresh vials and pyridine (containing DMAP (10 mg/ml), 20 µl) was added to each vial. Following conversion of DHP to the diacetate, enantiomeric purity was assessed by gas chromatography on a Perkin-Elmer Autosystem XL Gas chromatograph with a β-Dex 225 chiral column (Supelco, 30 m x 0.25 mm). GC conditions: injection volume, 1 µl; carrier gas, He; carrier gas pressure, 15 psi; injector temperature, 250 °C; oven temperature, 60 °C then increased at 3 °C/min; detector temperature, 300 °C; detection, flame ionised detector (FID). Retention times: (3R)-pentan-2-one diacetate, 29.9 min; (3S)-pentan-2-one diacetate, 30.3 min.

4.3 Results and Discussion

4.3.1 Library design strategy

We described the use of SCA to identify co-evolving networks of residues in TPP-dependent enzymes. The identification of such networks enables the selection of residues for mutagenesis that are functionally linked to the active site and the rational design of multiple-mutant libraries with synergistic potential. We have identified a nine residue network within the Pyr domain of TPP-dependent enzymes that includes the functional Asp 469 residue in *E. coli* TK (Figure 4.2). This network was comprised of two structurally contiguous groups of residues with a proline residue forming a hairpin turn at a midway point between the two groups. Here we describe the creation of variant libraries by site directed mutagenesis focused on this nine residue network.

The nine positions identified in the Pyr cluster included Gly 467, Asp 469, Gly 470, Thr 472, Pro 486, Pro 493, Asp 495, Glu 498, and Arg 520. Of these nine residues, we selected seven to target for mutagenesis. The two proline residues, Pro 486 and Pro 493, were excluded as mutagenesis of these residues is likely to disrupt the tertiary structure of the TK enzyme limiting the potential to identify active variants. A second strategy adopted to avoid the introduction of destabilising mutants was to limit mutations to the most frequently observed natural variants in the TPP-dependent enzyme MSA (Table 4.2). The exceptions to this strategy included Asp 469 for which mutations were selected that were known to improve activity towards PA, and Gly 467 for which one of the common residues is a proline which was avoided for the stability issues discussed above.

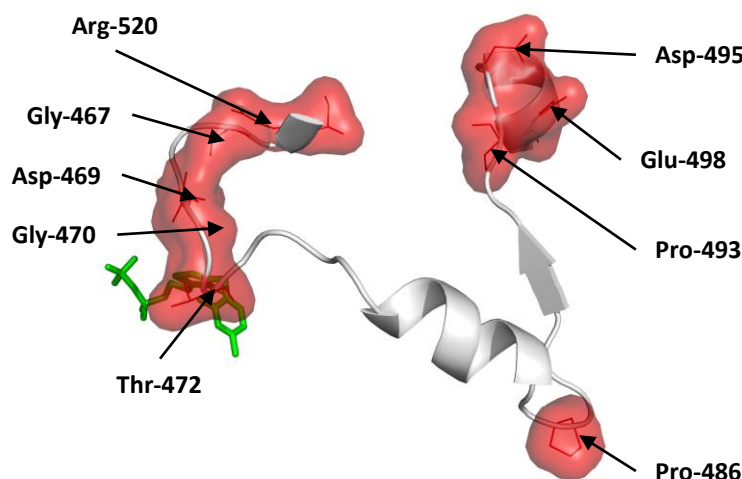


Figure 4.2 Cluster of co-evolving residues in *E. coli* TK identified by SCA of the PYR domain of TPP dependent enzymes. Two structurally contiguous groups of residues are linked by a proline residue (Pro 486) forming a hairpin turn in the tertiary structure. Arg 520, Gly 467, Asp 469, Gly 470, and Thr 472 make up the contiguous group proximal to the active site.

Residue	Position							Average in all proteins
	467	469	470	472	495	498	520	
A	2.1	3.7	2.1	10.7	3.7	8.9	6.3	7.3
C	0	0	0	0	0	0.5	2.6	2.5
D	10.7	45.3	2.9	9.7	44.5	11.3	1.0	5
E	1.8	<u>3.9</u>	0.3	0.5	12.8	42.1	1.8	6.1
F	0.3	1.6	0	1.8	2.4	1.0	2.6	4.2
G	43.5	3.7	47.9	0.5	0.5	1.0	6.0	7.2
H	1.3	1.3	0.3	0	0.3	0	0.3	2.3
I	1.0	2.4	9.4	0	1.0	6.0	3.9	5.3
K	0.5	2.1	2.6	0	0.3	1.0	19.6	6.4
L	0	7.1	13.4	9.7	8.4	4.5	0.3	8.9
M	0.3	1.8	2.6	0.3	0.8	1.8	1.0	2.3
N	0.3	<u>8.6</u>	4.2	0	6.3	0.5	2.1	4.3
P	<u>3.4</u>	1.3	0	1.8	0.8	0	0.3	5.2
Q	0	2.1	1.6	3.9	6.5	2.6	6.8	4
R	0	<u>6.8</u>	3.7	0.5	0.8	1.3	39.3	5.2
S	2.9	0.5	2.9	5.8	2.4	1.6	0.5	7.3
T	0	1.0	3.9	47.4	4.5	3.1	1.6	5.6
V	2.4	1.3	1.6	2.6	0.3	11.3	2.9	6.3
W	0	1.0	0	0	0	0.5	0.3	1.3
Y	0.3	0.5	0.3	4.2	0.3	0.5	0.5	3.3

Table 4.2 Frequency (%) of residues at selected positions within the TPP-dependent enzyme multiple sequence alignment. Chosen variants (bold), including the wild type residues (italic) were the most frequently occurring natural variants from an alignment of 382 homologous TPP-dependent enzymes. Exceptions (underlined) are G467P and D469E/N/R.

4.3.2 Construction and screening of SCA directed single mutant library

The twenty-eight single point mutations were introduced into *E. coli* TK by site directed mutagenesis and verified by sequencing. 96-well microplates were prepared as described in section 4.2.3 containing the single-point mutants of TK in triplicate. Measurement of cell density across microplates indicated successful growth with no positional effects on growth across the plate. Eight wells displayed unusually low cell density (Figure 4.3) but comparison with equivalent variants displaying high cell density demonstrated no correlation between activity of TK and cell density. Cell density does not therefore appear to correlate closely with the active enzyme concentration and cannot be used as a surrogate measure for protein concentration for the determination of specific activities.

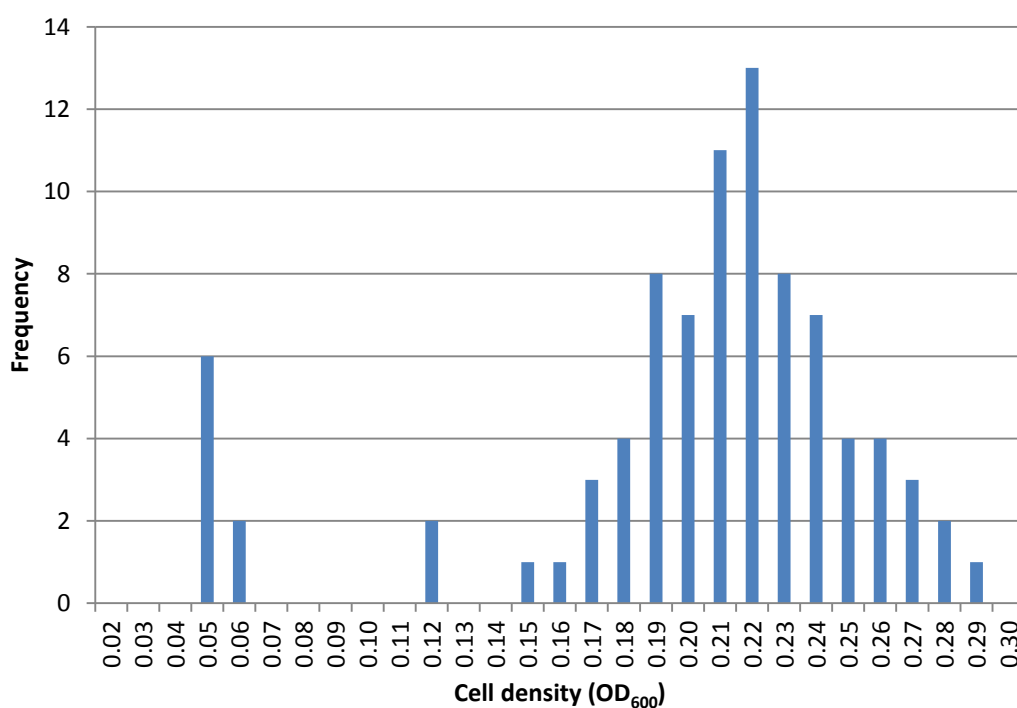


Figure 4.3 Distribution of cell growth across the microplate as measured by cell density (OD₆₀₀).

Activity of variants was determined for the TK catalysed conversion of propionaldehyde and HPA into DHP. The absolute activity of each variant was measured in triplicate using the colorimetric screen developed by Smith *et al* [89], relative activities were calculated using internal wild type standards (hextuplet). Good consistency was observed across the replicates giving confidence in the colorimetric screen to determine relative activities (Figure 4.4).

Variants with improved activities relative to wild type TK were identified for five of the seven residues in the SCA directed single mutant library, these sites included Gly 467, Asp 469, Gly 470, Tyr 472 and Arg 520 (Figure 4.4). Notably, these residues make up the structurally contiguous group of residues proximal to the active site (Figure 4.2). In contrast the two residues belonging to the more distant structural group failed to produce any variants with activity increases, all mutations at these positions resulted in seriously compromised TK enzyme with approximately 30% wild type activity (Figure 4.4).

As expected, all the Asp 469 variants displayed increased activity relative to the wild-type transketolase standards contained in the same plate (Figure 4.4). Asp 469 interacts with the α -hydroxyl group of natural aldehyde acceptors providing a possible explanation for the improved activity of variants at this position with non- α -hydroxylated substrates. Mutation of Arg 520 also produced a high proportion of variants with increased activity on PA relative to wild type TK. The Arg 520 variant R520V has previously been identified as improving TK activity on GA [9] and PA [10]. Arg 520 interacts with the phosphate group of natural TK substrates, it has been hypothesised that the reduction in steric bulk associated with variation of this

residue could explain the enhanced activity seen on non-phosphorylated substrates. Although R520G has been previously identified by directed evolution for enhanced activity on GA, R520K, R520Q and R520A have not.

The other three residues for which variants displayed improved activity- Gly 467, Gly 470, and Tyr 472- have not been previously targeted for saturation mutagenesis. The identification of variants at these positions with enhanced activity on PA demonstrates the potential of SCA to identify new positions which can be targeted for site directed mutagenesis. In this study only four variants were introduced at each site, the high frequency of improved variants despite the small sample size suggests that further improvements may be identified by full saturation mutagenesis of these SCA identified sites.

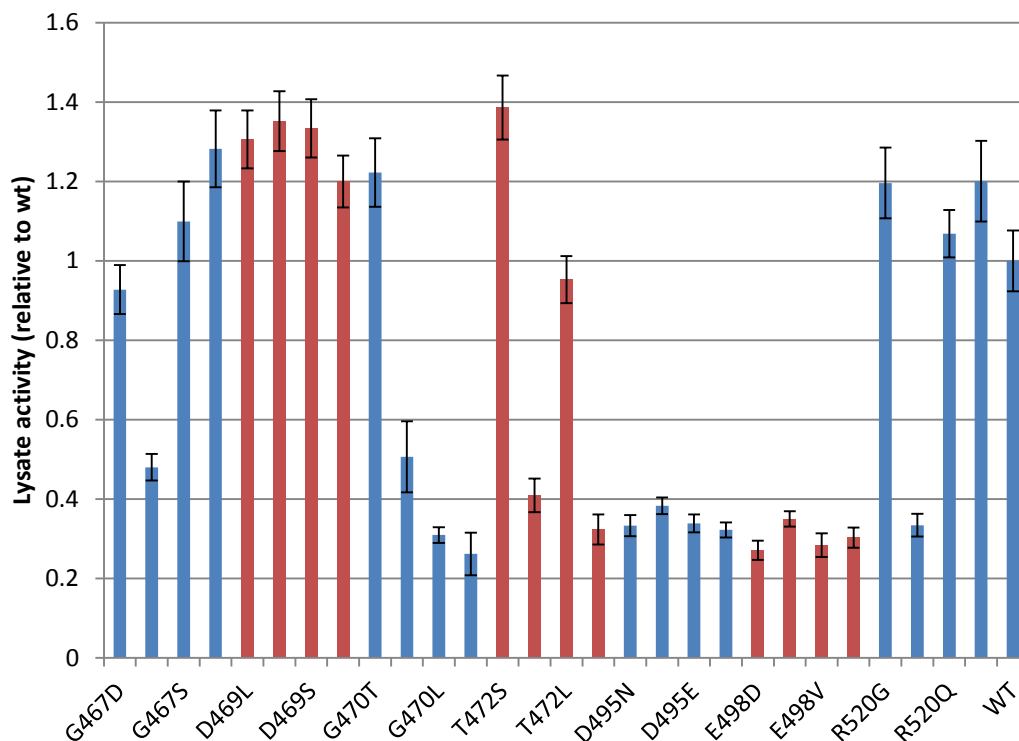


Figure 4.4 Relative lysate activities of single mutants relative to wild type on the reaction between HPA and propionaldehyde. Total activity determined using the high-throughput colorimetric assay with 50 mM LiHPA, 50 mM PA, and 50 mM Tris-HCl, 2.4 mM TPP, 9 mM MgCl₂, 250 mM NaCl, pH 7.0 to produce 1,3-dihydroxypentan-2-one (DHP).

4.3.3 Construction of SCA directed double and triple mutant libraries

Analysis of the single variant SCA directed library identified variants with both improved and impaired activity on PA relative to wild type TK. If the residues of this co-evolving cluster are synergistically coupled we could expect to see non-additive effects when the different mutations are combined in multiple mutants. Even with such a focussed and restricted library of twenty eight variants, the theoretical total number of mutant combinations at these seven sites would be 78,124. Libraries of this size are not out of the range of screening by high-throughput techniques but this is beyond the scope of the current study.

In order to test the hypothesis of synergy between the seven residues identified by SCA we have selected two groups of residues from which to produce smaller, more manageable multiple mutant libraries. Each of these groups includes residues from both the proximal contiguous cluster and the distal group of residues, for which no single variants with improved were identified.

The first group selected for combined mutagenesis includes residues Asp 469, Glu 498 and Arg 520. Variation at two of these residues has demonstrated improvements in activity on both GA and PA. In contrast, all four variants of Glu 498 resulted in severely impaired catalytic activity on PA. If synergistic relationships exist between these residues we could hope to see non-additive effects when the mutations are combined.

The second group selected for combined mutagenesis includes residues Gly 467 and Asp 495. Again this group includes positions from both contiguous structural clusters, one of which has produced improved variants (Gly 467), and one from which no improvements have been identified (Asp 495).

Following creation of the 540 variant triple mutant library (D469X/E498X/R520X) and the 90 variant double library (G467X/D495X), sample sequencing of ten wells from each was carried out to verify coverage of sequence space (Table 4.3). Full representation was established for the G467X/D495X library with all variants represented at each position. Although we did not attain full representation of variants across all the three sites of the D469X/E498X/R520X library, site D469 and E498 displayed three of the four possible variants, and at site R520 all four possible variants were observed. The diversity of the libraries therefore appeared very good, giving confidence that full coverage of the sequence space was attained. Notably, no wild type residues were identified in either library. In order to ensure that double as well as triple variants were captured for the D469X/E498X/R520X library, three further double mutant libraries were created for D469X/E498X, D469X/R520X, and E498X/R520X, each with 90 variants.

Sequencing sample	G467X/D495X Library	D469X/E498X/R520X Library
1	G467V/D495N	D469S/E498V/R520A
2	G467D/D495L	D469A/E498V/R520G
3	G467D/D495Q	D469A/E498I/R520K
4	G467D/D495N	D469T/E498D/R520A
5	G467A/D495E	D469T/E498D/R520A
6	G467D/D495E	Bad signal
7	G467S/D495N	D469S/E498V/R520Q
8	G467D/D495N	D469A/E498V/R520G
9	G467D/D495Stop	D469T/E498I/R520Q
10	Bad signal	D469T/E498V/R520Q

Table 4.3 Sample sequencing results of G467X/D495X and D469X/E498X/R520X libraries.

4.3.4 Double and triple mutant library colorimetric screen of activity

Libraries were screened in duplicate using the colorimetric screen developed by Smith *et al* [89]. For each plate, relative activity on the propionaldehyde HPA reaction was determined using internal wild type standards. The well-defined nature of the libraries produced (as determined by sample sequencing) presents an opportunity for analysis of variant libraries as a whole, comparing the average relative activities and the ranges of relative activities across libraries. The D469X/R520X library produced the highest mean relative activity. Average activities across the other four libraries were lower than wild type and the G467X/D495X library demonstrated the lowest average relative activity (Table 4.4).

Library	Experimental		Expected	
	Mean	Range	Mean	Range
G467X/D495X	0.73	0.42 – 1.04	0.33	0.15 – 0.49
D469X/E498X	0.68	0.36 – 0.95	0.39	0.33 – 0.47
D469X/R520X	1.07	0.64 – 1.49	1.23	0.40 – 1.62
E498X/R520X	0.92	0.65 – 1.16	0.29	0.09 – 0.42
D469X/E498X/R520X	0.90	0.25 – 1.91	0.37	0.11 – 0.57

Table 4.4 Mean relative activities and ranges of activities across libraries compared with that expected in unbiased libraries based on single variant relative activities. Total activity determined with 50 mM LiHPA, 50 mM PA, and 50 mM Tris-HCl, 2.4 mM TPP, 9 mM MgCl₂, 250 mM NaCl, pH 7.0 to produce 1,3-dihydroxypentan-2-one (DHP).

If we assume no bias in the libraries, we can calculate the expected mean activities based on the activities of single mutants. Comparing such expected mean relative activities with the experimental mean relative activities indicates generally higher than expected activity across the libraries. The exception to this is the D469X/R520X library which produced a lower experimental mean relative activity than that expected (Table 4.4).

Comparison of the ranges of expected and experimental activities alleviates the requirement for a lack of bias, assuming full coverage of variants has been attained. The experimental range of activities across the G467X/D495X library was 2.1- to 2.8-fold greater than expected. Relative activities at the low end of the range were as expected for the D469X/E498X library but activities at the high end were 2.0-fold greater than expected. The range of activities for the D469X/R520X library was as expected although the experimental range was slightly tighter than the expected range. The experimental ranges for both the E498X/R520X and D469X/E498X/R520X were markedly higher than expected. For the E498X/R520X library this was most apparent at the low end with 7.2-fold greater activity than expected, and for the D469X/E498X/R520X library the biggest increase was at the high end with a 3.4-fold increase over that expected.

These data suggest potential synergy between the mutations in the multiple mutant libraries. General synergy is apparent between Gly 467 and Asp 495 variants with higher than expected activities. Variants of residue Glu 498 display potential synergy with both the Asp 469 (D469X/E498X library) and the Arg 520 variants (E498X/R520X library). However, no double variants have been identified in either

library with superior activity to the single variants of Asp 469 or Arg 520. This suggests that the synergistic relationships in both combinations are acting to alleviate the negative impact of variants at Glu 498. In the triple library, D469X/E498X/R520X, variants have been identified that display a synergistic increase in activity over and above the activity seen with any single variant. Although no synergy is apparent between Asp 469 and Arg 520 variants in the D469X/R520X library, these residues may have a more complex synergistic relationship incorporating the third variant, Glu 498.

The most active variant following the primary colorimetric screen was found in the triple mutant library D469X/E498X/R520X which displayed a relative activity of 1.91. Sequencing identified this variant as D469S/E498D/R520Q. Such a triple variant is of particular interest because the E498D mutation results in an inactive enzyme in isolation. In order to explore the synergistic relationships between these residues further, a mutant cycle of this triple variant was created and purified for further detailed analysis.

4.3.5 Total lysate activities of the triple mutant cycle D469S/E498D/R520Q

In order to produce a full triple mutant cycle, constructs were created by site directed mutagenesis for the double variants D469S/E498D, D469S/R520Q, and E498D/R520Q. Single variants had been produced previously by site directed mutagenesis. Following production and verification of the triple cycle, total lysate relative activities were determined for the triple mutant cycle using the colorimetric assay (Figure 4.5).

The D469S mutation and the R520Q mutation independently alleviate the negative impact of E498D, producing higher relative activities than expected with the D469S/E498D and the E498D/R520Q combinations. When all three mutations are combined, a triple variant is produced with considerably higher relative activity than could be expected taking each of the variants in isolation. These results suggest significant synergy between the three variants. However, in the absence of the E498D mutation, no synergy is apparent between D469S and R520Q based on this analysis.

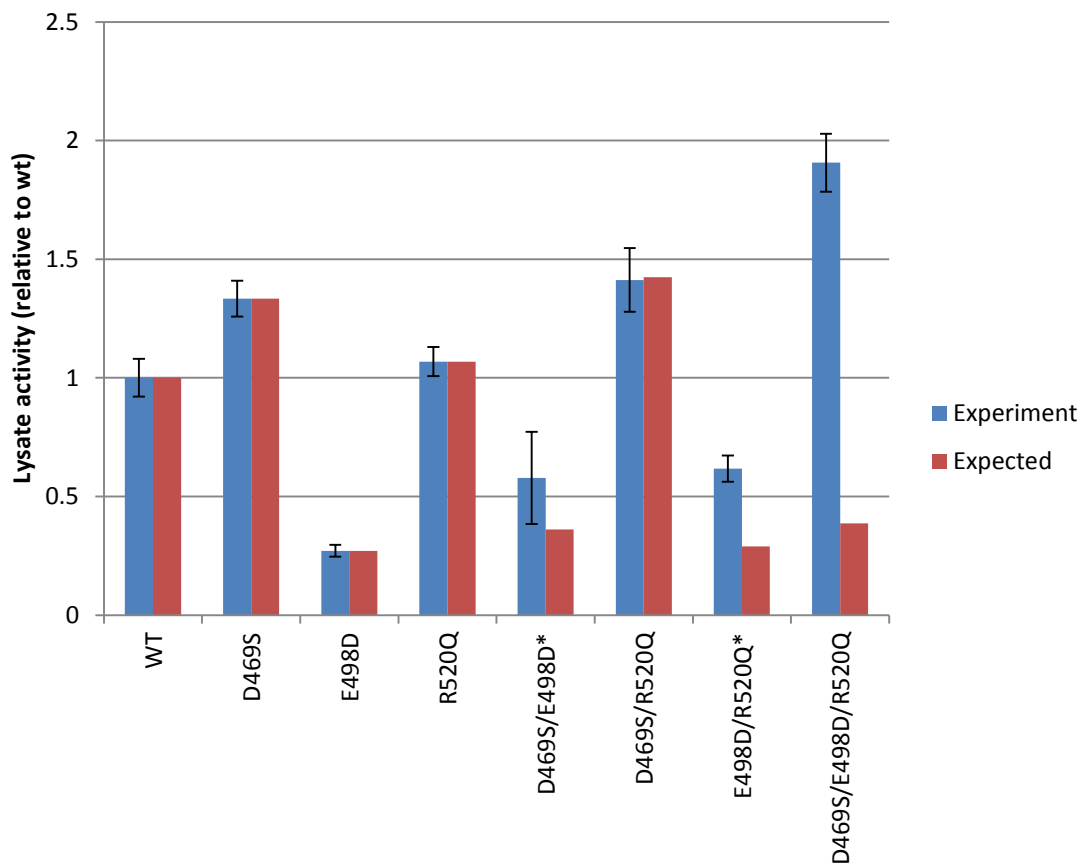


Figure 4.5 Experimental and expected total lysate activities of Transketolase variants in the triple mutant cycle of D469S/E498D/R520Q. Total activity determined with 50 mM LiHPA, 50 mM PA, and 50 mM Tris-HCl, 2.4 mM TPP, 9 mM MgCl₂, 250 mM NaCl, pH 7.0 to produce 1,3-dihydroxypentan-2-one (DHP).

4.3.6 Purification of the triple mutant cycle D469S/E498D/R520Q

Lab scale fermentations of the selected triple mutant cycle variants were carried out and variants were purified on Ni-NTA columns as described in section 4.2.2.9. Expression and solubility profiles of the different variants provide information on the potential origin of synergy between these three positions.

Expression of the single variant D469S was marginally decreased relative to wild type TK, in contrast the expression of the R520Q variant was increased by 40%. However, introduction of the E498D variant reduced expression to less than 40% that of wild type TK (Figure 4.6). All of the codons introduced by site directed mutagenesis were the optimal for expression in *E. coli* TK so the decrease in expression with E498D is likely to be due to a loss of protein stability or solubility.

The combination of the two variants in the double mutant E498D/R520Q recovers the total protein expression of TK but the protein remains insoluble as determined from the concentration of TK in the soluble fraction. The double mutant D469S/E498D fails to improve total protein expression but does marginally improve the solubility relative to the E498D mutation in isolation. The triple mutant D469S/E498D/R520Q increases the total protein expression relative to E498D but no increase in the soluble fraction of TK is observed (Figure 4.6).

Based on the results above the enhanced activity of the triple mutant cannot be explained by an increase in expression or stability relative to wild type TK. Instead, the improvement appears to be due to enhanced specific activity or better folding of the soluble fraction to the active native state.

For all variants, a similar percentage of soluble protein was His-tag purified using His affinity resin. In all variants containing the E498D variant, the yield was too low to carry out further analysis on purified enzyme.

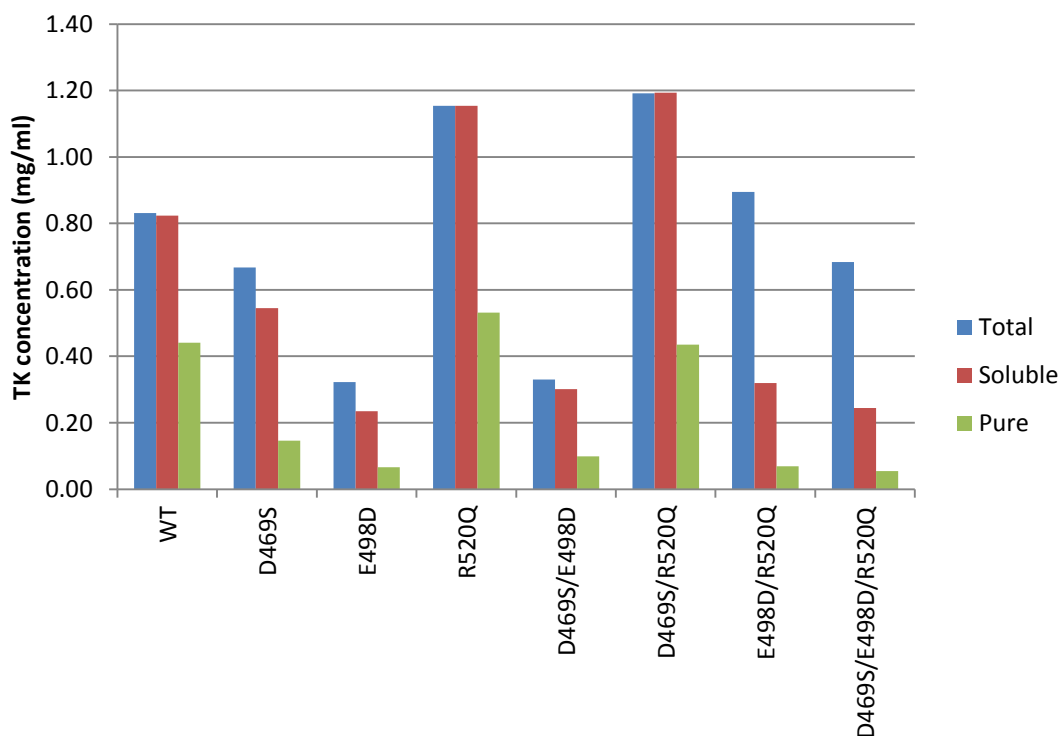


Figure 4.6 Total, soluble and pure concentrations of TK variants.

4.3.7 Solubilisation and refolding of E498D containing variants

In order to obtain kinetic data on the full triple mutant cycle of D469S/E498D/R520Q, efforts were made to obtain purified variants in suitable quantities for the three members of the cycle containing the E498D variant. New 100 ml fermentations were produced for each variant and prior to purification, TK was solubilised with Guanidine HCl as described in section 4.2.5.1. Following purification, refolding was attempted by both dialysis and drip-feeding. Unfortunately neither technique resulted in active protein as determined by HPLC

kinetic analysis. Further analysis of purified variants has therefore been limited to those variants that did not contain the E498D mutation.

4.3.8 Enzyme kinetics of purified variants

The enzyme kinetics of His-tag purified wild type TK and variants D469S, R520Q, and D469S/R520Q were established by HPLC with 50 mM HPA over a range of propionaldehyde concentrations (8 mM-60 mM) (Table 4.6 and Figure 4.7). At 50 mM, the concentration of HPA used here is approximately 10 times the K_m of wild type TK.

The single mutation D469S produced a variant of TK with a 60% increase in k_{cat} (57 min^{-1} vs 35 min^{-1}) and a 40% decrease in K_m (71 mM vs 181 mM) relative to wild type. The combined effect of these changes was a 4-fold increase in the corresponding k_{cat}/K_m (0.81 $\text{min}^{-1} \text{mM}^{-1}$ vs 0.19 $\text{min}^{-1} \text{mM}^{-1}$) relative to wild type. The R520Q variant resulted in a similar k_{cat} to wild type TK (31 min^{-1} vs 35 min^{-1}). However, the K_m for this variant almost doubled relative to wild type (329 mM vs 181 mM) resulting in a k_{cat}/K_m approximately half that of the wild type enzyme (0.10 $\text{min}^{-1} \text{mM}^{-1}$ vs 0.19 $\text{min}^{-1} \text{mM}^{-1}$).

The most dramatic changes in kinetic parameters were observed with the double mutant D469S/R520Q. This variant displayed a 20-fold increase in k_{cat} (700 min^{-1} vs 35 min^{-1}) which more than compensated for a 3.5-fold increase in K_m (628 mM vs 181 mM). The resulting k_{cat}/K_m was 6-fold higher than that of wild type (1.11 $\text{min}^{-1} \text{mM}^{-1}$ vs 0.19 $\text{min}^{-1} \text{mM}^{-1}$), representing the highest turnover constant determined for the production of DHP with any TK variant to date (Figure 4.7).

Although the high K_m of D469S/R520Q indicates poor substrate affinity for PA, the concentration of PA used in screens (50 mM) is at the lower end of the spectrum of what is used in cost-effective industrial biocatalytic transformations. Such low substrate affinity would be acceptable in industrial biocatalysis as long as substrate concentration could be increased sufficiently to achieve reasonable saturation of the active site. It is also important that enantioselectivity is not negatively impacted by a reduction in the number, or strength, of enzyme substrate interactions.

Previous kinetic analysis of purified wild type TK with propionaldehyde determined the K_m to be 140 ± 50 mM [10]. The K_m determined here is higher than that previously determined but is within the calculated error. The source of this discrepancy could also be the reaction conditions. In the current study, reactions were carried out in a salt concentration of 250 mM NaCl which was included to aid in the purification of the enzyme. Hibbert *et al* carried out their kinetic analysis in the absence of NaCl. The salt concentration used here was the same for each variant so we do not expect this discrepancy to adversely impact on our analysis of relative kinetic parameters.

Enzyme	Initial Velocities (mM min ⁻¹)						
	[PA] 8mM	[PA] 16mM	[PA] 24mM	[PA] 32mM	[PA] 40mM	[PA] 48mM	[PA] 60mM
WT	0.005	0.008	0.014	0.018	0.017	0.021	0.034
D469S	0.006	0.011	0.015	0.020	0.020	0.022	0.028
R520Q	0.003	0.006	0.007	0.012	0.011	0.016	^a
D469S/R520Q	0.027	0.051	0.077	0.120	0.099	0.153	0.236

Table 4.5 Initial velocities of a triple mutant cycle of TK mutants at varying propionaldehyde (PA) concentrations. Initial velocities were determined from the concentration of DHP using HPLC and purified protein. Activity determined with 50 mM LiHPA, and 50 mM Tris-HCl, 2.4 mM TPP, 9 mM MgCl₂, 250 mM NaCl, pH 7.0 to produce 1,3-dihydroxypentan-2-one (DHP). ^aNot available.

Enzyme	Lysate activity relative to wt	[Protein] ^b (mg/ml)	k_{cat} (min ⁻¹)	K_m (mM)	k_{cat}/K_m (min ⁻¹ mM ⁻¹)	e.e. ^c
Wt ^a	1.00 ± 0.08	0.44	35	181	0.19	57.1
D469S	1.33 ± 0.08	0.15	57	71	0.81	64.9
E498D	0.27 ± 0.02	0.07	n.a. ^d	n.a.	n.a.	n.a.
R520Q	1.07 ± 0.06	0.53	31	330	0.10	57.8
D469S/E498D	0.58 ± 0.19	0.10	n.a.	n.a.	n.a.	n.a.
D469S/R520Q	1.41 ± 0.13	0.44	700	628	1.11	67.4
E498D/R520Q	0.62 ± 0.06	0.07	n.a.	n.a.	n.a.	n.a.
D469S/E498D/R520Q	1.91 ± 0.12	0.05	n.a.	n.a.	n.a.	n.a.

Table 4.6 Kinetic parameters and enantioselectivities of a triple mutant cycle of TK mutants for the propionaldehyde (PA) substrate. Lysate activities were determined using a colorimetric assay; kinetic parameters were determined using HPLC and purified protein. Total activity determined with 50 mM LiHPA, 50 mM PA, and 50 mM Tris-HCl, 2.4 mM TPP, 9 mM MgCl₂, 250 mM NaCl, pH 7.0 to produce 1,3-dihydroxypentan-2-one (DHP). ^aWild-type specific activity is 0.029 ± 0.001 μmol/mg/min [10]. ^bObtained after purification. ^cEnantiomeric excess of the S-isomer. ^dNot available.

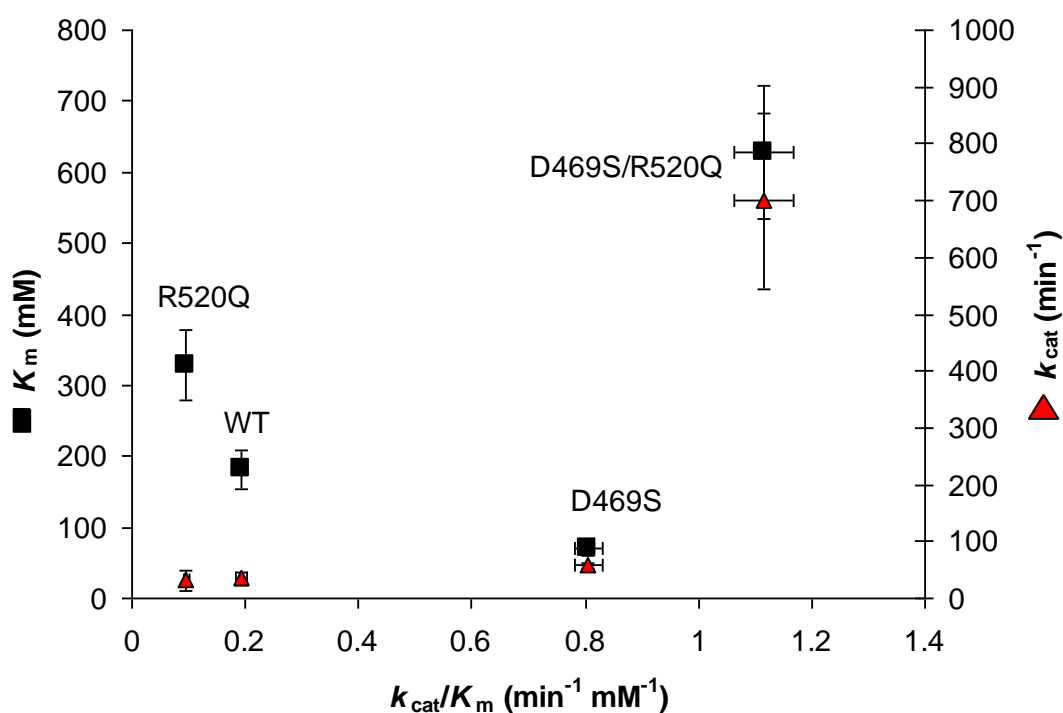


Figure 4.7 Comparison of k_{cat} , K_m and k_{cat}/K_m for the double mutant cycle of D469S and R520Q.

4.3.9 Enantioselectivity of variants compared to wild type

In addition to potential synergistic effects on turnover rate and substrate affinity, the synergy between co-evolving residues may also be associated with effects on enantioselectivity. Mutants of TK have been identified in previous work that have both improved and reversed enantioselectivity [11], evaluation of such changes in

the SCA directed variants may indicate further synergistic associations between the residues. It is also important to ascertain whether the reduced substrate affinity of the double mutant D469S/R520Q has negatively impacted on the enantioselectivity of the enzyme.

Gas chromatography was used to determine the stereospecificity of wild type TK and the three purified TK variants (Table 4.6). Wild type TK catalysed the production of S-DHP in 57% enantiomeric excess, this correlates closely with previous results for wild type TK [11]. The D469S variant demonstrated improved enantioselectivity relative to wild type, producing S-DHP in 65% enantiomeric excess, this is similar to the increase observed with another variant, D469T [11]. In contrast, the R520Q variant appeared to have no effect on enantioselectivity (S-DHP 58% ee). When the two variations were combined in the double mutant D469S/R520Q, enantioselectivity was increased to the same extent observed with D469S in isolation (S-DHP 67% ee).

These results suggest that the co-evolutionary pressures on Asp 469 and Arg 520 are more likely to be associated with turnover rate and substrate affinity rather than enantioselectivity. However, these particular variants were selected based on increased activity on the PA substrate. Full evaluation of the enantioselective pressures on co-evolution would require the enantiomeric evaluation of many more variants. Importantly for the use of the D469S/R520Q variant in industrial biocatalytic transformations, we have demonstrated that the potential decrease in substrate affinity has not negatively impacted on enantioselectivity and in fact the enantioselectivity has been improved relative to wild type TK.

4.4 Conclusions

We have produced libraries of single, double and triple TK variants based around a cluster of co-evolving residues identified by SCA. The cluster of residues comprised of two structurally contiguous groups of residues and although only a small number of variants were introduced at each position, single variants with improved activity on the non-natural substrate propionaldehyde were identified for each of the positions in the structural group proximal to the active site. This demonstrates the potential of the SCA technique to identify new residues to target for site directed mutagenesis in directed evolution experiments.

Although the distal residues did not confer enhanced activity in isolation, the co-evolution of such residues suggests potential synergistic relationships between these and the proximal residues. We sought to identify evidence of such synergistic relationships by producing targeted double and triple variant libraries. The positions selected for combination were chosen such that negatively impacting variants would be combined with positively impacting variants. Synergistic relationships are expected to be more easily identifiable for such combinations.

Across the double and triple libraries, one variant was identified as significantly increasing activity on PA. This variant was D469S/E498D/R520Q and a triple mutant cycle was created to further investigate the synergistic relationships between each mutation. Analysis of the total lysate activities of the members of this triple mutant cycle indicates a lack of synergy between D469S and R520Q, however, both of these mutations alleviate the negative impact of the E498D mutation and when all three mutations are present together, activity is increased considerably. Analysis of the

total concentration and soluble fraction for each member of the cycle suggested that the combined effect of these mutations on activity cannot be explained by synergistic effects on protein expression or solubility. Instead, synergy is likely to be related to specific activity or better folding of the soluble fraction to the active native state.

Although negative effects on expression and solubility meant that no E498D containing variants could be purified for further kinetic analysis, the kinetic analysis of the other members of the triple mutant cycle has revealed some interesting relationships between the residues. Both the D469S and the R520Q mutations have a relatively small impact on enzyme kinetic parameters in isolation. However, when these two mutations are combined in D469S/R520Q, the turnover constant is increased by 20-fold relative to wild type TK. The K_m is also increased relative to wild type for the double mutant, suggesting a decrease in substrate affinity, but the combined effects of these changes still resulted in a 5.8-fold increase in k_{cat}/K_m .

In contrast with earlier results based on total lysate relative activity, enzyme kinetic analysis indicates significant synergy between the two mutations D469S and R520Q. However, total lysate relative activities were determined at the low PA concentration of 50 mM. This is 12.6-fold lower than the K_m of D469S/R520Q and as such, any synergistic increase in turnover constant could be masked by the concurrent increase in K_m . The synergy between these residues may have been apparent in earlier screens had a higher concentration of propionaldehyde been adopted.

Without the enzyme kinetics for the triple mutant D469S/E498D/R520Q it is difficult to explain why combination of all three mutations had such a beneficial effect in earlier screens. However, we can hypothesize that by altering the conformation of the active site, the E498D mutation may have alleviated the increase in K_m , thereby unmasking the synergistic increase in turnover constant at the lower substrate concentration used in the screen.

Interestingly, the D469S/R520Q double mutant of TK obtained by this approach is not found in any of the natural 382 TPP-dependent enzyme sequences used to perform the SCA. For the naturally occurring R520Q mutation, the equivalent to Asp 469 in TK is found to be Lys in various PDC and PPDC enzymes, Tyr in one TK and Asp in several ALS enzymes, but not Ser. When D469S occurs, the equivalent to Arg 520 in TK is found to be Glu or Arg. However, as the natural variants have been selected during their evolution for a particular range of functions useful to the cell, it is not necessarily expected that precisely the same natural combinations would be found when selecting for a new property such as altered substrate specificity.

In D469S/R520Q we have identified a multiple variant of TK that has a higher turnover constant for the PA reaction than any other TK variant discovered to date. Although the K_m was also increased, this does not pose a problem as long as substrate concentration can be increased sufficiently and the lower substrate affinity doesn't negatively impact on enantioselectivity. We have shown that enantioselectivity is actually increased with PA relative to wild type. The D469S/R520Q mutant is therefore very promising for the synthesis of chiral aliphatic keto diols, particularly (3S)-1,3-dihydroxy pentan-2-one.

5 General Discussion

5.1 Overall summary of thesis

The aim of this thesis was to utilise computational and bioinformatics tools to further our understanding of the transketolase enzyme and to aid in our efforts to engineer transketolase for biocatalytic applications.

Initially we sought to increase our mechanistic understanding by modelling substrate binding in the active site of *E. coli* transketolase and the results presented in chapter 2 add to the growing body of structural information on this enzyme. Computational docking allowed us to model transient, reactive structures that would be very difficult or impossible to determine experimentally. Of particular interest was the docked conformation of the ketol donor HPA, which indicates an interesting angle of nucleophilic attack, which although unusual, is supported by recently published structural data.

In chapter 3 we have used a powerful bioinformatics approach termed Statistical Coupling Analysis to identify networks of co-evolving residues across the TPP-dependent enzymes. Statistical Coupling Analysis or SCA, provides an opportunity to delve deeper than the structure of proteins, uncovering networks of residues within the protein fold that have acted together through evolution to maintain or improve fitness. The identification of such networks in transketolase has important implications for enzyme engineering, suggesting combinations of residues to target simultaneously and identifying distant residues that are functionally coupled to key residues within the active site.

In chapter 4 we used the results of statistical coupling analysis to guide the design of multiple transketolase mutants with improved activity on non-natural substrates. Small targeted mutagenesis libraries were produced based on one of the co-evolving networks and simultaneous mutation of multiple residues within this cluster displayed synergistic effects on enzyme activity that would not have been anticipated from the effects of each mutation in isolation. Notably, we have identified a double mutant with a 20-fold improvement in turnover-rate using the non-natural substrate propionaldehyde. This is greater than any improvement identified to date from single variant libraries.

In these three results chapters we have achieved the main aims of this thesis. However, many questions have been raised in the course of this work and considerable additional work is required to test and expand on the conclusions presented here. In the following discussion we will review the conclusions presented and propose further work which we hope will develop these conclusions further.

5.2 Computational automated docking in transketolase

Computational automated docking is a powerful technique that can help to rationalise experimental observations from enzyme kinetics, identify potential transient structures along an enzyme reaction pathway that are difficult to obtain by experimental structure determination, and generate hypotheses to test further by experiment. Although computational docking is not completely accurate, the most obvious errors can be eliminated by visual inspection, as was the case with DX5P in this work. Transketolase is a particularly good model for this approach as

the lack of induced-fit side chain movements within the enzyme eliminates the requirement to model side chain flexibility and the potential errors this can introduce.

We used the AutoDock algorithm to reproduce the structures of known substrate-protein complexes as an initial benchmark for using the docking technique in other transketolase complexes. AutoDock was able to accurately reproduce the complex of DE4P bound in yeast TK within 1.65 Å RMSD of the crystal structure. Although AutoDock appears to struggle with explicitly charged groups, resulting in a slight shift of DE4P towards the phosphate interacting residues, the hydrogen bond network of the docked substrate was predicted accurately. Docking DE4P in *E. coli* TK demonstrated a near identical binding conformation to that for yeast TK. This is an important finding as it allows us to confidently apply the knowledge gained from studies of yeast TK to *E. coli* TK and *visa versa*.

Having demonstrated the accuracy of AutoDock we docked a series of known ketol donor and aldehyde acceptor substrates into the active site of *E. coli* TK and calculated their K_m values from AutoDock reported binding energies. Comparison with experimentally determined K_m values demonstrated good correlation, further validating the approach. Although AutoDock appears to systemically underestimate K_m values, this is not of concern for the use of the binding energy calculation to select minimal energy docked conformations.

Examination of the structures of TK-substrate complexes reveals a common binding conformation, similar to that of DE4P. Phosphorylated substrates all bound with the phosphate group interacting with a conserved group of residues near the entrance

to the active site. This strong interaction leads to low experimental and calculated K_m values, and the phosphate groups anchoring effects result in an inversely proportional relationship between substrate chain length and the distance of docked substrate to the ThDP cofactor. Both phosphorylated and non-phosphorylated aldol acceptor substrates bound with the Re-face of the aldehyde carbonyl exposed to nucleophilic attack by the α -carbon of the enamine to give S-enantiomer products.

Two of the aldehyde acceptors, glycolaldehyde and DE4P were docked both in the presence and absence of the reactive ThDP-enamine intermediate. Such structures would be difficult or impossible to determine experimentally given their reactive nature. Binding of the two substrates was found to be identical in the presence and absence of the enamine, indicating that formation of the enamine is not a prerequisite for binding in a reactive conformation.

Comparison of docked acyclic DR5P with the recently published structure of the bound cyclic form suggests a potential mechanism for the ring opening of the substrate. A series of C-C bond rotations and hydrogen bond exchanges has been proposed that collectively unwind the DR5P molecule and orientate the newly acyclic form in readiness for nucleophilic attack by the enamine.

We have also docked ketol donors HPA and DX5P in the active site of *E. coli* TK. Unfortunately the docked conformation of DX5P was orientated unfavourably relative to the ThDP thiazolium ring; however, examination of the docked conformation of HPA has produced some of the most interesting findings of the computational docking work. HPA was docked in such a conformation that the

angle of nucleophilic attack by the deprotonated ThDP cofactor is 68° rather than the favoured Bergi-Dunitz angle of 107° . This unusual result correlates well with the observed strain in the covalent ThDP-DX5P intermediate and supports the growing picture of a catalytically poised active site in which substrate interaction energies are converted into the strained high-energy intermediate. The identical docking conformation of unreactive FPA provides an opportunity to test this conclusion experimentally.

5.3 Statistical coupling analysis of transketolase

Computational modelling of substrate binding in the active site of TK has provided evidence supporting our previous assumptions that substrate binding is equivalent between *E. coli* and yeast TK, and that many of the natural substrate interactions are maintained with non-natural substrates. This work gives us confidence in using such assumptions and has also increased our mechanistic understanding of TK. However, we have not identified any new residue functions that could lead to the improved engineering of TK for biocatalytic applications. This is perhaps unsurprising given the extensive mutagenesis and enzyme engineering work which has focussed on the residues around the active site.

When it comes to engineering TK, and enzyme engineering as a whole, the main gaps in our experience relate to multiple mutation of residues and the identification of target residues distant to the active site. These types of interactions are governed by complex relationships that are difficult to tease apart with structural data alone. The relatively new technique of statistical coupling analysis (SCA) has the potential to guide our search for beneficial multiple and distant mutations and

the following results describe the application of this technique to the transketolase enzyme.

Using a multiple sequence alignment of the PP and Pyr domains of TPP-dependent enzymes we initially used SCA to identify residues evolutionarily coupled to the important functional residue Asp 469. Seven residues were identified, five of which made up a structurally contiguous group around the active site. The other two residues were more distant; however, both were positioned on elements of secondary structure leading directly to the active site and may play a role in fine adjustment of the tertiary structure. Two of the five identified active site residues, Arg 520 and His 26, had been previously identified in enzyme engineering for modified substrate specificity. The improved activity of Arg 520 variants on non-natural substrates has been rationalised by the role of Arg 520 in phosphate binding. Evolutionary coupling between Arg 520 and Asp 469 suggests there may be a subtler synergistic explanation for the increased activity of these variants.

In order to expand our analyses of evolutionary coupling we carried out global SCA on the whole PP and Pyr domain MSA. Focusing on areas of high coupling energy we identified thirty highly coupled residues, six of which have a known function in TK. Evolutionary conservation is often used to determine potentially functional residues; SCA offers an alternative approach to the identification of functional residues. Notably, only three of the thirty SCA identified residues would also have been identified in a cut of the thirty most conserved residues. Two of these residues, His 26 and His 66, are well characterised but the third, Gly 117, would

make an interesting target for saturation mutagenesis given its redundant identification by both approaches.

Residues identified by global SCA were dominated by those located at the subunit interface between the PP and Pyr domains; this is not unexpected as changes on one side are likely to require changes on the other side to maintain interactions. However, one small sub-cluster did stand out among the thirty residues, this sub-cluster included residues His 26, His 66 and Tyr 72. This group of residues displayed a distinct coupling pattern and was also identified among the residues coupled to Asp 469. Leu 62 was also of interest, displaying the highest levels of coupling in the whole alignment and clustered independently of the other twenty-nine residues.

In order to identify subtler networks of coupled residues that may have been masked by strong subunit interaction networks, SCA global analysis was repeated with just the Pyr domains. This strategy identified an interesting network of nine coupled residues which included Asp 469. When mapped onto the structure of TK, this cluster forms two structurally contiguous groups of residues linked midway by the ninth residue which forms a hairpin turn. Confidence in this cluster is enhanced by its self-consistency and its structurally contiguous nature.

The identification of co-evolving networks of residues in the TPP-dependent enzymes demonstrates the power of this technique. The co-dependence of variation through evolution is likely to mirror co-dependency between newly introduced mutations. As such, the networks identified provide a new recourse of residues to target both in isolation and in combination for the engineering of *E. coli* TK. The secondary finding of this work, that many coupling interactions are located

on the interface between interacting subunits, suggests a new application of the SCA technique. Determination of the sites of protein-protein interaction in multi-protein complexes is an experimentally intensive process. Multiple sequence alignments of two proteins known to interact could be concatenated prior to statistical coupling analysis, providing a new method for the prediction of protein-protein interaction sites.

5.4 SCA guided library design for engineering transketolase activity

Based on the networks of evolutionarily coupled residues identified by SCA, we sought to engineer the activity of transketolase by targeting multiple residues for combined mutagenesis. As a high-throughput colorimetric screen had been developed in the lab for activity on the propionaldehyde substrate, improved activity on this reaction was selected as an engineering goal. We chose the nine residue Pyr domain cluster of co-evolving residues to target for mutagenesis as it displays self-consistency, is structurally contiguous, and contains the Asp 469 residue (mutation of which has improved activity on PA).

Single point mutation of seven of the nine cluster residues to common variants in the multiple sequence alignment produced interesting results, with all residues proximal to the active site producing improvements in activity on PA. In contrast, mutation of the two residues making up the structural group distal to the active site did not confer any improvement in activity, in fact activity was severely impaired. The identification of improved activity in a relatively small library of single variants demonstrates the potential of SCA to select residues for mutagenesis. Full

saturation mutagenesis at all the sites identified by SCA could be a productive strategy in the future directed evolution of TK.

The potential for SCA to identify combinations of residues for multiple mutation was investigated by selecting two groups of residues from the cluster for combined mutagenesis. Average relative activities and ranges of relative activities across these libraries indicated synergy between the residues. This synergy was also displayed in a triple mutant cycle of a particularly active triple mutant, D469S/E498D/R520Q. Although these results support our assumption of synergistic relationships between co-evolving residues, we have not investigated whether any synergy is present between non-co-evolving residues. Such an experiment would represent an important negative control for further work.

Detailed kinetic analysis of a subset of the triple mutant cycle demonstrated that the double mutant D469S/R520Q increases turnover rate of TK on the PA reaction by 20-fold. Although K_m was also increased for this variant, the combined effect of these changes was a 5.8-fold increase in k_{cat}/K_m . The apparent synergy between these mutations contradicts earlier results which suggested the requirement of a third mutation at Glu 498 to unlock the synergistic relationships at these sites. However, it is likely that the increase in K_m , combined with the relatively low PA concentration used in earlier analyses, masked the synergistic increase in turnover rate of this double mutant.

Using SCA to guide the selection of multiple variants we have identified a double variant in D469S/R520Q that has a higher turnover rate for the production of DHP than any other TK variant discovered to date. These residues were known

previously for their propensity to improve activity on the PA reaction following mutagenesis. Based on this prior knowledge it could be argued that combination of such variants would be expected to give further improvements in activity. However, the improvements demonstrated here are greater than could be expected based on the effects of each mutation in isolation and earlier work to combine beneficial mutations in *E. coli* TK has failed to produce any improvements in activity.

Given the promising results obtained from targeting the nine residue Pyr domain cluster, further work should be carried out to investigate the potential of the other SCA clusters for mutagenesis. Of particular interest would be the small PP domain cluster of His 26, His 66 and Tyr 72, which demonstrated very robust clustering in our analysis. Many other residues were identified by SCA that have not been targeted for mutagenesis to date. The results presented here suggest that the creation of saturation mutagenesis libraries of these positions could be a very promising strategy for the future engineering of TK.

6 References

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7 Appendix

7.1 Example Autodock input files

7.1.1 DPF example file format

```

outlev 1 # diagnostic output level
intelec # calculate internal electrostatics
seed pid time # seeds for random generator
ligand_types C HD OA # atoms types in ligand
fld lQGD.maps.fld # grid_data_file
map lQGD.C.map # atom-specific affinity map
map lQGD.HD.map # atom-specific affinity map
map lQGD.OA.map # atom-specific affinity map
elecmap lQGD.e.map # electrostatics map
desolvmap lQGD.d.map # desolvation map
move HPA.pdbqt # small molecule
about 909.9433 895.6123 902.4398 # small molecule center
tran0 random # initial coordinates/A or random
quat0 random # initial quaternion
ndihe 4 # number of active torsions
dihe0 random # initial dihedrals (relative) or random
tstep 2.0 # translation step/A
qstep 50.0 # quaternion step/deg
dstep 50.0 # torsion step/deg
torsdof 2 0.274000 # torsional degrees of freedom and coefficient
rmstol 0.5 # cluster_tolerance/A
extrng 1000.0 # external grid energy
e0max 0.0 10000 # max initial energy; max number of retries
ga_pop_size 150 # number of individuals in population
ga_num_evals 50000000 # maximum number of energy evaluations
ga_num_generations 50000 # maximum number of generations
ga_elitism 1 # number of top individuals to survive to next generation
ga_mutation_rate 0.02 # rate of gene mutation
ga_crossover_rate 0.8 # rate of crossover
ga_window_size 10 #
ga_cauchy_alpha 0.0 # Alpha parameter of Cauchy distribution
ga_cauchy_beta 1.0 # Beta parameter Cauchy distribution
set_ga # set the above parameters for GA or LGA
sw_max_its 300 # iterations of Solis & Wets local search
sw_max_succ 4 # consecutive successes before changing rho
sw_max_fail 4 # consecutive failures before changing rho
sw_rho 1.0 # size of local search space to sample
sw_lb_rho 0.01 # lower bound on rho
ls_search_freq 0.06 # probability of performing local search on individual
set_sw1 # set the above Solis & Wets parameters
compute_unbound_extended # compute extended ligand energy
ga_run 50 # do this many hybrid GA-LS runs
analysis # perform a ranked cluster analysis

```

7.1.2 GPF example file format

```

npts 64 64 64 # num.grid points in xyz
gridfld lQGD.maps.fld # grid_data_file
spacing 0.2027777777778 # spacing(A)
receptor_types A C HD N OA P SA # receptor atom types
ligand_types C HD OA # ligand atom types
receptor lQGD.pdbqt # macromolecule
gridcenter -11.806 24.875 37.708 # xyz-coordinates or auto
smooth 0.5 # store minimum energy w/in rad(A)
map lQGD.C.map # atom-specific affinity map
map lQGD.HD.map # atom-specific affinity map
map lQGD.OA.map # atom-specific affinity map
elecmap lQGD.e.map # electrostatic potential map
dsolvmap lQGD.d.map # desolvation potential map
dielectric -0.1465 # <0, AD4 distance-dep.diel;>0, constant

```

7.1.3 PDBQT 1QGD file input

REMARK	4	XXXX	COMPLIES	WITH	FORMAT	V.	2.	0.				
ATOM	1	N	SER	A	2	-20.614	26.561	2.838	1.00	40.61	-0.064	N
ATOM	2	HN1	SER	A	2	-20.435	27.122	2.005	1.00	0.00	0.275	HD
ATOM	3	HN2	SER	A	2	-21.375	25.894	2.709	1.00	20.08	0.275	HD
ATOM	4	HN3	SER	A	2	-19.868	25.890	3.021	1.00	0.00	0.275	HD
ATOM	5	CA	SER	A	2	-20.857	27.468	4.000	1.00	41.29	0.297	C
ATOM	7	C	SER	A	2	-19.652	28.413	2.244	1.00	36.13	0.297	C
ATOM	8	O	SER	A	2	-18.548	28.052	3.949	1.00	40.50	-0.271	OA
ATOM	9	CB	SER	A	2	-21.127	26.624	5.254	1.00	37.74	0.208	C
ATOM	12	CG	SER	A	2	-22.465	25.785	6.976	1.00	50.58	-0.199	OA
ATOM	13	HG	SER	A	2	-22.418	25.263	5.791	1.00	0.00	0.209	HD
ATOM	14	N	SER	A	3	-19.983	29.605	4.754	1.00	31.54	-0.344	N
ATOM	15	HN	SER	A	3	-20.390	29.894	4.700	1.00	20.50	0.163	HD
ATOM	16	CA	SER	A	3	-18.897	30.515	5.117	1.00	28.90	0.200	C
ATOM	18	C	SER	A	3	-18.118	29.888	6.289	1.00	24.09	0.243	C
ATOM	19	O	SER	A	3	-18.654	29.043	7.067	1.00	25.26	-0.271	OA
ATOM	20	CB	SER	A	3	-19.424	31.872	5.573	1.00	28.68	0.199	C
ATOM	23	CG	SER	A	3	-20.260	31.729	6.723	1.00	28.67	-0.199	OA
ATOM	24	HG	SER	A	3	-20.888	32.574	7.000	1.00	0.00	0.209	HD
ATOM	25	N	ARG	A	4	-16.925	30.428	6.498	1.00	23.06	-0.346	N
ATOM	26	HN	ARG	A	4	-16.977	31.140	5.856	1.00	20.00	0.163	HD
ATOM	27	CA	ARG	A	4	-16.086	30.017	7.639	1.00	21.92	0.176	C
ATOM	29	C	ARG	A	4	-16.850	30.305	8.917	1.00	22.03	0.241	C
ATOM	30	O	ARG	A	4	-16.837	29.457	9.145	1.00	20.94	-0.271	OA
ATOM	31	CB	ARG	A	4	-14.735	30.712	7.632	1.00	22.03	0.036	C
ATOM	34	CG	ARG	A	4	-13.929	30.273	6.420	1.00	22.16	0.023	C
ATOM	37	CD	ARG	A	4	-12.451	29.521	6.582	1.00	23.62	0.138	C
ATOM	40	NE	ARG	A	4	-11.829	29.397	7.276	1.00	33.17	-0.227	N
ATOM	41	HE	ARG	A	4	-12.270	29.073	8.137	1.00	0.00	0.177	HD
ATOM	42	CE	ARG	A	4	-12.739	29.768	8.974	1.00	24.97	0.665	C
ATOM	43	NH1	ARG	A	4	-10.121	29.133	5.572	1.00	29.86	-0.235	N
ATOM	44	NH2	ARG	A	4	-9.279	28.647	5.442	1.00	0.00	0.174	HD
ATOM	45	ZNH1	ARG	A	4	-10.459	29.177	5.474	1.00	20.00	0.174	HD
ATOM	46	NH2	ARG	A	4	-10.316	27.760	7.607	1.00	34.82	-0.235	N
ATOM	47	NH3	ARG	A	4	-9.474	27.274	7.297	1.00	0.00	0.174	HD
ATOM	48	ZNH2	ARG	A	4	-8.789	27.480	8.140	1.00	0.00	0.174	HD
ATOM	49	N	LYS	A	5	-17.539	31.451	8.998	1.00	19.31	-0.346	N
ATOM	50	HN	LYS	A	5	-17.552	32.106	8.216	1.00	0.00	0.163	HD
ATOM	51	CA	LYS	A	5	-17.747	31.747	9.169	1.00	21.43	0.241	C
ATOM	53	C	LYS	A	5	-19.350	30.740	10.537	1.00	18.28	0.241	C
ATOM	54	O	LYS	A	5	-19.581	30.418	11.708	1.00	15.31	-0.271	OA
ATOM	55	CB	LYS	A	5	-18.926	33.157	10.133	1.00	25.44	0.138	C
ATOM	58	CG	LYS	A	5	-17.810	34.179	10.373	1.00	39.34	0.004	C
ATOM	61	CD	LYS	A	5	-18.308	35.562	10.737	1.00	50.52	0.027	C
ATOM	64	CE	LYS	A	5	-18.391	36.450	10.939	1.00	51.53	0.027	C
ATOM	67	NE	LYS	A	5	-18.180	37.877	9.865	1.00	66.78	-0.079	N
ATOM	68	H21	LYS	A	5	-17.567	38.604	9.437	1.00	0.00	0.274	HD
ATOM	69	H22	LYS	A	5	-18.717	38.204	10.000	1.00	0.00	0.274	HD
ATOM	70	H23	LYS	A	5	-18.956	37.677	9.233	1.00	0.00	0.274	HD
ATOM	71	N	GLU	A	6	-20.077	30.251	9.524	1.00	18.06	-0.346	N
ATOM	72	HN	GLU	A	6	-19.900	30.563	8.468	1.00	16.63	0.163	HD
ATOM	73	CA	GLU	A	6	-21.114	29.266	9.810	1.00	16.83	0.177	C
ATOM	75	C	GLU	A	6	-20.499	27.956	10.283	1.00	15.27	0.241	C
ATOM	76	O	GLU	A	6	-21.077	27.321	11.073	1.00	17.44	-0.271	OA
ATOM	77	CB	GLU	A	6	-21.921	28.946	8.519	1.00	33.90	0.045	C
ATOM	80	CG	GLU	A	6	-23.396	28.818	8.839	1.00	55.63	0.116	C
ATOM	83	CD	GLU	A	6	-24.237	27.827	8.127	1.00	43.07	0.027	C
ATOM	84	OE1	GLU	A	6	-23.806	27.388	6.979	1.00	65.99	-0.648	OA
ATOM	85	OE2	GLU	A	6	-25.343	27.492	8.560	1.00	61.69	-0.648	OA
ATOM	86	N	LEU	A	7	-18.421	29.598	9.347	1.00	15.93	-0.177	N
ATOM	87	HN	LEU	A	7	-18.918	28.165	8.964	1.00	0.00	0.163	HD
ATOM	88	CA	LEU	A	7	-18.669	26.383	10.169	1.00	16.12	0.177	C
ATOM	90	C	LEU	A	7	-18.421	26.616	11.013	1.00	17.03	-0.271	OA
ATOM	91	O	LEU	A	7	-18.395	25.623	12.395	1.00	16.21	-0.271	OA
ATOM	92	CB	LEU	A	7	-17.443	26.031	9.288	1.00	12.40	0.038	C
ATOM	95	CG	LEU	A	7	-17.443	25.706	8.977	1.00	12.40	0.038	C
ATOM	97	CD1	LEU	A	7	-16.660	25.489	6.977	1.00	18.92	0.009	C
ATOM	101	CD2	LEU	A	7	-16.660	25.531	6.977	1.00	17.59	0.009	C
ATOM	105	N	ALA	A	8	-17.667	27.702	11.980	1.00	14.64	-0.346	N
ATOM	106	HN	ALA	A	8	-17.543	28.436	11.282	1.00	0.00	0.163	HD
ATOM	107	CA	ALA	A	8	-17.543	27.944	12.013	1.00	17.03	-0.271	OA
ATOM	109	C	ALA	A	8	-18.425	27.999	14.298	1.00	17.03	0.240	C
ATOM	110	O	ALA	A	8	-18.399	27.557	15.471	1.00	14.80	-0.271	OA
ATOM	111	CB	ALA	A	8	-19.659	29.650	10.408	1.00	11.08	0.045	C
ATOM	115	N	ASN	A	9	-19.595	28.483	13.823	1.00	15.31	-0.346	N
ATOM	116	HN	ASN	A	9	-19.645	28.803	12.868	1.00	0.00	0.163	HD
ATOM	117	CA	ASN	A	9	-19.595	28.204	14.498	1.00	14.80	0.240	C
ATOM	119	C	ASN	A	9	-21.314	27.123	15.058	1.00	15.44	0.241	C
ATOM	120	O	ASN	A	9	-21.992	26.975	16.106	1.00	17.34	-0.271	OA
ATOM	121	CB	ASN	A	9	-20.659	29.377	10.213	1.00	16.63	0.045	C
ATOM	124	CG	ASN	A	9	-21.640	30.809	14.053	1.00	26.48	0.217	C
ATOM	125	ND2	ASN	A	9	-22.302	31.618	13.221	1.00	19.99	-0.370	N
ATOM	126	HD2	ASN	A	9	-22.052	32.616	12.203	1.00	17.03	0.177	HD
ATOM	127	HD2	ASN	A	9	-23.040	31.242	12.626	1.00	0.00	0.159	HD
ATOM	128	OD1	ASN	A	9	-20.747	31.264	14.772	1.00	20.85	-0.274	OA
ATOM	129	N	GLY	A	10	-20.987	26.095	10.317	1.00	15.11	-0.271	OA
ATOM	130	HN	GLY	A	10	-20.462	26.263	13.457	1.00	0.00	0.163	HD
ATOM	131	CA	GLY	A	10	-21.325	24.723	14.665	1.00	17.57	0.177	C
ATOM	133	C	GLY	A	10	-20.659	24.377	16.040	1.00	16.63	0.045	C
ATOM	134	O	GLY	A	10	-21.378	23.787	16.866	1.00	16.07	-0.271	OA
ATOM	135	CB	GLY	A	10	-20.844	23.743	13.627	1.00	15.95	0.042	C
ATOM	139	N	LEU	A	11	-18.459	24.780	16.468	1.00	14.63	-0.346	N
ATOM	140	HN	LEU	A	11	-18.934	25.258	15.528	1.00	0.00	0.163	HD
ATOM	141	CA	LEU	A	11	-18.809	24.573	17.577	1.00	11.15	0.180	C
ATOM	143	C	LEU	A	11	-18.559	25.294	18.976	1.00	15.89	-0.271	OA
ATOM	144	O	LEU	A	11	-19.772	24.795	19.812	1.00	13.22	-0.271	OA
ATOM	145	CB	LEU	A	11	-17.354	25.114	17.496	1.00	15.56	0.013	C
ATOM	147	CG1	LEU	A	11	-16.924	24.389	16.126	1.00	15.02	0.002	C
ATOM	150	CG2	LEU	A	11	-16.660	24.997	18.881	1.00	11.38	0.012	C
ATOM	154	CD1	LEU	A	11	-15.164	24.882	16.196				

ATOM	540	C	VAL	A	39	-7.818	26.900	11.134	1.00	12.19	0.241	C
ATOM	541	O	VAL	A	39	-6.251	25.251	11.134	1.00	12.19	-0.241	CA
ATOM	542	CB	VAL	A	39	-8.300	29.360	11.033	1.00	15.21	0.009	C
ATOM	544	CO1	VAL	A	39	-6.965	29.509	10.316	1.00	12.73	0.012	C
ATOM	548	CO2	VAL	A	39	-9.280	30.422	10.316	1.00	12.73	-0.012	CA
ATOM	552	N	LEU	A	40	-7.450	26.764	12.415	1.00	10.84	-0.346	N
ATOM	553	HN	LEU	A	40	-7.926	27.277	13.157	1.00	0.00	0.163	HD
ATOM	554	CA	LEU	A	40	-6.335	25.847	12.415	1.00	10.84	0.346	N
ATOM	556	C	LEU	A	40	-6.631	24.416	12.245	1.00	10.31	0.241	C
ATOM	557	O	LEU	A	40	-5.802	23.764	11.603	1.00	11.29	-0.271	OA
ATOM	558	CB	LEU	A	40	-6.064	25.851	14.239	1.00	15.52	0.038	C
ATOM	561	CG	LEU	A	40	-5.041	24.788	14.701	1.00	15.11	-0.020	C
ATOM	563	OD1	LEU	A	40	-3.701	25.040	14.043	1.00	13.43	0.009	C
ATOM	567	OD2	LEU	A	40	-4.863	24.795	16.029	1.00	14.98	-0.346	N
ATOM	571	N	TRF	A	41	-7.793	23.896	12.681	1.00	9.86	-0.346	N
ATOM	572	HN	TRF	A	41	-8.460	24.478	13.188	1.00	0.00	0.163	HD
ATOM	573	CA	TRF	A	41	-6.261	24.494	12.681	1.00	9.86	0.346	N
ATOM	575	C	TRF	A	41	-8.437	22.241	10.951	1.00	12.76	0.241	C
ATOM	576	O	TRF	A	41	-7.979	21.228	10.441	1.00	14.33	-0.271	OA
ATOM	577	CB	TRF	A	41	-6.929	23.994	14.043	1.00	14.33	0.009	C
ATOM	580	CG	TRF	A	41	-8.799	21.653	14.681	1.00	12.27	-0.028	A
ATOM	581	OD1	TRF	A	41	-8.185	22.493	15.574	1.00	9.73	0.096	A
ATOM	583	OD2	TRF	A	41	-6.857	20.370	15.306	1.00	11.27	-0.002	A
ATOM	584	CE2	TRF	A	41	-8.295	20.495	16.586	1.00	9.35	0.042	A
ATOM	585	CE3	TRF	A	41	-9.349	19.125	14.883	1.00	14.55	0.014	A
ATOM	587	HE1	TRF	A	41	-7.411	22.186	17.560	1.00	0.00	0.165	HD
ATOM	588	HE1	TRF	A	41	-7.411	22.186	17.560	1.00	0.00	0.165	HD
ATOM	589	CE2	TRF	A	41	-8.185	19.403	17.458	1.00	15.48	0.030	A
ATOM	591	CH2	TRF	A	41	-8.249	18.047	15.760	1.00	15.23	0.003	A
ATOM	593	CH2	TRF	A	41	-8.678	18.195	17.043	1.00	16.09	0.002	A
ATOM	595	N	ARG	A	42	-6.355	23.105	10.305	1.00	11.74	-0.346	N
ATOM	596	HN	ARG	A	42	-6.974	24.015	10.750	1.00	0.00	0.163	HD
ATOM	597	CA	ARG	A	42	-9.464	22.836	8.883	1.00	12.96	0.176	C
ATOM	599	C	ARG	A	42	-8.322	23.356	7.322	1.00	17.14	0.000	C
ATOM	600	O	ARG	A	42	-8.471	22.994	6.952	1.00	15.58	-0.271	OA
ATOM	601	CB	ARG	A	42	-10.764	23.549	8.450	1.00	11.69	0.036	C
ATOM	604	CG	ARG	A	42	-9.052	22.023	13.006	1.00	10.69	0.044	C
ATOM	607	CD	ARG	A	42	-13.315	23.594	8.759	1.00	20.64	0.138	C
ATOM	610	NE	ARG	A	42	-13.485	23.315	7.355	1.00	27.01	-0.227	C
ATOM	611	HE	ARG	A	42	-7.355	22.339	7.127	1.00	20.00	0.177	HD
ATOM	612	CG	ARG	A	42	-13.424	24.111	6.307	1.00	28.86	0.665	C
ATOM	613	NH1	ARG	A	42	-13.199	25.409	6.333	1.00	21.29	-0.235	N
ATOM	614	NH1	ARG	A	42	-13.199	25.409	6.333	1.00	21.29	-0.235	N
ATOM	615	2HH1	ARG	A	42	-13.070	25.834	7.251	1.00	0.00	0.174	HD
ATOM	616	NH2	ARG	A	42	-13.592	23.556	5.108	1.00	21.95	-0.235	N
ATOM	617	NH2	ARG	A	42	-13.592	23.556	5.108	1.00	21.95	-0.235	N
ATOM	618	2HH2	ARG	A	42	-13.766	22.551	5.088	1.00	10.00	0.174	HD
ATOM	619	N	ASP	A	43	-7.527	24.170	8.172	1.00	14.09	-0.346	N
ATOM	620	HN	ASP	A	43	-6.442	24.724	9.421	1.00	16.29	0.000	HD
ATOM	621	CA	ASP	A	43	-6.496	24.063	7.220	1.00	16.29	0.186	C
ATOM	623	C	ASP	A	43	-5.089	24.519	7.572	1.00	18.44	0.241	C
ATOM	624	O	ASP	A	43	-6.247	24.044	6.627	1.00	18.71	-0.271	OA
ATOM	625	CB	ASP	A	43	-6.401	26.031	6.990	1.00	15.39	0.147	C
ATOM	628	CG	ASP	A	43	-7.587	26.010	6.240	1.00	25.46	-0.175	C
ATOM	629	OD1	ASP	A	43	-6.027	26.027	6.078	1.00	23.52	0.048	A
ATOM	630	OD2	ASP	A	43	-7.463	27.775	5.853	1.00	30.01	-0.648	OA
ATOM	631	N	PHE	A	44	-4.729	23.760	8.812	1.00	11.35	-0.346	N
ATOM	632	HN	PHE	A	44	-5.423	23.729	9.358	1.00	0.00	0.163	HD
ATOM	633	CA	PHE	A	44	-3.339	23.474	9.900	1.00	9.86	0.180	C
ATOM	635	C	PHE	A	44	-3.101	22.124	9.783	1.00	14.56	0.241	C
ATOM	636	O	PHE	A	44	-4.059	21.528	9.100	1.00	14.78	-0.271	OA
ATOM	637	CB	PHE	A	44	-2.761	24.562	10.050	1.00	9.51	0.073	C
ATOM	640	CG	PHE	A	44	-2.713	25.903	9.337	1.00	11.66	-0.056	A
ATOM	641	CH1	PHE	A	44	-3.682	26.133	8.145	1.00	14.86	0.007	A
ATOM	643	CH2	PHE	A	44	-3.733	26.829	9.512	1.00	10.16	0.007	A
ATOM	645	CE1	PHE	A	44	-1.653	27.359	7.739	1.00	19.74	0.001	A
ATOM	647	CE2	PHE	A	44	-4.718	28.038	8.406	1.00	15.93	-0.174	HD
ATOM	649	CZ	PHE	A	44	-2.665	28.276	7.929	1.00	14.01	0.000	A
ATOM	651	N	LEU	A	45	-4.012	21.732	10.662	1.00	8.62	-0.346	N
ATOM	652	HN	LEU	A	45	-4.718	22.028	10.766	1.00	0.00	0.163	HD
ATOM	653	CA	LEU	A	45	-3.615	20.537	11.450	1.00	10.36	0.177	C
ATOM	655	C	LEU	A	45	-3.438	19.302	10.605	1.00	11.33	0.241	C
ATOM	656	O	LEU	A	45	-4.621	18.902	9.806	1.00	12.71	-0.271	OA
ATOM	657	CB	LEU	A	45	-4.635	20.369	12.568	1.00	9.60	0.038	C
ATOM	660	CG	LEU	A	45	-4.332	19.348	13.676	1.00	13.24	-0.020	C
ATOM	662	CD1	LEU	A	45	-6.929	19.958	10.000	1.00	20.00	0.000	C
ATOM	666	CD2	LEU	A	45	-5.639	19.014	14.318	1.00	14.26	0.009	C
ATOM	670	N	LYS	A	46	-2.356	18.587	10.939	1.00	11.39	-0.346	N
ATOM	671	HN	LYS	A	46	-1.712	18.893	10.305	1.00	11.74	0.000	HD
ATOM	672	CA	LYS	A	46	-2.123	17.312	10.156	1.00	10.46	0.176	C
ATOM	674	C	LYS	A	46	-3.223	16.163	10.189	1.00	11.89	0.241	C
ATOM	675	O	LYS	A	46	-1.798	15.952	12.069	1.00	11.51	-0.271	OA
ATOM	676	CB	LYS	A	46	-0.650	17.214	9.822	1.00	10.22	0.035	C
ATOM	679	CG	LYS	A	46	-1.251	15.967	9.040	1.00	12.57	-0.020	C
ATOM	682	CD	LYS	A	46	-1.242	15.945	8.727	1.00	18.12	0.027	C
ATOM	685	CE	LYS	A	46	2.046	15.477	9.932	1.00	20.62	0.229	C
ATOM	688	HE	LYS	A	46	-1.424	14.734	10.147	1.00	14.72	0.000	HD
ATOM	689	H21	LYS	A	46	4.027	15.028	10.358	1.00	0.00	0.274	HD
ATOM	690	H22	LYS	A	46	3.862	16.202	9.149	1.00	0.00	0.274	HD
ATOM	691	H31	LYS	A	46	-1.424	14.734	10.147	1.00	14.72	0.000	HD
ATOM	692	N	HIS	A	47	-3.617	15.478	10.815	1.00	12.22	-0.346	N
ATOM	693	HN	HIS	A	47	-4.142	15.666	9.961	1.00	0.00	0.163	HD
ATOM	694	CA	HIS	A	47	-4.437	14.437	11.000	1.00	10.85	0.000	HD
ATOM	696	C	HIS	A	47	-4.953	13.460	10.946	1.00	11.16	0.241	C
ATOM	697	O	HIS	A	47	-5.327	13.768	9.802	1.00	11.54	-0.271	OA
ATOM	698	CB	HIS	A	47	-4.904	15.017	10.663	1.00	10.87	0.000	HD
ATOM	701	CG	HIS	A	47	-6.159	15.697	12.460	1.00	11.91	0.028	A
ATOM	702	CD2	HIS	A	47	-6.357	16.669	11.526	1.00	10.87	0.114	A
ATOM	704	NH1	HIS	A	47	-4.412	13.396	12.967	1.00	11.92	-0.037	A
ATOM	705	NH1	HIS	A	47	-7.598	14.833	13.672	1.00	0.00	0.166	HD
ATOM	706	CE1	HIS	A	47	-8.336	16.166	12.407	1.00	10.88	0.180	A
ATOM	708	HE2	HIS	A	47	-4.718	16.950	12.000	1.00	13.33	-0.346	N
ATOM	709	HE2	HIS	A	47	-8.163	17.463	10.921	1.00	0.00	0.166	HD
ATOM	710	N	ASN	A	48	-5.150	12.300	11.524	1.00	10.52	-0.346	N
ATOM	711	HN	ASN	A	48	-6.029	12.927	10.900	1.00	0.00	0.163	HD
ATOM	712	CA	ASN	A	48	-6.029	11.297	10.891	1.00	13.33	0.185	C
ATOM	714	C	ASN	A	48	-6.990	10.850	11.966	1.00	12.07	0.243	C
ATOM	715	O	ASN	A	48	-6.459	12.083	9.739	1.00	15.53	-0.271	OA
ATOM	716	CB	ASN	A	48	-5.205	10.118	10.358	1.00	13.02	0.137	C
ATOM	719	CG	ASN	A	48	-6.086	9.043	9.733	1.00	13.41	0.217	C
ATOM	720	ND2	ASN	A	48	-5.522	8.877	9.449	1.00	10.83	-0.037	A
ATOM	721	H2D2	ASN	A	48	-4.542	7.665	6.003	1.00	0.00	0.159	HD
ATOM	722	H2D2	ASN	A	48	-6.118	7.151					

AT009	1098	HN	TYR	A	72	-13.049	15.109	27.921	1.00	0.000	0.217	HD
AT010	1099	N	TYR	A	73	-16.352	19.944	30.111	1.00	0.000	-0.242	N
AT011	1100	HN	SER	A	73	-16.133	19.824	21.181	1.00	0.000	0.163	HD
AT012	1101	CA	SER	A	73	-16.973	19.507	19.213	1.00	11.778	0.200	C
AT013	1103	C	SER	A	73	-16.050	19.399	16.026	1.00	0.000	-0.175	C
AT014	1104	O	SER	A	73	-16.454	18.604	16.972	1.00	13.778	-0.271	OA
AT015	1105	CB	SER	A	73	-17.251	21.029	19.235	1.00	11.113	0.199	C
AT016	1108	CG	SER	A	73	-18.329	21.468	20.210	1.00	0.000	-0.209	CG
AT017	1109	HG	SER	A	73	-18.495	22.202	20.166	1.00	0.000	0.209	HD
AT018	1110	N	LEU	A	74	-14.786	19.645	18.135	1.00	10.553	-0.346	N
AT019	1111	HN	LEU	A	74	-14.487	20.115	18.989	1.00	0.000	0.163	HD
AT020	1112	CA	LEU	A	74	-13.813	19.442	17.040	1.00	9.996	0.177	C
AT021	1114	C	LEU	A	74	-13.612	17.970	16.711	1.00	12.888	0.241	C
AT022	1115	O	LEU	A	74	-19.575	17.986	15.584	1.00	11.293	-0.273	OA
AT023	1116	CB	LEU	A	74	-12.454	20.075	17.428	1.00	8.884	0.038	C
AT024	1119	CG	LEU	A	74	-12.399	21.615	17.414	1.00	15.553	-0.020	C
AT025	1121	CD1	LEU	A	74	-12.373	22.113	15.950	1.00	10.917	0.009	C
AT026	1125	CD2	LEU	A	74	-13.465	17.103	17.710	1.00	11.111	-0.346	N
AT027	1130	HN	LEU	A	75	-19.491	17.435	18.674	1.00	10.000	0.163	HD
AT028	1131	CA	LEU	A	75	-13.265	15.670	17.440	1.00	11.887	0.177	C
AT029	1133	C	LEU	A	75	-14.492	15.068	16.761	1.00	11.009	0.241	C
AT030	1134	O	LEU	A	75	-14.385	14.268	15.798	1.00	15.552	-0.273	OA
AT031	1135	CB	LEU	A	75	-12.979	14.919	18.773	1.00	9.446	0.038	C
AT032	1138	CG	LEU	A	75	-11.661	15.358	19.449	1.00	13.336	-0.020	C
AT033	1140	CD1	LEU	A	75	-10.415	14.874	18.718	1.00	10.311	0.009	C
AT034	1144	CD2	LEU	A	75	-10.415	14.874	18.718	1.00	10.311	0.009	C
AT035	1148	N	HIS	A	76	-15.886	15.420	17.271	1.00	11.007	-0.346	N
AT036	1149	HN	HIS	A	76	-14.728	16.944	18.879	1.00	0.000	0.163	HD
AT037	1150	CA	HIS	A	76	-16.939	14.909	16.673	1.00	13.774	-0.182	C
AT038	1152	C	HIS	A	76	-15.385	15.385	15.271	1.00	16.007	0.241	C
AT039	1153	O	HIS	A	76	-17.254	14.562	14.315	1.00	14.446	-0.271	OA
AT040	1154	CB	HIS	A	76	-18.147	15.437	17.448	1.00	14.677	0.093	C
AT041	1155	CG	HIS	A	76	-12.448	15.448	16.717	1.00	10.000	0.163	HD
AT042	1158	CD2	HIS	A	76	-20.307	15.810	16.074	1.00	17.881	-0.114	C
AT043	1160	ND1	HIS	A	76	-19.885	13.734	16.717	1.00	17.336	-0.354	N
AT044	1161	ND2	HIS	A	76	-12.448	15.448	16.717	1.00	10.000	0.163	HD
AT045	1162	CE1	HIS	A	76	-21.007	13.728	16.016	1.00	25.772	0.180	C
AT046	1164	NE2	HIS	A	76	-21.272	14.961	15.616	1.00	18.487	-0.360	N
AT047	1165	HE2	HIS	A	76	-22.166	15.455	15.455	1.00	15.000	-0.273	OA
AT048	1166	N	LEU	A	77	-16.991	16.700	14.998	1.00	12.005	-0.346	N
AT049	1167	HN	LEU	A	77	-16.853	17.337	15.782	1.00	0.000	0.163	HD
AT050	1168	CA	LEU	A	77	-15.723	17.231	16.171	1.00	14.463	-0.271	C
AT051	1170	C	LEU	A	77	-16.089	16.725	12.649	1.00	16.689	0.241	C
AT052	1171	O	LEU	A	77	-16.492	16.575	11.468	1.00	15.111	-0.271	OA
AT053	1172	CB	LEU	A	77	-16.492	16.575	11.468	1.00	15.111	-0.271	OA
AT054	1175	CG	LEU	A	77	-18.243	19.921	14.780	1.00	15.228	-0.020	C
AT055	1177	CD1	LEU	A	77	-17.864	20.921	14.717	1.00	16.336	0.009	C
AT056	1181	CD2	LEU	A	77	-17.451	19.937	14.717	1.00	16.336	0.009	C
AT057	1185	N	THR	A	78	-14.807	16.513	13.984	1.00	11.445	-0.346	N
AT058	1186	HN	THR	A	78	-14.517	16.650	12.952	1.00	0.000	0.163	HD
AT059	1187	CA	THR	A	78	-13.822	16.093	12.952	1.00	14.482	-0.271	C
AT060	1189	C	THR	A	78	-13.823	14.571	11.736	1.00	13.886	0.243	C
AT061	1190	O	THR	A	78	-13.133	14.129	10.814	1.00	13.588	-0.271	OA
AT062	1191	CB	THR	A	78	-15.371	16.548	16.867	1.00	14.463	0.038	C
AT063	1193	CG2	THR	A	78	-12.275	16.015	13.642	1.00	10.999	0.042	C
AT064	1197	OG1	THR	A	78	-12.075	16.015	13.642	1.00	13.770	-0.393	OA
AT065	1198	HG1	THR	A	78	-11.159	16.291	18.978	1.00	0.000	0.163	HD
AT066	1199	N	GLY	A	79	-14.586	13.788	12.471	1.00	16.449	-0.350	N
AT067	1200	HN	GLY	A	79	-15.104	14.172	13.261	1.00	0.000	0.163	HD
AT068	1201	CA	GLY	A	79	-14.686	12.359	13.146	1.00	9.999	-0.271	C
AT069	1204	C	GLY	A	79	-14.101	11.406	13.144	1.00	18.511	0.236	C
AT070	1205	O	GLY	A	79	-14.027	10.217	12.809	1.00	14.222	-0.272	OA
AT071	1206	N	TYR	A	80	-14.668	11.485	14.144	1.00	12.881	-0.299	N
AT072	1207	HN	TYR	A	80	-13.714	12.850	14.551	1.00	0.000	0.163	HD
AT073	1208	CA	TYR	A	80	-13.134	10.911	15.307	1.00	11.552	0.180	C
AT074	1210	C	TYR	A	80	-14.281	10.919	13.919	1.00	10.655	0.241	C
AT075	1211	O	TYR	A	80	-15.444	10.445	15.700	1.00	16.008	-0.271	OA
AT076	1212	CB	TYR	A	80	-12.521	11.614	16.545	1.00	11.000	0.073	C
AT077	1215	CG	TYR	A	80	-12.521	12.213	16.545	1.00	11.000	0.073	C
AT078	1216	CD1	TYR	A	80	-11.100	13.355	15.432	1.00	10.611	0.010	C
AT079	1218	CD2	TYR	A	80	-9.995	11.592	16.617	1.00	14.773	0.010	C
AT080	1220	CE1	TYR	A	80	-9.862	10.911	16.617	1.00	14.773	0.010	C
AT081	1222	CE2	TYR	A	80	-8.752	12.130	16.298	1.00	13.800	0.037	C
AT082	1224	CE	TYR	A	80	-8.710	13.271	15.528	1.00	14.533	0.065	C
AT083	1225	OB	TYR	A	80	-7.458	13.844	15.800	1.00	13.844	0.037	C
AT084	1226	HH	TYR	A	80	-7.463	14.828	14.653	1.00	10.000	0.217	HD
AT085	1227	N	ASP	A	81	-13.900	8.945	16.528	1.00	14.641	-0.345	N
AT086	1228	HN	ASP	A	81	-12.717	10.577	16.528	1.00	0.000	0.163	HD
AT087	1229	CA	ASP	A	81	-14.931	8.081	17.113	1.00	20.334	-0.186	C
AT088	1231	C	ASP	A	81	-13.519	8.586	15.984	1.00	21.338	0.241	C
AT089	1232	O	ASP	A	81	-15.190	8.223	19.501	1.00	19.200	-0.271	OA
AT090	1233	CB	ASP	A	81	-14.329	6.701	17.378	1.00	25.778	0.147	C
AT091	1236	CG	ASP	A	81	-15.527	6.701	17.378	1.00	25.778	0.147	C
AT092	1237	OD1	ASP	A	81	-16.554	5.831	17.722	1.00	40.552	-0.648	OA
AT093	1238	OD2	ASP	A	81	-14.880	4.691	18.535	1.00	41.689	-0.648	OA
AT094	1239	N	GLY	A	82	-14.880	4.691	18.535	1.00	41.689	-0.648	OA
AT095	1240	HN	GLY	A	82	-16.589	10.001	17.330	1.00	0.000	0.163	HD
AT096	1241	CA	GLY	A	82	-16.998	10.359	19.397	1.00	15.770	-0.177	C
AT097	1243	C	GLY	A	82	-16.683	12.299	19.397	1.00	16.683	0.241	C
AT098	1244	O	GLY	A	82	-16.803	11.865	18.361	1.00	16.443	-0.271	OA
AT099	1245	CB	GLY	A	82	-16.130	11.577	19.784	1.00	15.772	0.038	C
AT100	1248	CG	GLY	A	82	-16.488	11.367	22.240	1.00	18.522	0.009	C
AT101	1250	CD1	GLY	A	82	-15.911	13.582	21.274	1.00	18.584	0.009	C
AT102	1254	CD2	GLY	A	82	-15.911	13.582	21.274	1.00	18.584	0.009	C
AT103	1258	N	PRO	A	83	-20.685	10.166	18.512	1.00	19.339	-0.177	C
AT104	1259	CA	PRO	A	83	-21.472	11.187	19.208	1.00	18.772	0.241	C
AT105	1262	C	PRO	A	83	-19.137	8.515	19.208	1.00	21.010	-0.271	OA
AT106	1263	CB	PRO	A	83	-21.378	8.735	18.616	1.00	20.773	0.037	C
AT107	1266	CG	PRO	A	83	-20.556	7.910	19.522	1.00	21.066	0.022	C
AT108	1269	CD	PRO	A	83	-19.137	8.515	19.208	1.00	21.010	-0.271	OA
AT109	1272	N	MET	A	84	-22.527	11.678	18.547	1.00	16.133	-0.346	N
AT110	1273	HN	MET	A	84	-22.746	11.315	17.619	1.00	0.000	0.163	HD
AT111	1274	CA	MET	A	84	-23.367	12.323	17.120	1.00	17.120	-0.271	C
AT112	1276	C	MET	A	84	-23.967	12.323	20.452	1.00	18.776	0.241	C
AT113	1277	O	MET	A	84	-24.087	13.143	21.374	1.00	18.322	-0.271	OA
AT114	1278	CB	MET	A	84	-24.547	13.097	18.377	1.00	17.717	0.042	C
AT115	1281	CG	MET	A	84	-25.288	14.341	18.730</				

AT0M	1675	CG2	VAL	A	109	-9.564	14.151	24.403	1.00	18.888	0.012	C	
AT0M	1679	N	GLD	A	110	-4.900	14.970	24.000	1.00	8.744	-0.463	N	
AT0M	1680	HN	GLD	A	110	-5.272	15.474	23.571	1.00	0.000	0.163	HD	
AT0M	1681	CA	GLD	A	110	-3.551	14.225	23.868	1.00	8.239	0.177	C	
AT0M	1683	C	GLD	A	110	-3.034	14.297	HN	GLD	A	110	0.000	0.000
AT0M	1684	C	GLD	A	110	-1.928	13.774	25.531	1.00	8.933	-0.271	OA	
AT0M	1685	CB	GLD	A	110	-2.698	15.211	23.051	1.00	10.773	0.045	C	
AT0M	1688	CG	GLD	A	110	-3.043	15.176	21.000	1.00	0.000	0.163	HD	
AT0M	1691	CD	GLD	A	110	-2.464	13.956	20.849	1.00	15.338	0.172	C	
AT0M	1692	OEI	GLD	A	110	-1.694	13.147	21.416	1.00	17.177	-0.648	OA	
AT0M	1693	OE2	GLD	A	110	-2.720	13.803	19.636	1.00	15.922	-0.648	OA	
AT0M	1694	N	THR	A	111	-3.707	15.013	26.179	1.00	11.558	-0.344	N	
AT0M	1695	HN	THR	A	111	-4.544	15.518	25.889	1.00	0.000	0.163	HD	
AT0M	1696	CA	THR	A	111	-3.241	15.083	27.981	1.00	7.520	-0.205	C	
AT0M	1698	C	THR	A	111	-4.454	15.424	28.434	1.00	12.885	0.243	C	
AT0M	1699	C	THR	A	111	-5.477	15.864	27.864	1.00	12.920	-0.271	OA	
AT0M	1700	CB	THR	A	111	-4.181	15.396	24.328	1.00	0.000	0.163	HD	
AT0M	1702	CG2	THR	A	111	-2.687	17.562	27.286	1.00	12.620	0.042	C	
AT0M	1705	OG1	THR	A	111	-1.724	16.247	29.071	1.00	9.338	-0.393	OA	
AT0M	1707	HG1	THR	A	112	-4.181	15.396	24.328	1.00	0.000	0.163	HD	
AT0M	1708	N	THR	A	112	-4.359	15.293	29.743	1.00	11.011	-0.344	N	
AT0M	1709	HN	THR	A	112	-3.488	14.937	30.137	1.00	0.000	0.163	HD	
AT0M	1710	CA	THR	A	112	-5.453	15.638	30.655	1.00	6.841	-0.205	C	
AT0M	1712	C	THR	A	112	-5.205	17.049	31.181	1.00	10.002	0.243	C	
AT0M	1713	C	THR	A	112	-4.117	17.290	31.729	1.00	10.779	-0.271	OA	
AT0M	1714	CB	THR	A	112	-5.489	14.621	31.100	1.00	11.001	-0.271	OA	
AT0M	1716	CG2	THR	A	112	-6.590	14.930	32.853	1.00	9.011	0.042	C	
AT0M	1720	OG1	THR	A	112	-5.714	13.335	31.219	1.00	11.337	-0.393	OA	
AT0M	1721	HG1	THR	A	112	-4.731	12.700	31.928	1.00	6.000	0.210	HD	
AT0M	1722	N	THR	A	113	-6.085	18.001	30.923	1.00	9.233	-0.344	N	
AT0M	1723	HN	THR	A	113	-4.917	17.791	30.731	1.00	0.000	0.163	HD	
AT0M	1724	CA	THR	A	113	-5.864	14.363	31.432	1.00	10.449	-0.205	C	
AT0M	1726	C	THR	A	113	-7.007	19.744	32.363	1.00	14.337	0.243	C	
AT0M	1727	C	THR	A	113	-6.229	19.229	32.768	1.00	12.315	-0.271	OA	
AT0M	1728	CB	THR	A	113	-5.717	20.424	30.314	1.00	16.553	0.042	C	
AT0M	1730	CG2	THR	A	113	-4.436	20.167	33.530	1.00	18.600	0.163	HD	
AT0M	1734	OG1	THR	A	113	-6.033	20.222	32.438	1.00	11.167	-0.393	OA	
AT0M	1735	HG1	THR	A	113	-7.616	20.425	29.927	1.00	0.000	0.210	HD	
AT0M	1736	N	GLY	A	114	-6.945	20.984	32.892	1.00	11.331	-0.350	N	
AT0M	1737	HN	GLY	A	114	-5.259	21.030	32.548	1.00	0.000	0.163	HD	
AT0M	1738	CA	GLY	A	114	-7.892	21.253	34.014	1.00	11.224	-0.225	C	
AT0M	1741	C	GLY	A	114	-7.105	22.094	35.021	1.00	11.330	0.238	C	
AT0M	1742	C	GLY	A	114	-7.403	23.242	34.262	1.00	15.512	-0.271	OA	
AT0M	1743	N	PRO	A	115	-6.001	21.583	35.508	1.00	11.222	-0.337	N	
AT0M	1744	CA	PRO	A	115	-5.127	22.357	36.400	1.00	9.733	-0.179	C	
AT0M	1746	C	PRO	A	115	-4.820	23.348	36.996	1.00	16.148	0.243	C	
AT0M	1747	C	PRO	A	115	-3.598	23.011	34.614	1.00	9.233	-0.271	OA	
AT0M	1748	CB	PRO	A	115	-4.167	21.344	36.996	1.00	13.448	0.037	C	
AT0M	1751	CG	PRO	A	115	-4.820	23.348	36.996	1.00	16.148	0.243	C	
AT0M	1754	CD	PRO	A	115	-5.481	20.132	35.377	1.00	13.811	-0.127	C	
AT0M	1757	N	GLY	A	116	-4.710	24.698	35.694	1.00	7.566	-0.346	N	
AT0M	1758	HN	GLY	A	116	-5.229	24.200	35.000	1.00	0.000	0.163	HD	
AT0M	1759	CA	GLY	A	116	-6.125	25.640	34.774	1.00	9.433	-0.177	C	
AT0M	1761	C	GLY	A	116	-2.419	25.701	34.754	1.00	12.004	0.240	C	
AT0M	1762	C	GLY	A	116	-9.201	25.502	35.100	1.00	9.811	-0.271	OA	
AT0M	1763	CB	GLY	A	116	-4.666	27.044	35.107	1.00	12.113	0.038	C	
AT0M	1766	CG	GLY	A	116	-6.177	27.172	34.897	1.00	11.440	-0.202	C	
AT0M	1768	CD1	GLY	A	116	-6.708	28.453	35.100	1.00	14.119	-0.271	OA	
AT0M	1772	CD2	GLY	A	116	-6.517	27.214	33.397	1.00	18.226	0.009	C	
AT0M	1776	N	GLY	A	117	-2.061	25.971	33.560	1.00	7.732	-0.351	N	
AT0M	1777	HN	GLY	A	117	-6.691	26.093	32.761	1.00	0.000	0.163	HD	
AT0M	1778	CA	GLY	A	117	-6.055	26.094	33.311	1.00	7.733	-0.225	C	
AT0M	1781	C	GLY	A	117	0.018	24.786	32.884	1.00	9.008	0.236	C	
AT0M	1782	C	GLY	A	117	-0.142	22.366	32.884	1.00	7.463	-0.179	C	
AT0M	1783	N	GLM	A	118	-0.537	23.616	33.255	1.00	8.447	-0.346	N	
AT0M	1784	HN	GLM	A	118	-1.424	23.603	33.758	1.00	0.000	0.163	HD	
AT0M	1785	CA	GLM	A	118	-0.142	22.366	32.884	1.00	7.463	-0.179	C	
AT0M	1787	C	GLM	A	118	-0.012	21.975	31.466	1.00	9.233	0.240	C	
AT0M	1788	C	GLM	A	118	0.894	21.305	30.983	1.00	7.119	-0.271	OA	
AT0M	1789	CB	GLM	A	118	-0.259	21.829	31.748	1.00	8.447	0.038	C	
AT0M	1792	CG	GLM	A	118	0.278	21.528	35.261	1.00	12.833	0.105	C	
AT0M	1795	CD	GLM	A	118	0.101	20.359	36.227	1.00	10.113	0.215	C	
AT0M	1796	HE2	GLM	A	118	-0.469	19.776	37.943	1.00	0.000	0.159	HD	
AT0M	1797	HE2	GLM	A	118	-0.758	19.776	37.943	1.00	0.000	0.159	HD	
AT0M	1798	HE2	GLM	A	118	-0.075	21.455	37.508	1.00	0.000	0.159	HD	
AT0M	1799	OE1	GLM	A	118	-0.800	19.855	37.396	1.00	0.000	0.174	HD	
AT0M	1800	N	GLY	A	119	-1.099	22.368	30.100	1.00	8.220	-0.351	N	
AT0M	1801	HN	GLY	A	119	-1.822	22.893	31.288	1.00	0.000	0.163	HD	
AT0M	1802	CA	GLY	A	119	-1.253	23.044	30.731	1.00	8.447	-0.225	C	
AT0M	1805	C	GLY	A	119	-0.070	22.595	28.546	1.00	8.446	0.236	C	
AT0M	1806	C	GLY	A	119	-0.070	22.595	28.546	1.00	8.446	0.236	C	
AT0M	1807	N	LIE	A	120	0.202	23.989	28.681	1.00	5.996	-0.346	N	
AT0M	1808	HN	LIE	A	120	-0.377	24.476	29.291	1.00	0.000	0.163	HD	
AT0M	1809	CA	LIE	A	120	-0.377	24.476	29.291	1.00	0.000	0.163	HD	
AT0M	1811	C	LIE	A	120	2.638	23.857	28.379	1.00	8.558	0.241	C	
AT0M	1812	C	LIE	A	120	3.504	23.597	27.499	1.00	7.118	-0.271	OA	
AT0M	1813	CB	LIE	A	120	2.638	26.043	28.103	1.00	10.113	0.038	C	
AT0M	1815	CG1	LIE	A	120	2.539	26.666	27.295	1.00	10.522	0.002	C	
AT0M	1818	CG2	LIE	A	120	1.583	26.443	29.579	1.00	12.228	0.012	C	
AT0M	1822	CD1	LIE	A	120	2.638	26.043	28.103	1.00	10.113	0.038	C	
AT0M	1826	N	ALA	A	121	2.815	24.443	29.650	1.00	5.665	-0.346	N	
AT0M	1827	HN	ALA	A	121	2.104	23.645	30.353	1.00	0.000	0.163	HD	
AT0M	1828	CA	ALA	A	121	4.013	22.710	30.353	1.00	6.841	-0.205	C	
AT0M	1830	C	ALA	A	121	4.122	23.888	29.255	1.00	5.933	0.240	C	
AT0M	1831	C	ALA	A	121	5.222	20.986	28.795	1.00	6.665	-0.271	OA	
AT0M	1832	CB	ALA	A	121	3.069	19.356	28.795	1.00	4.930	0.037	C	
AT0M	1836	N	ASN	A	122	3.037	22.611	29.154	1.00	5.884	-0.346	N	
AT0M	1837	HN	ASN	A	122	2.167	20.927	29.589	1.00	0.000	0.163	HD	
AT0M	1838	CA	ASN	A	122	3.069	19.356	28.795	1.00	4.930	-0.205	C	
AT0M	1840	C	ASN	A	122	3.373	19.611	26.939	1.00	5.117	0.241	C	
AT0M	1841	C	ASN	A	122	4.079	18.853	26.302	1.00	10.007	-0.271	OA	
AT0M	1842	CB	ASN	A	122	2.167	20.927	29.589	1.00	0.000	0.163	HD	
AT0M	1845	CG	ASN	A	122	1.184	18.300	29.877	1.00	8.447	0.217	C	
AT0M	1846	ND2	ASN	A	122	2.030	18.159	30.874	1.00	8.006	-0.370	N	
AT0M	1847	HE2	ASN	A	122	2.809	18.000	30.353	1.00	0.000	0.163	HD	
AT0M	1848	HE2	ASN	A	122	1.695	17.876	31.795	1.00	0.000	0.159	HD	
AT0M	1849	OD1	ASN	A	122	-0.053	18.109	30.339	1.00	10.998	-0.274	OA	
AT0M	1850	N	ALA	A	123	2.809	20.710	26.666	1.00	8.446			

ATOM	2246	O	THR	A	149	2.632	23.548	18.714	1.00	10.36	-0.271	OA
ATOM	2247	CB	THR	A	149	-5.747	23.048	19.714	1.00	10.43	-0.163	HD
ATOM	2248	CG2	THR	A	149	2.889	20.836	20.964	1.00	7.31	0.042	C
ATOM	2253	OG1	THR	A	149	4.581	19.903	19.470	1.00	10.23	-0.393	DA
ATOM	2254	HG1	THR	A	149	1.932	20.836	20.964	1.00	7.31	0.042	C
ATOM	2255	N	THR	A	150	0.678	10.241	18.704	1.00	8.05	-0.346	N
ATOM	2256	HN	THR	A	150	0.212	21.538	18.613	1.00	0.00	0.163	HD
ATOM	2257	CA	THR	A	150	-0.132	23.627	18.714	1.00	6.58	-0.073	C
ATOM	2259	C	THR	A	150	-0.819	23.663	20.233	1.00	9.96	0.241	C
ATOM	2260	O	THR	A	150	-1.335	22.640	20.649	1.00	12.27	-0.271	OA
ATOM	2261	CB	THR	A	150	-1.267	23.636	17.814	1.00	9.94	0.010	A
ATOM	2264	CG	THR	A	150	-0.793	23.663	16.365	1.00	6.77	-0.056	A
ATOM	2265	CD1	THR	A	150	-0.331	24.845	15.824	1.00	9.15	0.010	A
ATOM	2267	CD2	THR	A	150	-0.847	22.533	15.568	1.00	10.00	0.010	A
ATOM	2269	CE1	THR	A	150	0.095	24.906	14.498	1.00	10.46	0.037	A
ATOM	2271	CE2	THR	A	150	-0.433	22.595	14.234	1.00	8.14	0.037	A
ATOM	2273	CP	THR	A	150	0.437	23.797	13.949	1.00	10.04	0.065	A
ATOM	2274	OH	THR	A	150	0.450	23.868	12.389	1.00	13.21	-0.361	OA
ATOM	2275	HN	THR	A	150	0.779	24.688	12.040	1.00	0.00	0.217	HD
ATOM	2276	N	THR	A	151	-0.138	22.640	18.714	1.00	8.05	-0.346	N
ATOM	2277	HN	ALA	A	151	-0.514	25.658	20.469	1.00	0.00	0.163	HD
ATOM	2278	CA	ALA	A	151	-1.492	24.886	22.225	1.00	7.70	0.172	C
ATOM	2280	C	ALA	A	151	-2.455	26.757	22.313	1.00	9.14	0.010	A
ATOM	2281	O	ALA	A	151	-2.087	27.135	21.798	1.00	11.01	-0.271	OA
ATOM	2282	CB	ALA	A	151	-0.374	25.174	23.232	1.00	9.82	0.042	C
ATOM	2286	N	PHE	A	152	-3.882	25.962	22.955	1.00	5.22	-0.346	N
ATOM	2287	HN	PHE	A	152	-3.859	25.054	23.353	1.00	0.00	0.163	HD
ATOM	2288	CA	PHE	A	152	-4.434	27.117	23.193	1.00	4.33	0.180	C
ATOM	2290	C	PHE	A	152	-5.281	27.466	24.887	1.00	8.43	0.241	C
ATOM	2291	O	PHE	A	152	-4.242	26.493	25.468	1.00	9.85	-0.271	OA
ATOM	2292	CB	PHE	A	152	-4.584	26.698	22.517	1.00	9.57	0.073	C
ATOM	2295	CG	PHE	A	152	-6.353	26.750	21.521	1.00	13.77	-0.056	A
ATOM	2296	CD1	PHE	A	152	-5.474	26.972	20.461	1.00	7.04	0.010	A
ATOM	2298	CD2	PHE	A	152	-6.347	26.553	20.461	1.00	12.59	0.010	A
ATOM	2300	CE1	PHE	A	152	-5.980	27.015	19.168	1.00	15.90	0.001	A
ATOM	2302	CE2	PHE	A	152	-8.212	26.648	19.980	1.00	13.27	0.001	A
ATOM	2304	CG	PHE	A	152	-7.347	26.850	18.410	1.00	14.66	0.010	A
ATOM	2306	N	MET	A	153	-4.202	28.769	25.023	1.00	7.33	-0.346	N
ATOM	2307	HN	MET	A	153	-4.230	29.523	24.337	1.00	0.00	0.163	HD
ATOM	2308	CA	MET	A	153	-5.099	29.059	24.924	1.00	7.31	0.172	C
ATOM	2310	C	MET	A	153	-6.484	30.275	26.843	1.00	12.05	0.240	C
ATOM	2311	O	MET	A	153	-5.098	31.046	25.924	1.00	10.77	-0.271	OA
ATOM	2312	CB	MET	A	153	-5.002	29.629	26.944	1.00	13.68	0.010	A
ATOM	2315	CG	MET	A	153	-1.397	28.605	26.370	1.00	33.05	0.076	C
ATOM	2318	SD	MET	A	153	-0.052	29.696	26.244	1.00	21.02	-0.173	SA
ATOM	2319	CE	MET	A	153	-0.269	30.077	27.978	1.00	19.04	0.010	A
ATOM	2323	N	GLY	A	154	-5.124	30.487	28.135	1.00	11.30	-0.351	N
ATOM	2324	HN	GLY	A	154	-4.801	30.941	28.856	1.00	0.00	0.163	HD
ATOM	2325	CA	GLY	A	154	-5.922	31.698	29.152	1.00	8.67	0.172	C
ATOM	2328	C	GLY	A	154	-5.203	32.325	29.559	1.00	9.58	0.236	C
ATOM	2329	O	GLY	A	154	-4.037	32.279	29.885	1.00	8.44	-0.272	OA
ATOM	1440	N	ASP	A	155	-5.922	32.325	29.559	1.00	9.58	0.236	C
ATOM	1441	HN	ASP	A	155	-6.903	33.681	29.855	1.00	0.00	0.163	HD
ATOM	1442	CA	ASP	A	155	-5.231	34.413	31.063	1.00	7.42	0.186	C
ATOM	1443	C	ASP	A	155	-6.737	33.632	32.355	1.00	11.05	-0.163	DA
ATOM	1444	O	ASP	A	155	-3.775	33.994	32.901	1.00	11.07	-0.271	OA
ATOM	1445	CB	ASP	A	155	-6.169	35.008	31.565	1.00	10.56	0.147	C
ATOM	1446	CG	ASP	A	155	-6.633	36.410	30.473	1.00	11.13	0.172	C
ATOM	1447	OD1	ASP	A	155	-5.800	37.037	29.779	1.00	14.23	-0.648	OA
ATOM	1448	OD2	ASP	A	155	-7.849	36.452	30.185	1.00	12.16	-0.648	OA
ATOM	2342	N	GLY	A	156	-6.465	32.766	32.766	1.00	6.97	-0.163	HD
ATOM	2343	HN	GLY	A	156	-6.465	32.766	32.766	1.00	0.00	0.163	HD
ATOM	2344	CA	GLY	A	156	-5.248	31.876	33.930	1.00	11.15	0.225	C
ATOM	2347	C	GLY	A	156	-4.888	31.623	33.730	1.00	9.78	0.186	C
ATOM	2348	O	GLY	A	156	-3.064	31.272	34.639	1.00	10.57	-0.272	OA
ATOM	2349	N	CYS	A	157	-3.622	30.983	32.595	1.00	9.21	-0.346	N
ATOM	2350	HN	CYS	A	157	-4.365	30.977	31.949	1.00	0.00	0.163	HD
ATOM	2351	CA	CYS	A	157	-2.352	29.959	32.276	1.00	7.21	0.186	C
ATOM	2353	C	CYS	A	157	-1.203	30.981	32.300	1.00	10.21	0.241	C
ATOM	2354	O	CYS	A	157	-4.152	30.922	32.840	1.00	8.70	-0.271	OA
ATOM	2355	CB	CYS	A	157	-2.416	29.368	30.826	1.00	5.81	0.121	C
ATOM	2358	CG	CYS	A	157	-3.583	27.906	30.832	1.00	15.40	-0.095	SA
ATOM	2359	N	MET	A	158	-4.437	32.952	30.473	1.00	8.10	-0.163	HD
ATOM	2360	HN	MET	A	158	-2.378	32.421	31.494	1.00	0.00	0.163	HD
ATOM	2361	CA	MET	A	158	-6.425	33.235	31.741	1.00	4.66	0.177	C
ATOM	2362	C	MET	A	158	-6.086	33.235	31.741	1.00	4.66	0.177	C
ATOM	2364	O	MET	A	158	1.066	34.083	33.386	1.00	8.89	-0.271	OA
ATOM	2365	CB	MET	A	158	-0.863	34.000	30.933	1.00	5.32	0.045	C
ATOM	2366	CG	MET	A	158	-1.047	34.154	30.473	1.00	8.66	0.076	C
ATOM	2371	SD	MET	A	158	0.626	33.885	28.786	1.00	15.28	-0.173	SA
ATOM	2372	CE	MET	A	158	-0.137	34.154	30.473	1.00	8.85	0.010	A
ATOM	2376	N	ASP	A	159	-1.069	33.831	34.045	1.00	7.64	-0.346	N
ATOM	2377	HN	ASP	A	159	-2.030	33.605	33.789	1.00	0.00	0.163	HD
ATOM	2378	CA	ASP	A	159	-3.278	34.218	35.127	1.00	5.13	0.186	C
ATOM	2380	C	ASP	A	159	-0.039	34.243	36.104	1.00	9.29	0.241	C
ATOM	2381	O	ASP	A	159	0.834	33.608	37.085	1.00	7.47	-0.271	OA
ATOM	2382	CB	ASP	A	159	-4.848	34.617	34.147	1.00	9.45	0.045	C
ATOM	2385	CG	ASP	A	159	-2.596	35.981	35.721	1.00	9.65	0.076	C
ATOM	2388	SD	ASP	A	159	-4.092	36.381	36.663	1.00	11.04	-0.173	SA
ATOM	2389	CE	ASP	A	159	-1.077	36.116	35.448	1.00	14.48	0.010	A
ATOM	2393	N	GLY	A	160	-0.194	31.949	35.956	1.00	6.99	-0.346	N
ATOM	2394	HN	GLY	A	160	-0.846	31.679	35.220	1.00	0.00	0.163	HD
ATOM	2395	CA	GLY	A	160	-1.007	30.912	37.423	1.00	7.06	0.186	C
ATOM	2397	C	GLY	A	160	1.985	30.915	36.539	1.00	8.94	0.240	C
ATOM	2398	O	GLY	A	160	2.473	31.061	35.430	1.00	8.02	-0.271	OA
ATOM	2399	CB	GLY	A	160	-0.402	30.502	37.392	1.00	7.60	0.045	C
ATOM	2402	CG	GLY	A	160	-1.453	29.215	36.881	1.00	5.36	0.116	C
ATOM	2405	CD	GLY	A	160	-1.505	28.896	38.382	1.00	8.33	0.172	C
ATOM	2406	OD1	GLY	A	160	-4.457	28.872	39.025	1.00	6.16	-0.648	OA
ATOM	2407	OD2	GLY	A	160	-2.597	28.627	38.945	1.00	10.57	-0.648	OA
ATOM	2408	N	GLY	A	161	2.709	30.715	37.628	1.00	4.71	-0.351	N
ATOM	2409	HN	GLY	A	161	-2.229	30.606	38.522	1.00	0.00	0.163	HD
ATOM	2410	CA	GLY	A	161	4.175	30.644	37.602	1.00	6.84	0.225	C
ATOM	2413	C	GLY	A	161	4.708	29.578	36.644	1.00	7.08	0.236	C
ATOM	2414	O	GLY	A	161	-7.769	29.578	36.644	1.00	7.85	-0.648	OA
ATOM	2415	N	ILE	A	162	3.999	28.459	36.490	1.00	4.85	-0.346	N
ATOM	2416	HN	ILE	A	162	3.102	28.317	36.954	1.00	0.00	0.163	HD
ATOM	2417	CA	ILE	A	162	4.598	28.977	35.920	1.00	3.59	0.186	C
ATOM	2419	C	ILE	A	162	4.716	27.975	34.188	1.00	7.48	0.241	C
ATOM	2420	O	ILE	A	162	5.580	27.453	33.472	1.00	7.29	-0.271	OA
ATOM	2421	CB	ILE	A	162	3.818	26.102	35.379	1.00	8.12	0.045	C
ATOM	2423											

ATOM 2824 CB ILE A 189	-13.316	33.202	38.083	1.00	13.95	0.013	C
ATOM 2825 CG1 ILE A 189	-13.216	33.202	38.083	1.00	13.95	0.013	C
ATOM 2826 CG2 ILE A 189	-14.302	33.085	39.214	1.00	15.79	0.012	C
ATOM 2827 CD1 ILE A 189	-13.158	30.686	37.913	1.00	22.82	0.005	C
ATOM 2828 CD2 ILE A 189	-13.888	38.449	38.180	1.00	16.43	0.015	C
ATOM 2829 HN ASP A 190	-11.901	36.124	38.466	1.00	0.00	0.163	HD
ATOM 2830 CA ASP A 190	-13.274	37.476	39.537	1.00	13.42	0.186	C
ATOM 2831 C ASP A 190	-13.888	38.449	38.180	1.00	16.43	0.015	C
ATOM 2842 O ASP A 190	-14.387	39.601	39.406	1.00	15.09	-0.271	OA
ATOM 2843 CB ASP A 190	-12.086	37.991	40.362	1.00	14.80	0.147	C
ATOM 2844 CG ASP A 190	-11.758	37.171	41.607	1.00	15.62	0.175	C
ATOM 2847 OD1 ASP A 190	-12.486	36.171	41.875	1.00	13.77	-0.648	OA
ATOM 2848 OD2 ASP A 190	-10.793	37.531	42.335	1.00	14.26	-0.648	OA
ATOM 2849 N ASP A 190	-13.658	38.764	37.439	1.00	14.84	-0.351	N
ATOM 2850 HN GLY A 191	-13.168	38.036	36.932	1.00	0.00	0.163	HD
ATOM 2851 CA GLY A 191	-14.186	39.932	36.695	1.00	18.53	0.225	C
ATOM 2854 C GLY A 191	-13.168	38.036	36.932	1.00	0.00	0.163	HD
ATOM 2855 O GLY A 191	-12.427	39.572	35.095	1.00	14.77	-0.272	OA
ATOM 2856 N HIS A 192	-13.120	41.690	35.372	1.00	15.68	-0.346	N
ATOM 2857 HN HIS A 192	-11.762	42.330	36.938	1.00	0.00	0.163	HD
ATOM 2858 CA HIS A 192	-12.180	42.236	34.395	1.00	17.32	0.182	C
ATOM 2860 C HIS A 192	-10.775	42.130	34.962	1.00	19.80	0.241	C
ATOM 2861 O HIS A 192	-10.552	42.658	36.493	1.00	13.79	-0.271	OA
ATOM 2862 CB HIS A 192	-12.547	43.699	34.103	1.00	22.19	0.093	C
ATOM 2865 CD HIS A 192	-13.821	43.732	33.297	1.00	27.67	0.028	A
ATOM 2866 CD2 HIS A 192	-15.721	43.661	33.978	1.00	31.95	-0.107	A
ATOM 2868 ND1 HIS A 192	-13.793	43.835	31.923	1.00	36.45	-0.354	N
ATOM 2869 HD1 HIS A 192	-12.951	43.900	31.251	1.00	0.00	0.166	HD
ATOM 2870 CE1 HIS A 192	-15.026	43.834	31.248	1.00	41.32	0.180	A
ATOM 2872 NE2 HIS A 192	-15.841	43.737	32.497	1.00	38.93	-0.360	N
ATOM 2873 HE1 HIS A 192	-16.859	43.712	32.161	1.00	46.60	0.166	HD
ATOM 2874 N VAL 193	-9.857	41.539	34.197	1.00	15.81	-0.346	N
ATOM 2875 HN VAL A 193	-10.091	41.285	33.237	1.00	0.00	0.163	HD
ATOM 2876 CA VAL 193	-11.841	42.326	34.841	1.00	16.80	0.166	HD
ATOM 2878 C VAL 193	-7.616	42.411	35.006	1.00	17.56	0.241	C
ATOM 2879 O VAL A 193	-6.663	42.173	35.754	1.00	15.80	-0.271	OA
ATOM 2880 CB VAL 193	-11.729	43.740	35.216	1.00	20.24	0.093	C
ATOM 2882 CG1 VAL A 193	-8.501	39.042	33.530	1.00	15.39	0.012	C
ATOM 2886 CO2 VAL A 193	-7.384	41.016	32.434	1.00	19.49	0.012	C
ATOM 2890 N THR A 194	-7.863	43.568	34.401	1.00	12.45	-0.346	N
ATOM 2891 HN GLD A 194	-8.701	43.819	34.018	1.00	0.00	0.163	HD
ATOM 2892 CA GLD A 194	-6.937	44.730	34.874	1.00	14.96	0.177	C
ATOM 2894 C GLD A 194	-6.785	44.966	34.735	1.00	16.69	0.241	C
ATOM 2895 O GLD A 194	-5.758	45.575	36.754	1.00	18.19	-0.271	OA
ATOM 2896 CB GLD A 194	-7.356	46.067	34.269	1.00	23.10	0.045	C
ATOM 2899 CG GLD A 194	-7.724	46.067	34.794	1.00	42.42	0.045	C
ATOM 2902 CD GLD A 194	-9.248	45.649	32.827	1.00	52.11	0.172	C
ATOM 2903 OE1 GLD A 194	-9.943	46.496	33.437	1.00	64.18	-0.648	OA
ATOM 2904 OE2 GLD A 194	-11.825	44.596	32.827	1.00	72.46	-0.648	OA
ATOM 2905 N GLY A 195	-7.696	44.561	37.227	1.00	17.11	-0.351	N
ATOM 2906 HN GLY A 195	-8.548	44.113	36.889	1.00	0.00	0.163	HD
ATOM 2907 CA GLY A 195	-4.659	44.741	36.678	1.00	13.54	0.225	C
ATOM 2910 C GLY A 195	-6.333	43.994	39.231	1.00	14.76	0.236	C
ATOM 2911 O GLY A 195	-5.864	44.135	40.365	1.00	17.01	-0.272	OA
ATOM 2912 N THR A 196	-8.941	42.821	38.540	1.00	11.59	-0.346	N
ATOM 2913 HN TRP A 196	-6.337	42.664	37.617	1.00	0.00	0.163	HD
ATOM 2914 CA TRP A 196	-4.952	41.893	39.087	1.00	8.91	0.181	C
ATOM 2916 C TRP A 196	-6.014	41.337	38.394	1.00	11.48	0.241	C
ATOM 2917 O TRP A 196	-3.140	40.532	38.398	1.00	12.37	-0.271	OA
ATOM 2918 CB TRP A 196	-5.691	40.711	39.797	1.00	9.20	0.075	C
ATOM 2921 CG TRP A 196	-6.788	40.103	39.981	1.00	11.33	-0.271	OA
ATOM 2922 CD1 TRP A 196	-6.711	39.074	38.085	1.00	12.60	0.096	A
ATOM 2924 CD2 TRP A 196	-8.177	40.468	39.039	1.00	11.05	-0.002	A
ATOM 2925 CE2 TRP A 196	-8.868	39.678	38.124	1.00	14.41	-0.346	N
ATOM 2926 CE3 TRP A 196	-8.862	41.476	39.151	1.00	12.02	0.014	A
ATOM 2928 NE1 TRP A 196	-7.956	38.794	37.567	1.00	13.50	-0.365	N
ATOM 2929 HE1 TRP A 196	-8.172	38.937	36.898	1.00	11.00	-0.346	N
ATOM 2930 C22 TRP A 196	-10.241	39.796	37.929	1.00	13.27	0.030	A
ATOM 2932 CD3 TRP A 196	-10.226	41.601	39.549	1.00	15.36	0.001	A
ATOM 2934 CH2 ARG A 196	-8.760	40.760	38.497	1.00	11.00	0.163	HD
ATOM 2936 N PHE A 197	-4.128	41.769	36.759	1.00	9.41	-0.346	N
ATOM 2937 HN PHE A 197	-4.811	42.486	36.515	1.00	0.00	0.163	HD
ATOM 2938 CA PHE A 197	-1.241	41.191	37.928	1.00	11.00	0.241	C
ATOM 2940 C PHE A 197	-2.940	42.302	34.718	1.00	11.77	-0.271	OA
ATOM 2941 O PHE A 197	-3.809	42.544	33.872	1.00	10.25	-0.271	OA
ATOM 2942 CB PHE A 197	-1.839	39.967	36.410	1.00	9.62	0.163	HD
ATOM 2945 CG PHE A 197	-3.056	39.310	34.008	1.00	7.95	-0.056	A
ATOM 2946 CD1 PHE A 197	-1.701	39.330	34.160	1.00	10.124	0.007	A
ATOM 2948 CD2 PHE A 197	-3.718	38.880	32.412	1.00	16.47	0.007	A
ATOM 2950 CE1 PHE A 197	-0.990	38.481	33.144	1.00	15.51	0.001	A
ATOM 2952 CE2 PHE A 197	-1.000	40.170	34.160	1.00	17.10	0.001	A
ATOM 2954 CZ PHE A 197	-1.654	38.068	32.001	1.00	13.18	0.000	A
ATOM 2956 N THR A 198	-1.773	42.949	34.837	1.00	11.46	-0.344	N
ATOM 2957 HN THR A 198	-1.868	43.378	34.124	1.00	0.00	0.163	HD
ATOM 2958 CA THR A 198	-1.503	44.100	33.928	1.00	12.30	0.205	C
ATOM 2960 C THR A 198	-0.225	43.929	33.128	1.00	15.26	0.243	C
ATOM 2961 O THR A 198	-0.414	44.921	32.877	1.00	12.48	-0.593	OA
ATOM 2962 CB THR A 198	-1.380	45.414	34.771	1.00	10.42	0.146	C
ATOM 2964 CG2 THR A 198	-2.727	45.772	35.424	1.00	13.25	0.042	C
ATOM 2968 OE1 THR A 198	-4.055	45.237	35.981	1.00	15.93	0.146	C
ATOM 2969 OE1 THR A 198	-3.323	44.956	35.447	1.00	0.00	0.210	HD
ATOM 2970 N ASP A 199	0.312	42.715	33.017	1.00	8.03	-0.345	N
ATOM 2971 HN ASP A 199	0.237	42.725	32.877	1.00	8.52	-0.345	N
ATOM 2972 CA ASP A 199	1.543	42.868	32.294	1.00	5.89	0.186	C
ATOM 2974 C ASP A 199	1.445	42.971	33.036	1.00	9.74	0.241	C
ATOM 2975 O ASP A 199	3.149	42.922	34.411	1.00	11.19	-0.271	OA
ATOM 2976 CB ASP A 199	1.857	40.969	32.074	1.00	8.71	0.147	C
ATOM 2979 CG ASP A 199	2.198	40.279	33.401	1.00	14.80	0.175	C
ATOM 2980 OD1 ASP A 199	2.474	39.077	33.001	1.00	10.65	-0.648	OA
ATOM 2981 OD2 ASP A 199	2.550	43.395	30.297	1.00	9.43	-0.345	N
ATOM 2983 HN ASP A 200	4.371	43.688	30.784	1.00	0.00	0.163	HD
ATOM 2984 CA ASP A 200	2.591	43.601	28.807	1.00	7.81	0.186	C
ATOM 2986 C ASP A 200	3.045	42.209	28.348	1.00	7.92	0.241	C
ATOM 2987 O ASP A 200	3.193	43.287	28.348	1.00	8.52	-0.271	OA
ATOM 2988 CB ASP A 200	3.615	44.698	28.480	1.00	9.00	0.147	C
ATOM 2991 CG ASP A 200	3.765	44.880	26.982	1.00	19.15	0.175	C
ATOM 2992 OD1 ASP A 200	4.317	45.861	26.492	1.00	13.69	-0.648	OA
ATOM 2993 OD2 ASP A 200	4.317	45.861	26.492	1.00	13.69	-0.648	OA
ATOM 2994 N THR A 201	2.053	41.312	28.078	1.00	8.15	-0.344	N
ATOM 2995 HN THR A 201	4.078	41.640	28.078	1.00	0.00	0.163	HD
ATOM 2996 CA THR A 201	2.400	39.905	27.782	1.00	10.33	0.205	C
ATOM 2998 C THR A 201	3.253	39.778	26.751	1.00	9.36	0.243	C
ATOM 2999 O THR A 201	4.155	39.841	26.244	1.00	8.11	-0.271	OA
ATOM 3000 CB THR A 201	1.071	39.121	27.526	1.00	9.60	0.146	C
ATOM 3002 CG2 THR A 201	1.240	37.670	27.275	1.00	13.49	0.042	C
ATOM 3006 OE1 THR A 201	4.828	39.117	29.075	1.00	10.70	-0.271	OA
ATOM 3007 HG1 THR A 201	0.493	40.019	29.359	1.00	0.00	0.210	HD
ATOM 3008 N ALA A 202	3.019	40.605	25.502	1.00	8.62	-0.346	N
ATOM 3009 HN ALA A 202	2.253	41.279	25.518	1.00	0.00	0.163	HD
ATOM 3010 CA ALA A 202	3.910	40.421	24.345	1.00	5.63	0.172	C
ATOM 3012 C ALA A 202	5.338	40.764	24.693	1.00	9.70	0.240	C
ATOM 3013 O ALA A 202	6.271	40.064	24.244	1.00	9.40	-0.271	OA
ATOM 3014 CB ALA A 202	3.424	41.486	23.241	1.00	9.95	0.042	C
ATOM 3018 N MET A 203	5.609	43.837	25.498	1.00	8.33	-0.346	N
ATOM 3019 HN MET A 203	4.851	42.400	25.701	1.00	0.00	0.163	HD
ATOM 3020 CA MET A 203	7.002	42.165	25.795	1.00	8.60	0.177	C
ATOM 3022 C MET A 203	6.624	43.187	26.684	1.00	9.21	0.241	C
ATOM 3023 O MET A 203	8.795	40.751	26.547	1.00	8.22	-0.271	OA
ATOM 3024 CB MET A 203	7.011	43.528	26.537	1.00	7.52	0.045	C
ATOM 3027 CG MET A 203	8.433	43.991	27.014	1.00	20.46	0.076	C
ATOM 3030 SD MET A 203	8.251	45.600	27.899	1.00	24.61	-0.173	SA
ATOM 3031 CE MET A 203	7.376	46.597	26.688	1.00	22.75	0.089	C
ATOM 3035 N ARG A 204	6.805	40.513	27.566	1.00	8.34	-0.346	N
ATOM 3036 HN ARG A 204	5.843	40.831	27.683	1.00	0.00	0.163	HD
ATOM 3037 CA ARG A 204	7.360	39.385	28.379	1.00	10.62	0.176	C
ATOM 3039 C ARG A 204	7.819	38.240	27.510	1.00	7.08	0.241	C
ATOM 3040 O ARG A 204	8.874	37.613	27.740	1.00	7.40	-0.271	

AT001	3381	NZ	LVS	A	225	-4.984	30.488	3.507	1.00	48.07	-0.079	N
AT002	3382	H21	LVS	A	225	-3.555	29.869	3.191	1.00	47.00	-0.274	N
AT003	3383	H22	LVS	A	225	-4.414	31.301	3.739	1.00	0.00	0.274	H
AT004	3384	H23	LVS	A	225	-5.781	30.990	3.116	1.00	0.00	0.274	H
AT005	3385	N	ARG	A	226	-1.351	37.336	7.387	1.00	0.06	-0.079	N
AT006	3386	HN	ARG	A	226	-4.251	34.832	8.326	1.00	0.00	0.163	H
AT007	3387	CA	ARG	A	226	-2.139	34.913	7.981	1.00	12.36	0.176	C
AT008	3388	CA	ARG	A	226	-1.169	34.843	9.149	1.00	0.00	0.176	C
AT009	3389	O	ARG	A	226	0.034	34.696	8.975	1.00	11.49	-0.271	O
AT010	3390	O	ARG	A	226	-2.475	36.362	7.612	1.00	18.74	0.036	C
AT011	3391	CG	ARG	A	226	-1.351	37.336	7.387	1.00	0.06	0.079	C
AT012	3392	CD	ARG	A	226	-1.939	38.725	7.087	1.00	63.09	0.138	C
AT013	3400	NE	ARG	A	226	-3.107	38.682	6.212	1.00	70.56	-0.227	N
AT014	3401	HE	ARG	A	226	-3.032	38.087	9.397	1.00	10.00	0.179	H
AT015	3402	CZ	ARG	A	226	-4.259	39.331	6.368	1.00	72.30	0.665	C
AT016	3403	NH1	ARG	A	226	-4.446	40.127	7.413	1.00	74.21	-0.235	N
AT017	3404	NH2	ARG	A	226	-3.697	40.237	8.097	1.00	0.00	0.174	H
AT018	3405	2HH1	ARG	A	226	-5.235	39.189	5.476	1.00	70.97	-0.235	N
AT019	3406	NH2	ARG	A	226	-2.455	39.685	6.596	1.00	0.00	0.174	H
AT020	3407	2HH2	ARG	A	226	-5.091	38.576	4.673	1.00	0.00	0.174	H
AT021	3409	N	ALA	A	227	-1.690	34.984	10.395	1.00	11.19	-0.348	N
AT022	3410	HN	ALA	A	227	-2.695	35.126	10.536	1.00	0.00	-0.169	H
AT023	3411	CA	ALA	A	227	-0.762	34.915	11.538	1.00	13.39	0.172	C
AT024	3413	CA	ALA	A	227	-0.204	33.506	11.676	1.00	9.11	0.240	C
AT025	3414	O	ALA	A	227	-1.548	35.275	12.807	1.00	15.44	0.042	O
AT026	3415	CB	ALA	A	227	-1.041	32.177	11.387	1.00	8.98	-0.180	C
AT027	3419	N	VAL	A	228	-2.455	32.628	11.930	1.00	0.00	0.163	H
AT028	3421	CA	VAL	A	228	-0.462	31.118	11.528	1.00	6.37	0.180	C
AT029	3423	CA	VAL	A	228	0.604	31.471	10.635	1.00	0.00	0.172	C
AT030	3424	O	VAL	A	228	1.659	30.273	10.653	1.00	10.87	-0.271	O
AT031	3425	CB	VAL	A	228	-1.572	30.065	11.462	1.00	12.58	0.009	C
AT032	3427	CG1	VAL	A	228	-1.572	30.065	11.462	1.00	12.58	0.009	C
AT033	3431	CG2	VAL	A	228	-2.518	30.213	12.654	1.00	12.58	0.012	C
AT034	3435	N	GLD	A	229	0.324	31.335	9.211	1.00	9.12	-0.346	N
AT035	3436	HN	GLD	A	229	-0.582	31.898	10.260	1.00	0.00	-0.163	H
AT036	3437	CA	GLD	A	229	1.334	31.202	8.179	1.00	7.77	0.177	C
AT037	3439	C	GLD	A	229	2.648	31.842	8.558	1.00	11.66	0.241	C
AT038	3440	O	GLD	A	229	-1.717	32.292	9.582	1.00	0.00	-0.271	O
AT039	3441	CB	GLD	A	229	0.884	31.915	8.861	1.00	13.73	0.045	C
AT040	3444	CG	GLD	A	229	-0.287	31.059	6.369	1.00	13.76	0.116	C
AT041	3447	CD	GLD	A	229	-0.913	31.470	11.200	1.00	32.43	0.172	C
AT042	3448	OE1	GLD	A	229	-0.724	32.864	4.810	1.00	31.38	-0.648	O
AT043	3449	OE2	GLD	A	229	-1.638	30.916	4.454	1.00	33.36	-0.648	O
AT044	3450	N	GLD	A	230	1.632	33.523	9.260	1.00	0.00	0.163	H
AT045	3451	HN	GLD	A	230	1.370	33.710	9.524	1.00	14.36	0.177	C
AT046	3454	C	GLD	A	230	4.519	32.941	10.627	1.00	16.51	-0.253	C
AT047	3455	O	GLD	A	230	5.763	32.866	10.441	1.00	14.82	-0.271	O
AT048	3456	CB	GLD	A	230	3.466	35.148	9.984	1.00	17.83	0.045	C
AT049	3459	CG	GLD	A	230	4.716	35.202	10.200	1.00	13.31	-0.174	C
AT050	3462	CD	GLD	A	230	4.455	37.254	11.050	1.00	45.25	0.172	C
AT051	3463	OE1	GLD	A	230	3.449	37.920	10.758	1.00	41.66	-0.648	O
AT052	3464	OE2	GLD	A	230	4.229	37.615	10.248	1.00	45.80	-0.271	O
AT053	3465	N	ALA	A	231	3.785	32.441	11.619	1.00	11.60	-0.346	N
AT054	3466	HN	ALA	A	231	2.773	32.569	11.626	1.00	0.00	0.163	H
AT055	3467	CA	ALA	A	231	4.453	31.497	12.712	1.00	62.65	0.172	C
AT056	3469	C	ALA	A	231	5.109	30.946	12.245	1.00	13.24	0.240	C
AT057	3470	O	ALA	A	231	6.203	29.987	12.665	1.00	12.57	-0.271	O
AT058	3471	CB	ALA	A	231	4.359	31.332	13.597	1.00	11.70	0.172	C
AT059	3475	N	ARG	A	232	4.464	29.770	11.226	1.00	11.61	-0.346	N
AT060	3476	HN	ARG	A	232	3.593	30.150	10.854	1.00	0.00	0.163	H
AT061	3477	CA	ARG	A	232	4.009	28.538	11.000	1.00	15.51	0.176	C
AT062	3479	C	ARG	A	232	6.323	28.784	9.958	1.00	15.11	0.241	C
AT063	3480	O	ARG	A	232	7.204	27.918	9.915	1.00	17.10	-0.271	O
AT064	3481	CB	ARG	A	232	2.987	27.806	9.719	1.00	15.48	0.172	C
AT065	3484	CG	ARG	A	232	2.986	27.009	10.598	1.00	13.61	0.023	C
AT066	3487	CD	ARG	A	232	1.777	26.632	9.743	1.00	22.91	0.138	C
AT067	3490	HE	ARG	A	232	4.285	25.816	8.756	1.00	21.97	-0.271	O
AT068	3491	HE	ARG	A	232	2.525	26.269	7.729	1.00	0.00	0.177	H
AT069	3492	CG	ARG	A	232	2.426	24.504	8.737	1.00	38.77	0.685	C
AT070	3493	NH1	ARG	A	232	2.426	23.896	9.266	1.00	0.00	-0.163	H
AT071	3494	NH2	ARG	A	232	2.186	22.881	9.956	1.00	0.00	0.174	H
AT072	3495	2HH1	ARG	A	232	1.713	24.437	10.638	1.00	0.00	0.174	H
AT073	3496	2HH2	ARG	A	232	4.897	23.785	10.459	1.00	40.59	-0.271	O
AT074	3497	1HH2	ARG	A	232	3.005	22.786	7.627	1.00	0.00	0.174	H
AT075	3498	2HH2	ARG	A	232	3.165	24.267	6.867	1.00	0.00	0.174	H
AT076	3499	N	ALA	A	233	6.536	29.967	10.131	1.00	0.00	-0.346	N
AT077	3500	HN	ALA	A	233	5.824	30.688	9.537	1.00	0.00	0.163	H
AT078	3501	CA	ALA	A	233	7.411	29.296	10.897	1.00	0.00	0.172	C
AT079	3503	C	ALA	A	233	8.897	30.717	9.564	1.00	21.01	0.240	C
AT080	3504	O	ALA	A	233	10.004	30.830	9.060	1.00	16.18	-0.271	O
AT081	3505	CB	ALA	A	233	8.643	30.386	10.811	1.00	15.89	0.172	C
AT082	3509	N	VAL	A	234	8.648	30.951	10.851	1.00	15.89	-0.346	N
AT083	3510	HN	VAL	A	234	7.688	30.903	11.193	1.00	0.00	0.163	H
AT084	3511	CA	VAL	A	234	10.182	27.277	10.100	1.00	12.08	0.172	C
AT085	3513	C	VAL	A	234	10.181	29.941	12.343	1.00	17.76	0.241	C
AT086	3514	O	VAL	A	234	9.411	29.297	13.095	1.00	15.72	-0.271	O
AT087	3515	CB	VAL	A	234	9.210	31.218	13.095	1.00	15.72	0.172	C
AT088	3517	CG1	VAL	A	234	10.314	32.497	13.921	1.00	18.42	0.012	C
AT089	3521	CG2	VAL	A	234	8.725	33.539	12.280	1.00	16.05	0.012	C
AT090	3525	N	THR	A	235	11.433	27.443	12.170	1.00	14.81	-0.346	N
AT091	3526	HN	THR	A	235	12.006	29.979	11.358	1.00	0.00	0.163	H
AT092	3527	CA	THR	A	235	11.703	28.097	12.410	1.00	15.57	0.205	C
AT093	3529	C	THR	A	235	12.617	27.840	13.610	1.00	14.40	0.172	C
AT094	3530	O	THR	A	235	12.796	26.869	14.068	1.00	18.75	-0.271	O
AT095	3531	CB	THR	A	235	12.427	27.350	11.233	1.00	22.89	0.146	C
AT096	3533	CG2	THR	A	235	11.433	27.443	12.170	1.00	14.81	-0.346	N
AT097	3537	CG1	THR	A	235	13.559	28.157	10.908	1.00	23.12	-0.393	O
AT098	3538	HG1	THR	A	235	14.182	28.221	11.623	1.00	0.00	0.210	H
AT099	3539	N	ASP	A	236	12.123	29.111	11.104	1.00	12.22	-0.346	N
AT100	3540	HN	ASP	A	236	12.880	30.020	13.710	1.00	0.00	0.163	H
AT101	3541	CA	ASP	A	236	14.045	29.001	15.232	1.00	17.12	0.186	C
AT102	3543	C	ASP	A	236	15.185	31.320	16.536	1.00	16.65	0.241	C
AT103	3544	O	ASP	A	236	14.343	29.747	17.457	1.00	16.69	-0.271	O
AT104	3545	CB	ASP	A	236	15.358	29.683	14.830	1.00	22.44	0.147	C
AT105	3548	CG	ASP	A	236	15.185	31.320	16.536	1.00	16.65	0.241	C
AT106	3549	OD1	ASP	A	236	14.071	31.627	14.155	1.00	25.01	-0.648	O
AT107	3550	OD2	ASP	A	236	16.216	31.838	14.437	1.00	47.16	-0.648	O
AT108	3551	N	LVS	A	237	12.289	30.001	16.649	1.00	14.99	-0.346	N
AT109	3552	HN	LVS	A								

ATOM	3947	CG	PRO	A	264	-25.561	25.797	32.965	1.00	27.87	0.022	C
ATOM	3950	CD	LEU	A	265	-24.477	31.741	29.314	1.00	27.96	0.127	C
ATOM	3953	N	LEU	A	265	-25.834	29.271	29.904	1.00	23.39	-0.346	N
ATOM	3954	HN	LEU	A	265	-25.616	28.584	29.182	1.00	0.00	0.163	HD
ATOM	3955	CA	LEU	A	265	-26.359	30.662	29.090	1.00	24.46	-0.277	CA
ATOM	3957	C	LEU	A	265	-27.877	30.688	29.870	1.00	26.81	0.240	C
ATOM	3958	O	LEU	A	265	-28.321	31.815	30.137	1.00	30.52	-0.271	OA
ATOM	3959	CB	LEU	A	265	-26.280	30.732	29.416	1.00	26.82	-0.177	CB
ATOM	3962	CG	LEU	A	265	-24.821	30.713	27.491	1.00	30.48	-0.020	C
ATOM	3964	CD1	LEU	A	265	-24.793	30.815	25.990	1.00	25.52	0.009	C
ATOM	3968	CD2	LEU	A	265	-23.952	31.816	28.125	1.00	27.96	0.009	C
ATOM	3972	N	GLY	A	266	-28.627	29.591	29.814	1.00	27.05	-0.351	N
ATOM	3973	HN	GLY	A	266	-28.201	28.695	29.578	1.00	0.00	0.163	HD
ATOM	3974	CA	GLY	A	266	-30.072	29.668	30.993	1.00	24.05	0.275	CA
ATOM	3977	C	GLY	A	266	-30.838	29.555	28.768	1.00	27.93	0.236	C
ATOM	3978	O	GLY	A	266	-30.371	30.014	27.727	1.00	25.21	-0.272	OA
ATOM	3979	N	ASP	A	267	-32.380	28.619	29.704	1.00	0.00	0.163	HD
ATOM	3981	CA	ASP	A	267	-32.752	28.677	27.597	1.00	31.83	0.186	C
ATOM	3983	C	ASP	A	267	-32.055	28.942	26.809	1.00	31.58	0.175	C
ATOM	3984	O	ASP	A	267	-32.864	30.021	25.589	1.00	32.31	-0.271	OA
ATOM	3985	CB	ASP	A	267	-34.102	27.953	27.925	1.00	41.24	0.147	C
ATOM	3988	CG	ASP	A	267	-33.889	26.580	28.535	1.00	49.30	0.000	C
ATOM	3989	CD1	ASP	A	267	-33.100	25.766	28.009	1.00	49.09	-0.648	OA
ATOM	3990	CD2	ASP	A	267	-34.529	26.297	29.573	1.00	54.77	-0.648	OA
ATOM	3991	N	ALA	A	268	-33.753	30.844	28.521	1.00	0.00	0.163	HD
ATOM	3993	CA	ALA	A	268	-33.895	32.226	26.949	1.00	31.81	-0.172	CA
ATOM	3995	C	ALA	A	268	-32.683	32.923	26.170	1.00	28.62	0.210	C
ATOM	3996	O	ALA	A	268	-32.734	33.279	25.015	1.00	31.29	-0.271	OA
ATOM	4000	HN	LEU	A	269	-33.947	33.817	29.154	1.00	31.74	0.163	HD
ATOM	4001	N	GLU	A	269	-31.550	32.850	26.883	1.00	26.22	-0.346	N
ATOM	4002	HN	GLU	A	269	-31.539	32.477	27.832	1.00	0.00	0.163	HD
ATOM	4003	CA	GLU	A	269	-32.483	33.817	27.959	1.00	24.46	-0.277	CA
ATOM	4005	C	GLU	A	269	-29.875	32.585	25.089	1.00	25.09	0.241	C
ATOM	4006	O	GLU	A	269	-29.438	33.109	24.058	1.00	24.45	-0.271	OA
ATOM	4007	CB	GLU	A	269	-29.776	34.268	26.840	1.00	27.69	0.116	CB
ATOM	4010	CG	GLU	A	269	-28.263	35.700	26.441	1.00	36.08	0.172	C
ATOM	4013	CD1	GLU	A	269	-29.359	32.656	28.709	1.00	31.17	-0.648	OA
ATOM	4015	OE2	GLU	A	269	-27.394	36.363	25.843	1.00	33.18	-0.648	OA
ATOM	4016	N	ILE	A	270	-29.993	31.266	25.191	1.00	23.57	-0.346	N
ATOM	4017	HN	ILE	A	270	-30.369	30.843	26.103	1.00	22.00	0.163	HD
ATOM	4018	CA	ILE	A	270	-29.578	30.428	24.059	1.00	24.00	0.180	C
ATOM	4020	C	ILE	A	270	-30.429	30.760	22.844	1.00	24.56	0.241	C
ATOM	4021	O	ILE	A	270	-29.496	30.962	27.520	1.00	27.47	-0.346	N
ATOM	4022	CB	ILE	A	270	-29.639	28.938	24.385	1.00	26.50	0.013	C
ATOM	4024	CG1	ILE	A	270	-28.365	28.556	25.169	1.00	22.90	0.002	C
ATOM	4027	CG2	ILE	A	270	-29.758	28.923	24.615	1.00	23.62	0.013	C
ATOM	4031	CD1	ILE	A	270	-28.487	27.212	25.859	1.00	25.01	0.005	C
ATOM	4035	N	ALA	A	271	-31.733	30.980	23.065	1.00	24.71	-0.346	N
ATOM	4036	HN	ALA	A	271	-32.121	30.216	24.016	1.00	27.96	0.163	HD
ATOM	4037	CA	ALA	A	271	-32.585	31.314	23.926	1.00	25.39	-0.172	CA
ATOM	4039	C	ALA	A	271	-32.107	32.619	21.298	1.00	24.20	0.240	C
ATOM	4040	O	ALA	A	271	-32.032	32.644	20.480	1.00	27.14	-0.271	OA
ATOM	4041	CB	ALA	A	271	-34.069	31.383	22.276	1.00	27.17	0.042	C
ATOM	4045	N	LEU	A	272	-31.807	33.655	22.066	1.00	25.07	-0.346	N
ATOM	4046	HN	LEU	A	272	-29.949	33.577	23.424	1.00	24.43	0.163	HD
ATOM	4047	CA	LEU	A	272	-31.284	34.907	21.521	1.00	24.66	-0.177	CA
ATOM	4049	C	LEU	A	272	-29.932	34.794	20.840	1.00	27.16	0.241	C
ATOM	4050	O	LEU	A	272	-29.598	34.564	19.644	1.00	25.91	-0.271	OA
ATOM	4051	CB	LEU	A	272	-31.136	35.943	22.667	1.00	32.34	0.038	C
ATOM	4054	CG	LEU	A	272	-32.475	36.264	23.366	1.00	38.20	-0.020	C
ATOM	4056	CD1	LEU	A	272	-30.267	37.963	24.459	1.00	41.33	0.013	C
ATOM	4060	CD2	LEU	A	272	-33.380	37.018	22.398	1.00	36.38	-0.009	C
ATOM	4064	N	THR	A	273	-29.091	33.864	21.312	1.00	24.67	-0.346	N
ATOM	4065	HN	THR	A	273	-29.385	33.469	22.522	1.00	24.44	0.163	HD
ATOM	4066	CA	THR	A	273	-27.771	33.632	20.740	1.00	21.05	0.205	C
ATOM	4068	C	THR	A	273	-27.918	32.979	19.378	1.00	22.84	0.243	C
ATOM	4069	O	THR	A	273	-26.428	33.366	18.298	1.00	24.71	-0.271	OA
ATOM	4070	CB	THR	A	273	-26.920	32.751	17.676	1.00	26.16	0.146	C
ATOM	4072	CG2	THR	A	273	-25.552	32.430	19.092	1.00	24.32	0.042	C
ATOM	4076	OG1	THR	A	273	-27.639	33.459	23.294	1.00	0.00	0.210	HD
ATOM	4077	OG1	THR	A	273	-27.639	33.459	23.294	1.00	0.00	0.210	HD
ATOM	4078	N	ARG	A	274	-28.870	32.036	19.255	1.00	24.26	-0.346	N
ATOM	4079	HN	ARG	A	274	-28.870	32.036	19.255	1.00	24.26	-0.346	N
ATOM	4080	CA	ARG	A	274	-29.073	31.460	17.930	1.00	27.15	0.176	C
ATOM	4082	C	ARG	A	274	-29.452	32.576	16.949	1.00	29.91	0.241	C
ATOM	4083	OE1	ARG	A	274	-31.020	40.571	17.843	1.00	24.73	-0.271	OA
ATOM	4084	CB	ARG	A	274	-30.127	30.364	17.898	1.00	23.31	0.036	C
ATOM	4087	CG	ARG	A	274	-29.932	30.616	18.298	1.00	27.99	0.042	C
ATOM	4090	CD	ARG	A	274	-30.896	28.049	18.552	1.00	23.49	0.138	C
ATOM	4093	NE	ARG	A	274	-30.397	26.991	18.236	1.00	24.12	-0.227	N
ATOM	4094	HE	ARG	A	274	-29.626	26.344	17.179	1.00	24.96	0.176	C
ATOM	4095	CZ	ARG	A	274	-29.705	25.930	19.075	1.00	26.01	0.665	C
ATOM	4096	NH1	ARG	A	274	-29.471	26.378	20.306	1.00	27.17	-0.235	N
ATOM	4097	NH2	ARG	A	274	-29.777	26.309	20.306	1.00	27.17	-0.235	N
ATOM	4098	ZH1	ARG	A	274	-28.939	25.793	20.951	1.00	0.00	0.174	HD
ATOM	4099	ZH2	ARG	A	274	-29.290	24.719	18.729	1.00	22.08	-0.235	N
ATOM	4100	H2H2	ARG	A	274	-28.758	24.314	19.374	1.00	0.00	0.174	HD
ATOM	4101	H2H2	ARG	A	274	-28.758	24.314	19.374	1.00	0.00	0.174	HD
ATOM	4102	N	GLU	A	275	-30.319	33.484	17.405	1.00	27.70	-0.346	N
ATOM	4103	HN	GLU	A	275	-30.809	34.553	16.356	1.00	31.94	0.177	C
ATOM	4104	CA	GLU	A	275	-29.672	35.478	16.155	1.00	30.08	0.241	C
ATOM	4107	O	GLU	A	275	-29.563	35.883	15.257	1.00	27.73	-0.271	OA
ATOM	4108	CB	GLU	A	275	-31.929	35.329	17.250	1.00	39.95	0.045	C
ATOM	4111	CG	GLU	A	275	-32.325	36.644	16.621	1.00	67.08	0.116	C
ATOM	4114	CD	GLU	A	275	-31.482	37.371	16.876	1.00	31.79	-0.271	OA
ATOM	4115	OE1	GLU	A	275	-33.778	37.180	18.476	1.00	81.75	-0.648	OA
ATOM	4116	OE2	GLU	A	275							

ATCOM 4524	CD	GLN	A	301	-8.099	5.835	4.083	1.00	77.58	0.215	C
ATCOM 4525	HE2	GLN	A	301	-4.735	11.224	4.600	1.00	10.78	-0.176	C
ATCOM 4526	HE2	GLN	A	301	-4.964	4.161	3.686	1.00	0.00	0.159	HD
ATCOM 4527	HE2	GLN	A	301	-8.734	4.060	3.431	1.00	0.00	0.159	HD
ATCOM 4528	CE1	LYS	A	303	-4.228	6.007	4.157	1.00	84.63	-0.221	CA
ATCOM 4529	N	ALA	A	302	-5.143	9.863	3.575	1.00	13.81	-0.346	N
ATCOM 4530	HN	ALA	A	302	-5.991	10.427	3.634	1.00	0.00	0.163	HD
ATCOM 4531	CA	ALA	A	302	-4.179	4.037	3.634	1.00	17.72	-0.240	CA
ATCOM 4533	C	ALA	A	302	-2.833	10.557	3.063	1.00	16.71	0.240	C
ATCOM 4534	O	ALA	A	302	-1.732	10.069	2.640	1.00	15.84	-0.271	OA
ATCOM 4535	CB	ALA	A	302	-4.475	11.224	1.577	1.00	19.09	0.042	C
ATCOM 4539	N	LYS	A	303	-2.732	10.069	4.000	1.00	10.77	-0.346	N
ATCOM 4540	HN	LYS	A	303	-3.703	11.892	4.391	1.00	0.00	0.163	HD
ATCOM 4541	CA	LYS	A	303	-1.548	11.892	4.600	1.00	10.78	0.176	C
ATCOM 4543	C	LYS	A	303	-0.890	10.746	5.411	1.00	9.86	0.241	C
ATCOM 4544	O	LYS	A	303	0.331	10.639	5.365	1.00	13.44	-0.271	OA
ATCOM 4545	CB	LYS	A	303	-1.763	13.827	3.964	1.00	33.95	0.004	C
ATCOM 4548	CG	LYS	A	303	-2.169	14.339	4.863	1.00	25.36	0.004	C
ATCOM 4551	CD	LYS	A	303	-2.544	15.395	5.903	1.00	38.00	0.027	C
ATCOM 4554	CB	LYS	A	303	-4.074	16.327	5.903	1.00	33.96	-0.079	N
ATCOM 4557	NZ	LYS	A	303	-4.074	17.328	6.355	1.00	31.96	-0.079	N
ATCOM 4558	H21	LYS	A	303	-4.841	18.076	5.955	1.00	0.00	0.274	HD
ATCOM 4558	H22	LYS	A	303	-4.454	17.493	7.936	1.00	0.00	-0.274	HD
ATCOM 4560	H23	LYS	A	303	-4.626	16.738	6.978	1.00	0.00	0.274	HD
ATCOM 4561	N	GLD	A	304	-1.655	9.953	6.124	1.00	8.84	-0.346	N
ATCOM 4562	HE2	GLD	A	304	-4.668	10.067	6.353	1.00	0.00	-0.162	HD
ATCOM 4563	CA	GLD	A	304	-0.958	8.864	6.901	1.00	11.72	0.177	C
ATCOM 4565	C	GLD	A	304	-0.433	7.809	5.989	1.00	12.64	0.241	C
ATCOM 4566	O	GLD	A	304	-4.638	7.231	6.130	1.00	15.44	-0.271	OA
ATCOM 4567	CB	GLD	A	304	-1.947	8.232	7.917	1.00	7.07	0.045	C
ATCOM 4570	CG	GLD	A	304	-1.844	8.646	8.758	1.00	7.49	0.116	CA
ATCOM 4573	CD	GLD	A	304	-2.035	6.690	9.941	1.00	16.93	0.172	C
ATCOM 4574	OE1	GLD	A	304	-3.035	7.333	10.300	1.00	21.10	-0.648	OA
ATCOM 4575	OE2	GLD	A	304	-4.848	10.254	10.254	1.00	17.59	-0.648	OA
ATCOM 4576	N	SER	A	305	-1.229	7.515	4.886	1.00	13.63	-0.346	N
ATCOM 4577	HN	SER	A	305	-2.135	7.974	4.788	1.00	0.00	0.163	HD
ATCOM 4578	CA	SER	A	305	-4.852	6.385	4.100	1.00	15.43	-0.176	C
ATCOM 4580	C	SER	A	305	0.527	6.941	3.247	1.00	13.23	0.243	C
ATCOM 4581	O	SER	A	305	1.432	6.110	3.115	1.00	12.77	-0.271	OA
ATCOM 4582	CB	SER	A	305	-2.785	6.385	3.100	1.00	15.43	-0.176	C
ATCOM 4585	OG	SER	A	305	-2.878	5.646	3.226	1.00	34.64	-0.398	OA
ATCOM 4586	HG	SER	A	305	-3.486	5.538	2.504	1.00	0.00	0.209	HD
ATCOM 4587	N	ALA	A	306	-1.852	8.245	6.100	1.00	13.00	-0.346	N
ATCOM 4588	HN	ALA	A	306	-0.052	8.905	3.174	1.00	0.00	0.163	HD
ATCOM 4589	CA	ALA	A	306	1.968	8.723	2.457	1.00	15.41	0.172	C
ATCOM 4591	C	ALA	A	306	-4.784	8.099	4.100	1.00	11.18	-0.176	C
ATCOM 4592	O	ALA	A	306	4.200	8.204	3.160	1.00	12.90	-0.271	OA
ATCOM 4593	CB	ALA	A	306	1.819	10.197	2.065	1.00	14.64	0.042	C
ATCOM 4597	N	TRF	A	307	4.189	9.124	5.061	1.00	7.02	0.163	HD
ATCOM 4598	HN	TRF	A	307	1.868	9.134	5.061	1.00	0.00	0.163	HD
ATCOM 4599	CA	TRF	A	307	3.824	8.612	5.061	1.00	7.62	0.181	C
ATCOM 4601	C	TRF	A	307	4.426	8.065	6.224	1.00	8.27	-0.176	C
ATCOM 4602	O	TRF	A	307	5.362	6.809	6.123	1.00	8.76	-0.271	OA
ATCOM 4603	CB	TRF	A	307	3.325	9.311	7.181	1.00	8.68	0.075	C
ATCOM 4606	CG	TRF	A	307	4.426	8.065	6.224	1.00	8.27	-0.176	C
ATCOM 4607	CD1	TRF	A	307	5.383	9.989	8.477	1.00	11.23	0.096	A
ATCOM 4609	CD2	TRF	A	307	4.683	9.978	9.128	1.00	13.89	-0.002	A
ATCOM 4610	CE2	TRF	A	307	4.808	8.304	9.830	1.00	11.33	0.042	A
ATCOM 4611	CE3	TRF	A	307	4.044	6.757	9.371	1.00	13.90	0.014	A
ATCOM 4613	NE1	TRF	A	307	6.222	9.540	9.497	1.00	11.68	-0.365	N
ATCOM 4614	NE1	TRF	A	307	6.014	9.533	9.600	1.00	11.68	-0.365	N
ATCOM 4615	CE2	TRF	A	307	6.334	7.477	10.881	1.00	14.26	0.030	A
ATCOM 4617	CE3	TRF	A	307	4.574	5.929	10.149	1.00	14.34	0.001	A
ATCOM 4619	CH2	TRF	A	307	6.189	6.283	11.056	1.00	15.06	-0.176	C
ATCOM 4621	N	ASN	A	308	3.183	6.236	5.837	1.00	9.72	-0.346	N
ATCOM 4622	HN	ASN	A	308	2.219	6.555	5.738	1.00	0.00	0.163	HD
ATCOM 4623	CA	ASN	A	308	1.484	6.902	5.834	1.00	12.09	0.004	C
ATCOM 4625	C	ASN	A	308	4.483	4.348	4.876	1.00	11.39	-0.241	C
ATCOM 4626	O	ASN	A	308	5.360	3.539	5.121	1.00	10.85	-0.271	OA
ATCOM 4627	CB	ASN	A	308	-1.175	4.992	5.787	1.00	11.03	0.004	C
ATCOM 4630	CG	ASN	A	308	1.300	4.007	7.026	1.00	14.71	0.217	C
ATCOM 4631	NE2	ASN	A	308	0.013	3.650	6.949	1.00	14.50	-0.370	OA
ATCOM 4632	H2D	ASN	A	308	-0.491	6.931	6.931	1.00	0.00	0.159	HD
ATCOM 4633	H2D	ASN	A	308	-0.576	3.660	7.782	1.00	0.00	0.159	HD
ATCOM 4634	OD1	ASN	A	308	1.826	4.418	8.069	1.00	11.31	-0.274	OA
ATCOM 4635	N	GLD	A	309	-1.256	4.918	4.918	1.00	0.00	-0.163	HD
ATCOM 4636	HN	GLD	A	309	3.506	5.435	3.440	1.00	0.00	0.163	HD
ATCOM 4637	CA	GLD	A	309	5.228	4.442	2.555	1.00	13.95	0.177	C
ATCOM 4639	C	GLD	A	309	6.588	6.039	6.588	1.00	14.15	0.241	C
ATCOM 4640	O	GLD	A	309	7.599	4.404	2.518	1.00	14.00	-0.271	OA
ATCOM 4641	CB	GLD	A	309	6.213	4.027	4.027	1.00	19.96	0.042	C
ATCOM 4644	CG	GLD	A	309	3.415	4.456	4.749	1.00	27.54	0.116	CA
ATCOM 4647	CD	GLD	A	309	2.977	5.010	-0.607	1.00	42.19	0.172	C
ATCOM 4648	OE1	GLD	A	309	-2.216	4.290	-1.289	1.00	50.85	-0.648	OA
ATCOM 4649	OE2	GLD	A	309	2.216	4.290	-1.289	1.00	50.85	-0.648	OA
ATCOM 4650	N	LYS	A	310	6.697	6.238	3.357	1.00	13.98	-0.346	N
ATCOM 4651	HN	LYS	A	310	6.597	6.238	3.357	1.00	13.98	-0.346	N
ATCOM 4652	CA	LYS	A	310	7.966	6.809	3.728	1.00	11.98	0.176	C
ATCOM 4654	C	LYS	A	310	8.692	6.001	4.803	1.00	14.42	0.241	C
ATCOM 4655	O	LYS	A	310	7.815	8.209	4.354	1.00	20.86	0.035	C
ATCOM 4656	CB	LYS	A	310	7.815	8.209	4.354	1.00	20.86	0.035	C
ATCOM 4659	CG	LYS	A	310	7.689	9.335	3.355	1.00	32.44	0.004	C
ATCOM 4662	CD	LYS	A	310	7.244	9.335	3.355	1.00	32.44	0.004	C
ATCOM 4665	CE	LYS	A	310	8.244	11.100	5.071	1.00	44.40	0.229	C
ATCOM 4668	NZ	LYS	A	310	8.234	12.597	5.138	1.00	59.95	-0.079	N
ATCOM 4669	H21	LYS	A	310	7.299	12.597	5.138	1.00	59.95	-0.079	N
ATCOM 4670	H22	LYS	A	310	7.299	12.597	5.138	1.00	59.95	-0.079	N
ATCOM 4671	H23	LYS	A	310	8.395	13.022	4.225	1.00	0.00	0.274	HD
ATCOM 4672	N	PHE	A	311	6.896	4.173	5.750	1.00	81.83	-0.163	HD
ATCOM 4673	HN	PHE	A	311	6.884	5.601	5.721	1.00	0.00	0.163	HD
ATCOM 4674	CA	PHE	A	311	8.532	4.710	6.822	1.00	11.77	0.180	C
ATCOM 4676	C	PHE	A	311	4.057	4.008	6.247	1.00	9.68	-0.176	C
ATCOM 4677	O	PHE	A	311	10.114	2.891	6.605	1.00	10.93	-0.271	OA
ATCOM 4678	CB	PHE	A	311	7.475	4.459	7.931	1.00	8.50	0.073	C
ATCOM 4681	CG	PHE	A	311	8.165	3.715	9.328	1.00	14.89	-0.163	HD
ATCOM 4682	CD1	PHE	A	311	8.975	4.382	9.971	1.00	14.09	0.007	A
ATCOM 4684	CD2	PHE	A	311	8.036	2.329	9.162	1.00	16.81	0.007	A
ATCOM 4686	CE1	PHE	A	311	9.433	3.701	10.982	1.00	14.89	-0.163	HD
ATCOM 4688	CE2	PHE	A	311	8.708	1.650	11.663	1.00	18.26	0.001	A
ATCOM 4690	CE3	PHE	A	311	9.495	2.331	11.075	1.00	18.11	0.000	A
ATCOM 4692	N	ALA	A	312	-3.309	2.802	5.398	1.00	19.97	-0.346	N
ATCOM 4693	HN	ALA	A	312	7.419	3.206	5.034	1.00	0.00	0.163	HD
ATCOM 4694	CA	ALA	A	312	8.800	1.538	4.742	1.00	13.32	0.172	C
ATCOM 4696	C	ALA	A	312	10.089	4.770	4.973	1.00	13.62	-0.271	OA
ATCOM 4697	O	ALA	A	312	11.034	0.973	4.043	1.00	13.62	-0.271	OA
ATCOM 4698	CB	ALA	A	312	7.724	0.925	3.882	1.00	13.48	0.042	C
ATCOM 4702	N</										

ATOM 5075 HN PHE A 337	16.568	-6.453	27.417	1.00	0.00	0.163	HD
ATOM 5076 CA PHE A 337	17.280	-5.769	29.290	1.00	14.63	0.163	HD
ATOM 5078 C PHE A 337	17.202	-6.750	30.592	1.00	18.92	0.241	C
ATOM 5079 O PHE A 337	17.986	-6.384	31.538	1.00	20.80	-0.271	OA
ATOM 5080 CB ASP A 338	16.365	-4.327	29.273	1.00	0.00	0.163	HD
ATOM 5083 CG PHE A 337	16.378	-3.791	30.438	1.00	15.89	-0.056	A
ATOM 5084 CD1 PHE A 337	17.340	-2.831	30.651	1.00	20.28	0.007	A
ATOM 5086 CD2 PHE A 337	15.425	-4.040	31.351	1.00	19.29	0.007	A
ATOM 5088 CE1 PHE A 337	17.347	-2.104	31.832	1.00	17.77	0.001	A
ATOM 5090 CE2 PHE A 337	15.426	-3.312	32.600	1.00	18.89	0.001	A
ATOM 5092 CZ PHE A 337	16.387	-2.339	32.795	1.00	20.67	0.000	A
ATOM 5094 N ASP A 338	16.177	-7.393	30.671	1.00	15.88	-0.345	N
ATOM 5095 HN ASP A 338	15.596	-7.570	29.852	1.00	0.00	0.163	HD
ATOM 5096 CA ASP A 338	15.983	-8.068	31.943	1.00	21.81	0.186	C
ATOM 5098 C ASP A 338	17.050	-8.902	32.428	1.00	21.98	0.241	C
ATOM 5099 O ASP A 338	17.480	-8.829	33.577	1.00	18.93	-0.271	OA
ATOM 5100 CB ASP A 338	16.445	-10.332	32.966	1.00	21.20	0.163	HD
ATOM 5103 CG ASP A 338	14.281	-9.711	33.026	1.00	32.40	0.175	C
ATOM 5104 OD1 ASP A 338	13.788	-9.193	34.043	1.00	27.39	-0.648	OA
ATOM 5105 OD2 ASP A 338	14.545	-10.332	32.966	1.00	21.20	0.163	HD
ATOM 5106 N ALA A 339	17.574	-9.714	31.525	1.00	22.62	-0.346	N
ATOM 5107 HN ALA A 339	17.159	-9.743	30.594	1.00	0.00	0.163	HD
ATOM 5108 CA ALA A 339	18.717	-10.566	31.906	1.00	46.00	0.152	C
ATOM 5110 C ALA A 339	19.938	-9.746	32.197	1.00	23.56	0.240	C
ATOM 5111 O ALA A 339	20.519	-10.034	33.234	1.00	21.22	-0.271	OA
ATOM 5112 CB ALA A 339	19.034	-11.407	30.768	1.00	23.65	0.092	C
ATOM 5116 N LYS A 340	20.247	-8.660	31.486	1.00	16.76	-0.346	N
ATOM 5117 HN LYS A 340	19.665	-8.390	30.693	1.00	0.00	0.163	HD
ATOM 5118 CA LYS A 340	21.404	-7.486	31.101	1.00	15.82	0.163	C
ATOM 5120 C LYS A 340	21.228	-7.139	33.172	1.00	16.68	0.241	C
ATOM 5121 O LYS A 340	22.387	-6.922	33.937	1.00	17.47	-0.271	OA
ATOM 5122 CB LYS A 340	21.676	-6.862	30.729	1.00	19.28	0.035	C
ATOM 5125 CG LYS A 340	22.359	-7.553	29.542	1.00	33.54	0.004	C
ATOM 5128 CD LYS A 340	24.340	-8.499	30.261	1.00	49.26	0.027	C
ATOM 5131 CE LYS A 340	23.658	-6.922	27.497	1.00	56.92	0.229	C
ATOM 5134 NZ LYS A 340	23.532	-8.272	26.889	1.00	56.32	-0.079	N
ATOM 5135 HZ1 LYS A 340	22.658	-8.272	26.889	1.00	56.32	-0.079	N
ATOM 5136 HZ2 LYS A 340	22.658	-8.272	26.889	1.00	56.32	-0.079	N
ATOM 5137 HZ3 LYS A 340	23.376	-8.989	27.597	1.00	0.00	0.274	HD
ATOM 5138 N ALA A 341	20.680	-6.838	33.309	1.00	16.12	-0.346	N
ATOM 5139 HN ALA A 341	19.244	-6.981	32.756	1.00	0.00	0.163	HD
ATOM 5140 CA ALA A 341	19.755	-5.978	34.695	1.00	16.66	0.172	C
ATOM 5142 C ALA A 341	19.826	-6.835	35.824	1.00	16.69	0.241	C
ATOM 5143 O ALA A 341	20.387	-6.646	36.877	1.00	17.11	-0.271	OA
ATOM 5144 CB ALA A 341	18.400	-5.269	34.665	1.00	15.86	0.042	C
ATOM 5148 N LYS A 342	24.344	-8.215	31.461	1.00	21.59	-0.346	N
ATOM 5149 HN LYS A 342	18.905	-8.451	34.704	1.00	0.00	0.163	HD
ATOM 5150 CA LYS A 342	19.462	-9.210	36.676	1.00	21.24	0.176	C
ATOM 5152 C LYS A 342	20.852	-8.451	36.676	1.00	21.24	0.241	C
ATOM 5153 O LYS A 342	21.260	-9.783	38.186	1.00	17.73	-0.271	OA
ATOM 5154 CB LYS A 342	18.678	-10.499	36.370	1.00	24.04	0.035	C
ATOM 5157 CG LYS A 342	19.183	-10.240	37.300	1.00	24.79	0.092	C
ATOM 5160 CD LYS A 342	16.380	-11.504	36.100	1.00	44.46	0.027	C
ATOM 5163 CE LYS A 342	14.889	-11.127	36.330	1.00	57.43	0.229	C
ATOM 5166 NZ LYS A 342	14.029	-11.989	37.340	1.00	57.39	-0.079	N
ATOM 5167 HZ1 LYS A 342	13.097	-11.798	35.457	1.00	0.00	0.274	HD
ATOM 5168 HZ2 LYS A 342	14.296	-12.988	35.367	1.00	0.00	0.274	HD
ATOM 5169 HZ3 LYS A 342	14.339	-11.809	35.009	1.00	0.00	0.274	HD
ATOM 5170 N GLU A 343	21.727	-9.670	35.994	1.00	18.07	-0.346	N
ATOM 5171 HN GLU A 343	21.363	-9.548	35.049	1.00	0.00	0.163	HD
ATOM 5172 CA GLU A 343	23.155	-8.346	36.169	1.00	21.59	0.163	C
ATOM 5174 C GLU A 343	23.831	-8.809	36.934	1.00	19.88	0.241	C
ATOM 5175 O GLU A 343	24.678	-9.036	37.809	1.00	16.78	-0.271	OA
ATOM 5176 CB GLU A 343	23.713	-10.455	36.100	1.00	21.59	0.092	C
ATOM 5179 CG GLU A 343	23.965	-10.846	34.434	1.00	51.50	0.116	C
ATOM 5182 CD GLU A 343	25.187	-10.793	32.914	1.00	56.71	0.172	C
ATOM 5183 CE1 GLU A 343	24.577	-11.189	32.914	1.00	56.71	0.172	C
ATOM 5184 CE2 GLU A 343	25.930	-9.888	32.478	1.00	55.86	-0.648	OA
ATOM 5185 N PHE A 344	23.504	-7.551	36.633	1.00	14.77	-0.346	N
ATOM 5186 HN PHE A 344	22.833	-7.419	35.939	1.00	0.00	0.163	HD
ATOM 5187 CA PHE A 344	24.082	-6.413	37.355	1.00	14.92	0.180	C
ATOM 5189 C PHE A 344	23.669	-6.437	38.839	1.00	14.38	0.241	C
ATOM 5190 O PHE A 344	24.871	-6.187	39.892	1.00	14.38	-0.271	OA
ATOM 5191 CB PHE A 344	23.563	-5.125	36.690	1.00	17.25	0.073	C
ATOM 5194 CD PHE A 344	23.915	-3.830	37.359	1.00	18.97	-0.056	A
ATOM 5199 CE1 PHE A 344	25.522	-2.219	38.058	1.00	19.33	0.007	A
ATOM 5201 CE2 PHE A 344	23.240	-4.826	36.461	1.00	24.01	0.001	A
ATOM 5203 CZ PHE A 344	24.555	-1.417	38.586	1.00	14.39	0.000	A
ATOM 5205 N LYS A 345	22.334	-6.882	36.100	1.00	19.28	0.035	C
ATOM 5206 HN LYS A 345	17.745	-6.802	38.323	1.00	0.00	0.163	HD
ATOM 5207 CA LYS A 345	21.884	-6.779	40.480	1.00	17.97	0.180	C
ATOM 5209 C LYS A 345	22.709	-7.848	41.240	1.00	17.24	0.241	C
ATOM 5210 O LYS A 345	23.007	-7.449	42.397	1.00	14.60	-0.271	OA
ATOM 5211 CB LYS A 345	20.363	-6.997	40.470	1.00	14.50	0.013	C
ATOM 5213 CG1 LYS A 345	21.509	-6.077	40.077	1.00	15.45	0.092	C
ATOM 5216 CG2 LYS A 345	19.775	-7.393	41.848	1.00	13.76	0.012	C
ATOM 5220 CD1 LYS A 345	18.278	-5.935	39.502	1.00	14.52	0.005	C
ATOM 5224 N ALA A 346	22.328	-9.172	39.174	1.00	14.67	-0.346	N
ATOM 5225 HN ALA A 346	22.328	-9.226	39.174	1.00	0.00	0.163	HD
ATOM 5226 CA ALA A 346	23.403	-10.172	41.361	1.00	17.88	0.172	C
ATOM 5228 C ALA A 346	23.814	-9.531	40.100	1.00	17.01	0.241	C
ATOM 5229 O ALA A 346	25.404	-10.188	42.689	1.00	16.35	-0.271	OA
ATOM 5230 CB ALA A 346	23.281	-11.474	40.579	1.00	19.21	0.042	C
ATOM 5234 N LYS A 347	25.002	-9.172	42.814	1.00	16.04	-0.346	N
ATOM 5235 HN LYS A 347	25.023	-8.948	39.806	1.00	0.00	0.163	HD
ATOM 5236 CA LYS A 347	26.888	-8.738	40.860	1.00	15.04	0.176	C
ATOM 5238 C LYS A 347	27.005	-7.782	42.512	1.00	15.52	0.241	C
ATOM 5239 O LYS A 347	27.968	-7.841	42.814	1.00	16.04	-0.271	OA
ATOM 5240 CB LYS A 347	27.419	-8.058	39.590	1.00	21.09	0.035	C
ATOM 5243 CG LYS A 347	28.850	-7.393	40.480	1.00	24.44	0.092	C
ATOM 5246 CD LYS A 347	29.140	-6.329	38.803	1.00	47.67	0.027	C
ATOM 5249 CE LYS A 347	28.522	-5.101	39.376	1.00	63.06	0.229	C
ATOM 5252 NZ LYS A 347	29.189	-4.833	38.690	1.00	61.22	-0.079	N
ATOM 5253 HZ1 LYS A 347	28.773	-2.985	39.352	1.00	0.00	0.274	HD
ATOM 5254 HZ2 LYS A 347	29.236	-3.771	37.949	1.00	0.00	0.274	HD
ATOM 5255 HZ3 LYS A 347	30.184	-3.869	39.187	1.00	0.00	0.274	HD
ATOM 5256 N LYS A 348	26.066	-6.845	42.184	1.00	12.99	-0.346	N
ATOM 5257 HN LYS A 348	25.317	-6.752	41.498	1.00	0.00	0.163	HD
ATOM 5258 CA LYS A 348	26.156	-5.843	43.765	1.00	11.11	0.176	C
ATOM 5260 C LYS A 348	25.946	-6.713	44.656	1.00	8.66	0.241	C
ATOM 5261 O LYS A 348	26.606	-6.386	45.650	1.00	13.45	-0.271	OA
ATOM 5262 CB LYS A 348	25.000	-4.960	43.765	1.00	21.80	0.092	C
ATOM 5265 CG LYS A 348	25.083	-3.913	42.063	1.00	12.89	-0.020	C
ATOM 5267 CD1 LYS A 348	23.976	-2.860	42.237	1.00	17.46	0.009	C
ATOM 5271 CD2 LYS A 348	26.433	-2.219	43.862	1.00	0.00	0.163	HD
ATOM 5275 N GLN A 349	25.065	-7.693	44.678	1.00	9.99	-0.346	N
ATOM 5276 HN GLN A 349	24.538	-7.945	43.862	1.00	0.00	0.163	HD
ATOM 5277 CA GLN A 349	24.860	-6.420	45.988	1.00	14.67	0.176	C
ATOM 5279 C GLN A 349	26.114	-9.206	46.337	1.00	19.41	0.241	C
ATOM 5280 O GLN A 349	26.452	-9.396	47.523	1.00	14.54	-0.271	OA
ATOM 5281 CB GLN A 349	23.668	-8.201	45.988	1.00	14.64	0.042	C
ATOM 5284 CG GLN A 349	23.230	-10.101	47.086	1.00	12.91	0.105	C
ATOM 5287 CD GLN A 349	22.923	-11.584	48.261	1.00	21.62	0.015	C
ATOM 5288 CE1 GLN A 349	23.713	-9.115	49.100	1.00	14.17	-0.370	N
ATOM 5289 CE2 GLN A 349	23.516	-8.479	50.101	1.00	0.00	0.159	HD
ATOM 5290 CE3 GLN A 349	24.812	-9.840	49.400	1.00	20.80	0.092	C
ATOM 5291 CE4 GLN A 349	21.935	-8.455	48.107	1.00	14.54	-0.274	OA
ATOM 5292 N ALA A 350	26.836	-6.983	45.327	1.00	16.34	-0.346	N
ATOM 5293 HN ALA A 350	26.512	-6.938	46.371	1.00	0.00	0.163	HD
ATOM 5294 CA ALA A 350	28.089	-10.415	45.588	1.00	18.16	0.172	C
ATOM 5296 C ALA A 350	29.264	-9.521	45.883	1.00	22.32	0.240	C
ATOM 5297 O ALA A 350	30.276	-10.024	46.399	1.00	22.56	-0.271	OA
ATOM 5298 CB ALA A 350	28.415	-11.253	44.304	1.00	15.80	0.042	C
ATOM 5302 N ASN A 351	29.280	-8.211	45.634	1.00	16.52	-0.346	N
ATOM 5303 HN ASN A 351	28.423	-7.804	45.245	1.00	0.00	0.163	HD
ATOM 5304 CA ASN A 351	30.390	-7.312	45.864	1.00	18.70	0.185	C

ATOM 5662	N	PHE	A	375	16.351	6.886	27.060	1.00	9.08	-0.346	N
ATOM 5663	N	ARG	A	375	16.469	2.949	26.976	1.00	8.20	0.279	DA
ATOM 5664	CA	PHE	A	375	16.408	7.843	28.191	1.00	8.01	0.180	C
ATOM 5666	C	PHE	A	375	16.460	9.298	27.751	1.00	11.26	0.241	C
ATOM 5667	O	PHE	A	375	17.137	9.955	26.770	1.00	10.71	-0.271	OA
ATOM 5668	CB	PHE	A	375	17.736	7.613	28.841	1.00	11.04	0.073	C
ATOM 5671	CG	PHE	A	375	17.697	6.665	30.094	1.00	16.40	-0.056	A
ATOM 5672	CD1	PHE	A	375	16.545	6.364	30.170	1.00	15.85	0.007	A
ATOM 5674	CD2	PHE	A	375	18.927	6.105	30.509	1.00	19.28	0.007	A
ATOM 5676	CE1	PHE	A	375	16.574	5.489	31.860	1.00	22.81	0.001	A
ATOM 5678	CE2	PHE	A	375	18.954	5.248	31.604	1.00	19.37	0.001	A
ATOM 5680	CZ	PHE	A	375	17.773	4.921	32.246	1.00	16.69	0.000	A
ATOM 5682	N	LEU	A	376	16.650	10.139	28.423	1.00	7.94	-0.346	N
ATOM 5683	HN	LEU	A	376	14.963	9.777	29.084	1.00	0.00	0.163	HD
ATOM 5684	CA	LEU	A	376	16.770	11.592	28.190	1.00	10.88	0.177	C
ATOM 5686	C	LEU	A	376	15.727	12.170	29.611	1.00	13.32	0.240	C
ATOM 5687	O	LEU	A	376	16.653	12.956	29.616	1.00	15.86	-0.249	OA
ATOM 5688	CB	LEU	A	376	14.652	12.092	27.276	1.00	11.91	0.038	C
ATOM 5691	CG	LEU	A	376	14.686	13.570	26.890	1.00	18.47	-0.020	C
ATOM 5693	CD1	LEU	A	376	13.716	13.847	25.735	1.00	13.52	0.009	C
ATOM 5697	CD2	LEU	A	376	16.860	10.312	30.294	1.00	10.18	-0.351	M
ATOM 5702	HN	LEU	A	377	17.753	12.182	29.829	1.00	16.00	0.163	HD
ATOM 5703	CA	LEU	A	377	16.843	12.650	31.706	1.00	11.95	0.225	C
ATOM 5706	C	LEU	A	377	17.254	14.050	32.032	1.00	10.06	0.235	C
ATOM 5707	O	LEU	A	377	14.620	14.868	31.358	1.00	12.52	-0.292	OA
ATOM 5708	N	LEU	A	378	17.171	14.443	33.323	1.00	9.75	-0.351	N
ATOM 5709	HN	LEU	A	378	16.853	13.814	34.081	1.00	0.00	0.163	HD
ATOM 5710	CA	LEU	A	378	17.465	15.029	33.978	1.00	10.03	0.245	C
ATOM 5713	C	LEU	A	378	17.406	16.085	35.095	1.00	11.81	0.236	C
ATOM 5714	O	LEU	A	378	18.943	15.241	35.163	1.00	12.53	-0.272	OA
ATOM 5715	N	LEU	A	379	17.311	17.237	35.492	1.00	8.19	-0.344	N
ATOM 5716	HN	LEU	A	379	18.350	17.871	34.825	1.00	0.00	0.163	HD
ATOM 5717	CA	LEU	A	379	16.585	18.244	35.124	1.00	6.20	0.249	C
ATOM 5719	C	LEU	A	379	17.536	19.099	36.910	1.00	7.95	0.243	C
ATOM 5720	O	LEU	A	379	18.004	19.877	36.014	1.00	8.76	-0.271	OA
ATOM 5721	CB	LEU	A	379	19.171	18.358	36.014	1.00	11.39	0.163	HD
ATOM 5724	CG	LEU	A	379	19.162	18.014	38.890	1.00	13.92	-0.398	OA
ATOM 5725	HN	LEU	A	379	19.996	17.873	39.323	1.00	0.00	0.209	HD
ATOM 5726	N	LEU	A	380	16.865	19.241	39.323	1.00	6.24	-0.346	N
ATOM 5727	HN	ALA	A	380	16.586	18.884	38.694	1.00	0.00	0.163	HD
ATOM 5728	CA	ALA	A	380	16.592	20.953	38.136	1.00	7.22	0.172	C
ATOM 5730	C	ALA	A	380	17.762	21.484	38.136	1.00	11.92	0.241	C
ATOM 5731	O	ALA	A	380	17.625	22.120	39.958	1.00	10.04	-0.271	OA
ATOM 5732	CB	ALA	A	380	16.075	21.194	38.882	1.00	4.08	0.042	C
ATOM 5736	N	ASP	A	381	18.920	21.149	39.630	1.00	6.59	-0.346	N
ATOM 5737	HN	ASP	A	381	18.791	21.495	37.088	1.00	0.00	0.163	HD
ATOM 5738	CA	ASP	A	381	20.013	22.603	38.435	1.00	6.53	0.186	C
ATOM 5740	C	ASP	A	381	20.700	23.939	39.630	1.00	12.52	0.240	C
ATOM 5741	O	ASP	A	381	21.352	22.626	40.414	1.00	14.79	-0.271	OA
ATOM 5742	CB	ASP	A	381	19.744	20.474	38.801	1.00	13.72	0.147	C
ATOM 5745	CG	ASP	A	381	20.029	24.439	38.755	1.00	20.88	-0.346	N
ATOM 5746	OD1	ASP	A	381	21.861	24.804	37.833	1.00	16.89	-0.648	OA
ATOM 5747	OD2	ASP	A	381	21.266	25.707	39.696	1.00	15.35	-0.648	OA
ATOM 5748	N	LEU	A	382	20.611	20.608	39.696	1.00	11.11	0.163	HD
ATOM 5749	HN	LEU	A	382	20.053	20.077	39.106	1.00	0.00	0.163	HD
ATOM 5750	CA	LEU	A	382	21.278	19.896	40.448	1.00	13.47	0.177	C
ATOM 5752	C	LEU	A	382	22.015	18.671	40.448	1.00	15.61	-0.241	C
ATOM 5753	O	LEU	A	382	22.172	17.473	41.034	1.00	14.40	-0.271	OA
ATOM 5754	CB	LEU	A	382	20.280	19.447	41.932	1.00	10.71	0.038	C
ATOM 5757	CG	LEU	A	382	19.747	20.639	42.142	1.00	11.82	-0.020	C
ATOM 5759	CD1	LEU	A	382	18.539	20.207	43.575	1.00	11.95	0.009	C
ATOM 5763	CD2	LEU	A	382	20.894	21.132	43.671	1.00	22.22	0.009	C
ATOM 5767	N	LEU	A	383	22.528	18.702	43.671	1.00	16.46	-0.346	N
ATOM 5768	HN	ALA	A	383	22.424	19.549	38.522	1.00	0.00	0.163	HD
ATOM 5769	CA	ALA	A	383	23.226	17.542	38.510	1.00	18.29	0.172	C
ATOM 5771	C	ALA	A	383	24.363	16.983	39.429	1.00	15.61	0.241	C
ATOM 5772	O	ALA	A	383	24.451	15.743	39.510	1.00	17.26	-0.271	OA
ATOM 5773	CB	ALA	A	383	23.812	17.849	37.110	1.00	10.31	0.042	C
ATOM 5777	N	PRO	A	384	21.871	21.871	39.009	1.00	11.11	-0.346	N
ATOM 5778	CA	PRO	A	384	26.349	17.335	40.734	1.00	20.92	0.179	C
ATOM 5780	C	PRO	A	384	25.946	16.838	42.123	1.00	19.91	0.241	C
ATOM 5781	O	PRO	A	384	27.029	16.133	42.123	1.00	21.47	-0.271	OA
ATOM 5782	CB	PRO	A	384	27.308	18.510	40.876	1.00	22.18	0.037	C
ATOM 5785	CG	PRO	A	384	26.486	19.748	40.567	1.00	26.53	0.022	C
ATOM 5788	CD	PRO	A	384	25.317	19.267	39.918	1.00	27.79	-0.020	C
ATOM 5791	N	SER	A	385	24.738	17.176	42.575	1.00	16.28	-0.344	N
ATOM 5792	HN	SER	A	385	24.135	17.791	42.017	1.00	0.00	0.163	HD
ATOM 5793	CA	SER	A	385	24.273	16.678	43.267	1.00	17.82	0.241	C
ATOM 5795	C	SER	A	385	23.313	15.504	43.707	1.00	19.50	0.243	C
ATOM 5796	O	SER	A	385	25.021	15.848	43.707	1.00	21.72	-0.271	OA
ATOM 5797	CB	SER	A	385	23.483	17.752	44.647	1.00	18.04	0.199	C
ATOM 5800	CG	SER	A	385	24.353	18.856	44.818	1.00	29.62	-0.398	OA
ATOM 5801	HD	SER	A	385	22.424	18.702	42.689	1.00	0.00	0.163	HD
ATOM 5802	N	ASN	A	386	22.442	15.614	42.689	1.00	15.34	-0.345	N
ATOM 5803	HN	ASN	A	386	22.426	16.451	42.106	1.00	0.00	0.163	HD
ATOM 5804	CA	ASN	A	386	21.513	14.509	42.423	1.00	15.91	0.241	C
ATOM 5806	C	ASN	A	386	22.208	13.337	41.711	1.00	14.22	0.241	C
ATOM 5807	O	ASN	A	386	21.677	12.212	41.790	1.00	15.20	-0.271	OA
ATOM 5808	CB	ASN	A	386	20.308	14.683	42.148	1.00	11.82	0.037	C
ATOM 5811	CG	ASN	A	386	19.357	15.884	42.148	1.00	17.30	0.217	C
ATOM 5812	HD2	ASN	A	386	19.178	15.981	43.464	1.00	12.08	-0.370	N
ATOM 5813	HD1	ASN	A	386	20.924	16.862	43.464	1.00	15.93	0.037	C
ATOM 5814	2HD2	ASN	A	386	19.647	15.363	44.126	1.00	0.00	0.159	HD
ATOM 5815	OD1	ASN	A	386	18.790	16.631	41.348	1.00	13.11	-0.274	OA
ATOM 5816	N	LEU	A	387	23.601	15.945	43.921	1.00	10.48	-0.346	N
ATOM 5817	HN	LEU	A	387	23.061	14.533	40.921	1.00	0.00	0.163	HD
ATOM 5818	CA	LEU	A	387	24.086	12.544	40.318	1.00	11.84	0.177	C
ATOM 5820	C	LEU	A	387	23.104	11.884	39.171	1.00	15.40	0.241	C
ATOM 5821	O	LEU	A	387	23.193	10.958	39.112	1.00	14.76	-0.271	OA
ATOM 5822	CB	LEU	A	387	24.637	11.471	41.279	1.00	11.94	0.038	C
ATOM 5825	CG	LEU	A	387	25.418	12.040	42.475	1.00	16.00	-0.020	C
ATOM 5827	CD1	LEU	A	387	25.847	10.962	43.465	1.00	17.23	0.009	C
ATOM 5831	CD2	LEU	A	387	26.668	12.791	42.016	1.00	19.63	0.009	C
ATOM 5835	N	THR	A	388	22.787	13.131	38.306	1.00	14.44	-0.346	N
ATOM 5836	HN	THR	A	388	22.976	13.726	38.427	1.00	0.00	0.163	HD
ATOM 5837	CA	THR	A	388	21.938	12.328	37.178	1.00	11.56	0.205	C
ATOM 5839	C	THR	A	388	22.702	12.225	38.486	1.00	14.42	0.001	C
ATOM 5840	O	THR	A	388	22.111	11.753	34.896	1.00	14.82	-0.271	OA
ATOM 5841	CB	THR	A	388	20.840	13.366	36.949	1.00	16.09	0.146	C
ATOM 5843	CG	THR	A	388	21.970	13.139	37.439	1.00	18.31	-0.020	C
ATOM 5847	OD1	THR	A	388	21.386	14.688	37.173	1.00	17.62	-0.393	OA
ATOM 5848	OD2	THR	A	388	20.703	15.333	37.031	1.00	0.00	0.210	HD
ATOM 5849	N	LEU	A	389	23.859	12.652	35.829	1.00	14.33	-0.346	N
ATOM 5850	HN	LEU	A	389	24.379	13.111	36.637	1.00	0.00	0.163	HD
ATOM 5851	CA	LEU	A	389	24.724	12.483	34.571	1.00	16.13	0.	

ATOM 6222 N MET A 414	9.710	21.390	35.047	1.00	4.76	-0.346	N
ATOM 6223 HN MET A 414	9.98	21.95	35.48	1.00	4.76	-0.163	HD
ATOM 6224 CA MET A 414	10.500	20.217	34.691	1.00	6.80	0.177	C
ATOM 6226 C MET A 414	9.609	18.967	34.709	1.00	8.43	0.241	C
ATOM 6227 O MET A 414	8.787	19.546	34.491	1.00	0.00	-0.173	OA
ATOM 6228 CB MET A 414	11.640	19.988	35.728	1.00	7.87	0.045	C
ATOM 6231 CG MET A 414	12.270	18.599	35.646	1.00	9.94	0.076	C
ATOM 6234 SD MET A 414	13.420	18.599	35.646	1.00	9.94	-0.076	C
ATOM 6235 CE MET A 414	12.726	18.170	38.356	1.00	11.22	0.089	C
ATOM 6239 N THR A 415	8.818	18.811	35.775	1.00	6.06	-0.344	N
ATOM 6240 HN THR A 415	8.787	19.546	34.491	1.00	0.00	0.163	HD
ATOM 6241 CA THR A 415	7.993	17.611	35.957	1.00	6.04	0.205	C
ATOM 6243 C THR A 415	6.836	17.542	34.977	1.00	7.40	0.243	C
ATOM 6244 O THR A 415	5.556	16.468	34.998	1.00	6.82	-0.271	OA
ATOM 6245 CB THR A 415	7.499	17.511	37.423	1.00	6.82	0.146	C
ATOM 6247 CO2 THR A 415	6.945	16.108	37.710	1.00	5.85	0.042	C
ATOM 6251 OG1 THR A 415	6.939	16.108	37.710	1.00	5.85	-0.205	OA
ATOM 6252 HG1 THR A 415	8.986	18.430	38.111	1.00	0.00	0.210	HD
ATOM 6253 N ALA A 416	6.159	18.658	34.690	1.00	5.04	-0.346	N
ATOM 6254 HN ALA A 416	6.339	18.658	34.690	1.00	5.04	0.163	HD
ATOM 6255 CA ALA A 416	5.082	18.666	33.695	1.00	7.97	0.172	C
ATOM 6257 C ALA A 416	5.614	18.658	32.262	1.00	6.70	0.240	C
ATOM 6258 O ALA A 416	4.832	18.108	31.996	1.00	6.41	-0.271	OA
ATOM 6259 CB ALA A 416	4.264	19.939	33.967	1.00	6.89	0.042	C
ATOM 6263 N LEU A 417	6.842	19.133	32.016	1.00	5.74	-0.346	N
ATOM 6264 HN LEU A 417	7.464	18.941	30.693	1.00	4.66	0.180	C
ATOM 6267 C LEU A 417	7.725	17.457	30.500	1.00	7.34	0.241	C
ATOM 6268 O LEU A 417	7.442	16.896	30.421	1.00	7.55	-0.271	OA
ATOM 6269 CB LEU A 417	8.588	19.760	30.549	1.00	6.77	0.013	C
ATOM 6271 CG1 LEU A 417	8.412	18.567	30.549	1.00	6.20	0.013	C
ATOM 6274 CO2 LEU A 417	9.725	19.205	29.481	1.00	4.39	0.012	C
ATOM 6278 CD1 LEU A 417	9.689	22.081	30.525	1.00	8.48	0.005	C
ATOM 6281 HN ALA A 418	8.506	17.269	32.385	1.00	0.00	0.163	HD
ATOM 6284 CA ALA A 418	8.526	15.344	31.360	1.00	6.42	0.172	C
ATOM 6286 C ALA A 418	7.258	13.491	30.462	1.00	9.00	-0.271	OA
ATOM 6287 O ALA A 418	7.258	13.491	30.462	1.00	9.00	-0.271	OA
ATOM 6288 CB ALA A 418	9.280	14.862	32.595	1.00	6.77	0.042	C
ATOM 6292 N ASN A 419	6.159	18.658	34.690	1.00	5.04	-0.346	N
ATOM 6293 HN ASN A 419	6.175	15.724	32.420	1.00	0.00	0.163	HD
ATOM 6294 CA ASN A 419	4.847	14.369	31.432	1.00	9.12	0.185	C
ATOM 6296 C ASN A 419	5.114	14.588	29.924	1.00	10.50	0.241	C
ATOM 6297 O ASN A 419	4.003	13.551	29.326	1.00	8.11	-0.271	OA
ATOM 6298 CB ASN A 419	3.703	15.046	32.215	1.00	3.70	0.137	C
ATOM 6301 CG1 ASN A 419	3.703	14.420	30.698	1.00	5.15	0.042	C
ATOM 6302 ND2 ASN A 419	2.943	15.414	34.518	1.00	7.06	-0.370	N
ATOM 6303 HD2 ASN A 419	2.941	15.162	35.506	1.00	0.00	0.159	HD
ATOM 6304 HD2 ASN A 419	2.939	16.200	34.174	1.00	0.00	0.159	HD
ATOM 6305 OD1 ASN A 419	4.363	13.726	34.101	1.00	10.74	-0.274	OA
ATOM 6306 N GLY A 420	4.832	15.662	29.339	1.00	7.79	-0.351	N
ATOM 6307 HN GLY A 420	5.250	16.410	29.339	1.00	0.00	0.163	HD
ATOM 6308 CA GLY A 420	4.584	15.470	27.900	1.00	5.43	0.225	C
ATOM 6311 C GLY A 420	5.945	14.983	27.109	1.00	7.04	0.236	C
ATOM 6312 O GLY A 420	4.120	14.133	26.266	1.00	7.79	-0.271	OA
ATOM 6313 N LEU A 421	6.789	14.810	27.555	1.00	6.34	-0.346	N
ATOM 6314 HN LEU A 421	7.107	15.295	28.394	1.00	0.00	0.163	HD
ATOM 6315 CA LEU A 421	7.707	13.907	26.483	1.00	6.42	0.241	C
ATOM 6317 C LEU A 421	7.155	12.474	26.863	1.00	10.44	0.241	C
ATOM 6318 O LEU A 421	7.155	11.775	25.845	1.00	10.77	-0.271	OA
ATOM 6319 CB LEU A 421	7.155	13.952	28.451	1.00	8.58	0.042	C
ATOM 6321 CG1 LEU A 421	9.729	15.343	27.253	1.00	8.45	0.002	C
ATOM 6324 CO2 LEU A 421	10.010	12.815	26.964	1.00	9.27	0.012	C
ATOM 6328 CD1 SER A 422	6.159	18.658	34.690	1.00	5.04	-0.346	N
ATOM 6332 N SER A 422	6.942	12.640	28.030	1.00	7.55	-0.344	N
ATOM 6333 HN SER A 422	6.741	12.640	28.030	1.00	0.00	0.163	HD
ATOM 6334 CA SER A 422	6.113	10.693	26.179	1.00	6.61	0.241	C
ATOM 6336 C SER A 422	4.900	10.523	27.226	1.00	10.79	0.243	C
ATOM 6337 O SER A 422	4.763	9.503	26.560	1.00	9.12	-0.271	OA
ATOM 6338 CB SER A 422	4.832	10.524	26.560	1.00	6.40	0.042	C
ATOM 6341 OG ASER A 422	6.724	10.504	30.445	0.50	21.65	-0.398	OA
ATOM 6342 HG ASER A 422	6.405	10.399	31.334	1.00	0.00	0.209	HD
ATOM 6343 N LEU A 423	7.963	14.810	27.555	1.00	6.34	-0.346	N
ATOM 6344 HN LEU A 423	4.086	12.301	27.811	1.00	0.00	0.163	HD
ATOM 6345 CA LEU A 423	2.787	11.387	26.872	1.00	7.84	0.177	C
ATOM 6347 C LEU A 423	3.103	10.911	26.483	1.00	10.44	0.241	C
ATOM 6348 O LEU A 423	2.361	10.790	24.083	1.00	10.55	-0.271	OA
ATOM 6349 CB LEU A 423	1.899	12.634	26.638	1.00	9.01	0.038	C
ATOM 6352 CG LEU A 423	1.899	12.634	26.638	1.00	9.01	0.038	C
ATOM 6354 CD1 LEU A 423	0.695	14.095	28.302	1.00	11.04	0.009	C
ATOM 6358 CO2 LEU A 423	1.899	12.634	26.638	1.00	9.01	0.038	C
ATOM 6362 N HIS A 424	6.415	11.998	24.451	1.00	8.70	-0.346	N
ATOM 6363 HN HIS A 424	4.823	12.436	25.115	1.00	0.00	0.163	HD
ATOM 6364 CA HIS A 424	4.449	10.271	23.222	1.00	6.47	0.241	C
ATOM 6366 C HIS A 424	4.799	10.981	22.480	1.00	12.30	0.241	C
ATOM 6367 O HIS A 424	4.018	10.384	21.335	1.00	12.72	-0.271	OA
ATOM 6368 CB HIS A 424	6.012	12.969	22.480	1.00	6.42	0.042	C
ATOM 6371 CG HIS A 424	6.415	11.577	21.347	1.00	11.33	0.028	A
ATOM 6372 CO2 HIS A 424	5.668	11.419	20.169	1.00	11.63	0.114	A
ATOM 6374 HD1 HIS A 424	6.415	11.577	21.347	1.00	11.33	0.028	A
ATOM 6375 HD1 HIS A 424	7.265	11.419	21.109	1.00	0.00	0.166	HD
ATOM 6376 CE2 HIS A 424	6.988	12.666	19.436	1.00	14.77	0.180	A
ATOM 6378 HE2 HIS A 424	6.988	12.666	19.436	1.00	14.77	0.180	A
ATOM 6379 HE2 HIS A 424	6.232	14.290	18.368	1.00	0.00	0.166	HD
ATOM 6380 N GLY A 425	5.600	9.950	23.227	1.00	9.26	-0.351	N
ATOM 6381 HN GLY A 425	6.159	10.242	23.227	1.00	9.26	0.163	HD
ATOM 6382 CA GLY A 425	6.051	8.649	22.756	1.00	8.96	0.225	C
ATOM 6385 C GLY A 425	7.488	8.757	22.210	1.00	12.50	0.235	C
ATOM 6386 O GLY A 425	7.963	8.144	21.812	1.00	11.84	-0.271	OA
ATOM 6387 N GLY A 426	8.261	7.659	22.047	1.00	8.40	-0.351	N
ATOM 6388 HN GLY A 426	7.867	6.871	22.921	1.00	0.00	0.163	HD
ATOM 6389 CA GLY A 426	7.580	6.231	21.915	1.00	8.55	0.235	C
ATOM 6392 C GLY A 426	10.673	6.994	23.013	1.00	13.11	0.236	C
ATOM 6393 O GLY A 426	11.828	7.400	22.747	1.00	11.46	-0.272	OA
ATOM 6394 N PHE A 427	10.308	8.134	23.013	1.00	13.11	0.236	C
ATOM 6395 HN PHE A 427	9.328	8.296	24.475	1.00	0.00	0.163	HD
ATOM 6396 CA PHE A 427	11.364	6.334	25.248	1.00	8.10	0.180	C
ATOM 6398 C PHE A 427	10.942	7.716	26.606	1.00	9.98	0.241	C
ATOM 6399 O PHE A 427	9.764	7.513	26.819	1.00	11.92	-0.271	OA
ATOM 6400 CB PHE A 427	11.594	9.873	25.475	1.00	11.66	0.073	C
ATOM 6403 CG PHE A 427	12.189	10.663	24.268	1.00	11.11	0.056	C
ATOM 6404 CD1 PHE A 427	13.571	10.463	24.003	1.00	10.02	0.007	A
ATOM 6406 CO2 PHE A 427	11.361	11.253	23.402	1.00	15.32	0.007	A
ATOM 6408 CE1 PHE A 427	14.078	11.697	22.832	1.00	9.11	0.019	A
ATOM 6410 CE2 PHE A 427	11.870	11.856	22.050	1.00	11.15	0.001	A
ATOM 6412 CE PHE A 427	13.213	11.751	21.985	1.00	6.93	0.000	A
ATOM 6414 N LEU A 428	11.917	10.920	27.555	1.00	6.34	-0.346	N
ATOM 6415 HN LEU A 428	12.889	7.789	27.285	1.00	0.00	0.163	HD
ATOM 6416 CA LEU A 428	11.562	7.258	28.911	1.00	5.87	0.177	C
ATOM 6418 C LEU A 428	12.162	8.981	29.746	1.00	10.80	0.243	C
ATOM 6419 O LEU A 428	13.393	8.416	29.840	1.00	8.31	-0.271	OA
ATOM 6420 CB LEU A 428	12.366	9.594	29.172	1.00	10.92	0.038	C
ATOM 6423 CG LEU A 428	11.839	5.148	30.357	1.00	11.66	0.028	A
ATOM 6425 CD1 LEU A 428	12.561	3.181	30.525	1.00	17.36	0.009	C
ATOM 6429 CO2 LEU A 428	12.119	3.989	31.681	1.00	20.81	0.019	A
ATOM 6433 N PRO A 429	11.405	9.353	30.900	1.00	8.99	-0.337	N
ATOM 6434 CA PRO A 429	12.038	10.147	30.911	1.00	10.20	0.179	C
ATOM 6436 C PRO A 429	12.363	10.198	32.361	1.00	14.52	0.205	C
ATOM 6437 O PRO A 429	11.627	9.432	33.002	1.00	10.53	-0.271	OA
ATOM 6438 CB PRO A 429	10.931	11.543	30.795	1.00	12.53	0.037	C
ATOM 6441 CG PRO A 429	9.630	10.770	30.800	1.00	14.51	0.028	C
ATOM 6444 CD PRO A 429	9.951	9.472	30.072	1.00	9.31	0.127	C
ATOM 6447 N TYR A 430	13.461	10.740	32.891	1.00	7.80	-0.346	N
ATOM 6448 HN TYR A 430	14.138	11.191	32.276	1.00	0.00	0.163	HD
ATOM 6449 CA TYR A 430	13.718	10.709	34.340	1.00	8.69	0.180	C
ATOM 6451 C TYR A 430	13.985	12.164	34.735	1.00	10.15	0.241	C
ATOM 6452 O TYR A 430	14.432	11.959	33.905	1.00	8.47	-0.271	OA
ATOM 6453 CB TYR A 430	14.834	9.809	34.791	1.00	9.37	0.073	C
ATOM 6456 CG TYR A 430	16.262	10.046	34.398	1.00	9.71	0.056	C
ATOM 6457 CD1 TYR A 430	17.101	10.946	34.986				

ATOM 6808	CA	LYS	A452	-0.816	6.655	33.880	1.00	11.22	0.176	C
ATOM 6810	CA	LYS	A452	-0.711	6.711	33.880	1.00	11.22	0.176	C
ATOM 6811	O	LYS	A452	0.635	5.827	32.180	1.00	10.94	-0.271	OA
ATOM 6812	CB	LYS	A452	-1.790	5.984	33.342	1.00	9.23	0.035	C
ATOM 6815	CG	LYS	A452	-5.499	5.954	34.249	1.00	8.09	0.049	C
ATOM 6818	CD	LYS	A452	-3.651	4.122	34.136	1.00	35.96	0.027	C
ATOM 6821	CE	LYS	A452	-5.162	4.275	34.129	1.00	45.91	0.229	C
ATOM 6824	NE	LYS	A452	-5.470	4.151	32.178	1.00	11.22	0.176	C
ATOM 6825	HE1	LYS	A452	-6.691	4.254	32.732	1.00	0.00	0.274	HD
ATOM 6826	HE2	LYS	A452	-5.377	3.282	32.294	1.00	0.00	0.274	HD
ATOM 6827	HE3	LYS	A452	-5.221	4.806	32.101	1.00	0.00	0.274	HD
ATOM 6828	N	GLN	A453	1.315	7.673	33.240	1.00	7.77	-0.346	N
ATOM 6829	HN	GLN	A453	1.165	8.304	34.027	1.00	0.00	0.163	HD
ATOM 6830	CA	ARG	A454	12.950	7.861	32.406	1.00	9.15	0.176	C
ATOM 6832	C	GLN	A453	3.766	7.439	33.113	1.00	11.08	0.241	C
ATOM 6833	O	GLN	A453	3.924	7.648	34.302	1.00	12.00	-0.271	OA
ATOM 6834	CB	ARG	A454	1.400	9.791	31.275	1.00	13.32	0.105	C
ATOM 6837	CG	GLN	A453	1.344	9.095	29.924	1.00	17.55	0.215	C
ATOM 6840	CD	GLN	A453	-0.545	8.406	30.209	1.00	0.00	0.159	HD
ATOM 6842	HE2	GLN	A453	0.223	7.978	28.879	1.00	0.00	0.159	HD
ATOM 6844	OE1	GLN	A453	4.615	6.735	32.369	1.00	8.20	-0.346	N
ATOM 6845	N	ARG	A454	6.373	6.498	31.407	1.00	0.00	0.163	HD
ATOM 6847	CA	ARG	A454	4.877	7.462	33.030	1.00	16.09	0.241	C
ATOM 6849	C	ARG	A454	7.182	8.198	32.071	1.00	13.92	-0.271	OA
ATOM 6851	CB	ARG	A454	5.235	31.926	1.00	11.59	0.036	C	
ATOM 6854	CG	ARG	A454	7.814	4.736	32.438	1.00	16.81	0.023	C
ATOM 6857	CD	ARG	A454	-2.258	3.455	31.686	1.00	27.36	0.138	C
ATOM 6860	NE	ARG	A454	8.328	3.790	30.263	1.00	17.17	-0.227	N
ATOM 6861	HE	ARG	A454	7.867	4.444	29.949	1.00	0.00	0.177	HD
ATOM 6862	HE1	ARG	A454	8.107	1.906	29.696	1.00	32.69	-0.235	N
ATOM 6863	HE2	ARG	A454	10.047	1.336	28.999	1.00	0.00	0.174	HD
ATOM 6865	HE3	ARG	A454	8.227	1.414	29.078	1.00	0.00	0.174	HD
ATOM 6866	HE4	ARG	A454	8.977	3.404	28.075	1.00	27.58	-0.235	N
ATOM 6867	HE5	ARG	A454	9.457	2.834	27.378	1.00	0.00	0.174	HD
ATOM 6868	HE6	ARG	A454	8.212	2.732	27.378	1.00	0.00	0.174	HD
ATOM 6869	N	GLN	A453	7.400	7.724	34.230	1.00	9.93	-0.346	N
ATOM 6870	HN	GLN	A453	7.046	7.234	35.052	1.00	0.00	0.163	HD
ATOM 6871	CA	ASP	A455	6.713	7.113	34.381	1.00	9.36	0.241	C
ATOM 6873	C	GLN	A453	9.225	8.248	35.638	1.00	10.44	0.241	C
ATOM 6874	O	GLN	A453	9.259	7.925	36.606	1.00	11.94	-0.271	OA
ATOM 6875	CB	ASP	A455	6.114	9.000	32.000	1.00	10.69	0.105	C
ATOM 6878	CG	GLN	A453	8.933	11.186	35.013	1.00	12.32	0.105	C
ATOM 6881	CD	GLN	A453	8.329	12.540	35.260	1.00	11.79	0.215	C
ATOM 6882	HE2	GLN	A453	6.713	12.212	34.313	1.00	0.00	0.159	HD
ATOM 6883	HE3	GLN	A453	6.573	12.212	34.313	1.00	0.00	0.159	HD
ATOM 6884	HE4	GLN	A453	6.704	13.119	35.006	1.00	0.00	0.159	HD
ATOM 6885	OE1	ASP	A455	8.979	13.367	35.991	1.00	12.20	-0.346	N
ATOM 6886	N	VAL	A456	10.540	8.286	35.672	1.00	10.36	-0.346	N
ATOM 6887	HN	VAL	A456	11.066	8.796	34.847	1.00	0.00	0.163	HD
ATOM 6888	CA	VAL	A456	11.910	16.909	35.920	1.00	6.57	0.241	C
ATOM 6890	C	VAL	A456	11.611	9.224	37.603	1.00	11.29	0.241	C
ATOM 6891	O	VAL	A456	12.243	10.054	36.977	1.00	7.97	-0.271	OA
ATOM 6892	CB	VAL	A456	12.459	7.046	36.469	1.00	8.60	0.012	C
ATOM 6894	CG1	VAL	A456	13.213	6.803	37.939	1.00	9.87	0.012	C
ATOM 6898	CG2	VAL	A456	12.046	5.717	35.946	1.00	12.08	0.012	C
ATOM 6902	N	VAL	A456	13.443	4.440	36.212	1.00	0.00	-0.183	N
ATOM 6903	HN	MET	A457	10.741	8.711	39.363	1.00	0.00	0.163	HD
ATOM 6904	CA	MET	A457	11.544	10.766	39.527	1.00	8.57	0.177	C
ATOM 6906	C	MET	A457	11.614	10.426	40.946	1.00	9.58	0.049	C
ATOM 6907	O	MET	A457	12.405	9.539	41.432	1.00	11.16	-0.271	OA
ATOM 6908	CB	MET	A457	10.238	11.178	40.178	1.00	9.54	0.045	C
ATOM 6911	CG	MET	A457	9.117	11.435	39.476	1.00	11.91	0.049	C
ATOM 6914	SD	MET	A457	7.548	11.360	39.747	1.00	23.30	-0.173	SA
ATOM 6915	CE	MET	A457	6.433	12.017	38.520	1.00	10.86	-0.089	C
ATOM 6918	N	VAL	A456	13.787	11.000	40.628	1.00	8.46	0.146	N
ATOM 6920	HN	VAL	A456	13.906	11.670	39.666	1.00	0.00	0.163	HD
ATOM 6921	CA	VAL	A456	14.915	10.692	41.314	1.00	7.03	0.180	C
ATOM 6923	C	VAL	A456	15.013	9.767	42.163	1.00	10.63	0.012	C
ATOM 6924	O	VAL	A456	15.419	12.891	42.130	1.00	10.83	-0.271	OA
ATOM 6925	CB	VAL	A456	16.167	10.643	40.440	1.00	12.37	0.009	C
ATOM 6927	CG1	VAL	A456	17.433	9.246	40.440	1.00	12.37	0.009	C
ATOM 6931	CG2	VAL	A456	15.994	9.597	39.325	1.00	12.65	0.012	C
ATOM 6935	N	TYR	A459	14.601	11.426	43.628	1.00	9.06	-0.346	N
ATOM 6936	HN	TYR	A459	14.288	11.479	43.688	1.00	9.23	0.012	C
ATOM 6937	CA	TYR	A459	14.614	12.481	44.670	1.00	5.44	0.180	C
ATOM 6939	C	TYR	A459	15.823	12.128	43.704	1.00	9.75	0.012	C
ATOM 6940	O	TYR	A459	15.562	11.097	43.997	1.00	13.37	-0.271	OA
ATOM 6941	CB	TYR	A459	13.221	12.530	43.504	1.00	7.31	0.073	C
ATOM 6944	CG	TYR	A459	12.111	12.914	44.159	1.00	11.09	-0.346	N
ATOM 6945	CD1	TYR	A459	12.302	14.097	45.365	1.00	13.65	0.010	A
ATOM 6947	CD2	TYR	A459	10.898	12.288	44.302	1.00	10.32	0.010	A
ATOM 6949	CE1	TYR	A459	11.010	14.512	42.100	1.00	14.22	0.010	A
ATOM 6951	CE2	TYR	A459	9.896	12.710	43.424	1.00	7.87	0.037	A
ATOM 6953	CE	TYR	A459	10.116	13.833	42.103	1.00	10.89	0.065	A
ATOM 6954	OH	TYR	A459	9.307	14.243	41.749	1.00	11.51	0.217	HD
ATOM 6955	HH	TYR	A459	9.307	15.008	41.263	1.00	0.00	0.217	HD
ATOM 6956	N	THR	A460	16.703	12.969	45.799	1.00	9.45	-0.344	N
ATOM 6957	HN	THR	A460	16.747	13.392	46.147	1.00	11.32	0.012	C
ATOM 6958	CA	THR	A460	17.786	12.469	46.741	1.00	9.06	0.205	C
ATOM 6960	C	THR	A460	17.654	13.512	47.987	1.00	9.70	0.243	C
ATOM 6961	O	THR	A460	16.038	12.342	48.107	1.00	11.32	-0.271	OA
ATOM 6962	CB	THR	A460	19.149	12.868	46.035	1.00	10.78	0.146	C
ATOM 6964	CG2	THR	A460	19.203	12.440	46.672	1.00	7.28	0.042	C
ATOM 6968	CG1	THR	A460	19.304	14.265	47.946	1.00	22.22	-0.343	N
ATOM 6969	OG1	THR	A460	20.139	14.399	48.380	1.00	0.00	0.210	HD
ATOM 6970	N	HIS	A461	18.529	13.318	48.953	1.00	9.25	-0.346	N
ATOM 6971	HN	HIS	A461	19.197	13.558	48.193	1.00	8.23	0.182	C
ATOM 6972	CA	HIS	A461	18.630	14.034	49.101	1.00	6.72	0.182	C
ATOM 6974	C	HIS	A461	17.321	13.891	50.935	1.00	9.96	0.241	C
ATOM 6975	O	HIS	A461	18.513	14.039	49.147	1.00	11.91	-0.346	N
ATOM 6976	CB	HIS	A461	18.895	15.584	48.860	1.00	9.28	0.093	C
ATOM 6979	CG	HIS	A461	20.284	15.769	49.267	1.00	14.92	0.028	A
ATOM 6980	CD2	HIS	A461	21.352	16.427	49.720	1.00	11.91	0.028	A
ATOM 6982	ND1	HIS	A461	20.701	16.155	48.088	1.00	13.74	-0.354	N
ATOM 6983	HD1	HIS	A461	20.085	14.557	47.505	1.00	0.00	0.166	HD
ATOM 6984	CE1	HIS	A461	21.949	15.847	48.844	1.00	12.29	0.028	A
ATOM 6986	NE2	HIS	A461	22.381	16.286	48.826	1.00	11.96	-0.360	N
ATOM 6987	HE2	HIS	A461	23.319	16.680	48.898	1.00	0.00	0.166	HD
ATOM 6988	N	ASP	A462	17.067	16.046	50.109	1.00	8.29	-0.271	OA
ATOM 6989	HN	ASP	A462	17.814	15.911	51.293	1.00	0.00	0.163	HD
ATOM 6990	CA	ASP	A462	15.720	12.254	51.730	1.00	8.68	0.186	C
ATOM 6992	C	ASP	A462	15.362	12.427	53.192	1.00	11.43	-0.271	OA
ATOM 6994	CB	ASP	A462	15.500	10.755	51.405	1.00	8.11	0.147	C
ATOM 6997	CG	ASP	A462	16.333	9.817	52.285	1.00	12.65	0.233	C
ATOM 6998	OD1	ASP	A462	17.485	10.935	52.680	1.00	9.70	-0.648	OA
ATOM 6999	OD2	ASP	A462	15.813	9.717	52.632	1.00	11.18	-0.648	OA
ATOM 7000	N	SER	A463	16.260	12.917	54.062	1.00	8.00	0.000	C
ATOM 7001	HN	SER	A463	17.180	13.239	53.740	1.00	0.00	0.163	HD
ATOM 7002	CA	SER	A463	15.832	12.964	55.448	1.00	8.61	0.200	C
ATOM 7004	C	SER	A463	16.762	13.906	56.215	1.00	8.00	0.243	C
ATOM 7005	O	SER	A463	17.643	1					

ATOM 7377 N SER A 489	4.130	4.908	42.541	1.00	8.98	-0.344	N
ATOM 7378 HN SER A 489	1.463	4.959	43.108	1.00	8.42	0.150	HD
ATOM 7379 CA SER A 489	5.019	3.898	43.108	1.00	8.42	0.200	C
ATOM 7381 C SER A 489	6.202	2.440	43.745	1.00	9.10	0.243	C
ATOM 7382 O SER A 489	1.981	5.192	43.745	1.00	9.10	-0.720	OA
ATOM 7383 CB SER A 489	5.451	2.854	42.089	1.00	8.24	0.199	C
ATOM 7386 OG SER A 489	6.277	1.903	42.735	1.00	12.47	-0.398	DA
ATOM 7387 HD SER A 489	6.547	4.292	45.002	1.00	7.65	-0.344	N
ATOM 7388 N THR A 490	6.517	4.296	45.002	1.00	7.65	-0.344	N
ATOM 7389 HN THR A 490	6.018	3.524	45.444	1.00	8.00	0.163	HD
ATOM 7390 CA THR A 490	7.559	4.999	45.760	1.00	6.18	0.205	C
ATOM 7392 C THR A 490	8.505	4.007	46.436	1.00	9.83	0.243	C
ATOM 7393 O THR A 490	7.994	3.050	47.089	1.00	8.43	-0.271	OA
ATOM 7394 CB THR A 490	16.830	5.762	46.931	1.00	12.98	0.146	C
ATOM 7396 CG2 THR A 490	7.785	6.658	47.693	1.00	15.34	0.042	C
ATOM 7400 OG1 THR A 490	8.553	6.646	46.333	1.00	14.77	-0.393	DA
ATOM 7401 HG1 THR A 490	10.426	6.646	46.333	1.00	14.77	-0.393	DA
ATOM 7402 N TRP A 491	9.784	4.135	46.120	1.00	7.18	-0.346	N
ATOM 7403 HN TRP A 491	10.075	4.903	45.515	1.00	8.00	0.163	HD
ATOM 7404 CA TRP A 491	10.426	3.193	46.025	1.00	8.06	0.075	C
ATOM 7406 C TRP A 491	11.809	3.886	47.499	1.00	6.75	0.241	C
ATOM 7407 O TRP A 491	12.328	4.939	47.071	1.00	7.74	-0.271	OA
ATOM 7408 CB TRP A 491	11.571	2.509	45.428	1.00	8.96	0.075	C
ATOM 7411 CG TRP A 491	10.645	1.550	44.714	1.00	8.62	-0.028	A
ATOM 7412 CD1 TRP A 491	9.596	1.873	43.907	1.00	9.85	0.096	A
ATOM 7414 CD2 TRP A 491	9.596	1.873	43.907	1.00	9.85	0.096	A
ATOM 7415 CE2 TRP A 491	9.579	-0.356	44.002	1.00	9.02	0.042	A
ATOM 7416 CE3 TRP A 491	11.496	-0.806	45.110	1.00	8.34	0.014	A
ATOM 7418 CE4 TRP A 491	8.927	4.792	43.498	1.00	8.58	-0.365	N
ATOM 7419 HD TRP A 491	8.088	0.737	42.918	1.00	0.00	0.165	HD
ATOM 7420 CD2 TRP A 491	9.596	1.873	43.907	1.00	9.85	0.096	A
ATOM 7422 CD3 TRP A 491	11.191	-2.152	45.288	1.00	9.07	0.001	A
ATOM 7424 CH2 TRP A 491	10.081	-2.587	44.553	1.00	7.92	0.002	A
ATOM 7425 HD TRP A 491	8.088	0.737	42.918	1.00	0.00	0.165	HD
ATOM 7427 HN ARG A 492	11.656	2.528	49.014	1.00	8.00	0.163	HD
ATOM 7428 CA ARG A 492	13.165	4.009	49.540	1.00	7.04	0.176	C
ATOM 7430 C ARG A 492	14.156	3.341	51.115	1.00	4.76	0.241	C
ATOM 7431 O ARG A 492	14.054	2.203	50.847	1.00	9.54	-0.271	OA
ATOM 7432 CB ARG A 492	12.491	4.497	50.857	1.00	4.46	0.036	C
ATOM 7435 CG ARG A 492	12.341	3.441	51.115	1.00	4.76	0.241	C
ATOM 7438 CD ARG A 492	12.858	6.074	52.814	1.00	8.31	0.138	C
ATOM 7441 NE ARG A 492	13.807	6.935	53.525	1.00	11.10	-0.227	N
ATOM 7442 HE ARG A 492	14.156	7.757	53.525	1.00	11.10	-0.227	N
ATOM 7443 CE ARG A 492	14.267	6.739	54.773	1.00	12.48	0.665	C
ATOM 7444 NH1 ARG A 492	13.877	5.694	55.476	1.00	8.07	-0.225	N
ATOM 7445 HH1 ARG A 492	15.445	4.488	56.423	1.00	8.00	0.163	HD
ATOM 7446 2HH1 ARG A 492	13.243	4.998	55.066	1.00	0.00	0.174	HD
ATOM 7447 NH2 ARG A 492	15.096	7.767	55.209	1.00	7.31	-0.225	N
ATOM 7448 HH2 ARG A 492	15.445	4.488	56.423	1.00	8.00	0.163	HD
ATOM 7449 2HH2 ARG A 492	15.396	8.468	57.786	1.00	0.00	0.174	HD
ATOM 7450 N PRO A 493	15.111	2.609	48.953	1.00	9.42	-0.337	N
ATOM 7451 CA PRO A 493	16.010	4.472	49.329	1.00	8.44	0.241	C
ATOM 7453 C PRO A 493	17.012	1.579	50.180	1.00	8.88	-0.241	C
ATOM 7454 O PRO A 493	17.439	2.699	50.506	1.00	10.07	-0.271	OA
ATOM 7455 CB PRO A 493	16.739	3.819	50.506	1.00	10.07	-0.271	OA
ATOM 7458 CG PRO A 493	16.612	2.878	47.150	1.00	12.03	0.022	C
ATOM 7461 CD PRO A 493	15.302	3.383	47.692	1.00	8.70	0.127	C
ATOM 7464 N CYS A 494	16.954	4.482	50.162	1.00	8.60	-0.346	N
ATOM 7465 HN CYS A 494	16.989	-0.149	50.503	1.00	0.00	0.163	HD
ATOM 7466 CA CYS A 494	18.250	0.536	52.077	1.00	11.16	0.186	C
ATOM 7468 C CYS A 494	19.731	3.391	53.107	1.00	9.98	0.241	C
ATOM 7469 O CYS A 494	20.605	0.523	52.509	1.00	10.97	-0.271	OA
ATOM 7470 CB CYS A 494	17.839	-0.529	53.058	1.00	9.42	0.121	C
ATOM 7473 SG CYS A 494	18.308	2.745	52.925	1.00	11.89	-0.346	N
ATOM 7474 N ASP A 495	20.075	0.009	50.432	1.00	6.59	-0.345	N
ATOM 7475 HN ASP A 495	19.330	-0.194	49.765	1.00	0.00	0.163	HD
ATOM 7476 CA ASP A 495	21.457	1.939	49.865	1.00	8.00	0.241	C
ATOM 7478 C ASP A 495	21.553	-0.167	48.467	1.00	8.95	0.241	C
ATOM 7479 O ASP A 495	20.537	0.043	47.808	1.00	8.25	-0.271	OA
ATOM 7480 CB ASP A 495	21.992	3.023	50.200	1.00	8.80	0.241	C
ATOM 7483 CG ASP A 495	21.368	-2.721	50.155	1.00	10.66	0.175	C
ATOM 7484 CD1 ASP A 495	20.664	-2.824	49.141	1.00	9.92	-0.648	DA
ATOM 7485 CD2 ASP A 495	21.373	-7.805	49.141	1.00	13.00	-0.648	DA
ATOM 7486 N GLN A 496	22.737	-0.426	47.894	1.00	7.78	-0.346	N
ATOM 7487 HN GLN A 496	23.519	-0.698	48.488	1.00	0.00	0.163	HD
ATOM 7488 CA GLN A 496	22.859	1.347	48.989	1.00	11.61	0.271	C
ATOM 7490 C GLN A 496	22.207	-1.402	45.669	1.00	11.04	0.241	C
ATOM 7491 O GLN A 496	21.853	-1.202	44.506	1.00	10.77	-0.271	OA
ATOM 7492 CB GLN A 496	24.408	3.360	43.821	1.00	8.61	-0.346	N
ATOM 7495 CG GLN A 496	25.091	-1.747	46.320	1.00	11.18	0.105	C
ATOM 7498 CD GLN A 496	26.179	-2.834	46.819	1.00	11.73	-0.346	N
ATOM 7499 NE2 GLN A 496	27.176	-2.962	45.880	1.00	10.55	-0.370	N
ATOM 7500 HE2 GLN A 496	28.102	-3.021	45.456	1.00	0.00	0.159	HD
ATOM 7501 2HE2 GLN A 496	26.839	-3.748	44.436	1.00	0.00	0.159	HD
ATOM 7502 OE1 GLN A 496	26.898	-0.889	45.010	1.00	12.81	-0.274	OA
ATOM 7503 N VAL A 497	21.915	-2.549	46.313	1.00	6.54	-0.346	N
ATOM 7504 HN VAL A 497	22.221	-4.644	47.281	1.00	0.00	0.163	HD
ATOM 7505 CA VAL A 497	21.176	-3.639	45.660	1.00	8.54	0.180	C
ATOM 7507 C VAL A 497	19.694	-3.291	45.547	1.00	9.64	0.241	C
ATOM 7508 O VAL A 497	19.788	-4.617	46.313	1.00	9.11	-0.271	OA
ATOM 7509 CB VAL A 497	21.343	-4.945	46.467	1.00	9.74	0.009	C
ATOM 7511 CG1 VAL A 497	20.624	-6.144	45.822	1.00	12.77	0.012	C
ATOM 7515 CG2 VAL A 497	22.848	-5.780	45.822	1.00	12.77	0.012	C
ATOM 7519 N GLU A 498	19.079	-2.874	46.677	1.00	7.53	-0.346	N
ATOM 7520 HN GLU A 498	19.564	-2.862	47.574	1.00	0.00	0.163	HD
ATOM 7521 CA GLU A 498	17.605	-1.427	46.467	1.00	8.42	0.241	C
ATOM 7523 C GLU A 498	16.540	-1.087	44.918	1.00	10.82	-0.271	OA
ATOM 7525 CB GLU A 498	17.058	-1.740	45.822	1.00	8.42	0.241	C
ATOM 7528 CG GLU A 498	16.940	-1.395	48.876	1.00	7.91	0.116	C
ATOM 7531 CD GLU A 498	16.079	-2.900	50.081	1.00	11.69	0.172	C
ATOM 7532 OE1 GLU A 498	15.707	-1.740	45.822	1.00	8.42	0.241	C
ATOM 7533 OE2 GLU A 498	15.707	-3.551	50.819	1.00	10.56	-0.648	DA
ATOM 7534 N SER A 499	18.595	-0.376	45.609	1.00	7.31	-0.346	N
ATOM 7535 HN SER A 499	19.369	-1.395	46.467	1.00	8.42	0.241	C
ATOM 7536 CA SER A 499	18.543	0.759	44.644	1.00	6.72	0.200	C
ATOM 7538 C SER A 499	18.469	0.302	43.198	1.00	10.98	0.243	C
ATOM 7539 O SER A 499	17.951	-1.951	44.644	1.00	11.42	-0.271	OA
ATOM 7540 CB SER A 499	19.637	1.756	44.814	1.00	7.95	0.199	C
ATOM 7543 OG SER A 499	19.781	2.245	46.150	1.00	12.91	-0.398	DA
ATOM 7544 HG SER A 499	20.492	2.866	46.150	1.00	12.91	-0.398	DA
ATOM 7545 N ALA A 500	19.254	-0.698	42.812	1.00	10.74	-0.346	N
ATOM 7546 HN ALA A 500	19.890	-1.129	43.483	1.00	0.00	0.163	HD
ATOM 7547 CA ALA A 500	19.212	-1.879	43.134	1.00	9.75	0.240	C
ATOM 7549 C ALA A 500	17.413	-1.772	39.982	1.00	10.78	-0.271	OA
ATOM 7551 CB ALA A 501	20.366	-2.183	43.134	1.00	9.77	0.075	C
ATOM 7555 N VAL A 501	17.308	-2.597	42.087	1.00	8.58	-0.346	N
ATOM 7556 HN VAL A 501	17.761	-2.692	42.996	1.00	0.00	0.163	HD
ATOM 7557 CA VAL A 501	16.028	-2.713	41.010	1.00	6.41	-0.346	N
ATOM 7559 C VAL A 501	14.929	-2.191	41.643	1.00	6.26	0.241	C
ATOM 7560 O VAL A 501	14.021	-3.983	40.821	1.00	8.26	-0.271	OA
ATOM 7561 CB VAL A 501	15.611	-4.168	43.028	1.00	13.89	-0.346	N
ATOM 7563 CG1 VAL A 501	14.189	-4.267	42.902	1.00	10.79	0.012	C
ATOM 7567 CG2 VAL A 501	16.562	-3.794	43.161	1.00	12.49	0.012	C
ATOM 7571 N THR A 502	14.977	-1.119	42.026	1.00	7.26	-0.346	N
ATOM 7572 HN THR A 502	15.700	-1.054	43.142	1.00	0.00	0.163	HD
ATOM 7573 CA THR A 502	13.007	-0.849	42.026	1.00	8.72	0.241	C
ATOM 7575 C THR A 502	14.107	0.662	40.786	1.00	8.83	0.240	C
ATOM 7576 O THR A 502	13.089	0.977	40.282	1.00	10.07	-0.271	OA
ATOM 7577 CB THR A 502	14.223	0.987	43.426	1.00	8.81	0.042	A
ATOM 7581 N TRP A 503	15.333	0.916	40.440	1.00	7.57	-0.346	N
ATOM 7582 HN TRP A 503	16.156	0.097	41.002	1.00	0.00	0.163	HD
ATOM 7583 CA TRP A 503	15.489	1.512	39.103	1.00	10.94	0.181	C
ATOM 7585 C TRP A 503	14.920	0.558	38.064	1.00	11.38	0.241	C
ATOM 7586 O TRP A 503	14.232	0.973	37.134	1.00	10.52	-0.271	OA
ATOM 7587 CB TRP A 503	16.974	1.751	38.814	1.00	9.77	0.075	C
ATOM 7590 CG TRP A 503	17.321	3.209	38.834	1.00	9.33	-0.028	A
ATOM 7591 CD1 TRP A 503	18.021	3.959	39.805	1.00	11.59	0.096	A
ATOM 7593 CD2 TRP A 503	16.989	4.178	37.837	1.00	11.99	-0.002	A

ATOM 7936 N	GLN A 525	25.536	-2.270	53.093	1.00	14.500	-0.346	N
ATOM 7937 HN	GLN A 525	26.553	-2.201	53.110	1.00	14.500	-0.346	N
ATOM 7938 CA	GLN A 525	24.834	-3.274	52.307	1.00	13.774	0.177	C
ATOM 7940 C	GLN A 525	24.401	-4.458	53.153	1.00	15.388	0.241	C
ATOM 7941 O	GLN A 525	25.199	-4.809	53.943	1.00	17.110	-0.178	O
ATOM 7942 CB	GLN A 525	25.744	-3.810	51.190	1.00	13.336	0.044	C
ATOM 7945 CG	GLN A 525	25.066	-4.897	50.344	1.00	12.008	-0.105	C
ATOM 7948 CD	GLN A 525	25.753	-5.425	49.543	1.00	10.720	-0.070	C
ATOM 7949 NE2	GLN A 525	25.993	-6.297	48.581	1.00	10.606	-0.370	N
ATOM 7950 HE2	GLN A 525	25.713	-7.136	49.089	1.00	0.000	0.159	HD
ATOM 7951 2HE2	GLN A 525	26.454	-6.396	47.677	1.00	0.000	0.158	HD
ATOM 7952 OE1	GLN A 525	26.091	-4.032	48.385	1.00	10.444	-0.274	O
ATOM 7953 N	GLN A 526	23.216	-4.998	52.926	1.00	11.555	-0.346	N
ATOM 7954 HN	GLN A 526	22.629	-6.007	52.189	1.00	10.000	-0.163	N
ATOM 7955 CA	GLN A 526	22.724	-6.126	53.688	1.00	14.411	0.177	C
ATOM 7957 C	GLN A 526	22.975	-7.422	52.906	1.00	15.311	0.241	C
ATOM 7958 O	GLN A 526	23.225	-8.383	54.683	1.00	17.000	-0.178	O
ATOM 7959 CB	GLN A 526	21.226	-6.048	53.986	1.00	12.559	0.044	C
ATOM 7962 CG	GLN A 526	20.792	-4.753	54.682	1.00	11.207	-0.105	C
ATOM 7965 CD	GLN A 526	21.457	-5.687	56.050	1.00	10.594	-0.070	C
ATOM 7966 NE2	GLN A 526	21.678	-3.510	56.629	1.00	10.106	-0.370	N
ATOM 7967 HE2	GLN A 526	22.060	-3.465	57.574	1.00	0.000	0.159	HD
ATOM 7968 2HE2	GLN A 526	21.745	-2.630	56.386	1.00	0.000	0.159	HD
ATOM 7969 OE1	GLN A 526	21.518	-5.749	56.680	1.00	10.975	-0.274	O
ATOM 7970 N	GLD A 527	23.195	-8.490	53.667	1.00	16.522	-0.346	N
ATOM 7971 HN	GLD A 527	23.225	-8.383	54.683	1.00	17.000	-0.178	N
ATOM 7972 CA	GLD A 527	23.396	-9.821	53.078	1.00	14.644	0.177	C
ATOM 7974 C	GLD A 527	22.068	-10.425	52.733	1.00	13.555	0.241	C
ATOM 7975 O	GLD A 527	23.225	-10.425	53.476	1.00	15.007	-0.178	O
ATOM 7976 CB	GLD A 527	23.960	-10.758	54.190	1.00	25.711	0.045	C
ATOM 7979 CG	GLD A 527	25.433	-10.534	54.160	1.00	41.007	0.116	C
ATOM 7982 CD	GLD A 527	25.866	-11.327	55.651	1.00	61.400	0.172	C
ATOM 7983 OE1	GLD A 527	25.211	-11.750	56.559	1.00	63.655	-0.648	O
ATOM 7984 OE2	GLD A 527	25.421	-12.604	56.559	1.00	66.654	-0.648	O
ATOM 7985 N	ARG A 528	21.930	-11.126	51.600	1.00	9.488	-0.346	N
ATOM 7986 HN	ARG A 528	22.757	-11.262	51.019	1.00	0.000	0.163	N
ATOM 7987 CA	ARG A 528	21.944	-12.816	52.926	1.00	11.555	-0.346	N
ATOM 7989 C	ARG A 528	20.948	-13.095	50.519	1.00	16.144	0.241	C
ATOM 7990 O	ARG A 528	21.944	-13.202	49.819	1.00	15.336	-0.274	O
ATOM 7991 CB	ARG A 528	19.522	-10.816	53.476	1.00	0.032	0.032	C
ATOM 7994 CG	ARG A 528	19.608	-9.469	50.565	1.00	12.339	0.023	C
ATOM 7997 CD	ARG A 528	18.847	-8.680	49.503	1.00	11.990	0.138	C
ATOM 8000 HE	ARG A 528	19.037	-7.370	49.503	1.00	0.000	0.163	N
ATOM 8001 HE	ARG A 528	17.607	-7.260	50.599	1.00	0.000	0.177	HD
ATOM 8002 CZ	ARG A 528	19.382	-6.367	50.244	1.00	10.334	0.665	C
ATOM 8003 NH1	ARG A 528	19.037	-5.259	49.503	1.00	0.000	0.163	N
ATOM 8004 1BH1	ARG A 528	21.267	-5.670	49.784	1.00	0.000	0.174	HD
ATOM 8005 2BH1	ARG A 528	20.889	-7.821	49.215	1.00	0.000	0.174	HD
ATOM 8006 NH2	ARG A 528	19.037	-5.259	49.503	1.00	0.000	0.163	N
ATOM 8007 1H2	ARG A 528	19.668	-4.482	50.982	1.00	0.000	0.174	HD
ATOM 8008 2H2	ARG A 528	18.071	-5.799	51.322	1.00	0.000	0.174	HD
ATOM 8009 N	THR A 529	19.012	-14.017	50.716	1.00	15.711	-0.346	N
ATOM 8010 HN	THR A 529	19.241	-13.850	51.362	1.00	0.000	0.163	N
ATOM 8011 CA	THR A 529	20.117	-15.287	49.977	1.00	20.339	0.205	C
ATOM 8013 C	THR A 529	19.522	-15.007	48.602	1.00	21.945	0.241	C
ATOM 8014 O	THR A 529	18.946	-13.950	48.355	1.00	18.774	-0.274	O
ATOM 8015 CB	THR A 529	19.253	-16.366	50.636	1.00	19.671	0.146	C
ATOM 8017 CG	THR A 529	18.688	-16.493	49.648	1.00	16.144	0.023	C
ATOM 8021 OG1	THR A 529	17.897	-15.868	50.000	1.00	16.711	-0.393	O
ATOM 8022 HG1	THR A 529	17.897	-16.537	51.191	1.00	0.000	0.210	HD
ATOM 8023 N	GLY A 530	18.566	-17.972	47.600	1.00	6.939	-0.346	N
ATOM 8024 HN	GLY A 530	20.021	-16.856	47.912	1.00	0.000	0.163	N
ATOM 8025 CA	GLY A 530	18.978	-15.801	46.360	1.00	16.998	0.177	C
ATOM 8027 C	GLY A 530	19.472	-15.539	46.442	1.00	14.457	0.241	C
ATOM 8028 O	GLY A 530	16.912	-14.852	45.606	1.00	15.117	-0.274	O
ATOM 8029 CB	GLY A 530	19.181	-17.115	45.584	1.00	29.771	0.045	C
ATOM 8032 CG	GLY A 530	18.183	-17.063	44.758	1.00	19.589	0.116	C
ATOM 8035 CD	GLY A 530	19.681	-18.416	43.548	1.00	62.001	-0.172	C
ATOM 8036 OE1	GLY A 530	20.897	-18.688	43.673	1.00	69.223	-0.648	O
ATOM 8037 OE2	GLY A 530	18.847	-19.127	43.058	1.00	61.447	-0.648	O
ATOM 8038 N	GLN A 531	16.785	-16.201	47.372	1.00	15.144	-0.346	N
ATOM 8039 HN	GLN A 531	17.272	-16.858	47.981	1.00	0.000	0.163	N
ATOM 8040 CA	GLN A 531	14.737	-17.009	48.469	1.00	15.642	0.177	C
ATOM 8042 C	GLN A 531	15.036	-14.577	48.034	1.00	17.335	0.241	C
ATOM 8043 O	GLN A 531	14.020	-14.011	47.604	1.00	14.778	-0.274	O
ATOM 8044 CB	GLN A 531	14.737	-17.009	48.469	1.00	15.642	0.177	C
ATOM 8047 CG	GLN A 531	13.297	-16.786	48.793	1.00	32.550	0.116	C
ATOM 8050 CD	GLN A 531	12.684	-17.606	49.903	1.00	47.577	0.172	C
ATOM 8051 OE1	GLN A 531	13.288	-17.870	48.644	1.00	54.848	-0.648	O
ATOM 8052 OE2	GLN A 531	11.511	-18.003	49.725	1.00	50.200	-0.648	O
ATOM 8053 N	GLY A 532	16.977	-14.037	46.100	1.00	12.888	-0.346	N
ATOM 8054 HN	GLY A 532	16.507	-14.618	49.437	1.00	0.000	0.163	N
ATOM 8055 CA	GLY A 532	15.551	-12.666	49.438	1.00	14.003	0.177	C
ATOM 8057 C	GLY A 532	15.320	-13.181	48.824	1.00	12.619	0.241	C
ATOM 8058 O	GLY A 532	15.122	-10.674	48.101	1.00	15.553	-0.274	O
ATOM 8059 CB	GLY A 532	16.487	-12.317	50.621	1.00	11.990	0.044	C
ATOM 8062 CG	GLY A 532	16.488	-13.181	49.824	1.00	12.619	0.116	C
ATOM 8065 CD	GLY A 532	17.099	-13.070	52.968	1.00	15.996	0.215	C
ATOM 8066 NE2	GLY A 532	16.946	-12.897	54.198	1.00	19.774	-0.370	N
ATOM 8067 HE2	GLY A 532	16.946	-12.897	54.198	1.00	19.774	-0.370	N
ATOM 8068 2HE2	GLY A 532	17.219	-12.823	53.921	1.00	0.000	0.159	HD
ATOM 8069 OE1	GLY A 532	18.292	-13.098	52.760	1.00	17.777	-0.274	O
ATOM 8070 N	LEU A 533	16.922	-11.947	47.574	1.00	14.649	-0.346	N
ATOM 8071 HN	LEU A 533	17.516	-12.731	47.814	1.00	0.000	0.163	N
ATOM 8072 CA	LEU A 533	17.255	-11.030	46.470	1.00	13.822	0.177	C
ATOM 8074 C	LEU A 533	16.102	-10.947	47.574	1.00	12.627	0.241	C
ATOM 8075 O	LEU A 533	15.741	-9.865	46.470	1.00	12.445	-0.274	O
ATOM 8076 CB	LEU A 533	18.536	-11.530	45.853	1.00	15.006	0.038	C
ATOM 8079 CG	LEU A 533	19.211	-10.796	45.853	1.00	25.777	0.146	C
ATOM 8081 CD1	LEU A 533	19.637	-9.404	45.115	1.00	19.688	0.009	C
ATOM 8085 CD2	LEU A 533	20.437	-11.637	44.296	1.00	23.388	0.009	C
ATOM 8089 N	ALA A 534	15.493	-12.073	45.320	1.00	12.237	-0.346	N
ATOM 8090 HN	ALA A 534	15.836	-12.966	45.484	1.00	0.000	0.163	N
ATOM 8091 CA	ALA A 534	14.332	-12.052	44.229	1.00	16.877	0.172	C
ATOM 8093 C	ALA A 534	14.127	-10.710	44.415	1.00	18.117	-0.274	O
ATOM 8094 O	ALA A 534	13.232	-10.710	44.415	1.00	18.117	-0.274	O
ATOM 8095 CB	ALA A 534	13.933	-13.485	43.889	1.00	22.440	0.042	C
ATOM 8099 N	ASN A 535	12.912	-11.424	47.113	1.00	11.333	-0.346	N
ATOM 8100 HN	ASN A 535	13.584	-11.937	46.748	1.00	0.000	0.163	N
ATOM 8101 CA	ASN A 535	11.740	-10.823	46.821	1.00	9.939	0.185	C
ATOM 8103 C	ASN A 535	12.859	-9.308	47.031	1.00	11.442	0.241	C
ATOM 8104 O	ASN A 535	10.869	-8.777	47.495	1.00	11.411	-0.274	O
ATOM 8105 CB	ASN A 535	11.549	-11.469	48.204	1.00	12.111	0.137	C
ATOM 8108 CG	ASN A 535	10.623	-12.863	48.149	1.00	10.000	0.023	C
ATOM 8109 ND2	ASN A 535	11.044	-13.075	47.100	1.00	17.114	-0.370	N
ATOM 8110 HD2	ASN A 535	11.562	-13.454	49.985	1.00	0.000	0.159	HD
ATOM 8111 2HD2	ASN A 535	10.623	-14.637	49.985	1.00	0.000	0.159	HD
ATOM 8112 OD1	ASN A 535	10.315	-13.163	47.120	1.00	17.775	-0.274	O
ATOM 8113 N	ILE A 536	13.010	-8.741	46.730	1.00	12.228	-0.346	N
ATOM 8114 HN	ILE A 536	13.811	-9.252	46.987	1.00	10.000	0.163	N
ATOM 8115 CA	ILE A 536	13.090	-7.242	46.877	1.00	11.201	0.180	C
ATOM 8117 C	ILE A 536	12.007	-6.598	46.021	1.00	10.333	0.241	C
ATOM 8118 O	ILE A 536	11.284	-5.683	46.021	1.00	10.715	-0.274	O
ATOM 8119 CB	ILE A 536	14.449	-6.656	46.506	1.00	9.911	0.013	C
ATOM 8121 CG1	ILE A 536	15.552	-7.822	47.469	1.00	13.934	0.026	C
ATOM 8122 CG2	ILE A 536	14.462	-5.110	46.491	1.00	11.714	0.013	C
ATOM 8128 CD1	ILE A 536	16.946	-6.866	46.898	1.00	17.000	0.026	C
ATOM 8132 N	ALA A 537	11.810	-10.811	44.80				

AT001	8505	CD	GLD	A	562	24.365	0.419	59.285	1.00	55.83	0.172	C
AT002	8506	OE1	ZEA	A	565	24.200	1.471	60.964	1.00	55.82	0.172	C
AT003	8507	OE2	GLD	A	562	25.459	-0.134	59.574	1.00	64.19	-0.648	OA
AT004	8508	N	LEU	A	563	19.747	0.225	61.456	1.00	9.95	-0.346	N
AT005	8509	HN	VAL	A	565	19.450	0.907	79.524	1.00	10.31	-0.177	C
AT006	8510	CB	ALA	A	563	19.343	0.335	62.844	1.00	10.31	0.177	C
AT007	8511	C	LEU	A	563	18.437	-0.725	63.292	1.00	13.90	0.241	C
AT008	8512	C	LEU	A	563	18.440	-1.321	64.071	1.00	13.60	0.241	C
AT009	8513	CB	LEU	A	563	18.546	1.713	62.911	1.00	13.60	0.038	C
AT010	8514	CB	LEU	A	563	18.421	2.513	64.165	1.00	32.79	-0.020	C
AT011	8515	CD1	LEU	A	563	19.802	2.589	64.015	1.00	23.75	0.009	C
AT012	8516	CD2	LEU	A	563	17.908	3.911	63.779	1.00	20.04	0.009	C
AT013	8517	N	ALA	A	564	17.451	-1.094	62.441	1.00	9.59	-0.346	N
AT014	8518	HN	ALA	A	564	17.321	-0.948	61.554	1.00	10.00	-0.183	HD
AT015	8519	CA	ALA	A	564	16.578	-2.201	62.817	1.00	11.81	0.172	C
AT016	8520	C	ALA	A	564	17.362	-3.501	62.933	1.00	13.59	0.240	C
AT017	8521	C	ALA	A	564	17.268	-4.447	64.016	1.00	11.20	-0.271	OA
AT018	8522	CB	ALA	A	564	15.436	-2.432	61.788	1.00	9.08	0.042	C
AT019	8523	N	VAL	A	565	18.276	-3.780	61.992	1.00	11.42	-0.346	N
AT020	8524	HN	VAL	A	565	18.425	-4.999	62.508	1.00	15.60	0.165	HD
AT021	8525	CA	VAL	A	565	19.059	-5.011	62.066	1.00	9.14	0.180	C
AT022	8526	C	VAL	A	565	19.910	-4.987	63.352	1.00	13.58	0.241	C
AT023	8527	C	VAL	A	565	20.048	-6.022	64.016	1.00	11.20	-0.271	OA
AT024	8528	CB	VAL	A	565	19.942	-5.183	60.835	1.00	9.69	0.009	C
AT025	8529	CG1	VAL	A	565	20.905	-6.382	61.012	1.00	12.85	0.012	C
AT026	8530	CG2	VAL	A	565	19.562	-5.540	59.568	1.00	8.42	0.012	C
AT027	8531	N	ALA	A	566	20.516	-3.869	63.730	1.00	12.78	-0.346	N
AT028	8532	HN	ALA	A	566	20.443	-3.030	63.154	1.00	9.00	0.163	HD
AT029	8533	CA	ALA	A	566	21.292	-7.657	64.408	1.00	14.73	-0.172	C
AT030	8534	C	ALA	A	566	20.406	-4.064	66.178	1.00	17.36	0.240	C
AT031	8535	O	ALA	A	566	20.812	-5.725	67.124	1.00	15.63	-0.271	OA
AT032	8536	CB	ALA	A	566	22.047	-7.493	65.091	1.00	17.56	0.042	C
AT033	8537	N	ALA	A	567	19.178	-3.543	66.205	1.00	11.68	-0.346	N
AT034	8538	HN	ALA	A	567	18.458	-4.871	66.436	1.00	12.00	-0.271	OA
AT035	8539	CA	ALA	A	567	18.251	-3.791	67.594	1.00	15.11	0.172	C
AT036	8540	C	ALA	A	567	17.833	-5.263	67.351	1.00	18.09	0.240	C
AT037	8541	CB	ALA	A	567	18.458	-6.871	68.436	1.00	13.84	-0.271	OA
AT038	8542	CB	ALA	A	567	17.030	-2.871	67.220	1.00	16.50	0.042	C
AT039	8543	N	YR	A	568	17.653	-5.910	66.198	1.00	12.23	-0.346	N
AT040	8544	HN	YR	A	568	17.753	-6.389	65.867	1.00	12.00	-0.183	HD
AT041	8545	CA	YR	A	568	17.322	-7.323	66.124	1.00	11.56	0.180	C
AT042	8546	C	YR	A	568	18.459	-8.159	66.740	1.00	13.08	0.241	C
AT043	8547	O	YR	A	568	18.188	-9.213	67.124	1.00	12.35	-0.183	HD
AT044	8548	CB	YR	A	568	17.113	-7.680	64.630	1.00	10.73	0.073	C
AT045	8549	CG	YR	A	568	17.295	-9.129	64.268	1.00	11.48	-0.056	A
AT046	8550	CG1	YR	A	568	18.545	-9.622	64.583	1.00	15.68	0.042	C
AT047	8551	CG2	YR	A	568	16.190	-10.003	63.283	1.00	11.13	0.010	A
AT048	8552	HR	YR	A	568	18.700	-10.962	63.580	1.00	19.33	0.037	A
AT049	8553	HR	YR	A	568	17.359	-11.338	63.580	1.00	15.68	0.042	C
AT050	8554	CZ	YR	A	568	17.603	-11.792	63.168	1.00	17.65	0.065	A
AT051	8555	OH	YR	A	568	17.764	-13.130	63.313	1.00	19.52	-0.361	OA
AT052	8556	HN	GLD	A	569	18.620	-13.444	63.658	1.00	16.80	-0.271	OA
AT053	8557	N	GLD	A	569	19.706	-7.793	66.432	1.00	12.50	-0.346	N
AT054	8558	HN	GLD	A	569	19.872	-6.993	65.822	1.00	9.00	0.163	HD
AT055	8559	CA	GLD	A	569	20.833	-8.859	65.978	1.00	16.80	-0.271	OA
AT056	8560	C	GLD	A	569	20.847	-8.435	68.505	1.00	17.76	0.241	C
AT057	8561	O	GLD	A	569	21.121	-9.469	69.127	1.00	18.66	-0.271	OA
AT058	8562	CB	GLD	A	569	22.164	-10.026	69.450	1.00	19.20	0.042	C
AT059	8563	CG	GLD	A	569	22.426	-8.200	64.984	1.00	31.34	0.116	C
AT060	8564	CG1	GLD	A	569	23.729	-7.703	64.942	1.00	50.17	0.172	C
AT061	8565	CG2	GLD	A	569	24.328	-6.888	64.942	1.00	46.46	-0.648	OA
AT062	8566	OE	GLD	A	569	24.171	-8.023	63.355	1.00	49.56	-0.648	OA
AT063	8567	N	LYS	A	570	20.551	-7.262	69.068	1.00	13.54	-0.346	N
AT064	8568	HN	LYS	A	570	20.313	-6.452	68.497	1.00	12.80	-0.271	OA
AT065	8569	CA	LYS	A	570	20.577	-7.174	70.543	1.00	16.64	0.176	C
AT066	8570	C	LYS	A	570	19.456	-7.948	71.202	1.00	21.25	0.241	C
AT067	8571	O	LYS	A	570	19.600	-8.909	72.018	1.00	21.71	-0.271	OA
AT068	8572	CB	LYS	A	570	20.446	-5.714	71.005	1.00	19.74	0.035	C
AT069	8573	CG	LYS	A	570	21.554	-4.825	70.487	1.00	30.58	0.004	C
AT070	8574	CG1	LYS	A	570	22.800	-4.901	71.356	1.00	30.58	0.004	C
AT071	8575	CG2	LYS	A	570	23.465	-3.527	71.426	1.00	53.36	0.229	C
AT072	8576	OE	LYS	A	570	22.599	-2.565	72.165	1.00	65.21	-0.079	N
AT073	8577	H21	LYS	A	570	23.048	-6.649	69.068	1.00	9.00	-0.271	OA
AT074	8578	H22	LYS	A	570	22.350	-2.913	73.091	1.00	0.00	0.00	0.00
AT075	8579	H23	LYS	A	570	21.662	-2.513	71.765	1.00	0.00	0.274	HD
AT076	8580	N	LEU	A	571	18.233	-7.862	71.137	1.00	13.60	-0.180	N
AT077	8581	HN	LEU	A	571	18.140	-7.323	69.749	1.00	0.00	0.163	HD
AT078	8582	CA	LEU	A	571	17.115	-8.546	71.191	1.00	12.28	0.177	C
AT079	8583	C	LEU	A	571	17.241	-10.062	71.191	1.00	15.65	0.241	C
AT080	8584	O	LEU	A	571	16.795	-10.797	71.943	1.00	17.66	-0.271	OA
AT081	8585	CB	LEU	A	571	18.450	-8.200	70.583	1.00	15.63	0.042	C
AT082	8586	CG	LEU	A	571	15.349	-6.657	70.916	1.00	25.82	-0.240	C
AT083	8587	CG1	LEU	A	571	14.098	-6.329	70.102	1.00	30.32	-0.009	C
AT084	8588	CG2	LEU	A	571	15.045	-6.436	70.102	1.00	27.80	0.042	C
AT085	8589	N	HR	A	572	17.725	-10.539	69.909	1.00	13.21	-0.346	N
AT086	8590	HN	HR	A	572	17.983	-9.882	69.173	1.00	0.00	0.163	HD
AT087	8591	CA	HR	A	572	18.914	-9.714	69.878	1.00	13.16	0.042	C
AT088	8592	C	HR	A	572	18.890	-12.507	70.758	1.00	17.64	0.243	C
AT089	8593	O	HR	A	572	18.717	-13.609	71.278	1.00	14.64	-0.271	OA
AT090	8594	CB	HR	A	572	18.458	-12.266	70.102	1.00	12.12	0.042	C
AT091	8595	CG	HR	A	572	19.111	-11.507	68.870	1.00	27.39	0.042	C
AT092	8596	CG1	HR	A	572	17.473	-11.868	67.304	1.00	21.69	-0.393	OA
AT093	8597	CG2	HR	A	572	18.200	-12.507	71.120	1.00	21.69	-0.393	OA
AT094	8598	N	ALA	A	573	19.925	-11.777	71.120	1.00	15.29	-0.346	N
AT095	8599	HN	ALA	A	573	20.075	-10.887	70.645	1.00	0		

ATOM 9071	CO1	VAL	A	600	6.815	-9.341	55.134	1.00	14.04	0.012	C
ATOM 9072	CG2	VAL	A	600	4.234	-10.447	57.267	1.00	7.75	-0.346	N
ATOM 9079	N	LEU	A	601	3.410	-10.701	56.501	1.00	0.00	0.163	HD
ATOM 9080	HN	LEU	A	601	-0.929	-9.727	57.049	1.00	0.00	0.000	C
ATOM 9081	CA	LEU	A	601	2.209	-10.495	58.891	1.00	8.51	0.243	C
ATOM 9083	C	LEU	A	601	1.057	-10.166	58.647	1.00	9.65	-0.271	DA
ATOM 9084	O	LEU	A	601	2.939	-8.396	57.138	1.00	0.00	0.163	HD
ATOM 9085	CB	LEU	A	601	4.097	-7.337	57.618	1.00	17.98	-0.020	C
ATOM 9088	CG	LEU	A	601	3.582	-5.963	57.149	1.00	14.72	0.009	C
ATOM 9090	CD1	LEU	A	601	5.010	-4.799	58.827	1.00	16.72	0.009	C
ATOM 9094	CD2	LEU	A	601	2.423	-11.591	59.615	1.00	9.14	-0.337	N
ATOM 9098	N	PRO	A	602	1.345	-12.421	60.092	1.00	12.08	0.179	C
ATOM 9099	CA	PRO	A	602	0.370	-11.606	60.926	1.00	10.76	0.000	C
ATOM 9101	C	PRO	A	602	0.756	-10.939	61.891	1.00	11.49	-0.271	DA
ATOM 9102	O	PRO	A	602	2.029	-13.532	60.902	1.00	12.32	0.037	C
ATOM 9103	CB	PRO	A	602	3.450	-12.007	60.976	1.00	12.63	0.000	C
ATOM 9109	CD	PRO	A	602	3.764	-12.165	59.929	1.00	13.02	0.127	C
ATOM 9112	N	LYS	A	603	-0.927	-11.826	60.686	1.00	12.41	-0.346	N
ATOM 9113	HN	LYS	A	603	-1.929	-12.941	60.900	1.00	0.00	0.163	HD
ATOM 9114	CA	LYS	A	603	-1.926	-11.024	61.410	1.00	16.35	0.176	C
ATOM 9116	C	LYS	A	603	-1.916	-11.396	62.896	1.00	14.74	0.241	C
ATOM 9117	O	LYS	A	603	-2.239	-10.460	63.156	1.00	16.51	-0.271	DA
ATOM 9118	CB	LYS	A	603	-3.284	-11.222	60.761	1.00	23.85	0.035	C
ATOM 9121	CG	LYS	A	603	-4.254	-12.143	61.478	1.00	36.12	0.004	C
ATOM 9124	CD1	LYS	A	603	-5.466	-12.007	60.876	1.00	52.03	0.037	C
ATOM 9127	CD2	LYS	A	603	-6.484	-11.052	61.048	1.00	58.66	0.229	C
ATOM 9130	N2	LYS	A	603	-6.632	-10.069	59.937	1.00	66.03	-0.079	N
ATOM 9131	H21	LYS	A	603	-7.243	-9.946	60.878	1.00	0.00	0.274	HD
ATOM 9132	H22	LYS	A	603	-6.941	-10.526	59.079	1.00	0.00	0.274	HD
ATOM 9133	H23	LYS	A	603	-5.425	-9.727	59.079	1.00	0.00	0.274	HD
ATOM 9134	N	ALA	A	604	-1.531	-12.556	63.268	1.00	13.72	-0.346	N
ATOM 9135	HN	ALA	A	604	-1.262	-13.254	62.574	1.00	0.00	0.163	HD
ATOM 9136	CA	ALA	A	604	2.303	-11.024	63.156	1.00	17.21	0.000	C
ATOM 9138	C	ALA	A	604	-0.407	-12.127	65.471	1.00	20.98	-0.240	C
ATOM 9139	O	ALA	A	604	-0.485	-12.094	66.723	1.00	17.28	0.271	DA
ATOM 9140	CB	ALA	A	604	1.710	-11.826	66.723	1.00	21.88	0.000	C
ATOM 9144	N	VAL	A	605	0.626	-11.620	64.800	1.00	10.94	-0.346	N
ATOM 9145	HN	VAL	A	605	0.668	-11.689	63.783	1.00	0.00	0.163	HD
ATOM 9146	CA	VAL	A	605	1.939	-9.486	64.589	1.00	14.07	0.000	C
ATOM 9148	C	VAL	A	605	1.391	-9.486	65.620	1.00	16.61	0.241	C
ATOM 9149	O	VAL	A	605	1.491	-8.794	64.589	1.00	16.70	-0.271	DA
ATOM 9150	CB	VAL	A	605	3.062	-11.188	64.589	1.00	12.47	0.000	C
ATOM 9152	CG1	VAL	A	605	4.212	-10.525	65.614	1.00	16.89	0.012	C
ATOM 9156	CG2	VAL	A	605	3.388	-12.698	64.741	1.00	20.23	0.012	C
ATOM 9160	N	THR	A	606	1.000	-9.440	63.156	1.00	12.00	0.000	C
ATOM 9161	HN	THR	A	606	1.050	-9.561	66.802	1.00	0.00	0.163	HD
ATOM 9162	CA	THR	A	606	0.732	-7.538	66.842	1.00	16.41	0.205	C
ATOM 9164	C	THR	A	606	-1.827	-6.667	67.414	1.00	18.89	-0.000	C
ATOM 9165	O	THR	A	606	1.692	-5.452	67.424	1.00	14.06	-0.271	DA
ATOM 9166	CB	THR	A	606	-0.545	-7.330	67.694	1.00	17.52	0.146	C
ATOM 9168	CG2	THR	A	606	-1.722	-6.202	67.694	1.00	22.63	0.000	C
ATOM 9172	CG1	THR	A	606	-0.248	-7.891	68.972	1.00	18.91	-0.393	DA
ATOM 9173	H01	THR	A	606	-1.031	-7.763	69.494	1.00	0.00	0.210	HD
ATOM 9174	N	ARG	A	607	-0.929	-7.287	69.494	1.00	0.00	0.163	HD
ATOM 9175	HN	ARG	A	607	3.007	-8.300	67.884	1.00	0.00	0.163	HD
ATOM 9176	CA	ARG	A	607	4.012	-6.458	68.475	1.00	13.72	0.172	C
ATOM 9178	C	ARG	A	607	4.879	-6.049	69.284	1.00	13.87	0.000	C
ATOM 9179	O	ARG	A	607	5.739	-6.807	66.860	1.00	12.08	-0.271	DA
ATOM 9180	CB	ARG	A	607	4.840	-7.220	69.502	1.00	14.27	0.042	C
ATOM 9184	N	ARG	A	608	3.828	-4.490	69.502	1.00	15.56	-0.346	N
ATOM 9185	HN	ARG	A	608	3.828	-4.490	67.079	1.00	0.00	0.163	HD
ATOM 9186	CA	ARG	A	608	5.263	-4.565	65.484	1.00	8.65	0.176	C
ATOM 9188	C	ARG	A	608	4.890	-3.069	69.502	1.00	15.94	0.000	C
ATOM 9189	O	ARG	A	608	5.225	-2.257	66.323	1.00	12.83	-0.271	DA
ATOM 9190	CB	ARG	A	608	4.272	-4.314	64.308	1.00	11.81	0.036	C
ATOM 9193	CG	ARG	A	608	12.470	-5.492	62.800	1.00	12.83	0.000	C
ATOM 9196	CD	ARG	A	608	2.616	-5.492	62.800	1.00	14.32	0.138	C
ATOM 9199	NE	ARG	A	608	1.909	-6.783	62.553	1.00	12.30	-0.227	N
ATOM 9200	HE	ARG	A	608	12.029	-5.492	63.156	1.00	0.00	0.163	HD
ATOM 9201	C	ARG	A	608	1.135	-6.941	61.492	1.00	22.08	0.065	C
ATOM 9202	NH1	ARG	A	608	0.986	-5.931	60.636	1.00	8.44	-0.225	N
ATOM 9203	NH2	ARG	A	608	1.415	-6.202	60.636	1.00	0.00	0.163	HD
ATOM 9204	NH3	ARG	A	608	0.389	-6.053	59.818	1.00	0.00	0.174	HD
ATOM 9205	NH4	ARG	A	608	0.578	-8.129	61.364	1.00	11.66	-0.235	N
ATOM 9206	NH5	ARG	A	608	1.430	-8.906	62.800	1.00	0.00	0.163	HD
ATOM 9207	NH6	ARG	A	608	-0.019	-8.251	60.546	1.00	0.00	0.174	HD
ATOM 9208	N	VAL	A	609	7.143	-2.895	65.244	1.00	10.12	-0.346	N
ATOM 9209	HN	VAL	A	609	7.462	-2.652	69.502	1.00	0.00	0.163	HD
ATOM 9210	CA	VAL	A	609	7.817	-1.602	65.453	1.00	8.93	0.180	C
ATOM 9212	C	VAL	A	609	8.302	-1.114	64.308	1.00	0.00	0.163	HD
ATOM 9213	O	VAL	A	609	9.053	-1.737	63.386	1.00	10.04	-0.271	DA
ATOM 9214	CB	VAL	A	609	9.082	-1.765	66.319	1.00	11.13	0.009	C
ATOM 9216	CG1	VAL	A	609	8.753	-0.417	67.694	1.00	16.84	-0.000	C
ATOM 9220	CG2	VAL	A	609	8.776	-2.394	67.694	1.00	10.86	0.012	C
ATOM 9224	N	ALA	A	610	7.797	0.010	63.690	1.00	9.86	-0.346	N
ATOM 9225	HN	ALA	A	610	7.243	-0.612	63.690	1.00	0.00	0.163	HD
ATOM 9226	CA	ALA	A	610	8.243	0.697	62.429	1.00	9.88	0.172	C
ATOM 9228	C	ALA	A	610	9.370	1.709	62.692	1.00	8.84	0.240	C
ATOM 9229	O	ALA	A	610	8.243	2.424	63.156	1.00	11.19	-0.271	DA
ATOM 9230	CB	ALA	A	610	7.046	1.507	61.861	1.00	10.52	0.042	C
ATOM 9234	N	VAL	A	611	10.412	1.695	61.888	1.00	6.48	-0.346	N
ATOM 9235	HN	VAL	A	611	12.505	2.691	63.156	1.00	0.00	0.163	HD
ATOM 9236	CA	VAL	A	611	11.545	2.564	62.055	1.00	6.87	0.180	C
ATOM 9238	C	VAL	A	611	11.800	3.316	60.738	1.00	7.50	0.241	C
ATOM 9239	O	VAL	A	611	12.825	2.691	62.429	1.00	9.84	0.009	C
ATOM 9240	CB	VAL	A	611	12.825	1.827	62.451	1.00	9.84	0.009	C
ATOM 9242	CG1	VAL	A	611	13.819	2.911	62.950	1.00	12.04	0.012	C
ATOM 9246	CG2	VAL	A	611	12.825	2.691	62.429	1.00	9.84	0.009	C
ATOM 9250	N	GLU	A	612	11.660	4.614	60.734	1.00	7.26	-0.346	N
ATOM 9251	HN	GLU	A	612	11.395	5.104	61.588	1.00	0.00	0.163	HD
ATOM 9252	CA	GLU	A	612	11.888	3.369	59.866	1.00	8.83	0.000	C
ATOM 9254	C	GLU	A	612	12.110	6.838	59.866	1.00	7.69	0.241	C
ATOM 9255	O	GLU	A	612	11.295	7.394	60.620	1.00	8.75	-0.271	DA
ATOM 9256	CB	GLU	A	612	10.460	8.910	60.620	1.00	0.00	0.163	HD
ATOM 9259	CG	GLU	A	612	10.829	6.221	57.295	1.00	4.90	0.176	C
ATOM 9262	CD	GLU	A	612	9.710	5.969	56.286	1.00	9.29	0.172	C
ATOM 9263	OE1	GLU	A	612	13.589	2.266	61.529	1.00	9.26	0.000	C
ATOM 9264	OE2	GLU	A	612	9.964	5.162	55.369	1.00	11.13	-0.648	DA
ATOM 9265	N	ALA	A	613	13.080	7.518	59.231	1.00	5.56	-0.346	N
ATOM 9266	HN	ALA	A	613	13.685	7.947	58.598	1.00	0.00	0.163	HD
ATOM 9267	CA	ALA	A	613	13.256	8.935	59.516	1.00	8.35	0.172	C
ATOM 9269	C	ALA	A	613	12.258	9.793	58.671	1.00	10.34	0.240	C
ATOM 9270	O	ALA	A	613	12.450	8.840	59.727	1.00	9.99	-0.271	DA
ATOM 9271	CB	ALA	A	613	14.680	9.361	59.248	1.00	8.08	0.042	C
ATOM 9275	N	GLY	A	614	11.011	9.670	59.013	1.00	9.83	-0.351	N
ATOM 9276	HN	GLY	A	614	10.776	9.940	59.780	1.00	0.00	0.163	HD
ATOM 9277	CA	GLY	A	614	9.909	10.470	58.337	1.00			

ATOM 9659 O	PRO A	640	24.251	16.037	62.376	1.00	13.50	-0.271	OA
ATOM 9660 CN	ERG A	644	25.157	17.167	62.363	1.00	18.17	0.022	C
ATOM 9663 CG	PRO A	640	27.553	18.562	62.363	1.00	18.17	0.022	C
ATOM 9666 CD	PRO A	640	26.246	19.176	62.800	1.00	16.57	0.127	C
ATOM 9669 N	ALA A	641	24.755	19.963	61.401	1.00	14.63	0.143	C
ATOM 9670 HN	ALA A	641	25.337	15.548	59.454	1.00	0.00	0.163	HD
ATOM 9671 CA	ALA A	641	23.780	14.313	60.324	1.00	16.60	0.172	C
ATOM 9673 C	ALA A	641	23.927	13.994	61.401	1.00	15.47	0.172	C
ATOM 9674 O	ALA A	641	22.932	12.964	62.096	1.00	17.29	-0.271	OA
ATOM 9675 CB	ALA A	641	23.839	13.508	59.019	1.00	18.97	0.042	C
ATOM 9679 N	ALA A	641	25.156	12.976	61.936	1.00	15.48	-0.346	N
ATOM 9680 HN	ALA A	642	25.976	13.296	61.324	1.00	0.00	0.163	HD
ATOM 9681 CA	GLU A	642	25.284	12.027	62.979	1.00	20.34	0.177	C
ATOM 9683 C	GLU A	642	24.638	12.447	64.249	1.00	18.20	0.185	C
ATOM 9684 O	GLU A	642	24.000	11.801	65.027	1.00	19.05	-0.271	OA
ATOM 9685 CB	GLU A	642	26.763	11.690	63.222	1.00	30.97	0.045	C
ATOM 9688 CG	GLU A	642	28.704	11.085	66.961	1.00	15.49	0.019	C
ATOM 9691 CD	GLU A	642	28.521	11.023	64.993	1.00	80.68	0.172	C
ATOM 9692 OE1	GLU A	642	29.480	11.037	64.187	1.00	88.22	-0.648	OA
ATOM 9693 OE2	GLU A	642	28.754	11.085	66.961	1.00	15.49	0.019	C
ATOM 9694 N	LEU A	643	24.796	13.844	64.529	1.00	11.42	-0.346	N
ATOM 9695 HN	LEU A	643	25.344	14.443	63.911	1.00	0.00	0.163	HD
ATOM 9696 CA	LEU A	643	24.182	14.396	65.727	1.00	11.41	0.177	C
ATOM 9698 C	LEU A	643	22.676	14.479	65.616	1.00	16.29	0.241	C
ATOM 9699 O	LEU A	643	22.003	14.399	66.839	1.00	14.17	-0.271	OA
ATOM 9700 CB	LEU A	643	24.715	15.842	65.937	1.00	15.20	0.052	C
ATOM 9703 CG	LEU A	643	26.217	15.978	66.206	1.00	22.96	-0.020	C
ATOM 9705 CD1	LEU A	643	26.560	14.400	66.648	1.00	29.45	0.009	C
ATOM 9709 CD2	LEU A	643	26.600	15.023	67.296	1.00	30.96	0.009	C
ATOM 9713 N	LEU A	644	22.105	14.779	64.412	1.00	12.35	-0.346	N
ATOM 9714 HN	LEU A	644	24.607	14.774	63.573	1.00	15.48	-0.346	N
ATOM 9715 CA	LEU A	644	20.637	14.757	64.334	1.00	10.70	0.177	C
ATOM 9717 C	LEU A	644	20.021	13.388	64.575	1.00	10.65	0.241	C
ATOM 9718 O	LEU A	644	19.153	13.483	65.143	1.00	11.71	-0.271	OA
ATOM 9719 CB	LEU A	644	20.212	15.302	62.953	1.00	11.88	0.038	C
ATOM 9722 CG	LEU A	644	20.683	16.736	62.647	1.00	12.62	-0.020	C
ATOM 9724 CD1	LEU A	644	21.129	17.433	63.519	1.00	11.34	0.009	C
ATOM 9728 CD2	LEU A	644	19.993	17.139	63.624	1.00	12.08	0.009	C
ATOM 9732 N	PHE A	645	20.675	12.324	64.036	1.00	12.01	-0.346	N
ATOM 9733 HN	PHE A	645	21.543	12.413	63.519	1.00	11.34	0.009	C
ATOM 9734 CA	PHE A	645	20.051	10.993	64.260	1.00	11.32	0.180	C
ATOM 9736 C	PHE A	645	19.984	10.692	65.758	1.00	12.17	0.241	C
ATOM 9737 O	PHE A	645	18.957	10.444	66.919	1.00	13.86	-0.271	OA
ATOM 9738 CB	PHE A	645	20.860	9.920	63.534	1.00	16.21	0.073	C
ATOM 9741 CG	PHE A	645	20.532	9.851	62.061	1.00	20.91	-0.056	A
ATOM 9742 CD1	PHE A	645	19.120	9.566	65.758	1.00	16.43	0.007	A
ATOM 9744 CD2	PHE A	645	21.520	9.984	61.104	1.00	24.81	0.007	A
ATOM 9746 CE1	PHE A	645	18.921	9.595	60.285	1.00	23.80	0.001	A
ATOM 9748 CE2	PHE A	645	20.622	9.566	65.758	1.00	16.43	0.007	A
ATOM 9750 CZ	PHE A	645	19.904	9.745	59.344	1.00	22.14	0.000	A
ATOM 9752 N	GLU A	646	21.035	11.902	66.457	1.00	13.59	-0.346	N
ATOM 9753 HN	GLU A	646	21.833	11.677	65.985	1.00	12.01	0.007	A
ATOM 9754 CA	GLU A	646	21.046	10.919	67.923	1.00	18.84	0.177	C
ATOM 9756 C	GLU A	646	19.985	11.747	68.594	1.00	19.01	0.241	C
ATOM 9757 O	GLU A	646	18.163	11.288	69.154	1.00	21.81	-0.271	OA
ATOM 9758 CB	GLU A	646	22.431	11.325	68.449	1.00	26.62	0.045	C
ATOM 9761 CG	GLU A	646	23.094	10.388	69.436	1.00	50.81	0.116	C
ATOM 9764 CD	GLU A	646	24.600	10.600	69.374	1.00	64.48	0.045	C
ATOM 9765 OE1	GLU A	646	25.025	11.731	69.714	1.00	57.86	-0.648	OA
ATOM 9766 OE2	GLU A	646	25.310	9.657	68.966	1.00	61.63	-0.648	OA
ATOM 9767 N	GLU A	646	19.933	10.446	66.465	1.00	16.50	-0.346	N
ATOM 9768 HN	GLU A	647	20.605	13.424	67.621	1.00	0.00	0.163	HD
ATOM 9769 CA	GLU A	647	18.943	13.945	68.883	1.00	20.20	0.177	C
ATOM 9771 C	GLU A	647	19.500	13.500	69.500	1.00	21.30	0.241	C
ATOM 9772 O	GLU A	647	16.635	13.734	69.543	1.00	18.30	-0.271	OA
ATOM 9773 CB	GLU A	647	19.143	15.305	68.166	1.00	24.68	0.045	C
ATOM 9776 CG	GLU A	647	18.300	16.933	68.166	1.00	18.30	0.045	C
ATOM 9779 CD	GLU A	647	18.985	17.201	69.807	1.00	46.20	0.172	C
ATOM 9780 OE1	GLU A	647	19.976	16.742	70.408	1.00	47.50	-0.648	OA
ATOM 9781 OE2	GLU A	647	19.468	18.939	69.444	1.00	46.44	0.045	C
ATOM 9782 N	PHE A	648	17.173	12.887	67.544	1.00	13.41	-0.346	N
ATOM 9783 HN	PHE A	648	17.910	12.626	66.889	1.00	0.00	0.163	HD
ATOM 9784 CA	PHE A	648	15.788	12.859	67.923	1.00	15.33	0.177	C
ATOM 9786 C	PHE A	648	15.445	11.104	67.519	1.00	17.37	0.241	C
ATOM 9787 O	PHE A	648	14.371	10.628	67.134	1.00	19.68	-0.271	OA
ATOM 9788 CB	PHE A	648	15.433	12.901	66.923	1.00	16.57	0.045	C
ATOM 9791 CG	PHE A	648	15.287	14.390	65.042	1.00	15.55	-0.056	A
ATOM 9792 CD1	PHE A	648	14.241	15.124	66.547	1.00	18.54	0.007	A
ATOM 9794 CD2	PHE A	648	16.285	15.036	66.040	1.00	15.16	0.007	A
ATOM 9796 CE1	PHE A	648	14.174	16.502	65.836	1.00	20.55	0.001	A
ATOM 9798 CE2	PHE A	648	16.280	16.396	65.836	1.00	19.66	0.001	A
ATOM 9800 CZ	PHE A	648	15.157	17.140	65.075	1.00	21.41	0.000	A
ATOM 9802 N	GLY A	649	16.358	10.373	68.162	1.00	19.72	-0.351	N
ATOM 9803 HN	GLY A	649	17.213	10.981	68.154	1.00	18.84	0.163	HD
ATOM 9804 CA	GLY A	649	16.088	9.106	68.596	1.00	16.29	0.225	C
ATOM 9807 C	GLY A	649	16.349	7.862	67.665	1.00	12.71	0.236	C
ATOM 9808 O	GLY A	649	15.911	7.113	67.957	1.00	14.24	-0.271	OA
ATOM 9809 N	PHE A	650	17.106	8.067	66.621	1.00	11.40	-0.346	N
ATOM 9810 HN	PHE A	650	17.474	8.995	66.465	1.00	0.00	0.163	HD
ATOM 9811 CA	PHE A	650	15.815	8.022	68.470	1.00	16.80	0.177	C
ATOM 9813 C	PHE A	650	18.765	6.421	66.179	1.00	13.37	0.241	C
ATOM 9814 O	PHE A	650	19.811	6.618	65.606	1.00	12.47	-0.271	OA
ATOM 9815 CB	PHE A	650	16.141	7.976	63.810	1.00	11.21	0.007	A
ATOM 9818 CG	PHE A	650	16.141	7.976	63.810	1.00	11.21	-0.056	A
ATOM 9819 CD1	PHE A	650	15.194	7.040	63.374	1.00	13.18	0.007	A
ATOM 9821 CD2	PHE A	650	15.194	7.040	63.374	1.00	13.18	0.007	A
ATOM 9823 CE1	PHE A	650	13.911	7.460	63.008	1.00	9.90	0.001	A
ATOM 9825 CE2	PHE A	650	14.523	9.735	63.526	1.00	13.85	0.001	A
ATOM 9827 CZ	PHE A	650	13.594	8.066	63.119	1.00	11.00	0.007	A
ATOM 9829 N	THR A	651	18.663	5.705	62.321	1.00	16.11	-0.346	N
ATOM 9830 HN	THR A	651	17.758	5.633	67.785	1.00	0.00	0.163	HD
ATOM 9831 CA	THR A	651	19.812	6.027	67.062	1.00	45.41	0.177	C
ATOM 9833 C	THR A	651	19.348	6.039	68.243	1.00	12.40	0.243	C
ATOM 9834 O	THR A	651	18.175	3.376	68.544	1.00	15.03	-0.271	OA
ATOM 9835 CB	THR A	651	21.333	6.039	67.062	1.00	45.41	0.177	C
ATOM 9837 CG2	THR A	651	20.671	7.159	69.071	1.00	17.61	0.042	C
ATOM 9841 CD1	THR A	651	19.277	5.601	70.173	1.00	15.28	-0.393	OA
ATOM 9842 H01	THR A	651	19.939	5.988	70.978	1.00	16.00	-0.393	OA
ATOM 9843 N	VAL A	652	20.292	2.881	68.188	1.00	15.16	-0.346	N
ATOM 9844 HN	VAL A	652	21.237	2.938	67.902	1.00	0.00	0.163	HD
ATOM 9845 CA	VAL A	652							

Table with columns for ID, State, Agency, Type, Value 1, Value 2, Value 3, Value 4, Value 5, Value 6, Value 7, Value 8, Value 9, Value 10, Value 11, Value 12, Value 13, Value 14, Value 15, Value 16, Value 17, Value 18, Value 19, Value 20, Value 21, Value 22, Value 23, Value 24, Value 25, Value 26, Value 27, Value 28, Value 29, Value 30, Value 31, Value 32, Value 33, Value 34, Value 35, Value 36, Value 37, Value 38, Value 39, Value 40, Value 41, Value 42, Value 43, Value 44, Value 45, Value 46, Value 47, Value 48, Value 49, Value 50, Value 51, Value 52, Value 53, Value 54, Value 55, Value 56, Value 57, Value 58, Value 59, Value 60, Value 61, Value 62, Value 63, Value 64, Value 65, Value 66, Value 67, Value 68, Value 69, Value 70, Value 71, Value 72, Value 73, Value 74, Value 75, Value 76, Value 77, Value 78, Value 79, Value 80, Value 81, Value 82, Value 83, Value 84, Value 85, Value 86, Value 87, Value 88, Value 89, Value 90, Value 91, Value 92, Value 93, Value 94, Value 95, Value 96, Value 97, Value 98, Value 99, Value 100.

ATOM 10821 CA SER B 53	11.043	44.555	66.777	1.00	14.78	0.200	C	
ATOM 10822 C SER B 53	10.529	37.951	56.140	1.00	14.78	-0.273	CA	
ATOM 10823 O SER B 53	9.316	46.079	66.390	1.00	14.79	-0.271	OA	
ATOM 10824 O SER B 53	10.329	44.127	68.096	1.00	18.86	0.199	C	
ATOM 10825 CG SER B 53	12.406	42.029	66.390	1.00	14.79	-0.273	CA	
ATOM 10826 HG SER B 53	10.083	42.463	68.907	1.00	0.00	0.209	HD	
ATOM 10830 N TRP B 54	11.401	46.969	66.400	1.00	10.52	-0.346	N	
ATOM 10831 HN TRP B 54	12.406	46.039	66.390	1.00	14.79	-0.273	CA	
ATOM 10832 CA TRP B 54	10.847	48.323	66.153	1.00	10.16	0.181	C	
ATOM 10833 C TRP B 54	9.888	48.269	64.973	1.00	8.83	0.241	C	
ATOM 10835 O TRP B 54	10.242	47.827	63.841	1.00	11.69	-0.271	OA	
ATOM 10836 CB TRP B 54	12.032	49.259	65.912	1.00	10.36	0.075	C	
ATOM 10839 CG TRP B 54	11.688	50.616	65.401	1.00	11.03	-0.028	A	
ATOM 10840 CD1 TRP B 54	12.586	51.363	65.792	1.00	10.37	0.096	A	
ATOM 10842 CD2 TRP B 54	12.524	51.427	64.580	1.00	8.52	-0.002	A	
ATOM 10843 CE2 TRP B 54	11.843	52.647	64.382	1.00	7.75	0.042	A	
ATOM 10844 CE3 TRP B 54	12.533	51.330	65.792	1.00	10.37	0.096	A	
ATOM 10846 NE1 TRP B 54	10.674	52.597	65.074	1.00	9.86	-0.365	N	
ATOM 10847 HE1 TRP B 54	9.973	53.337	65.109	1.00	0.00	0.165	HD	
ATOM 10848 CD2 TRP B 54	12.533	51.330	65.792	1.00	10.37	0.096	A	
ATOM 10850 CE3 TRP B 54	14.305	52.251	63.213	1.00	12.67	0.001	A	
ATOM 10852 CH2 TRP B 54	13.603	53.472	63.024	1.00	12.01	0.002	A	
ATOM 10854 N ALA B 55	10.470	48.796	65.196	1.00	9.06	-0.346	N	
ATOM 10855 HN ALA B 55	8.496	49.316	66.054	1.00	0.00	0.163	HD	
ATOM 10856 CA ALA B 55	7.587	48.834	64.215	1.00	10.38	0.172	C	
ATOM 10857 C ALA B 55	8.882	49.288	62.998	1.00	12.38	0.290	C	
ATOM 10859 O ALA B 55	7.428	48.772	61.829	1.00	11.82	-0.271	OA	
ATOM 10860 CB ALA B 55	6.294	49.248	64.770	1.00	10.00	0.042	C	
ATOM 10864 N SER B 56	10.836	30.435	62.924	1.00	8.44	-0.346	N	
ATOM 10865 HN SER B 56	8.964	50.793	63.832	1.00	0.00	0.163	HD	
ATOM 10866 CA SER B 56	8.017	51.144	61.674	1.00	11.65	0.241	C	
ATOM 10868 C SER B 56	10.142	50.865	61.039	1.00	11.47	0.241	C	
ATOM 10869 O SER B 56	10.514	51.634	60.122	1.00	9.72	-0.271	OA	
ATOM 10870 CB SER B 56	8.147	51.647	61.194	1.00	11.58	0.241	C	
ATOM 10873 CG SER B 56	8.189	53.378	60.603	1.00	14.01	0.175	C	
ATOM 10874 OD1 ASP B 56	7.480	52.814	59.729	1.00	10.22	-0.648	OA	
ATOM 10875 OD2 ASP B 56	8.618	51.566	60.439	1.00	12.89	0.444	CA	
ATOM 10876 N ARG B 57	10.895	49.819	61.391	1.00	10.03	-0.346	N	
ATOM 10877 HN ARG B 57	10.575	49.201	62.137	1.00	0.00	0.163	HD	
ATOM 10878 CA ARG B 57	11.147	49.543	60.782	1.00	9.94	0.241	C	
ATOM 10880 C ARG B 57	11.983	49.697	59.238	1.00	10.52	0.241	C	
ATOM 10881 O ARG B 57	10.898	48.898	58.828	1.00	9.82	-0.271	OA	
ATOM 10882 CB ARG B 57	12.074	49.244	60.782	1.00	7.41	0.023	C	
ATOM 10885 CG ARG B 57	12.079	47.027	61.159	1.00	7.41	0.023	C	
ATOM 10888 CD ARG B 57	12.568	45.922	62.107	1.00	11.61	0.138	C	
ATOM 10891 CE ARG B 57	11.896	44.632	61.039	1.00	8.54	0.241	C	
ATOM 10892 HE ARG B 57	12.365	43.976	61.259	1.00	0.00	0.177	HD	
ATOM 10893 CH ARG B 57	10.729	44.240	62.364	1.00	15.34	0.665	C	
ATOM 10894 NH1 ARG B 57	10.021	45.007	63.039	1.00	8.54	0.241	C	
ATOM 10895 NH1 ARG B 57	10.413	45.922	63.471	1.00	0.00	0.174	HD	
ATOM 10896 2NH1 ARG B 57	9.125	44.701	63.574	1.00	0.00	0.174	HD	
ATOM 10897 NH2 ARG B 57	10.211	43.030	62.137	1.00	11.71	0.138	C	
ATOM 10898 1H2 ARG B 57	10.753	42.443	61.434	1.00	0.00	0.174	HD	
ATOM 10899 2H2 ARG B 57	9.315	42.724	62.449	1.00	0.00	0.174	HD	
ATOM 10900 N ASP B 57	10.928	49.440	58.290	1.00	22.22	-0.346	N	
ATOM 10901 HN ASP B 58	13.775	50.007	57.968	1.00	0.00	0.163	HD	
ATOM 10902 CA ASP B 58	13.008	48.911	57.044	1.00	8.87	0.186	C	
ATOM 10904 C ASP B 58	13.083	47.378	57.120	1.00	13.89	-0.271	OA	
ATOM 10905 O ASP B 58	13.661	46.871	58.156	1.00	9.80	-0.271	OA	
ATOM 10906 CB ASP B 58	14.188	49.352	56.185	1.00	7.89	0.147	C	
ATOM 10909 CG ASP B 58	14.235	50.822	56.185	1.00	11.01	0.00	0.163	HD
ATOM 10910 OD1 ASP B 58	13.204	51.461	55.806	1.00	10.13	-0.648	OA	
ATOM 10911 OD2 ASP B 58	15.372	51.345	55.539	1.00	11.79	-0.648	OA	
ATOM 10912 N SER B 59	12.567	46.636	62.924	1.00	8.43	-0.346	N	
ATOM 10913 HN SER B 59	12.110	47.110	55.444	1.00	0.00	0.163	HD	
ATOM 10914 CA SER B 59	12.611	45.178	56.208	1.00	9.99	0.176	C	
ATOM 10916 C SER B 59	12.436	44.706	55.444	1.00	0.00	0.163	HD	
ATOM 10917 O SER B 59	13.576	45.180	53.995	1.00	14.96	-0.271	OA	
ATOM 10918 CB SER B 59	11.230	44.555	55.892	1.00	10.44	0.036	C	
ATOM 10921 CG SER B 59	13.345	44.555	55.892	1.00	10.44	0.036	C	
ATOM 10924 CD SER B 59	9.932	45.433	57.968	1.00	10.16	0.138	C	
ATOM 10925 CE SER B 59	8.962	44.995	59.031	1.00	11.55	-0.227	CA	
ATOM 10928 HE ARG B 59	12.079	47.027	61.159	1.00	7.41	0.023	C	
ATOM 10929 CH ARG B 59	8.364	45.851	59.830	1.00	16.08	0.665	C	
ATOM 10930 NH1 ARG B 59	8.634	47.147	59.764	1.00	11.54	0.241	C	
ATOM 10931 1H1 ARG B 59	10.329	47.827	63.841	1.00	11.69	-0.271	OA	
ATOM 10932 2H1 ARG B 59	8.170	47.811	60.384	1.00	0.00	0.174	HD	
ATOM 10933 NH2 ARG B 59	7.476	45.403	60.712	1.00	11.26	-0.235	CA	
ATOM 10934 1H2 ARG B 59	7.428	44.434	61.674	1.00	11.65	0.241	C	
ATOM 10935 2H2 ARG B 59	7.012	46.095	61.332	1.00	0.00	0.174	HD	
ATOM 10936 N TRP B 59	10.898	48.898	58.828	1.00	9.82	-0.271	OA	
ATOM 10937 HN TRP B 60	14.631	43.030	56.531	1.00	0.00	0.163	HD	
ATOM 10938 CA TRP B 60	15.526	43.347	54.563	1.00	6.88	0.180	C	
ATOM 10940 C TRP B 60	10.251	47.817	63.841	1.00	11.69	-0.271	OA	
ATOM 10941 O TRP B 60	15.094	41.168	55.412	1.00	9.54	-0.271	OA	
ATOM 10942 CB TRP B 60	16.969	43.512	55.127	1.00	8.83	0.073	C	
ATOM 10945 CG TRP B 60	16.969	43.512	55.127	1.00	8.83	0.073	C	
ATOM 10946 CD1 TRP B 60	18.364	43.419	53.038	1.00	16.64	0.007	A	
ATOM 10948 CD2 TRP B 60	18.597	41.639	54.643	1.00	10.36	0.001	A	
ATOM 10950 CE1 TRP B 60	19.524	41.045	53.818	1.00	19.82	0.007	A	
ATOM 10952 CE2 TRP B 60	19.524	41.045	53.818	1.00	19.82	0.007	A	
ATOM 10954 CH TRP B 60	19.881	41.605	52.593	1.00	14.09	0.000	A	
ATOM 10956 N TRP B 60	15.169	39.351	52.102	1.00	8.64	-0.346	N	
ATOM 10957 HN VAL B 61	15.473	41.940	52.376	1.00	0.00	0.163	HD	
ATOM 10958 CA VAL B 61	15.048	39.912	52.951	1.00	5.52	0.280	C	
ATOM 10960 C VAL B 61	15.048	39.912	52.951	1.00	5.52	0.280	C	
ATOM 10961 O VAL B 61	16.364	39.827	50.953	1.00	8.81	-0.271	OA	
ATOM 10962 CB VAL B 61	13.700	39.655	52.244	1.00	11.57	0.009	C	
ATOM 10964 CD1 VAL B 61	13.518	38.171	51.511	1.00	14.41	0.225	CA	
ATOM 10968 CD2 VAL B 61	12.516	40.060	53.182	1.00	8.80	0.012	C	
ATOM 10972 N LEU B 62	16.804	38.287	52.559	1.00	9.15	-0.346	N	
ATOM 10973 HN LEU B 62	16.588	37.264	53.178	1.00	0.00	0.163	HD	
ATOM 10974 CA LEU B 62	17.809	37.587	51.764	1.00	9.84	0.177	C	
ATOM 10976 C LEU B 62	17.120	36.433	51.041	1.00	11.71	0.241	C	
ATOM 10977 O LEU B 62	16.959	35.269	51.899	1.00	11.39	-0.271	OA	
ATOM 10978 CB LEU B 62	18.905	37.071	52.732	1.00	8.58	0.038	C	
ATOM 10981 CG LEU B 62	20.052	36.370	51.954	1.00	15.76	0.020	C	
ATOM 10983 CD1 LEU B 62	20.867	35.213	50.909	1.00	11.21	0.042	CA	
ATOM 10987 CD2 LEU B 62	20.942	35.570	52.901	1.00	12.82	0.009	C	
ATOM 10991 N SER B 63	16.665	36.632	49.795	1.00	10.01	-0.346	N	
ATOM 10992 HN SER B 63	16.866	37.931	49.795	1.00	10.01	-0.346	N	
ATOM 10993 CA SER B 63	15.886	35.600	49.115	1.00	7.91	0.200	C	
ATOM 10995 C SER B 63	16.767	34.441	48.652	1.00	13.42	0.243	C	
ATOM 10996 O SER B 63	16.333	33.980	48.575	1.00	13.33	-0.271	OA	
ATOM 10997 CB SER B 63	15.133	36.168	47.904	1.00	12.93	0.199	C	
ATOM 11000 CG SER B 63	14.075	35.310	47.505	1.00	12.40	-0.398	OA	
ATOM 11001 HG SER B 63	15.886	35.686	46.925	1.00	14.06	0.499	CA	
ATOM 11002 N ASN B 64	18.034	34.739	48.347	1.00	13.37	-0.345	N	
ATOM 11003 HN ASN B 64	18.332	35.718	48.390	1.00	0.00	0.163	HD	
ATOM 11004 CA ASN B 64	19.015	33.715	47.140	1.00	14.64	0.241	C	
ATOM 11006 C ASN B 64	19.652	33.278	49.289	1.00	12.63	0.241	C	
ATOM 11007 O ASN B 64	20.827	33.533	49.586	1.00	14.99	-0.271	OA	
ATOM 11008 CB ASN B 64	20.071	34.237	47.929	1.00	15.53	0.241	C	
ATOM 11011 CG ASN B 64	20.587	35.547	47.269	1.00	21.48	0.217	C	
ATOM 11012 HD2 ASN B 64	21.619	36.066	46.473	1.00	19.53	-0.370	CA	
ATOM 11013 HD1 ASN B 64	21.862	37.017	46.251	1.00	0.00	0.159	HD	
ATOM 11014 2HD2 ASN B 64	22.011	35.434	45.778	1.00	0.00	0.159	HD	
ATOM 11015 OD1 ASP B 67	20.117	36.417	51.000	1.00	14.93	0.225	CA	
ATOM 11016 N GLY B 65	18.859	32.581	50.981	1.00	14.17	-0.351	N	
ATOM 11017 HN GLY B 65	18.020	32.232	49.676	1.00	0.00	0.163	HD	
ATOM 11018 CA GLY B 65	19.129	32.241	51.474	1.00	12.62	0.225	CA	
ATOM 11021 C GLY B 65	20.283	31.347	51.774	1.00	15.72	0.236	C	
ATOM 11022 O GLY B 65	20.808	31.303	52.900	1.00	13.98	-0.272	OA	
ATOM 11023 N HIS B 66	20.769	30.597	50.755	1.00	11.24	-0.346	N	
ATOM 11024 HN HIS B 66	20.262	30.582	49.870	1.00	0.00	0.163	HD	
ATOM 11025 CA HIS B 66	21.985	29.815	50.878	1.00	12.88	0.182	C	
ATOM 11027 C HIS B 66	23.155	28.739	51.222	1.00	17.31	0.241	C	
ATOM 11028 O HIS B 66	24.058							

ATOM 11396 N ASN B 89	30.280	28.913	65.256	1.00	13.09	-0.346 N
ATOM 11397 HN ASN B 89	29.483	28.004	64.020	1.00	13.03	-0.163 HD
ATOM 11398 CA ASN B 89	29.484	27.697	65.330	1.00	12.14	0.185 C
ATOM 11400 C ASN B 89	28.434	27.652	64.202	1.00	12.51	0.241 C
ATOM 11401 O ASN B 89	27.340	27.686	64.844	1.00	12.71	0.171 OA
ATOM 11402 CB ASN B 89	28.771	27.629	66.695	1.00	16.50	0.137 C
ATOM 11405 CG ASN B 89	29.810	27.116	67.707	1.00	34.35	0.217 C
ATOM 11406 HD2 ASN B 89	29.830	27.460	68.150	1.00	37.89	-0.274 N
ATOM 11407 HD2 ASN B 89	30.519	27.400	69.532	1.00	0.00	0.159 HD
ATOM 11408 2HD2 ASN B 89	29.170	28.497	69.040	1.00	0.00	0.159 HD
ATOM 11409 CD1 ASN B 89	30.608	26.200	67.490	1.00	32.17	-0.274 OA
ATOM 11410 N PHE B 90	28.758	28.253	63.073	1.00	12.86	-0.346 N
ATOM 11411 HN PHE B 90	29.618	28.800	63.029	1.00	0.00	0.163 HD
ATOM 11412 CA PHE B 90	27.905	26.352	61.976	1.00	13.22	0.180 C
ATOM 11414 C PHE B 90	27.615	26.701	61.551	1.00	12.75	0.241 C
ATOM 11415 O PHE B 90	28.504	25.814	61.579	1.00	12.12	-0.271 OA
ATOM 11416 CB PHE B 90	28.028	28.833	60.932	1.00	13.88	-0.163 HD
ATOM 11419 CG PHE B 90	27.940	28.805	59.370	1.00	17.48	-0.056 A
ATOM 11420 CD1 PHE B 90	26.659	29.316	59.207	1.00	9.59	0.007 A
ATOM 11422 CD2 PHE B 90	28.650	28.298	58.948	1.00	16.63	0.009 A
ATOM 11424 CE1 PHE B 90	26.065	29.377	57.938	1.00	10.02	0.001 A
ATOM 11428 CE2 PHE B 90	28.022	28.347	57.006	1.00	13.51	0.001 A
ATOM 11428 CG PHE B 90	26.743	28.869	56.941	1.00	12.83	0.000 A
ATOM 11430 N ARG B 91	26.326	26.391	61.326	1.00	9.92	-0.346 N
ATOM 11431 HN ARG B 91	25.639	27.140	61.410	1.00	0.00	0.163 HD
ATOM 11432 CA ARG B 91	26.005	24.003	62.045	1.00	16.45	0.241 C
ATOM 11434 C ARG B 91	25.884	22.805	61.747	1.00	14.85	-0.271 OA
ATOM 11439 CG ARG B 91	25.828	25.347	58.479	1.00	12.62	0.023 C
ATOM 11442 CD ARG B 91	26.538	25.204	57.311	1.00	32.62	0.000 A
ATOM 11445 NE ARG B 91	25.893	25.719	56.099	1.00	15.10	-0.227 N
ATOM 11446 HE ARG B 91	25.439	26.727	56.073	1.00	0.00	0.177 HD
ATOM 11447 CE1 ARG B 91	25.439	26.727	56.073	1.00	15.45	-0.227 N
ATOM 11448 NH1 ARG B 91	25.790	26.378	55.006	1.00	13.33	-0.235 N
ATOM 11449 NH1 ARG B 91	25.759	23.199	55.609	1.00	0.00	0.174 HD
ATOM 11450 2NH1 ARG B 91	24.585	23.199	55.609	1.00	0.00	0.174 HD
ATOM 11451 NH2 ARG B 91	24.600	25.609	54.430	1.00	14.48	-0.235 N
ATOM 11452 HH2 ARG B 91	23.939	26.609	54.591	1.00	0.00	0.174 HD
ATOM 11453 2HH2 ARG B 91	23.939	26.609	54.591	1.00	0.00	0.174 HD
ATOM 11454 N GLN B 92	26.218	24.399	63.305	1.00	12.70	-0.346 N
ATOM 11455 HN GLN B 92	26.289	25.391	63.531	1.00	0.00	0.163 HD
ATOM 11456 CA GLN B 92	26.349	23.867	63.460	1.00	14.58	0.241 C
ATOM 11458 C GLN B 92	25.080	23.328	65.181	1.00	14.23	0.241 C
ATOM 11459 O GLN B 92	24.245	24.251	65.373	1.00	15.62	-0.271 OA
ATOM 11460 CB GLN B 92	27.602	23.742	65.223	1.00	17.68	0.000 A
ATOM 11463 CG GLN B 92	28.871	23.947	64.403	1.00	17.78	0.105 C
ATOM 11466 CD GLN B 92	29.144	22.721	63.484	1.00	19.94	0.215 C
ATOM 11467 HE2 GLN B 92	29.187	24.942	62.171	1.00	15.64	0.000 A
ATOM 11468 HE2 GLN B 92	29.371	25.151	61.553	1.00	0.00	0.159 HD
ATOM 11469 2HE2 GLN B 92	29.081	23.868	61.756	1.00	0.00	0.159 HD
ATOM 11470 OE1 PHE B 93	29.272	22.122	62.687	1.00	6.68	-0.346 N
ATOM 11471 N LEU B 93	24.930	22.712	65.848	1.00	16.05	-0.346 N
ATOM 11472 HN LEU B 93	25.667	21.469	65.794	1.00	0.00	0.163 HD
ATOM 11473 CA LEU B 93	23.741	21.950	66.458	1.00	14.43	0.241 C
ATOM 11475 C LEU B 93	23.426	22.991	67.646	1.00	15.95	0.241 C
ATOM 11476 O LEU B 93	24.246	23.382	68.177	1.00	19.28	-0.271 OA
ATOM 11477 CB LEU B 93	23.940	22.544	67.937	1.00	17.47	0.000 A
ATOM 11480 CG LEU B 93	23.697	20.059	68.158	1.00	25.17	-0.020 C
ATOM 11482 CD1 LEU B 93	21.577	19.679	67.222	1.00	25.52	0.009 C
ATOM 11486 CD2 LEU B 93	23.145	19.861	68.977	1.00	27.83	-0.346 N
ATOM 11490 N HIS B 94	24.227	23.515	67.578	1.00	14.96	0.000 A
ATOM 11491 HN HIS B 94	21.603	23.143	66.863	1.00	0.00	0.163 HD
ATOM 11492 CA HIS B 94	21.702	24.568	66.820	1.00	21.51	0.241 C
ATOM 11494 C HIS B 94	22.443	25.903	68.361	1.00	22.24	0.241 C
ATOM 11495 O HIS B 94	22.267	26.690	69.312	1.00	21.95	-0.271 OA
ATOM 11496 CB HIS B 94	21.650	24.959	69.494	1.00	16.69	0.000 A
ATOM 11499 CG HIS B 94	20.820	22.846	70.056	1.00	34.87	0.028 A
ATOM 11500 CD2 HIS B 94	19.664	22.485	69.435	1.00	33.71	0.114 A
ATOM 11502 HD1 HIS B 94	21.193	21.809	70.907	1.00	16.69	-0.346 N
ATOM 11503 HD1 HIS B 94	22.024	21.774	71.486	1.00	0.00	0.166 HD
ATOM 11504 CE1 HIS B 94	20.258	20.874	70.802	1.00	36.10	0.180 A
ATOM 11506 HE2 HIS B 94	19.339	21.247	69.656	1.00	0.00	0.166 HD
ATOM 11507 HE2 HIS B 94	18.513	20.702	69.656	1.00	0.00	0.166 HD
ATOM 11508 N SER B 95	23.226	22.216	67.331	1.00	18.98	-0.346 N
ATOM 11509 HN SER B 95	23.434	25.539	67.331	1.00	0.00	0.163 HD
ATOM 11510 CA SER B 95	23.842	22.551	67.306	1.00	17.52	0.200 C
ATOM 11512 C SER B 95	22.759	23.534	66.852	1.00	19.15	0.249 C
ATOM 11513 O SER B 95	21.680	23.116	67.172	1.00	17.21	-0.271 OA
ATOM 11514 CB SER B 95	24.994	27.601	66.320	1.00	18.81	0.199 C
ATOM 11517 CG SER B 95	24.349	27.584	66.989	1.00	18.81	0.199 C
ATOM 11518 HG SER B 95	25.171	27.567	64.369	1.00	0.00	0.209 HD
ATOM 11519 N LYS B 96	23.017	29.844	66.896	1.00	11.60	-0.346 N
ATOM 11520 HN LYS B 96	23.017	29.844	66.896	1.00	11.60	-0.346 N
ATOM 11521 CA LYS B 96	22.077	30.819	66.365	1.00	11.97	0.176 C
ATOM 11523 C LYS B 96	22.420	31.122	64.914	1.00	11.17	0.241 C
ATOM 11524 O LYS B 96	21.911	31.094	64.617	1.00	11.97	0.241 C
ATOM 11525 CB LYS B 96	22.250	32.144	65.145	1.00	14.37	0.035 A
ATOM 11528 CG LYS B 96	21.740	31.848	68.578	1.00	26.79	0.004 C
ATOM 11531 CD LYS B 96	20.581	31.221	68.389	1.00	45.32	0.029 C
ATOM 11534 CE LYS B 96	20.724	32.869	70.613	1.00	52.00	0.229 C
ATOM 11537 N2 LYS B 96	20.405	34.111	71.364	1.00	66.91	-0.079 N
ATOM 11538 HE1 LYS B 96	21.822	33.825	70.000	1.00	0.00	0.163 HD
ATOM 11539 HE2 LYS B 96	19.980	34.808	70.753	1.00	0.00	0.274 HD
ATOM 11540 HE3 LYS B 96	21.256	34.612	71.620	1.00	0.00	0.274 HD
ATOM 11541 N THR B 97	23.402	29.184	62.100	1.00	11.21	0.200 C
ATOM 11542 HN THR B 97	23.674	29.508	64.820	1.00	0.00	0.163 HD
ATOM 11543 CA THR B 97	23.617	30.479	62.884	1.00	10.33	0.205 C
ATOM 11545 C THR B 97	23.402	29.184	62.100	1.00	11.21	0.200 C
ATOM 11546 O THR B 97	24.356	28.590	61.547	1.00	11.30	-0.271 OA
ATOM 11547 CB THR B 97	25.098	30.885	62.746	1.00	17.29	0.146 C
ATOM 11549 CD2 THR B 97	25.373	32.833	63.255	1.00	32.51	0.000 A
ATOM 11553 OG1 THR B 97	25.867	29.959	63.562	1.00	14.26	-0.393 OA
ATOM 11554 HG1 THR B 97	26.780	30.209	63.477	1.00	0.00	0.210 HD
ATOM 11555 N PRO B 98	22.957	31.897	61.932	1.00	11.53	-0.346 N
ATOM 11556 CA PRO B 98	21.803	27.625	61.070	1.00	10.17	0.179 C
ATOM 11558 C PRO B 98	22.142	27.837	59.613	1.00	12.90	0.241 C
ATOM 11559 O PRO B 98	22.389	28.947	59.147	1.00	11.53	-0.271 OA
ATOM 11560 CB PRO B 98	20.294	27.422	61.267	1.00	13.03	0.037 C
ATOM 11563 CG PRO B 98	19.804	28.768	61.691	1.00	11.37	0.022 C
ATOM 11566 CD PRO B 98	20.939	29.967	62.904	1.00	18.29	-0.346 N
ATOM 11569 N GLY B 99	22.210	26.729	58.840	1.00	9.45	-0.351 N
ATOM 11570 HN GLY B 99	21.943	25.829	59.239	1.00	0.00	0.163 HD
ATOM 11571 CA GLY B 99	22.456	26.784	61.453	1.00	14.04	0.236 C
ATOM 11574 C GLY B 99	21.840	27.753	56.600	1.00	14.04	0.236 C
ATOM 11575 O GLY B 99	22.377	28.262	55.633	1.00	11.53	-0.271 OA
ATOM 11576 N HIS B 100	20.857	27.899	56.871	1.00	11.72	-0.346 N
ATOM 11578 HN HIS B 100	20.120	27.317	57.588	1.00	0.00	0.163 HD
ATOM 11578 CA HIS B 100	19.736	28.894	56.168	1.00	9.60	0.182 C
ATOM 11580 C HIS B 100	19.102	29.719	57.089	1.00	10.00	0.236 C
ATOM 11581 O HIS B 100	18.904	29.150	58.348	1.00	12.55	0.241 C
ATOM 11582 CB HIS B 100	18.674	28.925	55.275	1.00	10.40	0.093 C
ATOM 11585 CG HIS B 100	19.312	27.576	54.089	1.00	11.67	0.028 A
ATOM 11586 CD2 HIS B 100	19.583	28.020	52.836	1.00	7.14	0.114 A
ATOM 11588 HD1 HIS B 100	19.740	26.868	54.073	1.00	17.78	-0.346 N
ATOM 11589 HD1 HIS B 100	19.601	25.620	54.863	1.00	0.00	0.166 HD
ATOM 11590 CE1 HIS B 100	20.266	25.958	52.901	1.00	13.39	0.180 A
ATOM 11592 HE2 HIS B 100	20.139	26.987	52.129	1.00	15.30	-0.346 N
ATOM 11593 HE2 HIS B 100	20.500	27.059	51.162	1.00	0.00	0.166 HD
ATOM 11594 N PRO B 101	18.842	30.987	57.074	1.00	13.81	-0.337 N
ATOM 11595 CA PRO B 101	18.386	31.870	58.125	1.00	14.81	0.190 C
ATOM 11597 C PRO B 101	16.980	31.505	58.581	1.00	15.29	0.241 C
ATOM 11598 O PRO B 101	16.189	31.146	57.705	1.00	11.09	-0.271 OA
ATOM 11599 CB PRO B 101	18.448	33.273	57.543	1.00	13.23	0.037 C
ATOM 11602 CG PRO B 101	19.140	33.161	56.215	1.00	13.82	0.022 C
ATOM 11605 CD PRO B 101	18.997	31.714	55.552	1.00	13.11	0.127 C
ATOM 11608 N GLU B 102	16.755	31.511	59.898	1.00	12.54	-0.346 N
ATOM 11609 HN GLU B 102	17.518	31.770	60.529	1.00	0.00	0.163 HD
ATOM 11610 CA GLU B 102	15.472	31.160	60.451	1.00	13.24	0.177 C
ATOM 11612 C GLU B 102	14.854	32.282	61.292	1.00	14.99	0.241 C
ATOM 11613 O GLU B 102	15.512	32.740	62.218	1.00	15.22	-0.271 OA
ATOM 11614 CB GLU B 102	15.556	29.893	61.342	1.00	14.04	0.045 C
ATOM 11617 CG GLU B 102	16.019	28.676	60.503	1.00	14.94	0.116 C
ATOM 11620 CD GLU B 102	16.188	27.464	61.424	1.00	20.28	-0.142 C

ATOM 11969	C	ALA B 129	1.878	46.800	54.843	1.00	8.81	0.240	C
ATOM 11970	O	ALA B 129	2.137	54.458	57.121	1.00	11.12	-0.271	OA
ATOM 11971	CB	ALA B 129	2.155	44.415	55.532	1.00	10.66	0.042	C
ATOM 11975	N	GLU B 130	3.101	47.211	54.492	1.00	6.63	-0.346	N
ATOM 11976	HN	GLU B 130	3.730	42.257	56.234	1.00	6.80	-0.172	C
ATOM 11977	CA	GLU B 130	3.525	48.595	54.844	1.00	6.83	0.177	C
ATOM 11979	C	GLU B 130	2.594	49.604	54.166	1.00	9.21	0.241	C
ATOM 11980	O	GLU B 130	2.117	50.250	54.771	1.00	11.11	-0.271	OA
ATOM 11981	CB	GLU B 130	4.966	48.810	54.313	1.00	4.16	0.045	C
ATOM 11984	CG	GLU B 130	5.573	50.169	54.821	1.00	4.20	0.116	C
ATOM 11987	CD	GLU B 130	5.301	51.374	53.960	1.00	15.54	-0.346	N
ATOM 11988	OE1	GLU B 130	4.809	51.307	52.828	1.00	10.78	-0.648	OA
ATOM 11989	OE2	GLU B 130	5.576	52.533	54.352	1.00	8.77	-0.648	OA
ATOM 11990	N	LYS B 131	2.332	49.395	52.020	1.00	7.62	-0.346	N
ATOM 11991	HN	LYS B 131	2.722	48.580	52.380	1.00	0.00	0.163	HD
ATOM 11992	CA	LYS B 131	1.495	50.331	52.115	1.00	8.70	0.176	C
ATOM 11994	C	LYS B 131	2.177	50.509	52.115	1.00	11.11	-0.271	OA
ATOM 11995	O	LYS B 131	-0.537	51.500	52.718	1.00	8.83	-0.271	OA
ATOM 11996	CB	LYS B 131	1.466	49.883	50.625	1.00	7.60	0.035	C
ATOM 11999	CG	LYS B 131	0.435	50.330	49.746	1.00	11.48	0.046	C
ATOM 12002	CD	LYS B 131	0.535	50.203	48.309	1.00	13.02	0.027	C
ATOM 12005	CE	LYS B 131	-0.193	51.289	47.463	1.00	19.08	0.229	C
ATOM 12008	NE	LYS B 131	-0.868	50.710	46.380	1.00	20.30	-0.079	N
ATOM 12009	HE1	LYS B 131	-1.345	51.421	45.726	1.00	0.00	0.274	HD
ATOM 12010	HE2	LYS B 131	-1.506	49.960	46.548	1.00	0.00	0.274	HD
ATOM 12011	HE3	LYS B 131	-0.737	50.051	45.709	1.00	0.00	0.274	HD
ATOM 12012	N	THR B 132	-0.489	49.266	52.988	1.00	5.76	-0.346	N
ATOM 12013	HN	THR B 132	0.032	48.399	52.988	1.00	0.00	0.163	HD
ATOM 12014	CA	THR B 132	-1.845	53.498	53.366	1.00	8.97	0.025	C
ATOM 12016	C	THR B 132	-1.939	49.754	54.161	1.00	11.59	0.243	C
ATOM 12017	O	THR B 132	-2.971	50.599	55.277	1.00	11.11	-0.271	OA
ATOM 12018	CB	THR B 132	-2.328	47.719	53.473	1.00	14.76	0.146	C
ATOM 12020	CG2	THR B 132	-3.791	47.664	53.873	1.00	12.63	0.042	C
ATOM 12024	OG1	THR B 132	-4.237	48.235	52.123	1.00	12.48	-0.392	OA
ATOM 12025	HG1	THR B 132	-1.267	47.257	51.863	1.00	0.00	0.210	HD
ATOM 12026	N	LEU B 133	-0.935	49.454	55.743	1.00	8.95	-0.346	N
ATOM 12027	HN	LEU B 133	-1.433	52.234	57.913	1.00	9.79	-0.271	OA
ATOM 12028	CA	LEU B 133	-0.971	50.009	57.121	1.00	10.25	0.177	C
ATOM 12030	C	LEU B 133	-0.753	51.527	57.132	1.00	8.35	0.241	C
ATOM 12031	O	LEU B 133	-1.433	52.234	57.913	1.00	9.79	-0.271	OA
ATOM 12032	CB	LEU B 133	0.096	49.329	57.970	1.00	10.92	0.038	C
ATOM 12035	CG	LEU B 133	-0.017	49.253	59.485	1.00	22.01	-0.020	C
ATOM 12037	CD1	LEU B 133	-1.455	50.400	59.133	1.00	15.13	0.046	C
ATOM 12041	CD2	LEU B 133	1.309	48.658	60.000	1.00	21.48	0.009	C
ATOM 12045	N	ALA B 134	0.106	52.032	56.260	1.00	6.90	-0.346	N
ATOM 12046	HN	ALA B 134	-1.652	56.409	62.675	1.00	0.00	0.163	HD
ATOM 12047	CA	ALA B 134	0.281	53.448	56.120	1.00	11.95	0.172	C
ATOM 12049	C	ALA B 134	-1.031	54.589	55.687	1.00	11.31	0.240	C
ATOM 12050	O	ALA B 134	-1.451	55.201	56.213	1.00	9.48	-0.271	OA
ATOM 12051	CB	ALA B 134	1.431	53.788	55.152	1.00	6.40	0.042	C
ATOM 12055	N	ALA B 135	-1.732	53.548	54.694	1.00	9.46	-0.346	N
ATOM 12056	HN	ALA B 135	-4.372	52.698	54.000	1.00	0.00	0.163	HD
ATOM 12057	CA	ALA B 135	-3.001	54.108	54.236	1.00	9.93	0.172	C
ATOM 12059	C	ALA B 135	-4.033	54.087	55.361	1.00	16.23	0.240	C
ATOM 12060	O	ALA B 135	-4.860	54.937	56.107	1.00	11.11	-0.271	OA
ATOM 12061	CB	ALA B 135	-3.550	53.325	53.031	1.00	10.04	0.042	C
ATOM 12065	N	GLN B 136	-4.074	53.048	56.180	1.00	8.84	-0.346	N
ATOM 12066	HN	GLN B 136	-4.418	52.278	56.988	1.00	0.00	0.163	HD
ATOM 12067	CA	GLN B 136	-5.046	52.985	57.275	1.00	10.34	0.177	C
ATOM 12069	C	GLN B 136	-4.701	53.879	58.457	1.00	13.39	0.241	C
ATOM 12070	O	GLN B 136	-4.649	54.396	59.076	1.00	13.35	-0.271	OA
ATOM 12071	CB	GLN B 136	-5.238	51.553	57.784	1.00	11.42	0.045	C
ATOM 12074	CG	GLN B 136	-5.841	50.586	56.762	1.00	9.05	0.105	C
ATOM 12077	CD	GLN B 136	-6.911	49.685	55.100	1.00	42.45	-0.392	OA
ATOM 12078	NE2	GLN B 136	-6.498	48.255	56.488	1.00	10.29	-0.370	N
ATOM 12079	HE2	GLN B 136	-6.547	47.297	56.833	1.00	0.00	0.159	HD
ATOM 12080	HE3	GLN B 136	-2.845	48.356	62.159	1.00	14.41	0.046	C
ATOM 12081	OE1	GLN B 136	-5.498	48.807	58.396	1.00	13.46	-0.274	OA
ATOM 12082	N	PRO B 137	-3.437	54.102	58.789	1.00	9.37	-0.346	N
ATOM 12083	HN	PRO B 137	-4.484	53.765	59.420	1.00	0.00	0.163	HD
ATOM 12084	CA	PRO B 137	-3.130	54.837	60.022	1.00	12.05	0.180	C
ATOM 12086	C	PRO B 137	-2.515	56.209	59.870	1.00	13.32	0.241	C
ATOM 12087	O	PRO B 137	-4.672	56.099	60.821	1.00	13.32	-0.271	OA
ATOM 12088	CB	PRO B 137	-2.140	53.979	60.873	1.00	10.85	0.073	C
ATOM 12091	CG	PRO B 137	-2.869	52.833	61.522	1.00	12.45	-0.056	A
ATOM 12092	CD1	PRO B 137	-3.472	52.966	62.000	1.00	11.63	0.045	C
ATOM 12094	CD2	PRO B 137	-2.949	51.598	60.866	1.00	11.95	0.007	A
ATOM 12096	CE1	PRO B 137	-4.173	51.959	63.355	1.00	15.57	0.003	A
ATOM 12098	CE2	PRO B 137	-3.644	50.558	61.400	1.00	15.22	0.001	A
ATOM 12100	CD	PRO B 137	-4.256	50.731	62.707	1.00	11.30	0.000	A
ATOM 12102	N	ASN B 138	-1.801	54.502	58.113	1.00	12.04	-0.346	N
ATOM 12103	HN	ASN B 138	-1.740	55.844	58.011	1.00	0.00	0.163	HD
ATOM 12104	CA	ASN B 138	-1.101	57.795	58.735	1.00	11.73	0.185	C
ATOM 12106	C	ASN B 138	-2.222	58.903	59.680	1.00	15.48	-0.271	OA
ATOM 12107	O	ASN B 138	-3.070	59.420	60.000	1.00	14.41	0.177	C
ATOM 12108	CB	ASN B 138	-0.151	57.933	57.525	1.00	11.63	0.137	C
ATOM 12111	CG	ASN B 138	-1.418	57.687	57.640	1.00	11.41	0.046	C
ATOM 12112	ND2	ASN B 138	1.908	56.911	56.587	1.00	9.17	-0.370	N
ATOM 12113	HD2	ASN B 138	2.755	56.349	56.664	1.00	0.00	0.159	HD
ATOM 12114	HD3	ASN B 138	3.238	56.349	56.664	1.00	0.00	0.159	HD
ATOM 12115	OD1	ASN B 138	1.369	56.577	58.744	1.00	10.04	-0.274	OA
ATOM 12116	N	ARG B 139	-1.693	60.086	59.246	1.00	14.81	-0.346	N
ATOM 12117	HN	ARG B 139	-2.459	62.217	60.440	1.00	11.41	0.046	C
ATOM 12118	CA	ARG B 139	-2.432	61.379	59.194	1.00	22.95	0.177	C
ATOM 12120	C	ARG B 139	-1.385	62.428	58.835	1.00	23.50	0.243	C
ATOM 12121	O	ARG B 139	-4.169	62.944	60.210	1.00	21.44	-0.271	OA
ATOM 12122	CB	ARG B 139	-3.251	61.653	60.418	1.00	24.47	0.040	C
ATOM 12125	CG	ARG B 139	-3.680	60.572	61.410	1.00	22.51	0.047	C
ATOM 12128	CD	ARG B 139	-4.939	59.946	60.975	1.00	22.43	0.179	C
ATOM 12130	NE	ARG B 139	-6.001	59.172	61.490	1.00	21.44	-0.206	N
ATOM 12131	HE	ARG B 139	-6.447	59.386	62.382	1.00	0.00	0.183	HD
ATOM 12132	CG	ARG B 139	-6.389	58.135	60.720	1.00	20.22	0.046	C
ATOM 12133	NH1	ARG B 139	-5.743	57.931	59.570	1.00	32.96	-0.235	N
ATOM 12134	NH2	ARG B 139	-4.999	58.580	59.333	1.00	0.00	0.174	HD
ATOM 12135	NH3	ARG B 139	-6.037	57.218	59.688	1.00	32.96	-0.235	N
ATOM 12136	NH4	ARG B 139	-7.351	57.296	61.104	1.00	33.19	-0.235	N
ATOM 12137	NH5	ARG B 139	-7.844	57.452	61.933	1.00	0.00	0.174	HD
ATOM 12138	NH6	ARG B 139	-7.648	56.431	60.779	1.00	32.96	-0.235	N
ATOM 12139	N	PRO B 140	-1.795	63.520	58.163	1.00	36.44	-0.377	N
ATOM 12140	CA	PRO B 140	-0.866	64.499	57.581	1.00	33.63	0.179	C
ATOM 12142	C	PRO B 140	-1.071	65.909	58.216	1.00	36.44	-0.377	N
ATOM 12143	O	PRO B 140	-0.455	65.356	59.686	1.00	21.68	-0.271	OA
ATOM 12144	CB	PRO B 140	-1.811	65.547	58.994	1.00	41.57	0.037	C
ATOM 12147	CG	PRO B 140	-1.064	65.254	60.666	1.00	36.44	-0.377	N
ATOM 12150	CD	PRO B 140	-3.208	63.930	58.018	1.00	36.02	0.127	C
ATOM 12153	N	GLY B 141	1.399	64.940	58.531	1.00	28.85	-0.351	N
ATOM 12154	HN	GLY B 141	-1.804	64.745	57.671	1.00	24.63	0.180	C
ATOM 12155	CA	GLY B 141	2.280	65.391	59.631	1.00	27.92	0.225	C
ATOM 12158	CG	GLY B 141	2.434	64.296	60.722	1.00	30.19	0.236	C
ATOM 12159	O	GLY B 141	3.211	64.443	62.459	1.00	29.89	-0.271	OA
ATOM 12160	N	HIS B 142	1.660	63.197	60.586	1.00	18.68	-0.346	N
ATOM 12161	HN	HIS B 142	1.002	63.131	59.809	1.00	0.00	0.163	HD
ATOM 12162	CA	HIS B 142	1.763	62.100	61.360	1.00			

ATOM 12529 N	CYS B 167	6.133	39.067	41.218	1.00	5.41	-0.345	N
ATOM 12530 HN	CYS B 167	6.640	40.874	41.000	1.00	6.00	-0.245	HD
ATOM 12531 CA	CYS B 167	6.640	40.366	40.676	1.00	6.65	0.186	C
ATOM 12532 C	CYS B 167	5.643	40.992	39.705	1.00	11.75	0.242	C
ATOM 12533 O	CYS B 167	4.804	42.923	39.035	1.00	7.14	-0.245	O
ATOM 12534 O	CYS B 167	8.012	40.134	40.035	1.00	8.00	0.121	C
ATOM 12535 SG	CYS B 167	9.262	39.697	41.320	1.00	10.32	-0.095	SA
ATOM 12536 N	HR SER B 168	6.860	42.933	39.035	1.00	7.14	-0.245	N
ATOM 12540 HN	SER B 168	5.017	39.090	39.138	1.00	0.00	0.163	HD
ATOM 12541 CA	SER B 168	3.798	40.612	38.168	1.00	7.79	0.200	C
ATOM 12542 C	SER B 168	2.794	41.359	39.035	1.00	7.78	0.243	C
ATOM 12544 O	SER B 168	2.387	42.511	38.738	1.00	8.86	-0.271	O
ATOM 12545 CB	SER B 168	3.125	39.415	37.464	1.00	12.42	0.199	C
ATOM 12548 CG	SER B 168	2.813	39.850	36.897	1.00	12.92	-0.196	O
ATOM 12549 HG	SER B 168	1.459	39.113	36.464	1.00	0.00	0.209	HD
ATOM 12550 N	LEU B 169	2.304	40.763	40.141	1.00	6.51	-0.346	N
ATOM 12551 HN	LEU B 169	4.627	38.828	39.866	1.00	10.60	-0.163	HD
ATOM 12552 CA	LEU B 169	1.323	41.426	40.989	1.00	6.95	0.177	C
ATOM 12553 C	LEU B 169	1.869	42.671	41.675	1.00	6.73	0.241	C
ATOM 12555 O	LEU B 169	0.825	40.465	42.103	1.00	7.79	0.038	O
ATOM 12556 CB	LEU B 169	-0.327	40.909	43.013	1.00	11.39	-0.020	C
ATOM 12559 CD1	LEU B 169	-1.561	41.313	42.266	1.00	13.25	-0.009	C
ATOM 12565 CD2	LEU B 169	-0.707	39.725	43.940	1.00	15.85	0.009	C
ATOM 12569 N	ALA B 170	3.152	42.664	42.051	1.00	10.03	-0.346	N
ATOM 12570 HN	ALA B 170	7.768	41.822	41.906	1.00	10.00	0.163	HD
ATOM 12571 CA	ALA B 170	3.784	43.838	42.674	1.00	9.42	0.172	C
ATOM 12573 C	ALA B 170	3.755	45.507	41.709	1.00	10.20	0.240	C
ATOM 12574 O	ALA B 170	4.665	46.157	42.138	1.00	10.10	-0.271	O
ATOM 12575 CB	ALA B 170	5.255	43.023	43.051	1.00	9.38	0.042	C
ATOM 12579 N	GLY B 171	3.903	44.816	40.100	1.00	10.45	-0.271	N
ATOM 12580 HN	GLY B 171	0.163	43.865	40.096	1.00	0.00	0.163	HD
ATOM 12581 CA	GLY B 171	3.842	45.862	39.320	1.00	11.58	0.225	C
ATOM 12584 C	GLY B 171	2.842	46.822	39.937	1.00	11.34	-0.163	C
ATOM 12585 O	GLY B 171	2.218	47.675	39.263	1.00	12.66	-0.272	O
ATOM 12586 N	THR B 172	1.441	45.551	39.328	1.00	7.06	-0.346	N
ATOM 12587 HN	THR B 172	4.627	38.828	39.866	1.00	10.60	-0.163	HD
ATOM 12588 CA	THR B 172	0.048	45.984	39.346	1.00	8.88	0.205	C
ATOM 12590 C	THR B 172	-0.251	46.852	40.559	1.00	16.14	0.243	C
ATOM 12591 O	THR B 172	-0.378	47.889	40.127	1.00	16.14	-0.271	O
ATOM 12592 CB	THR B 172	-0.919	44.778	39.387	1.00	15.42	0.146	C
ATOM 12594 CD2	THR B 172	-2.360	45.223	39.507	1.00	14.19	0.042	C
ATOM 12598 OG1	THR B 172	-4.720	44.026	40.100	1.00	14.00	-0.271	O
ATOM 12599 HG1	THR B 172	0.185	43.747	38.071	1.00	0.00	0.210	HD
ATOM 12600 N	LEU B 173	0.260	46.460	41.744	1.00	6.61	-0.346	N
ATOM 12601 HN	LEU B 173	4.627	38.828	39.866	1.00	10.60	-0.163	HD
ATOM 12602 CA	LEU B 173	-0.089	47.218	42.955	1.00	10.17	0.177	C
ATOM 12604 C	LEU B 173	0.850	48.381	43.240	1.00	12.92	0.241	C
ATOM 12605 O	LEU B 173	-0.720	49.036	42.717	1.00	12.58	-0.271	O
ATOM 12606 CB	LEU B 173	-0.036	46.246	44.177	1.00	10.27	0.038	C
ATOM 12609 CG	LEU B 173	-1.138	45.460	44.157	1.00	15.89	-0.020	C
ATOM 12611 CD1	LEU B 173	-0.897	44.155	45.005	1.00	15.81	-0.038	C
ATOM 12615 CD2	LEU B 173	-2.520	45.736	44.320	1.00	11.69	0.009	C
ATOM 12619 N	LYS B 174	1.725	48.743	42.320	1.00	7.91	-0.346	N
ATOM 12620 HN	LYS B 174	5.828	47.929	43.000	1.00	10.00	-0.163	HD
ATOM 12621 CA	LYS B 174	2.558	49.923	42.354	1.00	7.85	0.176	C
ATOM 12623 C	LYS B 174	3.800	50.071	43.624	1.00	10.29	0.241	C
ATOM 12624 O	LYS B 174	4.285	51.744	44.000	1.00	14.42	-0.271	O
ATOM 12625 CB	LYS B 174	1.600	51.119	42.186	1.00	13.53	0.035	C
ATOM 12628 CG	LYS B 174	0.905	50.924	40.801	1.00	16.37	0.004	C
ATOM 12631 CD	LYS B 174	-1.145	52.307	42.000	1.00	20.45	-0.038	C
ATOM 12634 CE	LYS B 174	-0.409	52.342	39.192	1.00	51.45	0.229	C
ATOM 12637 NZ	LYS B 174	-0.365	53.716	38.621	1.00	61.99	-0.079	N
ATOM 12638 HZ1	LYS B 174	4.737	49.739	41.000	1.00	0.00	0.274	HD
ATOM 12638 HZ2	LYS B 174	0.576	54.107	38.663	1.00	0.00	0.274	HD
ATOM 12640 HZ3	LYS B 174	-0.842	54.386	39.225	1.00	0.00	0.274	HD
ATOM 12641 N	LEU B 175	4.090	49.493	44.028	1.00	10.49	-0.163	N
ATOM 12642 HN	LEU B 175	4.157	48.184	43.138	1.00	0.00	0.163	HD
ATOM 12643 CA	LEU B 175	4.790	49.050	45.438	1.00	12.54	0.177	C
ATOM 12645 C	LEU B 175	9.125	49.739	45.326	1.00	10.00	-0.271	O
ATOM 12646 O	LEU B 175	7.190	49.145	45.104	1.00	10.10	-0.271	O
ATOM 12647 CB	LEU B 175	4.991	47.581	45.780	1.00	10.90	0.038	C
ATOM 12650 CD	LEU B 175	4.657	46.942	45.284	1.00	10.90	-0.038	C
ATOM 12652 CD1	LEU B 175	3.919	45.367	46.266	1.00	9.21	0.009	C
ATOM 12656 CD2	LEU B 175	2.794	47.442	47.083	1.00	13.12	0.009	C
ATOM 12660 N	GLY B 176	0.036	51.066	45.000	1.00	9.51	-0.163	N
ATOM 12661 HN	GLY B 176	5.195	51.528	45.315	1.00	0.00	0.163	HD
ATOM 12662 CA	GLY B 176	7.282	51.887	45.020	1.00	9.29	0.225	C
ATOM 12665 C	GLY B 176	8.032	51.714	44.124	1.00	12.14	-0.236	C
ATOM 12666 O	GLY B 176	9.462	52.080	45.925	1.00	12.16	-0.272	O
ATOM 12667 N	THR B 176	0.893	52.028	42.000	1.00	9.42	-0.346	N
ATOM 12668 HN	LYS B 177	6.927	50.924	47.500	1.00	0.00	0.163	HD
ATOM 12669 CA	LYS B 177	8.942	51.078	48.353	1.00	9.88	0.176	C
ATOM 12671 C	LYS B 177	9.548	49.686	49.000	1.00	10.15	-0.271	O
ATOM 12672 O	LYS B 177	10.357	49.369	49.258	1.00	13.84	-0.271	O
ATOM 12673 CB	LYS B 177	8.375	51.475	49.720	1.00	13.53	0.035	C
ATOM 12676 CG	LYS B 177	5.292	52.946	49.717	1.00	15.00	-0.038	C
ATOM 12679 CD	LYS B 177	7.738	53.332	52.008	1.00	19.89	0.027	C
ATOM 12682 CE	LYS B 177	6.716	54.417	51.426	1.00	26.56	0.229	C
ATOM 12685 NE	LYS B 177	5.079	55.000	50.178	1.00	26.56	-0.079	N
ATOM 12686 HZ1	LYS B 177	5.866	55.412	53.036	1.00	0.00	0.274	HD
ATOM 12687 HZ2	LYS B 177	7.448	54.892	53.338	1.00	0.00	0.274	HD
ATOM 12688 HZ3	LYS B 177	6.332	54.969	53.338	1.00	0.00	0.274	HD
ATOM 12689 N	LEU B 178	9.253	48.849	47.351	1.00	8.69	-0.346	N
ATOM 12690 HN	LEU B 178	8.520	49.105	46.690	1.00	0.00	0.163	HD
ATOM 12691 CA	LEU B 178	11.138	47.719	46.288	1.00	11.03	0.241	C
ATOM 12693 C	LEU B 178	10.970	48.186	45.154	1.00	7.24	-0.271	O
ATOM 12695 CB	LEU B 178	8.575	44.248	46.138	1.00	9.44	-0.038	C
ATOM 12698 CG	LEU B 178	9.646	45.143	46.272	1.00	13.76	-0.020	C
ATOM 12700 CD1	LEU B 178	10.156	44.486	47.571	1.00	16.98	0.009	C
ATOM 12704 CD2	LEU B 178	8.575	44.248	46.138	1.00	9.44	-0.038	C
ATOM 12708 N	ILE B 179	12.348	47.471	46.763	1.00	5.51	-0.346	N
ATOM 12709 HN	ILE B 179	12.444	47.139	47.723	1.00	0.00	0.163	HD
ATOM 12710 CA	ILE B 179	13.950	47.859	47.000	1.00	11.39	-0.038	C
ATOM 12712 C	ILE B 179	14.339	46.355	45.998	1.00	12.27	0.241	C
ATOM 12713 O	ILE B 179	14.732	45.930	47.074	1.00	12.01	-0.271	O
ATOM 12714 CB	ILE B 179	14.417	46.893	46.448	1.00	11.39	0.038	C
ATOM 12716 CG1	ILE B 179	13.635	50.141	46.224	1.00	12.02	0.002	C
ATOM 12719 CG2	ILE B 179	15.759	48.906	45.799	1.00	8.23	0.012	C
ATOM 12723 CD1	ILE B 179	15.175	51.397	46.000	1.00	12.22	-0.038	C
ATOM 12727 N	ALA B 180	14.518	45.709	44.854	1.00	9.82	-0.346	N
ATOM 12728 HN	ALA B 180	14.602	46.126	43.979	1.00	0.00	0.163	HD
ATOM 12729 CA	ALA B 180	15.175	46.396	44.600	1.00	10.00	-0.271	O
ATOM 12731 C	ALA B 180	16.205	44.459	44.105	1.00	12.46	0.240	C
ATOM 12732 O	ALA B 180	16.569	45.153	43.103	1.00	13.36	-0.271	O
ATOM 12733 CB	ALA B 180	14.293	43.381	44.848	1.00	9.86	0.038	C
ATOM 12737 N	PHE B 181	17.491	43.782	44.646	1.00	9.98	-0.346	N
ATOM 12738 HN	PHE B 181	17.330	43.254	45.504	1.00	0.00	0.163	HD
ATOM 12739 CA	PHE B 181	18.827	43.717	44.400	1.00	11.10	-0.271	O
ATOM 12741 C	PHE B 181	19.057	42.375	43.475	1.00	12.05	0.241	C
ATOM 12742 O	PHE B 181	18.913	41.434	44.221	1.00	10.28	-0.271	O
ATOM 12743 CB	PHE B 181	19.302	44.026	45.000	1.00	11.39	0.037	C
ATOM 12746 CG	PHE B 181	19.829	45.414	45.721	1.00	13.26	-0.056	C
ATOM 12747 CD1	PHE B 181	18.855	45.649	46.726	1.00	14.39	0.007	C
ATOM 12749 CD2	PHE B 181	20.814	46.280	51.4	1.00	11.09	0.007	O
ATOM 12751 CE1	PHE B 181	18.840	46.886	47.373	1.00	16.15	0.001	C
ATOM 12753 CE2	PHE B 181	20.871	47.500	46.168	1.00	10.75	-0.163	O
ATOM 12755 CE	PHE B 181	19.849	47.765	47.110	1.00	16.43	0.000	C
ATOM 12757 N	TYR B 182	19.350	42.292	42.110	1.00	8.00	-0.346	N
ATOM 12758 HN	TYR B 182	19.430	43					

AT00M	13661	CG	LEU	B	241	19.000	48.838	41.998	1.00	22.217	-0.020	C
AT00M	13662	CD1	LEU	B	241	19.510	49.271	43.379	1.00	22.117	0.009	C
AT00M	13667	CD2	LEU	B	241	19.510	49.271	43.379	1.00	22.117	0.009	C
AT00M	13671	N	MEP	B	242	19.889	45.598	38.663	1.00	10.662	-0.346	N
AT00M	13672	HN	MEP	B	242	19.889	45.598	38.663	1.00	10.662	-0.346	N
AT00M	13673	CA	MEP	B	242	22.095	44.664	38.358	1.00	15.663	0.241	C
AT00M	13675	CG	MEP	B	242	22.095	44.664	38.358	1.00	15.663	0.241	C
AT00M	13676	CG	MEP	B	242	22.095	44.664	38.358	1.00	15.663	0.241	C
AT00M	13677	CB	MEP	B	242	20.292	44.347	36.620	1.00	11.778	0.045	C
AT00M	13680	CG	MEP	B	242	18.770	44.034	36.532	1.00	20.669	0.076	C
AT00M	13683	SD	MEP	B	242	18.323	43.806	34.800	1.00	23.338	-0.178	SA
AT00M	13684	CE	MEP	B	242	19.029	42.193	34.504	1.00	18.112	0.089	C
AT00M	13688	N	CYS	B	243	22.650	43.937	39.350	1.00	11.337	-0.345	N
AT00M	13689	HN	CYS	B	243	22.650	43.937	39.350	1.00	11.337	-0.345	N
AT00M	13690	CA	CYS	B	243	24.025	44.157	39.728	1.00	10.477	0.186	C
AT00M	13692	C	CYS	B	243	24.916	43.070	39.191	1.00	11.555	0.241	C
AT00M	13693	C	CYS	B	243	24.916	43.070	39.191	1.00	11.555	0.241	C
AT00M	13694	CB	CYS	B	243	24.151	44.117	41.283	1.00	12.110	0.121	C
AT00M	13697	SG	CYS	B	243	23.095	45.404	42.028	1.00	17.773	-0.095	SA
AT00M	13698	N	LVS	B	244	25.875	42.531	38.380	1.00	13.007	-0.163	HD
AT00M	13699	HN	LVS	B	244	25.954	44.508	38.180	1.00	0.000	0.163	HD
AT00M	13700	CA	LVS	B	244	26.831	42.569	37.771	1.00	14.006	0.176	C
AT00M	13702	C	LVS	B	244	27.885	42.051	38.789	1.00	14.004	0.176	C
AT00M	13703	O	LVS	B	244	28.572	43.216	39.253	1.00	13.551	-0.271	OA
AT00M	13704	CB	LVS	B	244	27.468	43.237	36.537	1.00	22.228	0.035	C
AT00M	13707	CG	LVS	B	244	26.585	42.885	35.388	1.00	33.335	0.074	C
AT00M	13710	CD	LVS	B	244	26.373	44.068	34.421	1.00	57.274	0.027	C
AT00M	13713	CE	LVS	B	244	26.991	43.876	33.047	1.00	63.665	0.229	C
AT00M	13716	NZ	LVS	B	244	27.456	45.193	42.332	1.00	71.977	-0.079	N
AT00M	13717	H21	LVS	B	244	27.571	45.064	41.438	1.00	0.000	0.274	HD
AT00M	13718	H22	LVS	B	244	27.571	45.064	41.438	1.00	0.000	0.274	HD
AT00M	13719	H23	LVS	B	244	26.277	45.707	42.313	1.00	0.000	0.274	HD
AT00M	13720	N	THR	B	245	27.964	41.037	39.283	1.00	13.665	-0.344	N
AT00M	13721	N	THR	B	245	29.305	41.037	39.283	1.00	13.665	-0.344	N
AT00M	13722	CA	THR	B	245	28.838	40.674	40.333	1.00	14.116	0.205	C
AT00M	13724	C	THR	B	245	29.752	39.941	39.854	1.00	19.008	0.243	C
AT00M	13725	O	THR	B	245	29.410	38.745	38.745	1.00	19.008	-0.271	OA
AT00M	13726	CB	THR	B	245	28.306	40.299	41.663	1.00	16.004	0.146	C
AT00M	13728	CG2	THR	B	245	27.399	41.375	42.247	1.00	16.000	0.042	C
AT00M	13732	CG1	THR	B	245	27.399	41.375	42.247	1.00	16.000	0.042	C
AT00M	13733	HG1	THR	B	245	27.137	38.862	42.271	1.00	0.000	0.210	HD
AT00M	13734	N	LIE	B	246	30.803	39.917	40.619	1.00	16.533	-0.346	N
AT00M	13735	HN	LIE	B	246	31.027	39.744	40.619	1.00	16.533	-0.346	N
AT00M	13736	CA	LIE	B	246	31.647	38.003	40.321	1.00	18.665	0.180	C
AT00M	13738	C	LIE	B	246	31.441	37.068	41.521	1.00	14.599	0.241	C
AT00M	13739	O	LIE	B	246	34.037	37.212	42.614	1.00	20.000	-0.163	HD
AT00M	13740	CB	LIE	B	246	33.123	38.340	40.233	1.00	21.800	0.012	C
AT00M	13742	CG1	LIE	B	246	33.363	39.309	39.981	1.00	21.200	0.003	C
AT00M	13745	CG2	LIE	B	246	34.037	37.212	42.614	1.00	20.000	-0.163	HD
AT00M	13749	CD1	LIE	B	246	34.758	39.932	39.047	1.00	25.995	0.005	C
AT00M	13753	N	LIE	B	247	30.898	35.892	41.304	1.00	16.461	-0.346	N
AT00M	13754	HN	LIE	B	247	30.633	35.600	40.835	1.00	16.461	-0.346	N
AT00M	13755	CA	LIE	B	247	30.684	34.998	42.458	1.00	14.777	0.180	C
AT00M	13757	C	LIE	B	247	32.040	34.634	43.036	1.00	15.445	0.241	C
AT00M	13758	O	LIE	B	247	33.049	34.477	42.332	1.00	21.800	-0.079	N
AT00M	13759	CB	LIE	B	247	29.889	33.753	42.059	1.00	14.200	0.013	C
AT00M	13761	CG1	LIE	B	247	29.376	33.009	43.111	1.00	18.115	0.002	C
AT00M	13764	CG2	LIE	B	247	30.688	32.821	43.111	1.00	18.115	0.002	C
AT00M	13768	CD1	LIE	B	247	28.410	31.884	42.940	1.00	21.118	0.005	C
AT00M	13772	N	LIE	B	248	32.123	34.605	44.377	1.00	13.449	-0.351	N
AT00M	13773	HN	LIE	B	248	31.308	34.808	44.924	1.00	13.449	-0.351	N
AT00M	13774	CA	LIE	B	248	33.044	34.278	44.995	1.00	15.119	0.225	C
AT00M	13777	C	LIE	B	248	34.442	35.378	44.820	1.00	17.440	0.236	C
AT00M	13778	O	LIE	B	248	35.628	35.086	44.932	1.00	24.555	-0.346	N
AT00M	13779	N	PHE	B	249	34.027	36.634	44.642	1.00	18.300	-0.346	N
AT00M	13780	HN	PHE	B	249	33.026	36.827	44.663	1.00	0.000	0.163	HD
AT00M	13781	CA	PHE	B	249	34.928	37.453	44.611	1.00	16.116	0.241	C
AT00M	13783	O	PHE	B	249	36.136	37.773	45.386	1.00	21.667	0.241	C
AT00M	13784	C	PHE	B	249	35.923	37.785	46.606	1.00	18.588	0.241	C
AT00M	13785	CB	PHE	B	249	34.188	39.057	44.308	1.00	17.778	0.176	C
AT00M	13788	CG	PHE	B	249	35.039	40.290	44.295	1.00	24.999	-0.056	A
AT00M	13789	CD1	PHE	B	249	35.807	40.810	45.325	1.00	24.300	0.007	C
AT00M	13791	CG2	PHE	B	249	35.807	40.810	45.325	1.00	24.300	0.007	C
AT00M	13793	CE	PHE	B	249	36.990	41.927	45.147	1.00	28.665	0.001	A
AT00M	13795	CG2	PHE	B	249	35.591	42.062	42.895	1.00	25.885	0.001	A
AT00M	13797	CG	PHE	B	249	36.838	42.568	43.496	1.00	28.665	0.001	A
AT00M	13799	N	LVS	B	250	37.350	37.869	44.822	1.00	17.776	-0.351	N
AT00M	13800	HN	LVS	B	250	37.439	37.944	43.808	1.00	17.776	-0.351	N
AT00M	13801	CA	LVS	B	250	38.846	37.870	43.828	1.00	21.800	0.241	C
AT00M	13804	C	LVS	B	250	39.334	36.567	45.584	1.00	22.339	0.236	C
AT00M	13805	O	LVS	B	250	40.302	36.567	45.584	1.00	22.440	0.236	C
AT00M	13806	N	SER	B	251	38.725	35.434	45.240	1.00	19.233	-0.344	N
AT00M	13807	HN	SER	B	251	37.728	35.462	45.025	1.00	19.233	-0.344	N
AT00M	13808	CA	SER	B	251	39.422	34.157	45.157	1.00	20.555	0.241	C
AT00M	13810	C	SER	B	251	40.109	34.010	43.821	1.00	23.110	0.163	HD
AT00M	13811	O	SER	B	251	39.434	34.007	42.779	1.00	22.663	-0.271	OA
AT00M	13812	CB	SER	B	251	38.441	34.981	45.377	1.00	14.449	0.146	C
AT00M	13815	CG	SER	B	251	38.978	31.792	44.843	1.00	22.444	-0.398	OA
AT00M	13816	HG	SER	B	251	38.373	31.072	44.979	1.00	0.000	0.209	HD
AT00M	13817	N	SER	B	251	38.373	31.072	44.979	1.00	0.000	0.209	HD
AT00M	13818	CA	PRO	B	252	42.163	38.847	42.547	1.00	25.116	0.176	C
AT00M	13820	C	PRO	B	252	41.746	32.657	41.708	1.00	25.336	0.241	C
AT00M	13821	O	PRO	B	252	41.827	32.753	40.923	1.00	25.116	0.241	C
AT00M	13822	CB	PRO	B	252	43.641	33.751	42.935	1.00	24.551	0.037	C
AT00M	13825	CG	PRO	B	252	43.716	33.992	44.384	1.00	25.663	0.022	C
AT00M	13828	CD	PRO	B	252	43.716	33.992	44.384	1.00	25.663	0.022	C
AT00M	13831	N	ASN	B	253	41.509	31.431	42.935	1.00	22.229	-0.346	N
AT00M	13832	HN	ASN	B	253	41.540	31.421	43.333	1.00	0.000	0.163	HD
AT00M	13833	CA	ASN	B	253	42.000	30.332	43.716	1.00	23.778	0.241	C
AT00M	13835	C	ASN	B	253	39.753	29.960	41.334	1.00	26.884	0.241	C
AT00M	13836	O	ASN	B	253	39.469	29.179	40.415	1.00	27.112	0.241	C
AT00M	13837	CB	ASN	B	253	41.923	29.108	42.123	1.00	24.137	0.176	C
AT00M	13840	CG	ASN	B	253	43.427	29.373	42.008	1.00	37.200	-0.217	OA
AT00M	13841	CD1	ASN	B	253	44.096	29.601	43.129	1.00	26.550	0.170	C
AT00M	13842	CD2	ASN	B	253	44.096	29.601	43.129	1.00	26.550	0.170	C
AT00M	13843											

ATOM	14238	H21	LVS	B	280	46.894	39.915	52.293	1.00	0.00	0.274	HD
ATOM	14239	H22	LVS	B	280	46.894	39.915	52.293	1.00	0.00	0.274	HD
ATOM	14240	H23	LVS	B	280	48.077	39.506	53.405	1.00	0.00	0.274	HD
ATOM	14241	N	TRP	B	281	43.954	40.279	58.628	1.00	25.25	-0.346	N
ATOM	14242	HN	TRP	B	281	43.954	40.279	58.628	1.00	25.25	-0.346	N
ATOM	14243	CA	TRP	B	281	43.554	39.724	59.908	1.00	21.51	-0.180	C
ATOM	14244	C	TRP	B	281	43.271	38.236	59.914	1.00	24.46	-0.241	C
ATOM	14245	CB	TRP	B	281	42.757	37.438	58.271	1.00	21.11	-0.271	CB
ATOM	14246	CD	TRP	B	281	42.310	40.510	60.406	1.00	23.59	-0.073	CD
ATOM	14247	CE	TRP	B	281	42.694	41.989	60.375	1.00	27.28	-0.056	A
ATOM	14248	CF	TRP	B	281	43.527	42.501	61.379	1.00	30.29	0.010	A
ATOM	14249	CG	TRP	B	281	42.280	42.817	59.351	1.00	28.09	0.010	A
ATOM	14250	CH	TRP	B	281	43.901	43.835	61.355	1.00	34.65	0.037	A
ATOM	14251	CI	TRP	B	281	42.647	44.150	59.326	1.00	34.25	0.086	A
ATOM	14252	CL	TRP	B	281	43.460	44.652	60.336	1.00	39.11	0.065	A
ATOM	14253	CM	TRP	B	281	43.834	45.979	60.303	1.00	40.18	-0.361	DA
ATOM	14254	CN	TRP	B	281	43.609	37.606	61.031	1.00	41.07	-0.346	N
ATOM	14255	CO	TRP	B	282	44.002	38.153	61.797	1.00	40.00	0.163	HD
ATOM	14256	CP	TRP	B	282	44.446	36.368	62.969	1.00	42.32	-0.232	C
ATOM	14257	CQ	TRP	B	282	41.958	35.828	61.259	1.00	49.80	0.243	C
ATOM	14258	CR	TRP	B	282	41.106	36.717	61.397	1.00	49.24	-0.271	DA
ATOM	14259	CS	TRP	B	282	44.104	35.805	62.947	1.00	49.71	0.042	C
ATOM	14260	CT	TRP	B	283	41.630	34.546	61.173	1.00	26.35	-0.337	N
ATOM	14261	CU	TRP	B	283	40.227	34.135	61.173	1.00	21.92	0.179	C
ATOM	14262	CV	TRP	B	283	39.482	34.658	62.368	1.00	21.87	-0.271	DA
ATOM	14263	CH	TRP	B	283	39.923	34.488	63.514	1.00	16.48	-0.271	DA
ATOM	14264	CI	TRP	B	283	40.303	33.600	61.182	1.00	23.75	0.037	C
ATOM	14265	CJ	TRP	B	283	41.785	32.311	60.750	1.00	26.89	0.022	C
ATOM	14266	CK	TRP	B	283	42.540	33.398	60.973	1.00	24.58	0.127	C
ATOM	14267	CL	TRP	B	283	43.252	34.665	62.189	1.00	19.57	-0.346	N
ATOM	14268	CM	TRP	B	284	37.844	35.342	61.230	1.00	0.00	0.163	HD
ATOM	14269	CN	TRP	B	284	37.411	35.671	63.251	1.00	21.26	0.180	C
ATOM	14270	CO	TRP	B	284	36.371	35.396	62.046	1.00	19.44	-0.271	DA
ATOM	14271	CP	TRP	B	284	37.578	37.025	65.187	1.00	24.72	-0.271	DA
ATOM	14272	CQ	TRP	B	284	36.963	34.562	64.238	1.00	18.32	0.073	C
ATOM	14273	CR	TRP	B	284	36.371	35.396	62.046	1.00	19.44	-0.271	DA
ATOM	14274	CS	TRP	B	284	35.172	33.507	62.801	1.00	21.71	0.007	A
ATOM	14275	CT	TRP	B	284	37.089	32.213	63.323	1.00	22.37	0.007	A
ATOM	14276	CU	TRP	B	284	34.601	31.448	62.571	1.00	24.79	0.001	A
ATOM	14277	CV	TRP	B	284	35.373	31.275	61.935	1.00	22.18	0.000	A
ATOM	14278	CH	TRP	B	285	36.927	34.583	63.471	1.00	23.24	0.086	A
ATOM	14279	CI	TRP	B	285	39.230	37.305	62.534	1.00	0.00	0.163	HD
ATOM	14280	CJ	TRP	B	285	39.517	38.721	64.107	1.00	21.20	0.177	C
ATOM	14281	CK	TRP	B	285	41.427	37.589	62.268	1.00	40.80	0.086	A
ATOM	14282	CL	TRP	B	285	39.356	40.083	62.160	1.00	20.12	-0.271	DA
ATOM	14283	CM	TRP	B	285	41.047	38.595	64.074	1.00	25.33	0.045	C
ATOM	14284	CN	TRP	B	285	41.427	37.589	62.268	1.00	40.80	0.086	A
ATOM	14285	CO	TRP	B	285	42.944	37.983	65.625	1.00	59.84	0.172	C
ATOM	14286	CP	TRP	B	285	43.439	39.109	65.386	1.00	74.19	-0.648	DA
ATOM	14287	CQ	TRP	B	285	41.570	38.153	61.797	1.00	40.00	-0.346	N
ATOM	14288	CR	TRP	B	286	38.393	40.873	64.056	1.00	16.86	-0.346	N
ATOM	14289	CS	TRP	B	286	38.154	40.712	65.034	1.00	0.00	0.163	HD
ATOM	14290	CT	TRP	B	286	39.966	42.109	63.929	1.00	28.27	-0.271	DA
ATOM	14291	CU	TRP	B	286	38.535	43.260	64.182	1.00	18.51	0.243	C
ATOM	14292	CV	TRP	B	286	38.255	43.269	65.380	1.00	19.21	-0.271	DA
ATOM	14293	CH	TRP	B	286	36.425	42.211	64.949	1.00	16.98	-0.271	DA
ATOM	14294	CI	TRP	B	286	35.727	41.089	62.573	1.00	21.39	0.002	C
ATOM	14295	CJ	TRP	B	286	36.018	43.583	62.803	1.00	17.10	0.012	C
ATOM	14296	CK	TRP	B	286	36.087	43.004	64.044	1.00	19.94	0.086	A
ATOM	14297	CL	TRP	B	286	39.343	44.213	63.618	1.00	21.84	-0.337	N
ATOM	14298	CM	TRP	B	287	39.917	45.135	64.423	1.00	21.50	0.179	C
ATOM	14299	CN	TRP	B	287	38.869	44.627	64.166	1.00	25.59	0.086	A
ATOM	14300	CO	TRP	B	287	37.854	46.537	64.279	1.00	17.06	-0.271	DA
ATOM	14301	CP	TRP	B	287	40.496	45.891	63.466	1.00	23.29	0.037	C
ATOM	14302	CQ	TRP	B	287	42.425	46.963	62.926	1.00	26.67	0.000	A
ATOM	14303	CR	TRP	B	287	39.727	44.176	62.205	1.00	19.56	0.127	C
ATOM	14304	CS	TRP	B	287	39.158	46.817	66.064	1.00	22.67	-0.346	N
ATOM	14305	CT	TRP	B	288	40.018	46.867	66.376	1.00	20.90	-0.346	N
ATOM	14306	CU	TRP	B	288	38.311	47.833	66.641	1.00	24.77	0.200	C
ATOM	14307	CV	TRP	B	288	37.842	48.909	65.684	1.00	23.27	0.243	C
ATOM	14308	CH	TRP	B	288	36.659	48.329	64.829	1.00	20.90	-0.271	DA
ATOM	14309	CI	TRP	B	288	39.114	48.549	67.767	1.00	30.43	-0.199	C
ATOM	14310	CJ	TRP	B	288	38.306	48.354	68.924	1.00	40.06	-0.398	DA
ATOM	14311	CK	TRP	B	288	38.789	48.289	69.140	1.00	40.45	-0.346	N
ATOM	14312	CL	TRP	B	288	38.739	49.428	64.870	1.00	23.43	-0.346	N
ATOM	14313	CM	TRP	B	289	39.693	49.428	64.870	1.00	23.43	-0.346	N
ATOM	14314	CN	TRP	B	289	38.468	50.474	62.899	1.00	27.07	0.000	A
ATOM	14315	CO	TRP	B	289	37.407	50.996	62.857	1.00	27.38	0.241	C
ATOM	14316	CP	TRP	B	289	36.872	51.219	64.044	1.00	21.85	-0.271	DA
ATOM	14317	CQ	TRP	B	289	39.805	50.818	63.247	1.00	37.22	0.045	C
ATOM	14318	CR	TRP	B	289	39.798	51.998	62.299	1.00	61.71	0.116	C
ATOM	14319	CS	TRP	B	289	37.416	48.834	64.476	1.00	22.44	-0.271	DA
ATOM	14320	CT	TRP	B	289	41.953	51.202	61.638	1.00	84.94	-0.648	DA
ATOM	14321	CU	TRP	B	289	41.123	52.926	60.543	1.00	81.85	-0.648	DA
ATOM	14322	CV	TRP	B	289	37.346	48.834	64.476	1.00	22.44	-0.271	DA
ATOM	14323	CH	TRP	B	290	38.007	48.168	62.862	1.00	0.00	0.163	HD
ATOM	14324	CI	TRP	B	290	36.337	48.335	61.531	1.00	20.77	0.180	C
ATOM	14325	CJ	TRP	B	290	38.018	47.662	60.919	1.00	17.24	0.086	A
ATOM	14326	CK	TRP	B	290	39.987	48.668	61.681	1.00	17.67	-0.271	DA
ATOM	14327	CL	TRP	B	290	36.765	46.988	60.931	1.00	19.67	0.013	C
ATOM	14328	CM	TRP	B	290	38.018	47.662	60.919	1.00	17.24	0.086	A
ATOM	14329	CN	TRP	B	290	35.625	46.360	60.170	1.00	17.48	0.012	C
ATOM	14330	CO	TRP	B	290	37.789	48.158	58.876	0.50	22.46	0.005	C
ATOM	14331	CP	TRP	B	290	34.252	46.863	60.919	1.00	14.22	-0.271	DA
ATOM	14332	CQ	TRP	B	291	35.783	47.237	63.853	1.00	0.00	0.163	HD
ATOM	14333	CR	TRP	B	291	33.679	49.670	64.175	1.00	18.04	0.180	C
ATOM	14334	CS	TRP	B	291	33.12	49.670	64.175	1.00	21.93	0.146	C
ATOM	14335	CT	TRP	B	291	31.907	49.245	64.277	1.00	18.09	-0.271	DA
ATOM	14336	CU	TRP	B	291	33.784	47.114	65.598	1.00	16.53	0.073	C
ATOM	14337	CV	TRP	B	291	33.788	45.601	65.161	1.00	16.56	0.086	A
ATOM	14338	CH	TRP	B	291	32.708	44.856	65.212	1.00	14.86	0.010	A
ATOM	14339	CI	TRP	B	291	34.880	44.899	66.166	1.00	16.76	0.010	A
ATOM	14340	CJ	TRP	B	291	36.425	44.899	66.166	1.00	16.76	0.010	A
ATOM	14341	CK	TRP	B	291	34.864	43.518	66.263	1.00	18.68	0.037	A
ATOM	14342	CL	TRP	B	291	33.740	42.811	65.854	1.00	19.81	0.065	A
ATOM	14343	CM	TRP	B	291	33.750	43.439	65.854	1.00	19.81	0.065	A
ATOM	14344	CN	TRP	B	291	34.488	40.979	66.283	1.00	0.00	0.217	HD
ATOM	14345	CO	TRP	B	292	33.958	50.075	64.669	1.00	20		

ATOM	14798	HE2	LVS	B	316	1.146	64.903	75.211	1.00	0.00	0.274	HD
ATOM	14799	HE2	LVS	B	316	-1.250	64.906	75.211	1.00	0.00	0.274	HD
ATOM	14800	N	ALA	B	317	-3.172	63.036	69.754	1.00	12.02	-0.346	N
ATOM	14801	HN	ALA	B	317	-2.279	62.774	69.336	1.00	0.00	0.163	HD
ATOM	14802	CA	ALA	B	317	-4.269	63.040	69.754	1.00	11.66	-0.346	N
ATOM	14804	C	ALA	B	317	-5.211	62.376	68.483	1.00	13.28	0.240	C
ATOM	14805	O	ALA	B	317	-6.411	62.567	68.304	1.00	13.32	-0.271	DA
ATOM	14806	CB	ALA	B	317	-3.610	64.070	67.471	1.00	11.66	0.163	HD
ATOM	14810	N	TYR	B	318	-4.746	61.126	68.351	1.00	7.56	-0.346	N
ATOM	14811	HN	TYR	B	318	-3.774	60.948	68.603	1.00	0.00	0.163	HD
ATOM	14812	CA	TYR	B	318	-5.556	59.990	67.842	1.00	8.29	0.183	C
ATOM	14814	C	TYR	B	318	-5.450	58.809	67.788	1.00	13.42	0.243	C
ATOM	14815	O	TYR	B	318	-4.825	57.780	68.421	1.00	14.01	-0.271	DA
ATOM	14816	CB	TYR	B	318	-4.894	59.593	66.479	1.00	9.00	-0.274	HD
ATOM	14819	CG	TYR	B	318	-4.894	60.742	65.483	1.00	15.44	-0.056	A
ATOM	14820	CD1	TYR	B	318	-6.047	61.239	64.891	1.00	20.17	0.010	A
ATOM	14822	CD2	TYR	B	318	-3.492	61.001	65.197	1.00	12.07	0.074	A
ATOM	14824	CE1	TYR	B	318	-6.003	62.323	63.990	1.00	17.15	0.037	A
ATOM	14826	CE2	TYR	B	318	-3.643	62.406	64.234	1.00	17.68	0.037	A
ATOM	14828	CF	TYR	B	318	-4.826	62.894	63.045	1.00	11.86	0.163	A
ATOM	14829	OH	TYR	B	318	-4.689	63.981	62.812	1.00	20.35	-0.361	OA
ATOM	14830	HH	TYR	B	318	-5.495	64.319	62.439	1.00	0.00	0.217	HD
ATOM	14831	N	PRO	B	319	-5.882	58.903	70.037	1.00	15.58	-0.337	N
ATOM	14832	CA	PRO	B	319	-5.699	57.846	70.107	1.00	16.12	0.179	C
ATOM	14834	C	PRO	B	319	-6.282	56.496	70.645	1.00	16.79	0.241	C
ATOM	14835	O	PRO	B	319	-5.466	55.432	70.956	1.00	15.00	-0.277	DA
ATOM	14836	CB	PRO	B	319	-6.322	58.367	72.328	1.00	18.55	0.037	C
ATOM	14839	CG	PRO	B	319	-7.081	59.609	71.948	1.00	19.63	0.022	C
ATOM	14842	CD	PRO	B	319	-4.788	60.673	70.986	1.00	17.77	0.147	C
ATOM	14845	N	GLN	B	320	-7.453	56.479	69.995	1.00	16.72	-0.346	N
ATOM	14846	HN	GLN	B	320	-6.322	57.521	69.743	1.00	16.00	0.163	HD
ATOM	14847	CA	GLN	B	320	-8.042	55.176	68.642	1.00	16.85	0.177	C
ATOM	14849	C	GLN	B	320	-7.224	54.461	68.555	1.00	18.34	0.241	C
ATOM	14850	O	GLN	B	320	-6.345	53.403	69.287	1.00	17.84	-0.271	DA
ATOM	14851	CB	GLN	B	320	-9.477	55.325	69.203	1.00	22.32	0.044	C
ATOM	14854	CG	GLN	B	320	-10.506	55.992	70.070	1.00	24.74	0.105	C
ATOM	14857	CD	GLN	B	320	-10.345	54.611	69.121	1.00	23.14	0.074	C
ATOM	14858	NE2	GLN	B	320	-10.471	56.788	72.369	1.00	27.59	-0.370	N
ATOM	14859	HE2	GLN	B	320	-10.363	56.617	73.369	1.00	0.00	0.159	HD
ATOM	14860	2HE2	GLN	B	320	-10.725	57.221	72.021	1.00	0.00	0.232	HD
ATOM	14861	OE1	GLN	B	320	-10.104	54.601	71.978	1.00	83.41	-0.274	OA
ATOM	14862	N	GLU	B	321	-6.783	55.225	67.563	1.00	12.91	-0.346	N
ATOM	14863	HN	GLU	B	321	-5.046	56.240	67.021	1.00	0.00	0.163	HD
ATOM	14864	CA	GLU	B	321	-5.941	54.716	66.486	1.00	11.77	0.177	C
ATOM	14866	C	GLU	B	321	-4.629	54.230	67.046	1.00	13.44	0.241	C
ATOM	14867	O	GLU	B	321	-5.157	53.148	67.819	1.00	13.85	-0.271	DA
ATOM	14868	CB	GLU	B	321	-5.718	55.325	65.998	1.00	13.76	0.045	C
ATOM	14871	CG	GLU	B	321	-7.036	56.055	64.654	1.00	22.52	0.116	C
ATOM	14874	CD	GLU	B	321	-7.922	57.127	64.398	1.00	27.01	0.074	C
ATOM	14875	OE1	GLU	B	321	-7.799	57.565	66.404	1.00	23.95	-0.648	OA
ATOM	14876	OE2	GLU	B	321	-8.814	57.589	66.491	1.00	49.48	-0.648	OA
ATOM	14877	N	ALA	B	322	-4.274	54.955	67.945	1.00	10.12	0.163	HD
ATOM	14878	HN	ALA	B	322	-4.356	55.854	68.235	1.00	0.00	0.163	HD
ATOM	14879	CA	ALA	B	322	-2.724	54.510	68.538	1.00	12.69	0.172	C
ATOM	14881	C	ALA	B	322	-3.886	53.167	69.129	1.00	15.43	0.241	C
ATOM	14882	O	ALA	B	322	-1.999	52.302	69.159	1.00	13.25	-0.271	DA
ATOM	14883	CB	ALA	B	322	-2.140	55.537	69.507	1.00	13.92	0.042	C
ATOM	14887	N	ALA	B	323	-4.962	53.000	70.163	1.00	14.53	-0.346	N
ATOM	14888	HN	ALA	B	323	-4.654	53.750	70.082	1.00	0.00	0.163	HD
ATOM	14889	CA	ALA	B	323	-4.179	51.727	70.741	1.00	14.94	0.172	C
ATOM	14891	C	ALA	B	323	-4.432	50.588	69.849	1.00	13.40	0.241	C
ATOM	14892	O	ALA	B	323	-3.970	49.465	69.989	1.00	13.47	-0.271	DA
ATOM	14893	CB	ALA	B	323	-5.349	51.835	71.925	1.00	14.37	0.042	C
ATOM	14897	N	GLU	B	324	-4.142	50.841	68.646	1.00	15.58	-0.346	N
ATOM	14898	HN	GLU	B	324	-5.532	51.771	68.497	1.00	0.00	0.163	HD
ATOM	14899	CA	GLU	B	324	-5.356	49.799	67.646	1.00	12.25	0.177	C
ATOM	14901	C	GLU	B	324	-7.042	49.823	66.476	1.00	12.42	0.241	C
ATOM	14902	O	GLU	B	324	-3.805	48.303	66.639	1.00	12.82	-0.271	DA
ATOM	14903	CB	GLU	B	324	-6.429	50.201	66.631	1.00	13.08	0.045	C
ATOM	14906	CG	GLU	B	324	-8.828	49.984	67.138	1.00	19.40	0.074	C
ATOM	14909	CD	GLU	B	324	-8.083	48.477	67.259	1.00	18.23	-0.172	C
ATOM	14910	OE1	GLU	B	324	-8.195	47.953	66.120	1.00	18.17	-0.648	OA
ATOM	14911	OE2	GLU	B	324	-8.492	47.882	66.198	1.00	26.33	-0.648	OA
ATOM	14912	N	PRO	B	325	-3.222	50.470	66.635	1.00	9.97	-0.346	N
ATOM	14913	HN	PRO	B	325	-3.489	51.433	66.838	1.00	0.00	0.163	HD
ATOM	14914	CA	PRO	B	325	-4.844	50.347	66.197	1.00	12.42	0.172	C
ATOM	14916	C	PRO	B	325	-1.125	49.246	66.100	1.00	14.58	0.241	C
ATOM	14917	O	PRO	B	325	-0.573	48.266	66.360	1.00	11.24	-0.271	DA
ATOM	14918	CB	PRO	B	325	-1.175	51.312	69.743	1.00	13.07	-0.373	C
ATOM	14921	CG	PRO	B	325	0.164	51.332	65.103	1.00	15.66	-0.056	A
ATOM	14922	CD	PRO	B	325	-0.230	50.026	64.731	1.00	17.78	0.074	C
ATOM	14924	CE2	PRO	B	325	1.316	51.269	65.874	1.00	15.59	0.007	A
ATOM	14926	CE1	PRO	B	325	1.516	51.156	63.121	1.00	13.96	0.001	A
ATOM	14928	CF	PRO	B	325	-0.230	50.434	65.044	1.00	15.13	0.074	C
ATOM	14930	CF	PRO	B	325	2.654	51.097	63.907	1.00	12.39	0.001	A
ATOM	14932	N	THR	B	326	-0.996	49.592	68.143	1.00	12.00	-0.346	N
ATOM	14933	HN	THR	B	326	-1.042	49.544	68.143	1.00	0.00	0.163	HD
ATOM	14934	CA	THR	B	326	-0.181	48.713	69.033	1.00	14.15	0.205	C
ATOM	14936	C	THR	B	326	-0.792	47.334	69.187	1.00	14.38	0.243	C
ATOM	14937	O	THR	B	326	-0.620	46.310	69.187	1.00	14.38	-0.271	DA
ATOM	14938	CB	THR	B	326	-0.206	49.487	70.399	1.00	18.96	0.146	C
ATOM	14940	CG	THR	B	326	0.314	48.651	71.530	1.00	29.33	0.042	C
ATOM	14944	CD	THR	B	326	-2.702	48.620	69.187	1.00	28.45	-0.271	DA
ATOM	14945	HG1	THR	B	326	0.616	51.122	71.005	1.00	0.00	0.210	HD
ATOM	14946	N	ARG	B	327	-2.110	47.227	69.306	1.00	11.34	-0.346	N
ATOM	14947	HN	ARG	B	327	-1.702	48.225	69.306	1.00	0.00	0.163	HD
ATOM	14948	CA	ARG	B	327	-2.707	45.865	69.456	1.00	11.23	0.176	C
ATOM	14950	C	ARG	B	327	-2.448	45.016	68.215	1.00	13.27	0.241	C
ATOM	14951	O	ARG	B	327	-3.431	44.816	68.290	1.00	14.48	-0.271	DA
ATOM	14952	CB	ARG	B	327	-4.213	46.301	69.685	1.00	10.67	0.036	C
ATOM	14955	CG	ARG	B	327	-5.012	44.725	69.866	1.00	12.02	0.023	C
ATOM	14958	CD	ARG	B	327	-4.489	43.998	69.876	1.00	14.44	0.074	C
ATOM	14961	NE	ARG	B	327	-7.043	45.377	68.407	1.00	12.85	-0.227	C
ATOM	14962	HE	ARG	B	327	-7.302	46.357	68.289	1.00	0.00	0.177	HD
ATOM	14963	CE	ARG	B	327	-7.243	45.941	68.167	1.00	12.63	-0.271	DA
ATOM	14964	NH1	ARG	B	327	-6.878	43.283	67.369	1.00	11.16	-0.235	N
ATOM	14965	NH1	ARG	B	327	-7.032	42.672	66.267	1.00	0.00	0.174	HD
ATOM	14966	NH2	ARG	B	327	-6.452	42.882	68.299	1.00	12.64	-0.346	N
ATOM	14967	NH2	ARG	B	327	-7.804	45.098	66.363	1.00	13.44	-0.235	N
ATOM	14968	NH2	ARG	B	327							

ATOM 15365	N	ALA B 353	-29.577	25.400	59.747	1.00	15.47	-0.346	N
ATOM 15366	HN	ALA B 353	-29.577	25.400	59.747	1.00	15.47	-0.346	N
ATOM 15367	CA	ALA B 353	-29.666	25.687	58.309	1.00	13.43	0.172	C
ATOM 15368	C	ALA B 353	-28.871	24.611	57.576	1.00	16.43	0.240	C
ATOM 15370	O	ALA B 353	-27.788	24.663	56.871	1.00	17.14	-0.278	O
ATOM 15371	CB	ALA B 353	-29.043	27.050	58.063	1.00	12.18	0.042	C
ATOM 15375	N	LYS B 354	-29.358	24.028	56.499	1.00	14.11	-0.346	N
ATOM 15376	HN	LYS B 354	-29.358	24.028	56.499	1.00	14.11	-0.346	N
ATOM 15377	CA	LYS B 354	-28.646	23.031	55.725	1.00	13.67	0.176	C
ATOM 15379	C	LYS B 354	-27.937	23.774	54.590	1.00	13.62	0.241	C
ATOM 15380	O	LYS B 354	-28.604	23.985	53.967	1.00	12.57	-0.271	O
ATOM 15381	CB	LYS B 354	-29.628	25.022	55.106	1.00	15.44	0.035	C
ATOM 15384	CG	LYS B 354	-28.874	20.826	54.486	1.00	16.23	0.004	C
ATOM 15387	CD	LYS B 354	-29.956	19.852	53.976	1.00	25.96	0.072	C
ATOM 15390	CE	LYS B 354	-29.335	18.755	53.128	1.00	41.05	0.229	C
ATOM 15393	NZ	LYS B 354	-30.343	18.185	52.180	1.00	45.81	-0.079	N
ATOM 15394	HE1	LYS B 354	-26.659	24.222	54.486	1.00	15.00	0.172	C
ATOM 15395	HE2	LYS B 354	-30.772	18.909	51.604	1.00	0.00	0.274	HD
ATOM 15396	HE3	LYS B 354	-31.179	17.862	52.667	1.00	0.00	0.274	HD
ATOM 15397	N	LYS B 354	-26.659	24.222	54.486	1.00	15.00	0.172	C
ATOM 15398	HN	LYS B 354	-26.659	24.222	54.486	1.00	15.00	0.172	C
ATOM 15399	CA	LYS B 354	-26.017	25.022	53.796	1.00	9.82	0.180	C
ATOM 15401	C	LYS B 354	-24.588	24.470	53.976	1.00	8.71	0.242	C
ATOM 15402	O	LYS B 354	-24.165	23.647	54.433	1.00	10.65	-0.271	O
ATOM 15403	CB	LYS B 354	-25.894	26.512	54.145	1.00	13.71	0.013	C
ATOM 15405	CG1	LYS B 354	-25.498	26.903	55.508	1.00	14.08	0.332	C
ATOM 15408	CG2	LYS B 354	-27.269	27.205	54.203	1.00	14.94	0.012	C
ATOM 15412	CD1	LYS B 354	-24.907	28.135	55.906	1.00	12.10	0.005	C
ATOM 15416	N	ALA B 356	-29.435	24.866	52.511	1.00	9.21	-0.376	N
ATOM 15417	HN	ALA B 356	-24.362	25.517	51.852	1.00	0.00	0.163	HD
ATOM 15418	CA	ALA B 356	-28.573	24.322	52.245	1.00	10.20	0.172	C
ATOM 15420	C	ALA B 356	-21.662	24.903	53.339	1.00	9.42	0.240	C
ATOM 15421	O	ALA B 356	-21.835	26.091	53.707	1.00	12.77	-0.271	O
ATOM 15422	CB	ALA B 356	-29.542	27.244	53.000	1.00	11.00	0.042	C
ATOM 15426	N	SER B 357	-20.579	24.218	57.720	1.00	9.12	-0.344	N
ATOM 15427	HN	SER B 357	-20.391	23.280	53.366	1.00	0.00	0.163	HD
ATOM 15428	CA	SER B 357	-18.779	23.734	54.673	1.00	11.00	0.006	C
ATOM 15430	C	SER B 357	-18.887	25.989	54.010	1.00	7.61	0.243	C
ATOM 15431	O	SER B 357	-18.407	26.878	54.715	1.00	9.80	-0.271	O
ATOM 15432	CB	SER B 357	-18.779	23.734	54.673	1.00	9.18	0.139	C
ATOM 15435	CG	SER B 357	-18.110	23.018	54.454	1.00	7.80	-0.398	O
ATOM 15436	HG	SER B 357	-17.555	22.395	54.909	1.00	0.00	0.209	HD
ATOM 15437	N	ARG B 358	-18.936	26.636	52.925	1.00	0.00	0.163	N
ATOM 15438	HN	ARG B 358	-19.204	25.335	52.094	1.00	0.00	0.163	HD
ATOM 15439	CA	ARG B 358	-18.211	27.274	52.106	1.00	10.50	0.176	C
ATOM 15441	C	ARG B 358	-18.126	28.477	52.000	1.00	12.94	0.243	C
ATOM 15442	O	ARG B 358	-18.606	29.602	52.568	1.00	9.92	-0.271	O
ATOM 15443	CB	ARG B 358	-17.985	27.150	50.597	1.00	8.98	0.036	C
ATOM 15446	CG	ARG B 358	-17.265	27.027	49.467	1.00	12.43	0.006	C
ATOM 15449	CD	ARG B 358	-18.858	27.120	48.292	1.00	15.01	0.138	C
ATOM 15452	NE	ARG B 358	-19.994	26.822	47.446	1.00	21.44	-0.227	N
ATOM 15453	HE	ARG B 358	-18.739	26.364	47.378	1.00	11.00	0.006	C
ATOM 15454	CA	ARG B 358	-20.152	27.062	46.159	1.00	26.59	0.665	C
ATOM 15455	NH1	ARG B 358	-19.204	27.634	45.424	1.00	18.58	-0.235	N
ATOM 15456	NH2	ARG B 358	-18.319	27.926	45.256	1.00	17.44	0.006	C
ATOM 15457	2NH1	ARG B 358	-19.326	27.820	44.429	1.00	0.00	0.174	HD
ATOM 15458	NH2	ARG B 358	-21.311	26.680	45.614	1.00	22.80	-0.235	N
ATOM 15459	H2	ARG B 358	-22.039	26.241	44.789	1.00	20.00	-0.174	HD
ATOM 15460	2H2	ARG B 358	-21.433	26.864	44.619	1.00	0.00	0.174	HD
ATOM 15461	N	LYS B 359	-20.435	28.253	52.449	1.00	9.73	-0.346	N
ATOM 15462	HN	LYS B 359	-20.435	28.253	52.449	1.00	9.73	-0.346	N
ATOM 15463	CA	LYS B 359	-21.315	29.403	52.742	1.00	10.41	0.176	C
ATOM 15465	C	LYS B 359	-21.251	29.693	54.238	1.00	10.00	0.241	C
ATOM 15466	O	LYS B 359	-21.262	29.869	54.000	1.00	9.78	-0.271	O
ATOM 15467	CB	LYS B 359	-22.745	29.115	52.292	1.00	8.48	0.035	C
ATOM 15470	CG	LYS B 359	-23.784	30.189	52.528	1.00	15.50	0.004	C
ATOM 15473	CD	LYS B 359	-24.359	31.966	51.923	1.00	17.47	0.027	C
ATOM 15476	CE	LYS B 359	-24.439	32.546	52.114	1.00	31.31	0.229	C
ATOM 15479	NE	LYS B 359	-23.947	33.876	52.512	1.00	20.00	-0.079	N
ATOM 15480	HE1	LYS B 359	-24.427	34.572	52.674	1.00	21.00	0.006	C
ATOM 15481	HE2	LYS B 359	-23.273	34.221	51.828	1.00	0.00	0.274	HD
ATOM 15482	HE3	LYS B 359	-23.345	33.798	53.332	1.00	0.00	0.274	HD
ATOM 15483	N	ALA B 360	-21.642	29.403	52.511	1.00	9.21	-0.376	N
ATOM 15484	HN	ALA B 360	-21.186	27.686	54.714	1.00	0.00	0.163	HD
ATOM 15485	CA	ALA B 360	-21.035	28.884	56.504	1.00	10.77	0.172	C
ATOM 15487	C	ALA B 360	-19.779	27.983	55.420	1.00	9.18	0.243	C
ATOM 15488	O	ALA B 360	-19.685	30.473	57.762	1.00	10.03	-0.271	O
ATOM 15489	CB	ALA B 360	-21.017	27.518	57.245	1.00	8.36	0.042	C
ATOM 15493	N	ARG B 361	-18.740	29.418	52.925	1.00	9.58	-0.344	N
ATOM 15494	HN	ARG B 361	-18.827	28.695	55.275	1.00	0.00	0.163	HD
ATOM 15495	CA	ARG B 361	-18.827	30.163	54.128	1.00	9.04	0.176	C
ATOM 15497	C	ARG B 361	-17.650	31.631	55.770	1.00	10.42	0.243	C
ATOM 15498	O	ARG B 361	-17.248	32.499	56.582	1.00	11.29	-0.271	O
ATOM 15499	CB	ARG B 361	-19.262	29.467	54.254	1.00	9.16	0.036	C
ATOM 15502	CG	ARG B 361	-15.265	30.335	55.150	1.00	11.24	-0.398	O
ATOM 15503	HG	ARG B 361	-14.609	29.907	54.613	1.00	0.00	0.209	HD
ATOM 15504	N	GLN B 362	-18.902	30.428	54.639	1.00	9.16	-0.346	N
ATOM 15505	HN	GLN B 362	-18.625	31.155	54.005	1.00	0.00	0.163	HD
ATOM 15506	CA	GLN B 362	-18.606	33.328	54.365	1.00	10.07	0.177	C
ATOM 15508	C	GLN B 362	-17.439	35.966	55.487	1.00	11.17	0.243	C
ATOM 15509	O	GLN B 362	-19.173	35.118	55.814	1.00	10.64	-0.271	O
ATOM 15510	CB	GLN B 362	-19.421	33.425	53.058	1.00	11.08	0.044	C
ATOM 15513	CG	GLN B 362	-19.544	34.901	52.100	1.00	15.27	0.012	C
ATOM 15516	CD	GLN B 362	-20.261	35.038	51.284	1.00	22.93	0.215	C
ATOM 15517	NE	GLN B 362	-19.736	35.816	50.333	1.00	21.54	-0.370	N
ATOM 15518	H2	GLN B 362	-18.937	36.364	50.400	1.00	19.39	0.006	C
ATOM 15519	2H2	GLN B 362	-20.216	35.903	49.437	1.00	0.00	0.159	HD
ATOM 15520	OE1	ASN B 363	-21.279	34.378	51.091	1.00	20.79	-0.274	O
ATOM 15521	N	ASN B 363	-20.427	33.827	50.400	1.00	19.39	0.006	C
ATOM 15522	HN	ASN B 363	-20.632	32.306	55.777	1.00	0.00	0.163	HD
ATOM 15523	CA	ASN B 363	-21.206	33.854	57.168	1.00	11.81	0.185	C
ATOM 15525	C	ASN B 363	-20.347	34.169	58.914	1.00	15.43	0.243	C
ATOM 15526	O	ASN B 363	-20.614	35.118	59.114	1.00	12.99	-0.271	O
ATOM 15527	CB	ASN B 363	-22.370	32.951	57.631	1.00	8.61	0.137	C
ATOM 15530	CG	ASN B 363	-24.433	32.647	56.486	1.00	15.27	0.017	C
ATOM 15531	ND2	ASN B 363	-24.212	31.710	56.642	1.00	14.84	-0.370	N
ATOM 15532	H2	ASN B 363	-24.926	31.591	55.923	1.00	0.00	0.159	HD
ATOM 15533	2H2	ASN B 363	-24.159	31.429	55.943	1.00	0.00	0.159	HD
ATOM 15534	OD1	ASN B 363	-23.506	33.589	55.643	1.00	14.10	-0.274	O
ATOM 15535	N	ALA B 364	-19.338	33.360	58.716	1.00	11.19	-0.346	N
ATOM 15536	HN	ALA B 364	-19.338	33.360	58.716	1.00	11.19	-0.346	N
ATOM 15537	CA	ALA B 364	-18.425	33.663	59.813	1.00	14.05	0.172	C
ATOM 15539	C	ALA B 364	-17.511	34.835	59.505	1.00	12.75	0.240	C
ATOM 15540	O	ALA B 364	-17.219	34.627	60.387	1.00	13.71	-0.271	O
ATOM 15541	CB	ALA B 364	-17.608	32.370	60.177	1.00	12.11	0.042	C
ATOM 15545	N	LIE B 365	-17.037	34.987	58.244	1.00	11.14	-0.346	N
ATOM 15546	HN	LIE B 365	-17.037	34.987	58.244	1.00	11.14	-0.346	N
ATOM 15547	CA	LIE B 365	-16.241	36.135	57.858	1.00	10.10	0.180	C
ATOM 15549	C	LIE B 365	-17.103	37.405	58.046	1.00	14.73	0.241	C
ATOM 15550	O	LIE B 365	-16.644	38.440	58.113	1.00	11.43	-0.271	O
ATOM 15551	CB	LIE B 365	-15.735	36.071	56.399	1.00	8.28	0.013	C
ATOM 15553	CG1	LIE B 365	-14.712	34.941	56.207	1.00	8.03	0.002	C
ATOM									

AT0015947	CA	SER	B	393	-14.472	43.937	51.881	1.00	11.223	0.200	C
AT0015948	CA	SER	B	396	-2.420	46.755	50.146	1.00	11.078	0.271	CA
AT0015950	CA	SER	B	393	-15.192	45.528	50.241	1.00	13.664	-0.271	OA
AT0015951	CB	SER	B	393	-14.546	42.737	50.903	1.00	15.711	0.199	C
AT0015954	CG	SER	B	393	-10.388	46.066	50.146	1.00	11.004	-0.271	CA
AT0015955	HG	SER	B	393	-14.602	40.813	51.101	1.00	0.000	0.209	HD
AT0015956	N	LVS	B	394	-13.067	45.664	50.922	1.00	12.779	-0.346	N
AT0015957	N	LVS	B	394	-12.372	45.226	51.396	1.00	12.000	0.176	N
AT0015958	CA	LVS	B	394	-12.670	46.713	49.980	1.00	12.668	0.176	C
AT0015960	C	LVS	B	394	-11.341	46.252	49.390	1.00	13.443	0.241	C
AT0015961	C	LVS	B	394	-10.388	46.066	50.146	1.00	11.004	-0.271	CA
AT0015962	CB	LVS	B	394	-12.406	48.040	50.687	1.00	18.223	0.035	C
AT0015965	CG	LVS	B	394	-13.658	48.656	51.299	1.00	33.211	0.004	C
AT0015968	CD	LVS	B	394	-13.295	49.297	52.672	1.00	51.226	0.076	C
AT0015971	CE	LVS	B	394	-14.427	50.154	53.167	1.00	62.779	0.229	C
AT0015974	NZ	LVS	B	394	-14.256	50.466	54.613	1.00	71.333	-0.079	N
AT0015975	HZ1	LVS	B	394	-13.349	50.896	54.794	1.00	0.000	0.274	HD
AT0015976	HZ2	LVS	B	394	-14.144	49.615	55.164	1.00	0.000	0.274	HD
AT0015978	N	ALA	B	395	-12.022	46.394	47.499	1.00	10.115	0.163	HD
AT0015979	HN	ALA	B	395	-10.043	45.576	47.484	1.00	10.322	0.172	C
AT0015982	C	ALA	B	395	-8.956	46.929	47.522	1.00	11.582	0.240	C
AT0015983	O	ALA	B	395	-9.247	47.764	47.031	1.00	12.028	-0.271	OA
AT0015984	CB	ALA	B	395	-10.381	45.110	46.027	1.00	13.255	0.042	C
AT0015988	N	LIE	B	396	-7.504	45.245	47.838	1.00	0.000	0.163	HD
AT0015990	CA	LIE	B	396	-6.820	47.204	47.787	1.00	10.877	0.180	C
AT0015992	C	LIE	B	396	-6.154	47.659	46.974	1.00	14.644	0.245	C
AT0015993	O	LIE	B	396	-5.394	48.642	46.328	1.00	14.442	-0.271	OA
AT0015994	CG	LIE	B	396	-4.722	48.577	48.939	1.00	11.778	0.172	C
AT0015996	CG1	LIE	B	396	-4.670	45.617	47.984	1.00	17.771	0.002	C
AT0015999	CG2	LIE	B	396	-5.922	46.382	50.070	1.00	7.511	0.012	C
AT0016003	CG3	LIE	B	396	-4.945	47.213	49.522	1.00	15.920	0.076	C
AT0016007	N	ASN	B	397	-6.600	47.056	45.302	1.00	16.114	-0.346	N
AT0016008	HN	ASN	B	397	-7.168	46.259	45.338	1.00	0.000	0.163	HD
AT0016009	CA	ASN	B	397	-6.222	46.717	44.923	1.00	22.455	0.172	C
AT0016011	C	ASN	B	397	-7.162	48.873	43.764	1.00	20.202	0.241	C
AT0016012	O	ASN	B	397	-6.867	49.701	42.884	1.00	25.444	-0.271	OA
AT0016013	CB	ASN	B	397	-6.867	49.701	42.884	1.00	25.444	-0.271	CA
AT0016016	CG	ASN	B	397	-7.571	46.096	42.739	1.00	21.994	0.217	C
AT0016017	ND2	ASN	B	397	-8.286	46.363	43.635	1.00	29.883	-0.370	N
AT0016018	2HD2	ASN	B	397	-7.945	45.289	43.635	1.00	0.000	0.159	HD
AT0016019	2HD2	ASN	B	397	-9.203	45.923	43.563	1.00	0.000	0.159	HD
AT0016020	OD1	ASN	B	397	-7.980	45.389	43.643	1.00	20.886	-0.274	OA
AT0016021	N	GLD	B	398	-4.988	47.152	47.152	1.00	14.266	0.163	HD
AT0016022	CA	GLD	B	398	-8.473	48.304	45.172	1.00	0.000	0.177	C
AT0016023	CA	GLD	B	398	-9.169	50.146	44.360	1.00	18.009	0.163	HD
AT0016025	C	GLD	B	398	-8.967	52.367	45.306	1.00	23.335	-0.271	OA
AT0016026	CB	GLD	B	398	-10.632	49.991	44.392	1.00	19.298	0.045	C
AT0016030	CG	GLD	B	398	-12.740	49.574	43.406	1.00	44.444	0.172	C
AT0016033	CD	GLD	B	398	-12.380	48.098	43.316	1.00	41.855	-0.172	C
AT0016034	OE1	GLD	B	398	-13.182	48.197	44.272	1.00	47.355	-0.648	OA
AT0016035	OE2	GLD	B	398	-7.405	47.576	42.231	1.00	49.119	0.172	C
AT0016036	N	ASF	B	399	-8.828	50.662	46.720	1.00	13.882	-0.346	N
AT0016037	HN	ASF	B	399	-8.810	49.991	46.852	1.00	0.000	0.163	HD
AT0016038	CA	ASF	B	399	-6.413	47.533	47.533	1.00	14.778	0.172	C
AT0016040	C	ASF	B	399	-7.471	51.042	48.670	1.00	15.922	0.241	C
AT0016041	O	ASF	B	399	-7.517	49.951	49.265	1.00	13.003	-0.271	OA
AT0016042	CB	ASF	B	399	-8.965	51.402	48.339	1.00	15.726	0.172	C
AT0016045	CG	ASF	B	399	-9.973	52.084	50.034	1.00	27.888	-0.175	C
AT0016046	OD1	ASF	B	399	-8.991	52.084	50.034	1.00	19.448	-0.648	OA
AT0016047	OD2	ASF	B	399	-11.039	52.929	49.674	1.00	32.689	0.172	C
AT0016048	N	ALA	B	400	-6.387	51.827	48.714	1.00	12.533	-0.346	N
AT0016049	HN	ALA	B	400	-6.411	52.734	48.248	1.00	0.000	0.163	HD
AT0016050	CA	ALA	B	400	-4.123	51.439	49.410	1.00	14.416	0.172	C
AT0016052	C	ALA	B	400	-5.407	51.106	50.878	1.00	13.229	0.240	C
AT0016053	O	ALA	B	400	-4.572	50.394	51.468	1.00	12.253	-0.271	OA
AT0016054	CB	ALA	B	400	-4.125	52.740	49.395	1.00	10.000	0.172	C
AT0016058	N	ALA	B	401	-6.457	51.628	51.506	1.00	10.588	-0.346	N
AT0016059	HN	ALA	B	401	-7.111	52.230	51.005	1.00	0.000	0.163	HD
AT0016060	CA	ALA	B	401	-6.668	51.924	52.934	1.00	16.454	0.172	C
AT0016062	C	ALA	B	401	-7.600	50.455	53.134	1.00	11.336	0.240	C
AT0016063	O	ALA	B	401	-8.158	49.959	54.224	1.00	12.774	-0.271	OA
AT0016064	CB	ALA	B	401	-7.288	52.642	51.461	1.00	10.000	0.172	C
AT0016068	N	GLV	B	402	-7.785	49.320	52.103	1.00	12.021	-0.351	N
AT0016069	HN	GLV	B	402	-7.247	49.477	51.251	1.00	0.000	0.163	HD
AT0016070	CA	GLV	B	402	-8.711	48.212	53.659	1.00	17.078	0.172	C
AT0016073	C	GLV	B	402	-8.270	46.963	52.872	1.00	16.111	0.236	C
AT0016074	O	GLV	B	402	-7.902	47.028	52.624	1.00	12.334	-0.271	OA
AT0016075	N	ASN	B	403	-8.973	45.949	52.621	1.00	9.881	-0.346	N
AT0016076	HN	ASN	B	403	-9.667	45.834	51.874	1.00	0.000	0.163	HD
AT0016077	CA	ASN	B	403	-7.131	44.646	53.437	1.00	9.719	0.172	C
AT0016079	C	ASN	B	403	-8.971	43.376	52.620	1.00	10.666	0.241	C
AT0016080	O	ASN	B	403	-9.212	42.302	53.187	1.00	10.335	-0.271	OA
AT0016081	CB	ASN	B	403	-8.747	43.200	53.598	1.00	10.000	0.172	C
AT0016084	CG	ASN	B	403	-11.175	44.757	54.150	1.00	18.129	0.217	C
AT0016085	ND2	ASN	B	403	-12.131	45.116	54.988	1.00	16.667	-0.370	N
AT0016086	2HD2	ASN	B	403	-11.872	45.323	55.953	1.00	0.000	0.159	HD
AT0016087	2HD2	ASN	B	403	-11.872	45.323	55.953	1.00	0.000	0.159	HD
AT0016088	OD1	ASN	B	403	-11.490	44.506	52.979	1.00	12.112	-0.274	OA
AT0016089	N	TYR	B	404	-8.759	43.402	50.146	1.00	12.111	-0.346	N
AT0016090	HN	TYR	B	404	-8.542	44.442	50.926	1.00	0.000	0.163	HD
AT0016091	CA	TYR	B	404	-8.994	42.530	50.444	1.00	8.333	0.180	C
AT0016093	C	TYR	B	404	-7.871	45.455	48.727	1.00	11.223	-0.271	OA
AT0016094	O	TYR	B	404	-10.499	42.324	50.009	1.00	7.775	0.073	C
AT0016098	CG	TYR	B	404	-10.863	39.731	47.270	1.00	8.444	0.172	C
AT0016099	CD1	TYR	B	404	-10.621	40.937	47.879	1.00	11.162	0.010	A
AT0016101	CD2	TYR	B	404	-11.223	39.911	49.945	1.00	12.663	0.010	A
AT0016103	CE1	TYR	B	404	-10.863	39.731	47.270	1.00	8.444	0.172	C
AT0016105	CE2	TYR	B	404	-11.488	38.715	49.299	1.00	12.116	0.037	A
AT0016107	CG	TYR	B	404	-11.305	38.648	47.939	1.00	9.005	0.065	A
AT0016108	OH	TYR	B	404	-11.549	37.837	48.284	1.00	14.887	0.172	C
AT0016109	HN	TYR	B	404	-11.422	37.377	46.383	1.00	0.000	0.217	HD
AT0016110	N	LIE	B	405	-7.555	41.271	48.812	1.00	8.667	-0.346	N
AT0016111	HN	LIE	B	405	-7.711	40.393	49.284	1.00	0.000	0.163	HD
AT0016112	CA	LIE	B	405	-6.690	41.289	47.639	1.00	9.688	0.180	C
AT0016114	C	LIE	B	405	-7.204	40.359	46.539	1.00	8.522	0.241	C
AT0016115	O	LIE	B	405	-7.388	39.499	46.817	1.00	9.829	-0.271	OA
AT0016116	CB	LIE	B	405	-5.245	40.824	47.994	1.00	11.667	0.013	C
AT0016118	CG1	LIE	B	405	-4.675	41.649	49.136	1.00	15.229	0.002	C
AT0016121	CG2	LIE	B	405	-4.332	40.669	49.395	1.00	16.466	0.163	HD
AT0016125	CD1	LIE	B	405	-3.376	41.305	49.799	1.00	19.883	0.005	C
AT0016129	N	HIS	B	406	-7.372	40.845	45.301	1.00	9.889	-0.346	N
AT0016130	HN	HIS	B								

ATOM	16515	OG1	THR	B	431	-4.326	31.548	52.015	1.00	9.95	-0.393	OA
ATOM	16516	HG1	THR	B	431	-1.404	21.771	47.191	1.00	0.04	0.210	HD
ATOM	16517	N	SER	B	432	-7.348	31.849	49.861	1.00	7.61	-0.344	N
ATOM	16518	HN	SER	B	432	-6.882	32.436	49.149	1.00	0.00	0.163	HD
ATOM	16519	CA	SER	B	432	-8.320	28.782	49.861	1.00	8.47	-0.344	N
ATOM	16520	C	SER	B	432	-8.163	30.427	48.031	1.00	8.65	-0.243	C
ATOM	16521	O	SER	B	432	-7.859	31.179	47.141	1.00	8.80	-0.271	OA
ATOM	16522	CB	SER	B	432	-9.711	28.132	49.149	1.00	8.19	-0.344	N
ATOM	16523	CG	SER	B	432	-10.772	30.536	49.522	1.00	13.98	-0.398	OA
ATOM	16524	OG	SER	B	432	-11.631	30.929	49.620	1.00	0.00	0.209	HD
ATOM	16525	HG	SER	B	432	-8.450	29.159	47.903	1.00	7.43	-0.344	N
ATOM	16526	NN	THR	B	433	-8.720	28.573	48.066	1.00	0.00	0.163	HD
ATOM	16527	CA	THR	B	433	-8.548	28.546	46.466	1.00	6.76	0.205	C
ATOM	16528	C	THR	B	433	-9.259	27.386	46.976	1.00	10.22	0.242	C
ATOM	16529	O	THR	B	433	-9.697	26.843	47.787	1.00	9.44	-0.271	OA
ATOM	16530	CB	THR	B	433	-7.155	28.361	45.839	1.00	8.91	0.146	C
ATOM	16531	CG	THR	B	433	-8.145	25.329	43.989	1.00	10.64	-0.056	A
ATOM	16540	OG1	THR	B	433	-7.365	27.775	44.534	1.00	9.37	-0.393	OA
ATOM	16541	HN	THR	B	433	-6.505	27.661	44.147	1.00	0.00	0.210	HD
ATOM	16542	N	THR	B	434	-9.211	26.445	45.109	1.00	7.48	-0.346	N
ATOM	16543	HN	PHE	B	434	-9.343	26.827	44.689	1.00	0.00	0.163	HD
ATOM	16544	CA	PHE	B	434	-10.046	25.089	45.743	1.00	9.24	0.180	C
ATOM	16545	C	PHE	B	434	-8.986	24.455	46.421	1.00	10.68	0.241	C
ATOM	16547	O	PHE	B	434	-7.791	24.411	46.190	1.00	9.51	-0.271	OA
ATOM	16548	CB	PHE	B	434	-10.301	24.461	44.552	1.00	7.81	0.073	C
ATOM	16551	CG	PHE	B	434	-11.445	25.789	44.217	1.00	12.06	0.007	A
ATOM	16552	CD1	PHE	B	434	-11.533	24.870	42.211	1.00	22.35	0.007	A
ATOM	16556	CE1	PHE	B	434	-12.455	26.344	43.486	1.00	13.59	0.073	A
ATOM	16558	CE2	PHE	B	434	-12.612	25.380	41.492	1.00	16.01	0.001	A
ATOM	16560	CE3	PHE	B	434	-13.450	24.627	40.187	1.00	16.60	0.001	A
ATOM	16562	N	LEU	B	435	-9.733	24.223	47.187	1.00	5.77	-0.346	N
ATOM	16563	HN	LEU	B	435	-10.434	23.036	47.257	1.00	0.00	0.163	HD
ATOM	16564	CA	LEU	B	435	-11.825	21.930	46.739	1.00	6.30	0.056	A
ATOM	16566	C	LEU	B	435	-7.459	21.721	47.011	1.00	7.73	0.241	C
ATOM	16567	O	LEU	B	435	-6.316	21.565	47.433	1.00	6.78	-0.271	OA
ATOM	16568	CB	LEU	B	435	-9.225	20.316	46.498	1.00	8.30	0.056	A
ATOM	16571	CG	LEU	B	435	-8.733	20.316	46.498	1.00	10.50	-0.020	C
ATOM	16573	CD1	LEU	B	435	-7.443	21.029	50.485	1.00	8.78	0.009	C
ATOM	16577	CD2	LEU	B	435	-7.829	21.930	49.239	1.00	9.36	0.056	A
ATOM	16581	N	MET	B	436	-7.870	21.299	45.798	1.00	6.74	-0.346	N
ATOM	16582	HN	MET	B	436	-8.829	21.432	45.478	1.00	0.00	0.163	HD
ATOM	16583	CA	MET	B	436	-6.943	20.627	47.187	1.00	9.13	0.241	C
ATOM	16585	C	MET	B	436	-5.620	21.517	44.786	1.00	8.30	0.241	C
ATOM	16586	O	MET	B	436	-4.507	21.011	44.767	1.00	9.71	-0.271	OA
ATOM	16587	CB	MET	B	436	-4.403	20.627	45.309	1.00	7.50	0.056	A
ATOM	16590	CG	MET	B	436	-6.473	19.559	42.641	1.00	10.09	0.076	C
ATOM	16593	SD	MET	B	436	-5.898	17.986	43.317	1.00	12.58	-0.173	SA
ATOM	16594	CE	MET	B	436	-4.749	17.825	42.641	1.00	13.59	0.056	A
ATOM	16598	N	PHE	B	437	-5.788	22.850	44.642	1.00	6.10	-0.346	N
ATOM	16599	HN	PHE	B	437	-6.716	23.244	44.779	1.00	0.00	0.163	HD
ATOM	16600	CA	PHE	B	437	-5.693	23.712	43.300	1.00	4.79	0.056	A
ATOM	16602	C	PHE	B	437	-3.899	24.135	45.526	1.00	9.89	0.241	C
ATOM	16603	O	PHE	B	437	-2.865	24.781	45.397	1.00	10.94	-0.271	OA
ATOM	16604	CB	PHE	B	437	-4.203	24.868	43.300	1.00	13.59	0.056	A
ATOM	16607	CG	PHE	B	437	-5.997	24.256	42.224	1.00	12.40	-0.056	A
ATOM	16608	CD1	PHE	B	437	-5.427	23.242	41.456	1.00	14.43	0.007	A
ATOM	16610	CD2	PHE	B	437	-7.272	24.680	41.930	1.00	13.89	0.007	A
ATOM	16612	CE1	PHE	B	437	-6.128	22.652	40.399	1.00	18.79	0.001	A
ATOM	16614	CE2	PHE	B	437	-7.986	24.120	40.886	1.00	20.75	0.001	A
ATOM	16616	CE3	PHE	B	437	-7.409	23.113	40.187	1.00	18.84	0.001	A
ATOM	16618	N	VAL	B	438	-4.093	23.474	46.673	1.00	7.72	-0.346	N
ATOM	16619	HN	VAL	B	438	-4.935	22.915	46.810	1.00	0.00	0.163	HD
ATOM	16620	CA	VAL	B	438	-4.063	23.964	47.750	1.00	7.70	0.056	A
ATOM	16622	C	VAL	B	438	-1.799	22.869	47.224	1.00	10.04	0.241	C
ATOM	16623	O	VAL	B	438	-0.673	23.244	47.604	1.00	8.09	-0.271	OA
ATOM	16624	CB	VAL	B	438	-1.423	22.869	46.498	1.00	10.73	0.056	A
ATOM	16626	CG1	VAL	B	438	-3.621	21.351	48.865	1.00	9.03	0.012	C
ATOM	16630	CG2	VAL	B	438	-2.562	23.189	50.189	1.00	11.87	0.012	C
ATOM	16634	N	GLU	B	439	-1.911	21.993	46.312	1.00	8.46	-0.346	N
ATOM	16635	HN	GLU	B	439	-2.840	21.637	45.946	1.00	0.00	0.163	HD
ATOM	16636	CA	GLU	B	439	-0.737	21.309	45.673	1.00	7.35	0.177	C
ATOM	16638	C	GLU	B	439	-0.145	22.349	46.906	1.00	9.50	0.241	C
ATOM	16639	O	GLU	B	439	1.317	22.169	44.960	1.00	10.66	-0.271	OA
ATOM	16640	CB	GLU	B	439	-1.112	20.136	44.731	1.00	3.49	0.045	C
ATOM	16643	CG	GLU	B	439	-1.749	19.621	45.197	1.00	6.16	0.056	A
ATOM	16646	CD	GLU	B	439	-1.642	17.648	44.793	1.00	13.89	0.172	C
ATOM	16647	CE1	GLU	B	439	-1.411	17.573	43.763	1.00	11.88	-0.648	OA
ATOM	16648	CE2	GLU	B	439	-1.758	16.659	42.768	1.00	9.48	-0.648	OA
ATOM	16649	N	TYR	B	440	-0.466	23.380	44.271	1.00	4.95	-0.346	N
ATOM	16650	HN	TYR	B	440	-1.423	23.457	44.271	1.00	0.00	0.163	HD
ATOM	16651	CA	TYR	B	440	0.374	24.240	43.648	1.00	6.84	0.241	C
ATOM	16653	C	TYR	B	440	1.117	25.194	44.730	1.00	8.23	0.241	C
ATOM	16654	O	TYR	B	440	-0.503	25.862	44.730	1.00	12.30	-0.271	OA
ATOM	16655	CB	TYR	B	440	-0.505	25.322	42.846	1.00	4.47	0.073	C
ATOM	16658	CG	TYR	B	440	-0.831	24.834	41.444	1.00	8.13	-0.056	A
ATOM	16659	CD1	TYR	B	440	-1.272	24.580	40.187	1.00	5.57	-0.271	OA
ATOM	16661	CD2	TYR	B	440	-0.606	25.667	40.399	1.00	6.25	0.010	A
ATOM	16663	CE1	TYR	B	440	-1.743	23.148	39.964	1.00	7.68	0.037	A
ATOM	16665	CE2	TYR	B	440	-1.450	23.260	38.885	1.00	8.33	0.065	A
ATOM	16667	CE3	TYR	B	440	-1.788	23.547	37.615	1.00	9.05	-0.361	OA
ATOM	16669	HN	TYR	B	440	-1.532	22.652	40.187	1.00	12.30	0.172	C
ATOM	16670	N	ALA	B	441	-0.489	25.375	45.882	1.00	6.30	-0.346	N
ATOM	16671	HN	ALA	B	441	-0.416	24.937	46.054	1.00	0.00	0.163	HD
ATOM	16672	CA	ALA	B	441	-1.211	25.629	47.111	1.00	7.13	0.172	C
ATOM	16674	C	ALA	B	441	1.764	25.411	48.038	1.00	8.99	0.240	C
ATOM	16675	O	ALA	B	441	1.999	26.028	49.080	1.00	7.12	-0.271	OA
ATOM	16676	CB	ALA	B	441	-0.017	23.637	47.534	1.00	6.32	-0.346	N
ATOM	16680	N	ARG	B	442	1.969	24.113	47.866	1.00	6.32	-0.346	N
ATOM	16681	HN	ARG	B	442	1.895	23.704	46.934	1.00	0.00	0.163	HD
ATOM	16682	CA	ARG	B	442	3.303	23.600	46.934	1.00	0.00	0.176	CD
ATOM	16684	C	ARG	B	442	3.441	23.693	49.891	1.00	6.21	0.241	C
ATOM	16685	O	ARG	B	442	3.352	23.674	45.132	1.00	8.35	-0.271	OA
ATOM	16686	CB	ARG	B	442	-1.562	21.838	48.193	1.00	7.23	0.176	CD
ATOM	16689	CG	ARG	B	442	2.542	20.741	49.490	1.00	14.23	0.023	C
ATOM	16692	CD	ARG	B	442	1.180	20.818	50.177	1.00	15.77	0.138	C
ATOM	16695	NE	ARG	B	442	-2.863	19.880	51.462	1.00	21.27	-0.271	OA
ATOM	16696	HE	ARG	B	442	0.118	19.201	51.155	1.00	0.00	0.177	HD
ATOM	16697	CG	ARG	B	442	1.559	19.944	52.422	1.00	31.73	0.665	C
ATOM	16698	HN1	ARG	B	442	-2.533	20.885	52.539	1.00	10.43	0.176	CD
ATOM	16699	HN1	ARG	B	442	3.064	20.847	53.411	1.00	0.00	0.174	HD

AT0M	17091	N	GLV	B	467	-18.553	17.212	42.321	1.00	12.86	-0.351	N
AT0M	17092	HN	GLV	B	467	-18.999	16.955	42.943	1.00	13.87	-0.225	C
AT0M	17093	CA	GLV	B	467	-19.208	16.697	41.142	1.00	13.87	0.225	C
AT0M	17096	C	GLV	B	467	-18.524	16.783	39.804	1.00	14.44	0.236	C
AT0M	17097	C	GLV	B	468	-18.408	16.982	39.804	1.00	14.44	-0.236	C
AT0M	17098	N	GLV	B	468	-19.228	17.383	38.825	1.00	9.86	-0.346	N
AT0M	17099	HN	GLV	B	468	-20.026	17.964	39.080	1.00	0.00	0.163	HD
AT0M	17100	CA	GLV	B	468	-18.488	17.981	37.000	1.00	12.01	0.045	C
AT0M	17102	C	GLV	B	468	-17.508	17.681	36.981	1.00	12.06	0.241	C
AT0M	17103	O	GLV	B	468	-17.014	17.377	35.946	1.00	12.82	-0.271	OA
AT0M	17104	CB	GLV	B	468	-19.973	18.011	36.605	1.00	12.01	0.045	C
AT0M	17107	CG	GLV	B	468	-21.340	17.322	36.729	1.00	14.55	0.116	C
AT0M	17110	CD	GLV	B	468	-22.264	17.917	37.760	1.00	23.66	0.172	C
AT0M	17111	OE1	GLV	B	468	-21.803	18.452	38.788	1.00	17.68	-0.648	OA
AT0M	17112	OE2	GLV	B	468	-23.506	17.835	37.559	1.00	18.84	-0.648	OA
AT0M	17113	N	ASP	B	469	-16.850	18.640	37.650	1.00	11.44	-0.346	N
AT0M	17114	HN	ASP	B	469	-17.461	18.893	38.769	1.00	0.00	0.648	HD
AT0M	17115	CA	ASP	B	469	-15.501	19.027	37.166	1.00	10.55	0.186	C
AT0M	17117	C	ASP	B	469	-14.563	17.817	37.078	1.00	14.20	0.241	C
AT0M	17118	O	ASP	B	469	-14.932	20.142	38.066	1.00	9.57	0.147	C
AT0M	17119	CB	ASP	B	469	-15.458	21.493	37.628	1.00	23.68	0.175	C
AT0M	17122	CG	ASP	B	469	-16.075	21.607	36.941	1.00	20.91	-0.676	OA
AT0M	17124	OD2	ASP	B	469	-15.303	22.468	38.359	1.00	19.78	-0.648	OA
AT0M	17125	N	GLV	B	470	-14.754	16.759	37.880	1.00	9.27	-0.351	N
AT0M	17126	HN	GLV	B	470	-15.441	16.893	38.769	1.00	0.00	0.163	HD
AT0M	17127	CA	GLV	B	470	-13.997	15.543	37.689	1.00	8.52	0.225	C
AT0M	17130	C	GLV	B	470	-12.666	15.414	38.388	1.00	9.61	0.238	C
AT0M	17131	O	GLV	B	470	-12.342	16.142	39.334	1.00	11.79	-0.272	OA
AT0M	17132	N	PRO	B	471	-11.919	14.358	38.043	1.00	9.72	-0.337	N
AT0M	17133	CA	PRO	B	471	-10.799	15.911	36.900	1.00	8.01	0.241	C
AT0M	17135	C	PRO	B	471	-9.834	14.818	38.915	1.00	10.65	0.241	C
AT0M	17136	O	PRO	B	471	-8.839	14.688	38.876	1.00	8.29	-0.271	OA
AT0M	17137	CB	PRO	B	471	-10.651	15.284	38.989	1.00	12.69	0.127	C
AT0M	17140	CG	PRO	B	471	-10.961	15.984	36.818	1.00	13.91	0.022	C
AT0M	17143	CD	PRO	B	471	-12.256	13.363	36.989	1.00	12.69	0.127	C
AT0M	17146	N	HRB	B	472	-9.458	15.779	38.989	1.00	14.44	-0.346	N
AT0M	17147	HN	HRB	B	472	-10.047	15.867	37.229	1.00	0.00	0.163	HD
AT0M	17148	CA	HRB	B	472	-8.300	16.706	38.157	1.00	6.31	0.205	C
AT0M	17150	C	HRB	B	472	-7.651	16.436	39.000	1.00	8.07	0.241	C
AT0M	17151	O	HRB	B	472	-7.671	18.336	39.746	1.00	11.62	-0.271	OA
AT0M	17152	CB	HRB	B	472	-8.023	17.363	36.903	1.00	9.39	0.146	C
AT0M	17154	CG2	HRB	B	472	-9.783	16.580	39.601	1.00	9.85	0.241	C
AT0M	17158	OG1	HRB	B	472	-9.205	18.286	36.552	1.00	13.52	-0.393	OA
AT0M	17159	HG1	HRB	B	472	-9.365	18.885	37.271	1.00	0.00	0.210	HD
AT0M	17160	N	HRB	B	472	-10.724	16.893	39.601	1.00	12.02	-0.346	N
AT0M	17161	HN	HRB	B	473	-10.608	17.463	39.082	1.00	0.00	0.163	HD
AT0M	17162	CA	HRB	B	473	-10.234	18.870	40.666	1.00	8.48	0.182	C
AT0M	17164	C	HRB	B	473	-9.785	18.870	40.666	1.00	8.48	0.182	C
AT0M	17165	O	HRB	B	473	-10.658	18.870	40.666	1.00	9.71	-0.271	OA
AT0M	17166	CB	HRB	B	473	-11.450	19.679	40.155	1.00	7.24	0.093	C
AT0M	17169	CG	HRB	B	473	-9.109	19.776	39.191	1.00	14.31	0.241	C
AT0M	17170	CD2	HRB	B	473	-11.471	22.082	39.063	1.00	14.64	0.114	C
AT0M	17172	HD1	HRB	B	473	-10.362	20.540	38.061	1.00	15.24	-0.354	N
AT0M	17173	HD1	HRB	B	473	-9.969	19.637	37.798	1.00	14.77	0.166	HD
AT0M	17174	OE1	HRB	B	473	-10.233	21.650	37.369	1.00	15.04	0.180	A
AT0M	17176	NE2	HRB	B	473	-10.881	22.592	38.108	1.00	21.02	-0.380	N
AT0M	17177	HE2	HRB	B	473	-10.942	23.865	37.018	1.00	0.00	0.241	C
AT0M	17178	N	GLM	B	474	-11.182	16.394	41.945	1.00	9.54	-0.346	N
AT0M	17179	HN	GLM	B	474	-11.173	16.394	41.977	1.00	0.00	0.163	HD
AT0M	17180	CA	GLM	B	474	-10.614	15.800	44.112	1.00	10.22	0.243	C
AT0M	17182	C	GLM	B	474	-9.882	14.848	43.808	1.00	11.19	-0.271	OA
AT0M	17184	CB	GLM	B	474	-12.879	16.066	42.821	1.00	9.46	0.022	C
AT0M	17187	CG	GLM	B	474	-13.821	15.521	42.009	1.00	9.55	0.105	C
AT0M	17190	CD	GLM	B	474	-14.402	14.377	41.192	1.00	16.85	0.215	C
AT0M	17191	HE2	GLM	B	474	-9.473	14.828	40.760	1.00	11.43	0.241	C
AT0M	17192	HE2	GLM	B	474	-16.242	15.353	41.000	1.00	0.00	0.159	HD
AT0M	17193	2HE2	GLM	B	474	-16.067	13.759	40.288	1.00	0.00	0.159	HD
AT0M	17194	OE1	GLM	B	474	-10.431	16.426	45.266	1.00	11.81	-0.337	N
AT0M	17195	N	PRO	B	475	-9.450	16.021	46.239	1.00	9.47	-0.179	C
AT0M	17196	CA	PRO	B	475	-9.695	16.885	46.100	1.00	13.51	0.241	C
AT0M	17199	O	PRO	B	475	-10.841	14.174	46.866	1.00	10.23	-0.271	OA
AT0M	17200	CB	PRO	B	475	-9.597	17.008	47.412	1.00	8.09	0.037	C
AT0M	17201	CG	PRO	B	475	-10.942	16.483	46.923	1.00	9.37	0.243	C
AT0M	17206	CD	PRO	B	475	-11.246	17.544	45.798	1.00	9.38	0.127	C
AT0M	17208	N	VAL	B	476	-8.601	13.864	46.953	1.00	8.10	-0.346	N
AT0M	17210	HN	VAL	B	476	-7.688	14.565	46.900	1.00	0.00	0.163	HD
AT0M	17211	CA	VAL	B	476	-8.696	12.519	47.516	1.00	7.61	0.180	C
AT0M	17213	C	VAL	B	476	-7.844	14.448	48.881	1.00	7.66	0.241	C
AT0M	17214	O	VAL	B	476	-7.981	12.113	48.835	1.00	10.06	-0.271	OA
AT0M	17215	CB	VAL	B	476	-8.387	11.389	46.510	1.00	12.38	0.009	C
AT0M	17217	CG1	VAL	B	476	-9.949	11.999	47.205	1.00	10.31	0.241	C
AT0M	17221	CG2	VAL	B	476	-9.499	11.378	45.436	1.00	9.59	0.012	C
AT0M	17225	N	GLU	B	477	-6.427	12.765	48.324	1.00	4.82	-0.346	N
AT0M	17226	HN	GLU	B	477	-6.427	12.765	48.324	1.00	4.82	-0.346	N
AT0M	17227	CA	GLU	B	477	-5.324	12.556	49.280	1.00	8.41	0.177	C
AT0M	17229	C	GLU	B	477	-4.770	13.848	49.875	1.00	10.26	0.241	C
AT0M	17230	O	GLU	B	477	-4.031	12.515	46.502	1.00	12.42	-0.271	OA
AT0M	17231	CB	GLU	B	477	-4.216	11.770	48.554	1.00	8.85	0.045	C
AT0M	17234	CG	GLU	B	477	-3.400	12.466	47.454	1.00	8.27	0.116	C
AT0M	17237	CD	GLU	B	477	-4.031	12.515	46.502	1.00	12.42	-0.271	OA
AT0M	17238	OE1	GLU	B	477	-5.281	12.593	45.916	1.00	10.86	-0.648	OA
AT0M	17239	OE2	GLU	B	477	-3.296	12.677	45.035	1.00	11.44	-0.648	OA
AT0M	17240	N	GLM	B	478	-4.366	14.987	47.449	1.00	7.49	-0.346	N
AT0M	17241	HN	GLM	B	478	-6.232	14.987	49.024	1.00	0.00	0.163	HD
AT0M	17242	CA	GLM	B	478	-4.765	16.273	50.000	1.00	8.14	0.177	C
AT0M	17244	C	GLM	B	478	-4.979	16.510	50.498	1.00	9.09	0.241	C
AT0M	17245	O	GLM	B	478	-4.099	17.052	52.154	1.00	8.76	-0.271	OA
AT0M	17246	CB	GLM	B	478	-5.414	17.418	49.212	1.00	10.55	0.044	C
AT0M	17249	CG	GLM	B	478	-6.030	17.437	47.786	1.00	7.59	0.180	C
AT0M	17252	CD	GLM	B	478	-5.341	16.315	46.919	1.00	12.04	0.215	C
AT0M	17253	NE2	GLM	B	478	-4.488	15.865	45.991	1.00	8.70	-0.370	N
AT0M	17254	HE2	GLM	B	478	-5.967	15.865	45.991	1.00	8.70	-0.370	N
AT0M	17255	2HE2	GLM	B	478	-4.848	15.106	45.407	1.00	0.00	0.159	HD
AT0M	17256	OE1	GLM	B	478	-6.457	15.823	47.125	1.00	7.84	-0.274	OA
AT0M	17257	N	VAL	B	479	-6.148	16.477	47.000	1.00	8.01	-0.346	N
AT0M	17258	HN	VAL	B	479	-6.864	15.715	41.419	1.00	0.00	0.163	HD
AT0M	17259	CA	VAL	B	479	-6.373	16.410	53.442	1.00	6.37	0.180	C
AT0M	172											

ATOM 17649	CG	LYS	B	504	-13.358	31.494	68.125	1.00	9.94	0.004	C
ATOM 17650	CD	LYS	B	504	-8.451	27.193	68.125	1.00	9.94	0.004	C
ATOM 17651	CE	LYS	B	504	-14.187	11.969	70.399	1.00	19.48	0.229	C
ATOM 17652	NE	LYS	B	504	-13.171	31.433	71.316	1.00	20.62	-0.079	N
ATOM 17653	H21	LYS	B	504	-10.252	32.938	70.240	1.00	10.00	0.000	HD
ATOM 17660	H22	LYS	B	504	-12.292	31.267	70.827	1.00	0.00	0.274	HD
ATOM 17661	H23	LYS	B	504	-13.395	30.476	71.587	1.00	0.00	0.274	HD
ATOM 17662	N	TYR	B	505	-11.171	29.946	66.461	1.00	8.41	0.163	N
ATOM 17663	HN	TYR	B	505	-11.966	28.589	65.767	1.00	0.00	0.163	HD
ATOM 17664	CA	TYR	B	505	-9.899	28.513	66.347	1.00	10.12	0.180	C
ATOM 17665	CB	TYR	B	505	-8.791	29.074	65.458	1.00	10.92	0.241	C
ATOM 17666	O	TYR	B	505	-7.728	29.298	66.017	1.00	11.04	-0.271	OA
ATOM 17668	CG	TYR	B	505	-10.005	26.993	66.112	1.00	7.95	0.073	C
ATOM 17671	CD	TYR	B	505	-8.860	26.331	66.650	1.00	10.00	-0.059	A
ATOM 17672	CE	TYR	B	505	-8.926	25.692	67.964	1.00	16.67	0.010	A
ATOM 17674	CD2	TYR	B	505	-7.733	25.845	65.910	1.00	12.19	0.010	A
ATOM 17676	CE1	TYR	B	505	-5.432	23.809	68.309	1.00	20.64	-0.158	OA
ATOM 17678	CE2	TYR	B	505	-6.713	25.071	66.462	1.00	13.60	0.037	A
ATOM 17680	CZ	TYR	B	505	-6.809	24.612	67.743	1.00	16.92	0.065	A
ATOM 17681	OH	TYR	B	505	-5.492	23.809	68.309	1.00	20.64	-0.158	OA
ATOM 17682	HH	TYR	B	505	-5.879	23.489	69.201	1.00	0.00	0.217	HD
ATOM 17683	N	GLY	B	506	-8.995	29.197	64.153	1.00	9.73	-0.351	N
ATOM 17684	HN	GLY	B	506	-9.067	28.955	63.766	1.00	0.00	0.165	HD
ATOM 17685	CA	GLY	B	506	-7.932	29.676	63.247	1.00	8.15	0.225	C
ATOM 17688	C	GLY	B	506	-7.549	31.136	63.544	1.00	9.78	0.236	C
ATOM 17689	O	GLY	B	506	-6.187	31.518	63.979	1.00	12.83	-0.272	OA
ATOM 17690	N	VAL	B	507	-8.501	32.033	63.829	1.00	9.62	-0.346	N
ATOM 17691	HN	VAL	B	507	-9.483	31.759	63.857	1.00	0.00	0.163	HD
ATOM 17692	CA	VAL	B	507	-8.101	33.649	60.101	1.00	12.28	0.180	C
ATOM 17694	C	VAL	B	507	-7.415	33.937	60.500	1.00	9.96	0.241	C
ATOM 17695	O	VAL	B	507	-5.217	34.074	60.500	1.00	12.82	-0.271	OA
ATOM 17696	CB	VAL	B	507	-9.243	34.459	63.976	1.00	13.42	-0.203	A
ATOM 17698	CG1	VAL	B	507	-9.855	34.420	62.574	1.00	13.38	0.012	C
ATOM 17702	CG2	VAL	B	507	-10.350	34.101	62.147	1.00	14.47	-0.137	A
ATOM 17706	N	GLU	B	508	-7.686	32.671	66.437	1.00	9.20	-0.346	N
ATOM 17707	HN	GLU	B	508	-8.478	32.969	66.269	1.00	0.00	0.163	HD
ATOM 17708	CA	GLU	B	508	-6.039	32.713	67.131	1.00	11.95	0.137	C
ATOM 17710	C	GLU	B	508	-5.728	33.966	67.826	1.00	17.47	0.241	C
ATOM 17711	O	GLU	B	508	-4.978	32.977	68.811	1.00	15.21	-0.271	OA
ATOM 17712	CB	GLU	B	508	-8.101	33.649	60.101	1.00	12.28	0.180	C
ATOM 17715	CG	GLU	B	508	-9.154	33.085	69.004	1.00	14.92	0.116	C
ATOM 17718	CD	GLU	B	508	-10.153	32.660	67.063	1.00	25.24	0.172	C
ATOM 17719	OE1	GLU	B	508	-10.318	31.465	66.350	1.00	18.11	-0.271	OA
ATOM 17720	OE2	GLU	B	508	-10.818	33.516	70.681	1.00	24.56	-0.648	OA
ATOM 17721	N	ARG	B	509	-5.407	31.229	66.749	1.00	9.18	-0.346	N
ATOM 17722	HN	ARG	B	509	-6.000	30.920	67.973	1.00	10.23	0.000	HD
ATOM 17723	CA	ARG	B	509	-4.185	30.634	66.799	1.00	12.08	0.216	C
ATOM 17725	C	ARG	B	509	-3.018	31.367	66.509	1.00	18.61	0.171	C
ATOM 17726	O	ARG	B	509	-4.831	32.693	67.043	1.00	21.46	-0.271	OA
ATOM 17727	CB	ARG	B	509	-4.321	29.252	65.831	1.00	15.86	0.036	C
ATOM 17730	CG	ARG	B	509	-3.175	28.298	65.875	1.00	19.38	0.023	C
ATOM 17733	CD	ARG	B	509	-1.302	27.064	64.945	1.00	18.71	-0.271	OA
ATOM 17736	NE	ARG	B	509	-2.170	26.300	65.480	1.00	24.51	-0.227	N
ATOM 17737	HE	ARG	B	509	-2.124	26.293	66.499	1.00	0.00	0.177	HD
ATOM 17738	CZ	ARG	B	509	-1.220	25.934	66.499	1.00	13.52	-0.271	OA
ATOM 17739	NH1	ARG	B	509	-1.214	25.931	63.005	1.00	13.04	-0.235	N
ATOM 17740	NH2	ARG	B	509	-1.934	25.921	63.605	1.00	0.00	0.174	HD
ATOM 17741	2NH1	ARG	B	509	-0.460	24.902	63.605	1.00	0.00	0.174	HD
ATOM 17742	NH2	ARG	B	509	-0.287	25.111	65.690	1.00	25.49	-0.235	N
ATOM 17743	1NH2	ARG	B	509	-0.292	25.201	66.706	1.00	0.00	0.174	HD
ATOM 17744	2NH2	ARG	B	509	-0.463	24.985	63.605	1.00	0.00	0.174	HD
ATOM 17745	N	GLN	B	510	-2.166	31.461	68.122	1.00	14.74	-0.346	N
ATOM 17746	HN	GLN	B	510	-2.276	31.209	68.399	1.00	0.00	0.163	HD
ATOM 17747	CA	GLN	B	510	-0.620	32.570	67.230	1.00	42.40	-0.271	OA
ATOM 17749	C	GLN	B	510	0.272	32.966	66.693	1.00	19.76	0.241	C
ATOM 17750	O	GLN	B	510	1.055	32.699	66.198	1.00	23.30	-0.271	OA
ATOM 17751	CB	GLN	B	510	-2.428	33.820	63.926	1.00	14.62	-0.271	OA
ATOM 17754	CG	GLN	B	510	-1.774	33.900	69.298	1.00	46.92	0.105	C
ATOM 17757	CD	GLN	B	510	-1.676	35.408	69.168	1.00	60.33	0.215	C
ATOM 17758	HE2	GLN	B	510	-0.478	36.937	70.281	1.00	16.26	-0.346	N
ATOM 17759	HE2	GLN	B	510	-2.004	37.117	70.147	1.00	0.00	0.159	HD
ATOM 17760	2HE2	GLN	B	510	-2.427	35.675	71.089	1.00	0.00	0.159	HD
ATOM 17761	OE1	GLN	B	510	-2.625	35.928	69.168	1.00	69.55	-0.271	OA
ATOM 17762	N	ASP	B	511	0.445	30.694	66.889	1.00	15.59	-0.346	N
ATOM 17763	HN	ASP	B	511	-0.294	30.113	67.285	1.00	0.00	0.163	HD
ATOM 17764	CA	ASP	B	511	-1.733	30.490	66.421	1.00	18.10	-0.271	OA
ATOM 17766	C	ASP	B	511	1.712	29.141	65.363	1.00	13.11	0.241	C
ATOM 17767	O	ASP	B	511	2.629	28.308	65.264	1.00	11.80	-0.271	OA
ATOM 17768	CB	ASP	B	511	2.467	29.343	63.005	1.00	16.26	-0.271	OA
ATOM 17771	CG	ASP	B	511	1.257	28.332	68.210	1.00	36.87	0.175	C
ATOM 17772	CD1	ASP	B	511	-0.780	28.862	67.768	1.00	39.21	-0.271	OA
ATOM 17773	OD2	ASP	B	511	1.576	27.575	69.285	1.00	42.63	-0.648	OA
ATOM 17774	N	GLY	B	512	0.736	29.309	64.466	1.00	11.53	-0.351	N
ATOM 17775	HN	GLY	B	512	-0.480	29.294	62.966	1.00	11.07	0.163	HD
ATOM 17776	CA	GLY	B	512	0.793	28.459	63.279	1.00	8.48	0.225	C
ATOM 17779	C	GLY	B	512	-0.502	28.660	62.486	1.00	10.73	0.238	C
ATOM 17780	O	GLY	B	512	-0.480	29.294	62.966	1.00	11.07	0.163	HD
ATOM 17781	N	PRO	B	513	-0.487	28.128	62.170	1.00	10.02	-0.337	N
ATOM 17782	CA	PRO	B	513	-1.608	28.294	60.370	1.00	8.63	0.179	C
ATOM 17784	C	PRO	B	513	-0.780	27.867	61.620	1.00	12.34	-0.271	OA
ATOM 17785	O	PRO	B	513	-2.590	26.291	61.264	1.00	10.43	-0.271	OA
ATOM 17786	CB	PRO	B	513	-0.934	28.159	68.992	1.00	6.70	0.037	C
ATOM 17789	CG	PRO	B	513	-0.866	28.483	69.320	1.00	10.62	0.127	C
ATOM 17792	CD	PRO	B	513	0.661	27.461	68.621	1.00	6.82	0.227	C
ATOM 17795	N	THR	B	514	-3.911	27.636	60.007	1.00	9.80	-0.346	N
ATOM 17796	HN	THR	B	514	-4.616	28.483	61.620	1.00	8.66	-0.271	OA
ATOM 17797	CA	THR	B	514	-5.094	26.755	60.152	1.00	7.80	0.205	C
ATOM 17799	C	THR	B	514	-5.791	27.602	58.801	1.00	9.64	0.243	C
ATOM 17800	O	THR	B	514	-6.113	27.647	58.801	1.00	9.64	-0.271	OA
ATOM 17801	CB	THR	B	514	-6.064	27.409	61.141	1.00	10.70	0.146	C
ATOM 17803	CG2	THR	B	514	-7.267	26.511	61.386	1.00	13.24	0.042	C
ATOM 17807	OH1	THR	B	514	-5.400	27.658	62.966	1.00	9.92	-0.271	OA
ATOM 17808	OH1	THR	B	514	-6.004	28.064	61.016	1.00	0.00	0.210	HD
ATOM 17809	N	ALA	B	515	-5.990	25.281	58.315	1.00	5.77	-0.346	N
ATOM 17810	HN	ALA	B	515	-6.639	25.314	57.009	1.00	6.95	0.172	C
ATOM 17811	CA	ALA	B	515	-6.639	25.214	57.009	1.00	6.95	0.172	C
ATOM 17813	C	ALA	B	515	-8.087	24.754	57.240	1.00	8.77	0.240	C
ATOM 17814	O	ALA	B	515	-6.303	23.893	58.989	1.00	11.81	-0.271	OA
ATOM 17815	CB	ALA	B	515	-5.897	24.077	56.259	1.00	8.38	0.042	C
ATOM 17819	N	LEU	B	516	-9.054	25.387	58.606	1.00	9.25	-0.346	N
ATOM 17820	HN	LEU	B	516	-8.820	24.688	54.339	1.00	10.00	0.163	HD
ATOM 17821	CA	LEU	B	516	-10.440	24.968	56.701	1.00	10.15	0.177	C
ATOM 17823	C	LEU	B	516	-10.819	24.203	55.435	1.00	11.18	0.241	C
ATOM 17824	O	LEU	B	516	-8.820	24.688	54.339	1.00	10.00	-0.271	OA
ATOM 17825	CB	LEU	B	516	-11						

ATOM 18206	C	GLY	B	539	-11.983	17.181	67.430	1.00	9.02	0.235	C
ATOM 18207	O	VAL	B	540	-12.551	16.268	67.139	1.00	-0.272	OA	
ATOM 18208	N	LEU	B	540	-12.353	16.350	68.423	1.00	9.61	-0.351	N
ATOM 18209	HN	GLY	B	540	-12.064	16.571	69.376	1.00	0.00	0.163	HD
ATOM 18210	CA	LEU	B	540	-11.410	16.066	69.198	1.00	9.25	0.241	C
ATOM 18211	C	GLY	B	540	-14.103	15.044	69.495	1.00	10.96	0.236	C
ATOM 18212	O	VAL	B	540	-13.526	15.368	70.591	1.00	9.28	-0.272	OA
ATOM 18213	N	LEU	B	541	-15.253	14.026	69.130	1.00	7.67	-0.351	N
ATOM 18214	HN	TYR	B	541	-15.564	14.313	68.397	1.00	0.00	0.163	HD
ATOM 18215	CA	TYR	B	541	-16.177	14.618	70.454	1.00	9.41	0.180	C
ATOM 18216	C	VAL	B	541	-17.314	13.437	70.137	1.00	8.13	0.241	C
ATOM 18217	O	VAL	B	541	-17.580	13.336	68.970	1.00	10.43	-0.271	OA
ATOM 18218	CB	TYR	B	541	-16.708	16.036	70.721	1.00	8.81	0.073	C
ATOM 18219	CG	VAL	B	542	-17.555	16.409	69.598	1.00	12.58	-0.006	N
ATOM 18220	CD1	TYR	B	541	-16.937	17.234	68.521	1.00	7.91	0.010	A
ATOM 18221	CD2	TYR	B	541	-18.944	16.536	69.614	1.00	12.39	0.010	A
ATOM 18222	CE1	TYR	B	541	-19.745	16.406	72.136	1.00	6.00	0.163	HD
ATOM 18223	CE2	TYR	B	541	-19.735	17.082	68.605	1.00	11.29	0.037	A
ATOM 18224	CZ	TYR	B	541	-19.077	17.670	67.515	1.00	8.46	0.065	A
ATOM 18234	OH	TYR	B	542	-19.021	18.200	66.183	1.00	22.68	-0.382	OA
ATOM 18235	HH	TYR	B	541	-20.758	18.124	66.534	1.00	0.00	0.217	HD
ATOM 18236	N	VAL	B	542	-17.946	13.150	71.196	1.00	9.37	-0.348	N
ATOM 18237	HN	VAL	B	542	-17.445	13.406	72.136	1.00	0.00	0.163	HD
ATOM 18238	CA	VAL	B	542	-19.095	12.230	71.004	1.00	7.36	0.180	C
ATOM 18240	C	VAL	B	542	-20.321	13.066	70.605	1.00	13.16	0.241	C
ATOM 18241	O	VAL	B	542	-20.789	13.939	69.360	1.00	11.88	-0.276	OA
ATOM 18242	CB	VAL	B	542	-19.412	11.517	72.335	1.00	12.76	0.009	C
ATOM 18244	CG1	VAL	B	542	-20.595	10.570	72.179	1.00	12.98	0.012	C
ATOM 18248	CG2	VAL	B	542	-18.444	10.722	72.903	1.00	12.60	0.012	C
ATOM 18252	N	LEU	B	543	-20.834	12.754	69.428	1.00	8.04	-0.346	N
ATOM 18253	HN	LEU	B	543	-20.412	12.549	68.904	1.00	0.00	0.163	HD
ATOM 18254	CA	LEU	B	543	-21.866	13.446	68.842	1.00	8.12	0.241	C
ATOM 18256	C	LEU	B	543	-23.244	12.633	69.105	1.00	13.86	0.177	C
ATOM 18257	O	LEU	B	543	-22.387	12.522	68.809	1.00	12.19	-0.276	OA
ATOM 18258	CB	LEU	B	543	-21.751	13.991	67.312	1.00	11.60	0.038	C
ATOM 18261	CG	LEU	B	543	-22.916	14.280	66.570	1.00	16.32	-0.020	C
ATOM 18263	CD1	LEU	B	543	-22.367	14.187	66.570	1.00	12.13	0.009	C
ATOM 18267	CD2	LEU	B	543	-22.747	14.187	66.509	1.00	17.28	0.009	C
ATOM 18271	N	LYS	B	544	-23.221	11.332	68.932	1.00	15.29	-0.346	N
ATOM 18272	HN	LYS	B	544	-22.387	11.224	68.932	1.00	0.00	0.163	HD
ATOM 18273	CA	LYS	B	544	-24.374	10.460	69.231	1.00	13.34	0.176	C
ATOM 18275	C	LYS	B	544	-23.834	9.325	70.106	1.00	14.63	0.241	C
ATOM 18276	O	LYS	B	544	-22.753	9.781	69.137	1.00	12.57	-0.276	OA
ATOM 18277	CB	LYS	B	544	-25.039	9.842	68.006	1.00	15.54	0.035	C
ATOM 18280	CG	LYS	B	544	-25.613	10.868	67.021	1.00	17.38	0.004	C
ATOM 18283	CD	LYS	B	544	-25.981	12.488	66.676	1.00	17.00	0.009	C
ATOM 18286	CE	LYS	B	544	-27.620	12.135	66.876	1.00	35.45	0.229	C
ATOM 18289	NZ	LYS	B	544	-28.460	13.278	66.876	1.00	37.10	-0.079	N
ATOM 18290	HZ1	LYS	B	544	-29.852	13.709	66.876	1.00	11.88	0.009	C
ATOM 18291	HZ2	LYS	B	544	-27.912	13.959	67.401	1.00	0.00	0.274	HD
ATOM 18292	HZ3	LYS	B	544	-29.101	12.925	67.618	1.00	0.00	0.274	HD
ATOM 18293	N	ASP	B	545	-24.571	9.343	71.141	1.00	14.59	-0.346	N
ATOM 18294	HN	ASP	B	545	-25.478	9.364	71.318	1.00	0.00	0.163	HD
ATOM 18295	CA	ASP	B	545	-24.067	7.869	72.033	1.00	16.70	0.186	C
ATOM 18297	C	ASP	B	545	-25.149	8.666	72.388	1.00	16.33	-0.276	OA
ATOM 18298	O	ASP	B	545	-26.307	8.533	71.102	1.00	21.16	-0.271	OA
ATOM 18299	CB	ASP	B	545	-23.534	8.533	73.306	1.00	15.70	0.147	C
ATOM 18302	CG	ASP	B	545	-22.445	8.465	73.374	1.00	21.41	0.146	C
ATOM 18303	CD1	ASP	B	545	-21.999	6.637	73.382	1.00	23.00	-0.648	OA
ATOM 18304	CD2	ASP	B	545	-21.986	8.004	70.048	1.00	25.94	-0.648	OA
ATOM 18305	N	GLN	B	546	-24.838	7.444	74.464	1.00	20.01	-0.346	N
ATOM 18306	HN	GLN	B	546	-23.852	5.561	73.161	1.00	0.00	0.163	HD
ATOM 18307	CA	CYS	B	546	-25.796	4.740	73.405	1.00	19.11	0.186	C
ATOM 18309	C	CYS	B	546	-25.828	7.441	74.464	1.00	22.49	-0.276	OA
ATOM 18310	O	CYS	B	546	-24.922	5.322	75.540	1.00	22.60	-0.271	OA
ATOM 18311	CB	CYS	B	546	-25.224	3.387	72.954	1.00	18.97	0.121	C
ATOM 18314	CG	CYS	B	546	-25.471	3.387	72.954	1.00	21.31	0.121	C
ATOM 18315	N	ALA	B	547	-26.822	4.135	75.562	1.00	19.81	-0.346	N
ATOM 18316	HN	ALA	B	547	-27.606	3.762	75.026	1.00	0.00	0.163	HD
ATOM 18317	CA	ALA	B	547	-26.802	3.151	74.464	1.00	24.49	-0.276	OA
ATOM 18319	C	ALA	B	547	-25.834	2.819	77.315	1.00	24.96	0.240	C
ATOM 18320	O	ALA	B	547	-25.944	1.863	76.578	1.00	33.35	-0.271	OA
ATOM 18321	CB	ALA	B	547	-26.169	3.437	73.306	1.00	23.31	0.146	C
ATOM 18325	N	GLY	B	548	-24.913	2.919	78.238	1.00	25.34	-0.351	N
ATOM 18326	HN	GLY	B	548	-24.825	3.750	78.823	1.00	0.00	0.163	HD
ATOM 18327	CA	GLY	B	548	-27.027	2.819	77.315	1.00	21.31	-0.276	OA
ATOM 18330	C	GLY	B	548	-22.751	1.961	77.589	1.00	26.17	0.236	C
ATOM 18331	O	GLY	B	548	-22.604	0.011	76.940	1.00	30.37	-0.276	OA
ATOM 18332	N	GLM	B	549	-21.856	0.986	77.306	1.00	23.02	-0.346	N
ATOM 18333	HN	GLM	B	549	-22.007	0.121	78.059	1.00	0.00	0.163	HD
ATOM 18334	CA	GLM	B	549	-23.164	0.725	77.315	1.00	27.72	-0.276	OA
ATOM 18336	C	GLM	B	549	-20.903	0.770	75.284	1.00	26.36	0.243	C
ATOM 18337	O	GLM	B	549	-21.653	-0.186	75.016	1.00	23.02	-0.271	OA
ATOM 18338	CB	GLM	B	549	-21.653	0.371	73.338	1.00	38.24	0.243	C
ATOM 18341	CG	GLM	B	549	-18.128	0.724	76.746	1.00	60.16	-0.175	OA
ATOM 18344	CD	GLM	B	549	-16.966	-0.167	77.114	1.00	71.93	0.215	C
ATOM 18345	HE2	GLM	B	549	-18.445	0.330	77.974	1.00	71.41	-0.276	OA
ATOM 18346	HE3	GLM	B	549	-15.779	1.276	76.625	1.00	0.00	0.159	HD
ATOM 18347	HE4	GLM	B	549	-14.946	-0.262	77.223	1.00	0.00	0.159	HD
ATOM 18348	OH1	GLM	B	549	-19.209	0.828	76.536	1.00	80.29	0.122	C
ATOM 18349	N	PRO	B	550	-20.312	1.469	74.308	1.00	23.45	-0.337	N
ATOM 18350	CA	PRO	B	550	-20.561	1.135	72.924	1.00	21.77	0.179	C
ATOM 18352	C	PRO	B	550	-19.209	0.828	76.536	1.00	80.29	0.122	C
ATOM 18353	O	PRO	B	550	-18.843	-0.322	72.985	1.00	19.07	-0.271	OA
ATOM 18354	CB	PRO	B	550	-19.936	2.299	72.138	1.00	22.73	0.037	C
ATOM 18357	CG	PRO	B	550	-19.442	2.656	74.464	1.00	27.08	0.122	C
ATOM 18360	CD	PRO	B	550	-20.545	-0.896	71.606	1.00	16.02	-0.346	N
ATOM 18364	HN	GLU	B	551	-21.454	0.606	71.335	1.00	20.65	0.163	HD
ATOM 18365	CA	GLU	B	551	-19.967	-2.086	70.976	1.00	20.45	0.177	C
ATOM 18367	C	GLU	B	551	-19.276	-1.667	69.658	1.00	18.66	0.241	C
ATOM 18368	O	GLU	B	551	-19.491	-1.410	69.658	1.00	18.39	-0.276	OA
ATOM 18369	CB	GLU	B	551	-21.110	-3.306	70.553	1.00	23.50	0.045	C
ATOM 18372	CG	GLU	B	551	-22.117	-3.286	71.652	1.00	45.05	0.116	C
ATOM 18375	CD	GLU	B	551	-22.269	-4.009	70.990	1.00	44.62	0.035	C
ATOM 18376	OE1	GLU	B	551	-23.943	-4.009	70.263	1.00	45.13	-0.648	OA
ATOM 18377	OE2	GLU	B	551	-23.517	-3.142	72.067	1.00	61.59	-0.648	OA
ATOM 18378	N	LEU	B	552	-19.669	0.695	69.137	1.00	14.41	-0.346	N
ATOM 18379	HN	LEU	B	552	-20.013	0.108	69.643	1.00	0.00	0.163	HD
ATOM 18380	CA	LEU	B	552	-19.170	-0.091	67.824	1.00	16.73	0.177	C
ATOM 18382	C	LEU	B	552	-19.071	0.688	68.932	1.00	17.28	-0.276	OA
ATOM 18383	O	LEU	B	552	-19.982	2.088	68.154	1.00	14.25	-0.271	OA
ATOM 18384	CB	LEU	B	552	-20.168	-0.550	66.769	1.00	19.73	0.038	C
ATOM 18387	CG	LEU	B	552	-19.785	-0.550	66.769	1.00	22.60	0.038	

AT0M	18798	1H1	ARG	B	579	-16.704	7.015	72.706	1.00	0.00	0.174	HD
AT0M	18799	2H1	ARG	B	579	-16.483	9.807	74.137	1.00	0.00	0.174	HD
AT0M	18800	NH2	ARG	B	579	-18.909	6.628	74.988	1.00	21.81	-0.235	N
AT0M	18801	1H2	ARG	B	579	-19.922	6.543	75.073	1.00	0.00	0.174	HD
AT0M	18802	2H2	ARG	B	579	-20.308	6.579	75.204	1.00	0.00	0.174	HD
AT0M	18803	N	VAL	B	580	-21.396	7.741	67.652	1.00	9.77	-0.346	N
AT0M	18804	HN	VAL	B	580	-21.977	8.228	68.334	1.00	0.00	0.163	HD
AT0M	18805	CA	VAL	B	580	-20.808	9.469	69.801	1.00	9.80	-0.241	C
AT0M	18807	C	VAL	B	580	-19.860	9.526	67.093	1.00	13.04	0.241	C
AT0M	18808	O	VAL	B	580	-20.283	10.304	67.949	1.00	11.19	-0.271	OA
AT0M	18809	CB	VAL	B	580	-21.863	9.167	65.643	1.00	11.16	0.009	C
AT0M	18811	CG1	VAL	B	580	-21.199	10.022	64.537	1.00	13.78	0.012	C
AT0M	18815	CG2	VAL	B	580	-22.748	8.065	65.015	1.00	11.14	0.012	C
AT0M	18819	N	VAL	B	581	-18.624	9.567	66.598	1.00	7.94	-0.348	N
AT0M	18820	HN	VAL	B	581	-18.327	8.862	65.924	1.00	0.00	0.163	HD
AT0M	18821	CA	VAL	B	581	-17.688	10.612	67.011	1.00	7.60	0.180	C
AT0M	18823	C	VAL	B	581	-18.428	9.812	66.528	1.00	11.33	0.012	C
AT0M	18824	O	VAL	B	581	-17.162	11.002	64.697	1.00	11.03	-0.271	OA
AT0M	18825	CB	VAL	B	581	-16.330	9.984	67.367	1.00	7.59	0.009	C
AT0M	18827	CG1	VAL	B	581	-15.229	11.042	67.938	1.00	14.68	0.012	C
AT0M	18831	CG2	VAL	B	581	-16.491	9.151	68.656	1.00	14.53	0.012	C
AT0M	18835	N	SER	B	582	-17.673	12.810	65.951	1.00	7.95	-0.344	N
AT0M	18836	HN	SER	B	582	-18.010	13.152	66.876	1.00	0.00	0.163	HD
AT0M	18837	CA	SER	B	582	-17.415	13.751	64.882	1.00	10.19	0.200	C
AT0M	18839	C	SER	B	582	-15.959	14.195	65.156	1.00	8.61	0.243	C
AT0M	18840	O	SER	B	582	-16.468	14.886	66.938	1.00	9.31	-0.271	OA
AT0M	18841	CB	SER	B	582	-18.329	14.978	64.988	1.00	10.24	0.199	C
AT0M	18844	CG	SER	B	582	-18.054	15.926	63.926	1.00	10.02	-0.398	OA
AT0M	18845	HC	SER	B	582	-18.620	16.896	63.985	1.00	0.00	0.209	HD
AT0M	18846	N	ME7	B	583	-15.107	14.202	64.152	1.00	6.17	-0.346	N
AT0M	18847	HN	ME7	B	583	-16.463	15.212	64.351	1.00	0.00	0.163	HD
AT0M	18848	CA	ME7	B	583	-13.712	14.571	64.311	1.00	5.00	0.177	C
AT0M	18850	C	ME7	B	583	-13.312	15.678	63.362	1.00	8.48	0.243	C
AT0M	18851	O	ME7	B	583	-14.828	16.472	62.759	1.00	10.24	-0.271	OA
AT0M	18852	CB	ME7	B	583	-12.814	13.305	64.092	1.00	13.13	0.045	C
AT0M	18855	CG	AMET	B	583	-11.487	13.540	64.836	0.50	21.88	0.076	C
AT0M	18858	SD	AMET	B	583	-10.395	13.743	65.192	0.50	19.82	-0.271	OA
AT0M	18859	CE	AMET	B	583	-10.714	11.154	66.125	0.50	17.86	0.089	C
AT0M	18863	N	PRO	B	584	-13.782	16.911	63.579	1.00	7.43	-0.337	N
AT0M	18864	CA	PRO	B	584	-13.446	18.248	64.246	1.00	7.92	-0.271	OA
AT0M	18866	C	PRO	B	584	-11.937	18.285	62.663	1.00	10.57	0.241	C
AT0M	18867	O	PRO	B	584	-11.477	18.730	61.615	1.00	9.17	-0.271	OA
AT0M	18868	CB	PRO	B	584	-12.662	19.448	62.294	1.00	7.43	0.241	C
AT0M	18871	CG	PRO	B	584	-14.569	18.857	64.658	1.00	8.53	0.022	C
AT0M	18874	CD	PRO	B	584	-14.707	17.331	64.658	1.00	11.90	0.127	C
AT0M	18877	N	SER	B	585	-11.923	19.159	64.378	1.00	11.12	-0.344	N
AT0M	18878	HN	SER	B	585	-11.630	17.743	64.660	1.00	0.00	0.163	HD
AT0M	18879	CA	SER	B	585	-9.760	18.703	63.654	1.00	9.58	0.200	C
AT0M	18881	C	SER	B	585	-9.923	19.149	64.378	1.00	11.12	-0.344	N
AT0M	18882	O	SER	B	585	-9.027	17.186	65.614	1.00	7.62	-0.271	OA
AT0M	18883	CB	SER	B	585	-9.371	19.676	64.262	1.00	9.26	0.199	C
AT0M	18886	CG	SER	B	585	-9.951	18.804	64.983	1.00	10.69	-0.346	N
AT0M	18887	HC	SER	B	585	-7.713	20.645	64.755	1.00	0.00	0.209	HD
AT0M	18888	N	THR	B	586	-8.203	16.627	63.633	1.00	6.01	-0.344	N
AT0M	18889	HN	THR	B	586	-9.160	16.543	62.621	1.00	0.00	0.163	HD
AT0M	18890	CA	THR	B	586	-7.384	15.300	64.319	1.00	7.03	0.205	C
AT0M	18892	C	THR	B	586	-6.272	16.009	65.132	1.00	12.00	0.243	C
AT0M	18893	O	THR	B	586	-6.913	15.476	66.063	1.00	10.05	-0.271	OA
AT0M	18894	CB	THR	B	586	-6.799	14.453	63.221	1.00	11.41	0.146	C
AT0M	18896	CG2	THR	B	586	-7.898	13.748	62.445	1.00	9.26	0.042	C
AT0M	18900	CG1	ASP	B	587	-6.047	14.262	62.810	1.00	7.99	-0.346	N
AT0M	18901	HG1	ASP	B	587	-5.357	15.703	62.810	1.00	0.00	0.210	HD
AT0M	18902	N	ASP	B	587	-5.654	17.135	64.689	1.00	9.58	-0.345	N
AT0M	18903	HN	ASP	B	587	-4.924	17.588	63.926	1.00	0.00	0.163	HD
AT0M	18904	CA	ASP	B	587	-4.606	17.727	65.503	1.00	8.26	0.186	C
AT0M	18906	C	ASP	B	587	-5.082	18.315	66.828	1.00	9.99	0.241	C
AT0M	18907	O	ASP	B	587	-4.337	18.687	66.328	1.00	11.48	-0.271	OA
AT0M	18908	CB	ASP	B	587	-3.824	18.791	64.700	1.00	10.21	0.147	C
AT0M	18911	CG	ASP	B	587	-4.758	19.834	64.137	1.00	8.24	0.175	C
AT0M	18912	CG1	ASP	B	587	-4.001	19.560	63.926	1.00	9.48	-0.346	N
AT0M	18913	CG2	ASP	B	587	-4.881	20.925	64.743	1.00	9.53	-0.648	OA
AT0M	18914	N	ALA	B	588	-6.267	18.921	66.883	1.00	11.41	-0.346	N
AT0M	18915	HN	ALA	B	588	-6.826	19.029	66.883	1.00	12.16	0.163	HD
AT0M	18916	CA	ALA	B	588	-6.793	19.462	68.134	1.00	12.49	0.146	C
AT0M	18918	C	ALA	B	588	-7.167	18.296	69.038	1.00	9.11	0.242	C
AT0M	18919	O	ALA	B	588	-6.839	18.421	69.038	1.00	11.41	-0.271	OA
AT0M	18920	CB	ALA	B	588	-8.014	20.360	67.893	1.00	7.85	0.042	C
AT0M	18924	N	PHB	B	589	-7.740	17.211	68.486	1.00	5.00	-0.346	N
AT0M	18925	HN	PHB	B	589	-7.975	17.224	67.489	1.00	0.00	0.163	HD
AT0M	18926	CA	PHB	B	589	-8.035	16.020	69.287	1.00	7.91	0.180	C
AT0M	18928	C	PHB	B	589	-8.011	16.376	69.287	1.00	11.75	-0.346	N
AT0M	18929	O	PHB	B	589	-6.748	14.987	71.074	1.00	10.86	-0.271	OA
AT0M	18930	CB	PHB	B	589	-8.736	14.946	68.996	1.00	9.13	0.073	C
AT0M	18933	CG	PHB	B	589	-10.172	13.700	69.188	1.00	8.48	-0.346	N
AT0M	18934	CD1	PHB	B	589	-10.174	13.764	70.078	1.00	9.59	0.007	C
AT0M	18936	CD2	PHB	B	589	-8.407	12.522	69.019	1.00	12.87	0.007	C
AT0M	18938	CG1	PHB	B	589	-10.522	12.611	70.803	1.00	12.94	-0.346	N
AT0M	18940	CG2	PHB	B	589	-8.747	11.382	69.747	1.00	12.49	0.001	C
AT0M	18942	CG	PHB	B	589	-9.823	11.443	70.638	1.00	10.44	0.000	C
AT0M	18944	N	ARG	B	590	-5.758	15.488	68.128	1.00	0.00	0.163	HD
AT0M	18945	HN	ARG	B	590	-5.758	15.488	68.128	1.00	0.00	0.163	HD
AT0M	18946	CA	ARG	B	590	-4.495	14.557	69.606	1.00	12.47	0.186	C
AT0M	18948	C	ARG	B	590	-4.354	14.734	71.589	1.00	12.68	-0.271	OA
AT0M	18950	CB	ARG	B	590	-3.438	14.437	68.504	1.00	11.73	0.147	C
AT0M	18953	CG	ARG	B	590	-4.766	13.371	67.826	1.00	11.41	-0.346	N
AT0M	18954	OD1	ASP	B	590	-4.726	12.574	67.632	1.00	11.89	-0.648	OA
AT0M	18955	OD2	ASP	B	590	-3.058	13.385	66.432	1.00	15.43	-0.648	OA
AT0M	18956	N	LYS	B	591	-4.633	17.067	70.100	1.00	0.00	0.163	HD
AT0M	18957	HN	LYS	B	591	-4.633	17.067	70.100	1.00	0.00	0.163	HD
AT0M	18958	CA	LYS	B	591	-3.533	17.458	71.892	1.00	12.36	0.176	C
AT0M	18960	C	LYS	B	591	-4.333	17.344	73.171	1.00	10.70	-0.271	OA
AT0M	18961	O	LYS	B	591	-3.815	17.910	74.191	1.00	15.83	-0.271	OA
AT0M	18962	CB	LYS	B	591	-3.520	18.949	71.443	1.00	15.67	0.035	C
AT0M	18965	CG	LYS	B	591	-4.493	18.248	70.449	1.00	10.04	-0.346	N
AT0M	18968	CD	LYS	B	591	-2.379	20.549	69.849	1.00	25.84	0.027	C
AT0M	18971	CG	LYS	B	591	-1.182	20.661	68.905	1.00	30.62	0.229	C
AT0M	18974	NE	LYS	B	591	-2.459	19.524	70.825	1.00	16.65	-0.346	N
AT0M	18975	H21	LYS	B	591	-0.699	21.567					

ATOM 19372	CE2	TYR	B 618	-3.787	11.344	59.239	1.00	10.27	0.037 A
ATOM 19374	CE2	TYR	B 618	-5.053	0.291	59.239	1.00	10.27	0.037 A
ATOM 19376	CB	TYR	B 618	-4.858	11.514	60.101	1.00	10.77	0.065 A
ATOM 19377	OH	TYR	B 618	-4.661	12.195	61.304	1.00	12.43	-0.361 DA
ATOM 19378	HN	TYR	B 618	-5.398	12.667	61.304	1.00	12.43	0.037 A
ATOM 19379	N	TYR	B 618	-6.684	6.721	57.953	1.00	9.72	-0.346 N
ATOM 19380	HN	TRP	B 619	-7.053	6.552	57.017	1.00	0.00	0.163 HD
ATOM 19381	CA	TRP	B 619	-7.518	6.135	59.314	1.00	10.77	0.065 A
ATOM 19383	C	TRP	B 619	-7.096	5.152	59.897	1.00	11.04	0.241 C
ATOM 19384	O	TRP	B 619	-7.626	5.072	61.024	1.00	10.35	-0.271 OA
ATOM 19385	CB	TRP	B 619	-9.012	6.201	58.625	1.00	6.37	0.075 C
ATOM 19388	CG	TRP	B 619	-9.486	7.614	58.382	1.00	6.75	-0.028 A
ATOM 19389	CD1	TRP	B 619	-9.626	8.162	57.150	1.00	9.64	0.096 A
ATOM 19391	CD2	TRP	B 619	-9.825	8.609	59.329	1.00	5.68	-0.002 A
ATOM 19392	CE2	TRP	B 619	-10.173	9.779	58.661	1.00	11.54	0.042 A
ATOM 19393	CE3	TRP	B 619	-9.873	8.590	60.776	1.00	11.30	0.014 A
ATOM 19395	HE1	TRP	B 619	-10.745	9.420	57.429	1.00	22.68	-0.205 DA
ATOM 19396	HE1	TRP	B 619	-10.231	10.137	56.595	1.00	0.00	0.165 HD
ATOM 19397	CD2	TRP	B 619	-10.559	10.960	59.313	1.00	11.00	0.030 A
ATOM 19399	CG3	TRP	B 619	-10.659	9.969	62.742	1.00	12.89	-0.166 DA
ATOM 19401	CH2	TRP	B 619	-10.563	10.942	60.698	1.00	12.89	0.002 A
ATOM 19403	N	TRP	B 620	-6.056	4.410	59.508	1.00	9.65	-0.346 N
ATOM 19404	HN	TRP	B 620	-5.619	4.582	58.602	1.00	0.00	0.163 HD
ATOM 19405	CA	TRP	B 620	-5.541	3.352	60.377	1.00	9.05	0.180 C
ATOM 19407	C	TRP	B 620	-4.985	3.928	61.681	1.00	10.40	0.241 C
ATOM 19408	O	TRP	B 620	-4.407	2.549	59.743	1.00	9.43	0.073 C
ATOM 19409	CB	TRP	B 620	-4.942	1.671	58.615	1.00	13.24	-0.056 A
ATOM 19413	CD1	TRP	B 620	-4.727	2.054	57.016	1.00	9.02	0.010 A
ATOM 19415	CD2	TRP	B 620	-4.727	2.054	57.016	1.00	9.02	0.010 A
ATOM 19417	CE1	TRP	B 620	-5.037	1.873	57.848	1.00	6.39	0.037 A
ATOM 19419	CE2	TRP	B 620	-5.219	1.256	56.267	1.00	15.53	0.037 A
ATOM 19421	CZ	TRP	B 620	-5.852	0.079	56.545	1.00	22.77	0.065 A
ATOM 19422	OH	TRP	B 620	-6.771	1.518	55.702	1.00	0.00	0.217 HD
ATOM 19423	HN	TRP	B 620	-6.771	1.518	55.702	1.00	0.00	0.217 HD
ATOM 19424	N	LYS	B 621	-4.608	5.199	61.724	1.00	7.38	-0.346 N
ATOM 19425	HN	LYS	B 621	-4.615	5.824	62.976	1.00	0.00	0.163 HD
ATOM 19426	CA	LYS	B 621	-4.183	5.824	62.976	1.00	0.00	0.163 HD
ATOM 19428	C	LYS	B 621	-5.253	5.749	64.057	1.00	11.58	0.241 C
ATOM 19429	O	LYS	B 621	-4.918	4.644	60.886	1.00	10.11	-0.271 OA
ATOM 19430	CB	LYS	B 621	-3.932	7.808	62.666	1.00	9.85	0.035 C
ATOM 19433	CG	LYS	B 621	-3.568	8.059	63.952	1.00	11.27	0.004 C
ATOM 19436	CD	LYS	B 621	-4.182	12.114	64.829	1.00	17.49	0.229 C
ATOM 19439	CE	LYS	B 621	-2.773	10.122	64.829	1.00	17.49	0.229 C
ATOM 19442	N2	LYS	B 621	-2.381	11.619	64.659	1.00	26.11	-0.079 N
ATOM 19443	HE1	LYS	B 621	-3.182	12.114	64.829	1.00	17.49	0.229 C
ATOM 19444	HE2	LYS	B 621	-3.085	12.123	64.120	1.00	0.00	0.274 HD
ATOM 19445	HE3	LYS	B 621	-1.588	11.691	64.021	1.00	0.00	0.274 HD
ATOM 19446	N	LYS	B 622	-6.923	5.824	62.976	1.00	6.68	0.163 HD
ATOM 19447	HN	TYR	B 622	-6.777	5.885	62.741	1.00	0.00	0.163 HD
ATOM 19448	CA	TYR	B 622	-7.625	5.815	64.694	1.00	9.56	0.180 C
ATOM 19450	C	TYR	B 622	-8.434	5.111	61.181	1.00	11.81	-0.271 OA
ATOM 19451	O	TYR	B 622	-9.177	4.317	65.610	1.00	14.63	-0.271 OA
ATOM 19452	CB	TYR	B 622	-8.620	7.004	64.488	1.00	7.76	0.073 C
ATOM 19455	CG	TYR	B 622	-8.871	8.304	66.224	1.00	11.40	-0.056 A
ATOM 19456	CD1	TYR	B 622	-7.579	8.744	66.012	1.00	9.53	0.010 A
ATOM 19458	CD2	TYR	B 622	-7.442	9.067	63.660	1.00	12.52	0.010 A
ATOM 19460	CE1	TYR	B 622	-6.852	8.909	66.216	1.00	8.70	0.037 A
ATOM 19462	CE2	TYR	B 622	-6.751	10.254	63.851	1.00	10.13	0.037 A
ATOM 19464	CZ	TYR	B 622	-6.482	10.672	65.145	1.00	13.36	0.065 A
ATOM 19465	OH	TYR	B 622	-6.788	1.856	62.666	1.00	0.00	-0.346 N
ATOM 19466	HN	TYR	B 622	-5.608	12.142	66.212	1.00	0.00	0.217 HD
ATOM 19467	N	VAL	B 623	-8.368	3.679	63.595	1.00	10.16	-0.346 N
ATOM 19468	HN	VAL	B 623	-8.758	3.869	62.318	1.00	10.69	0.180 C
ATOM 19469	CA	VAL	B 623	-9.226	2.472	63.631	1.00	11.59	0.180 C
ATOM 19471	C	VAL	B 623	-8.419	1.196	63.814	1.00	13.49	0.241 C
ATOM 19472	O	VAL	B 623	-9.018	2.979	64.420	1.00	17.68	-0.346 N
ATOM 19473	CB	VAL	B 623	-10.168	2.287	62.424	1.00	14.40	0.009 C
ATOM 19475	CD1	VAL	B 623	-10.929	3.592	62.113	1.00	14.55	0.012 C
ATOM 19479	CD2	VAL	B 623	-10.382	3.922	61.369	1.00	15.00	-0.166 DA
ATOM 19483	N	GLY	B 624	-7.130	1.228	63.616	1.00	14.08	-0.351 N
ATOM 19484	HN	GLY	B 624	-6.724	2.100	63.278	1.00	0.00	0.163 HD
ATOM 19485	CA	GLY	B 624	-6.236	1.935	63.278	1.00	0.00	0.163 HD
ATOM 19488	C	GLY	B 624	-6.391	-0.988	62.757	1.00	17.52	0.236 C
ATOM 19489	O	GLY	B 624	-6.976	-0.700	61.716	1.00	18.04	-0.272 OA
ATOM 19490	N	LEU	B 625	-5.847	6.829	63.112	1.00	14.40	-0.346 N
ATOM 19491	HN	LEU	B 625	-5.367	-3.238	63.897	1.00	0.00	0.163 HD
ATOM 19492	CA	LEU	B 625	-5.920	-3.208	62.013	1.00	13.78	0.177 C
ATOM 19494	C	LEU	B 625	-7.210	-1.008	62.178	1.00	19.24	-0.241 C
ATOM 19495	O	LEU	B 625	-7.456	-4.834	61.236	1.00	23.56	-0.271 OA
ATOM 19496	CB	LEU	B 625	-6.472	-1.612	62.013	1.00	15.93	0.037 A
ATOM 19499	CG	LEU	B 625	-3.359	-3.476	62.020	1.00	23.89	-0.020 C
ATOM 19501	CD1	LEU	B 625	-2.251	-4.410	62.456	1.00	24.81	0.009 C
ATOM 19505	CD2	LEU	B 625	-4.195	-3.556	60.616	1.00	30.23	-0.346 N
ATOM 19509	N	ASN	B 626	-8.019	-3.335	63.132	1.00	14.78	-0.346 N
ATOM 19510	HN	ASN	B 626	-7.800	-3.126	63.832	1.00	0.00	0.163 HD
ATOM 19511	CA	ASN	B 626	-8.289	-2.544	64.694	1.00	20.37	-0.271 OA
ATOM 19513	C	ASN	B 626	-10.469	-3.799	62.997	1.00	23.90	0.241 C
ATOM 19514	O	ASN	B 626	-11.545	-4.227	63.426	1.00	25.59	-0.271 OA
ATOM 19515	CB	ASN	B 626	-8.110	-6.139	64.970	1.00	33.05	-0.217 C
ATOM 19518	CG	ASN	B 626	-8.110	-6.139	64.970	1.00	33.05	-0.217 C
ATOM 19519	ND2	ASN	B 626	-7.286	-5.734	65.120	1.00	42.22	-0.370 N
ATOM 19520	HD2	ASN	B 626	-6.432	-6.259	64.694	1.00	42.22	-0.370 N
ATOM 19521	HD2	ASN	B 626	-6.477	-6.327	66.105	1.00	0.00	0.159 HD
ATOM 19522	OD1	ASN	B 626	-7.926	-7.147	64.294	1.00	41.40	-0.274 OA
ATOM 19523	N	GLY	B 627	-9.380	-2.417	63.812	1.00	14.40	-0.346 N
ATOM 19524	HN	GLY	B 627	-9.380	-2.417	63.812	1.00	14.40	-0.346 N
ATOM 19525	CA	GLY	B 627	-11.506	-1.802	62.142	1.00	18.49	0.225 C
ATOM 19528	C	GLY	B 627	-11.873	-3.659	60.669	1.00	21.22	-0.346 N
ATOM 19529	O	GLY	B 627	-11.385	-2.744	59.966	1.00	19.89	-0.272 OA
ATOM 19530	N	ALA	B 628	-12.673	-0.888	60.216	1.00	14.65	-0.347 N
ATOM 19531	HN	ALA	B 628	-12.673	-0.888	60.216	1.00	14.65	-0.347 N
ATOM 19532	CA	ALA	B 628	-13.006	-0.848	58.781	1.00	15.30	-0.172 C
ATOM 19534	C	ALA	B 628	-12.980	0.618	58.351	1.00	16.29	0.240 C
ATOM 19535	O	ALA	B 628	-13.807	1.935	61.181	1.00	18.80	-0.166 DA
ATOM 19536	CB	ALA	B 628	-14.441	-1.343	58.610	1.00	14.84	0.042 C
ATOM 19540	N	ILE	B 629	-12.736	0.907	57.081	1.00	10.96	-0.346 N
ATOM 19541	HN	ILE	B 629	-12.736	0.907	57.081	1.00	10.96	-0.346 N
ATOM 19542	CA	ILE	B 629	-12.655	2.294	56.652	1.00	9.70	0.180 C
ATOM 19544	C	ILE	B 629	-13.488	2.446	55.386	1.00	14.73	0.241 C
ATOM 19545	O	ILE	B 629	-11.218	1.940	55.729	1.00	14.40	0.073 C
ATOM 19546	CB	ILE	B 629	-11.209	2.721	56.287	1.00	14.23	0.013 C
ATOM 19548	CD1	ILE	B 629	-10.271	2.537	57.485	1.00	12.55	0.002 C
ATOM 19551	CD2	ILE	B 629	-11.218	1.940	55.729	1.00	14.40	0.073 C
ATOM 19555	CD1	ILE	B 629	-8.789	2.489	57.222	1.00	12.96	0.005 C
ATOM 19559	N	VAL	B 630	-14.446	3.336	55.400	1.00	11.44	-0.346 N
ATOM 19560	HN	VAL	B 630	-15.228	3.679	54.211	1.00	6.78	0.180 C
ATOM 19561	CA	VAL	B 630	-14.623	4.996	53.661	1.00	12.06	0.241 C
ATOM 19564	O	VAL	B 630	-14.928	0.070	54.128	1.00	11.30	-0.346 N
ATOM 19565	CB	VAL	B 630	-16.709	3.864	54.516	1.00	14.66	0.009 C
ATOM 19567	CD1	VAL	B 630	-17.489	4.185	53.248	1.00	14.17	0.012 C
ATOM 19571	CD2	VAL	B 630	-17.489	4.185	53.248	1.00	14.17	0.012 C
ATOM 19575	N	GLY	B 631	-13.694	4.815	52.739	1.00	10.55	-0.351 N
ATOM 19576	HN	GLY	B 631	-13.680	4.815	52.740	1.00	10.55	0.163 HD
ATOM 19577	CA	GLY	B 631	-12.860	5.991	52.404	1.00	11.83	0.225 C
ATOM 19580	C	GLY	B 631	-12.704	5.782	50.70			

ATCOM	19953	HN	ALA	B	657	-23.657	-1.586	55.677	1.00	0.00	0.163	RD
ATCOM	19954	CA	LYS	B	658	-22.789	-3.113	56.076	1.00	22.139	0.172	C
ATCOM	19956	C	ALA	B	657	-21.362	-3.652	56.539	1.00	21.74	0.240	C
ATCOM	19957	O	ALA	B	657	-20.986	-4.688	57.181	1.00	20.89	-0.271	OA
ATCOM	19958	CB	LYS	B	658	-22.852	-3.717	54.559	1.00	27.87	0.042	C
ATCOM	19962	N	LVS	B	658	-20.506	-2.688	56.302	1.00	21.36	-0.346	N
ATCOM	19963	HN	LVS	B	658	-20.805	-1.884	55.751	1.00	0.00	0.163	RD
ATCOM	19964	CA	LYS	B	658	-19.149	-2.743	56.529	1.00	16.66	0.176	C
ATCOM	19966	C	LYS	B	658	-19.129	-2.819	58.358	1.00	16.93	0.241	C
ATCOM	19967	O	LYS	B	658	-18.272	-3.484	58.926	1.00	18.74	-0.271	OA
ATCOM	19968	CB	LYS	B	658	-16.397	-1.440	56.529	1.00	22.21	0.035	C
ATCOM	19971	CG	LYS	B	658	-17.583	-1.284	55.303	1.00	32.54	0.004	C
ATCOM	19974	CD	LYS	B	658	-17.312	-2.489	54.459	1.00	36.82	0.027	C
ATCOM	19977	CE	LYS	B	658	-16.062	-2.393	53.151	1.00	50.03	0.239	C
ATCOM	19980	NZ	LYS	B	658	-17.393	-3.154	52.075	1.00	57.14	-0.079	N
ATCOM	19981	H21	LYS	B	658	-17.909	-3.090	51.198	1.00	0.00	0.274	HD
ATCOM	19982	H22	LYS	B	658	-17.228	-4.124	52.965	1.00	0.00	0.274	HD
ATCOM	19983	H23	LYS	B	658	-16.423	-2.857	51.965	1.00	0.00	0.274	HD
ATCOM	19984	N	ALA	B	659	-19.980	-2.046	59.040	1.00	15.19	-0.346	N
ATCOM	19985	HN	LYS	B	659	-20.615	-7.430	58.533	1.00	0.00	0.163	RD
ATCOM	19986	CA	ALA	B	659	-20.013	-2.071	60.492	1.00	19.10	0.172	C
ATCOM	19988	C	ALA	B	659	-20.416	-3.467	61.014	1.00	22.57	0.240	C
ATCOM	19989	O	ALA	B	659	-19.841	-4.048	61.940	1.00	20.85	-0.271	OA
ATCOM	19990	CB	ALA	B	659	-21.047	-1.062	61.025	1.00	19.39	0.042	C
ATCOM	19994	N	LYS	B	660	-21.449	-4.023	60.381	1.00	21.45	-0.346	N
ATCOM	19995	HN	LYS	B	660	-21.313	-3.509	59.632	1.00	0.00	0.163	RD
ATCOM	19996	CA	LYS	B	660	-21.933	-5.365	60.742	1.00	26.24	0.176	C
ATCOM	19998	C	LYS	B	660	-20.903	-6.465	60.591	1.00	27.01	0.241	C
ATCOM	19999	O	LYS	B	660	-20.03	-7.439	61.360	1.00	26.97	-0.271	OA
ATCOM	20000	CB	LYS	B	660	-23.180	-5.732	59.946	1.00	25.27	0.035	C
ATCOM	20003	CD	LYS	B	660	-24.464	-5.146	60.521	1.00	34.80	0.004	C
ATCOM	20006	CE	LYS	B	660	-25.612	-5.475	59.865	1.00	42.36	0.027	C
ATCOM	20009	CE	LYS	B	660	-26.953	-5.509	60.274	1.00	42.35	0.239	C
ATCOM	20012	NE	LYS	B	660	-28.076	-5.609	59.286	1.00	54.92	-0.079	N
ATCOM	20013	H21	LYS	B	660	-28.977	-5.632	59.763	1.00	0.00	0.274	HD
ATCOM	20014	H22	LYS	B	660	-27.959	-6.407	58.662	1.00	0.00	0.274	HD
ATCOM	20015	H23	LYS	B	660	-28.035	-4.863	58.592	1.00	0.00	0.274	HD
ATCOM	20016	N	LYS	B	660	-19.955	-6.360	60.678	1.00	26.91	-0.346	N
ATCOM	20017	HN	GLD	B	661	-19.972	-5.556	59.051	1.00	0.00	0.163	RD
ATCOM	20018	CA	GLD	B	661	-18.884	-7.338	59.523	1.00	26.27	0.177	C
ATCOM	20020	C	GLD	B	661	-17.974	-7.371	60.737	1.00	29.08	0.241	C
ATCOM	20021	O	GLD	B	661	-17.232	-8.344	60.962	1.00	30.19	-0.271	OA
ATCOM	20022	CB	GLD	B	661	-18.105	-7.074	58.243	1.00	33.00	0.045	C
ATCOM	20025	CG	GLD	B	661	-18.798	-7.423	56.937	1.00	46.16	0.116	C
ATCOM	20028	CD	GLD	B	661	-18.157	-6.746	55.734	1.00	57.89	0.172	C
ATCOM	20029	OE1	LYS	B	662	-16.954	-6.406	55.820	1.00	51.49	-0.648	OA
ATCOM	20030	OE2	LYS	B	662	-18.831	-6.538	54.696	1.00	60.71	-0.648	OA
ATCOM	20031	N	LEU	B	662	-17.957	-6.343	61.592	1.00	19.78	-0.346	N
ATCOM	20032	HN	LEU	B	662	-18.561	-5.542	61.406	1.00	0.00	0.163	RD
ATCOM	20033	CA	LEU	B	662	-17.122	-6.320	62.764	1.00	21.60	0.177	C
ATCOM	20035	C	LEU	B	662	-17.847	-6.901	63.987	1.00	28.52	0.241	C
ATCOM	20036	O	LEU	B	662	-17.242	-7.021	65.056	1.00	31.15	-0.271	OA
ATCOM	20037	CB	LEU	B	662	-16.726	-4.859	63.096	1.00	20.33	0.038	C
ATCOM	20040	CG	LEU	B	662	-16.026	-4.105	61.937	1.00	18.64	-0.020	C
ATCOM	20042	CD1	LEU	B	662	-15.685	-2.466	62.328	1.00	16.83	0.009	C
ATCOM	20046	CD2	LEU	B	662	-14.758	-4.813	61.507	1.00	22.60	0.009	C
ATCOM	20050	N	LEU	B	663	-19.153	-7.075	63.882	1.00	30.45	-0.345	N
ATCOM	20051	HN	LEU	B	663	-18.604	-6.900	62.982	1.00	0.00	0.163	RD
ATCOM	20052	CA	LEU	B	663	-19.967	-7.497	65.015	1.00	33.60	0.186	C
ATCOM	20054	C	LEU	B	663	-20.122	-9.017	65.120	1.00	40.57	0.196	C
ATCOM	20055	O	LEU	B	663	-20.203	-8.431	66.399	1.00	47.28	-0.646	OA
ATCOM	20056	CB	LEU	B	663	-21.338	-6.842	64.947	1.00	27.35	0.040	C
ATCOM	20059	CG	LEU	B	663	-21.392	-5.310	65.031	1.00	40.25	-0.020	C
ATCOM	20061	CD1	LEU	B	663	-22.825	-4.833	64.874	1.00	40.03	0.009	C
ATCOM	20065	CD2	LEU	B	663	-20.797	-4.842	66.354	1.00	38.68	0.009	C
ATCOM	20069	OXT	LEU	B	663	-19.647	-9.796	64.263	1.00	40.12	-0.646	OA
ATCOM	4	H1*	FFP	A	670	-5.600	27.610	40.336	1.00	0.00	0.166	HD
ATCOM	15	H3*	FFP	A	670	-6.093	24.915	37.319	1.00	0.00	0.168	HD
ATCOM	20	1H4*	FFP	A	670	-9.487	25.521	36.862	1.00	0.00	0.157	HD
ATCOM	21	2H4*	FFP	A	670	-8.083	24.575	36.312	1.00	0.00	0.157	HD
TER	22	FFP	A	670								
HEZATM	1	CM2	FFP	B	670	-4.284	25.837	38.892	1.00	12.30	0.089	C
HEZATM	2	CM4	FFP	B	670	-9.185	30.397	38.331	1.00	13.51	0.069	C
HEZATM	3	N1*	FFP	B	670	-6.215	27.154	39.661	1.00	11.68	-0.355	N
HEZATM	5	S1	FFP	B	670	-10.712	28.618	35.038	1.00	30.38	-0.087	SA
HEZATM	6	C2	FFP	B	670	-10.608	27.567	36.764	1.00	36.93	0.141	A
HEZATM	7	N3	FFP	B	670	-10.107	28.153	37.465	1.00	14.24	-0.349	N
HEZATM	8	C4	FFP	B	670	-9.768	29.477	37.318	1.00	11.95	0.021	A
HEZATM	9	C5	FFP	B	670	-10.041	28.900	36.035	1.00	24.73	0.008	A
HEZATM	10	C6	FFP	B	670	-9.745	31.317	35.653	1.00	26.35	0.075	C
HEZATM	11	C7	FFP	B	670	-10.442	31.600	34.338	1.00	31.82	0.186	C
HEZATM	12	O7	FFP	B	670	-9.507	32.396	33.647	1.00	31.26	-0.289	OA
HEZATM	13	C2*	FFP	B	670	-5.727	26.214	38.824	1.00	16.98	0.078	A
HEZATM	14	N3*	FFP	B	670	-6.486	25.622	37.940	1.00	10.00	-0.332	N
HEZATM	16	O1A	FFP	B	670	-8.853	33.580	31.697	1.00	16.83	-0.624	OA
HEZATM	17	O1B	FFP	B	670	-12.764	32.684	31.687	1.00	21.70	-0.616	OA
HEZATM	18	C4*	FFP	B	670	-7.833	25.960	37.850	1.00	13.02	0.109	A
HEZATM	19	N4*	FFP	B	670	-8.500	25.273	36.928	1.00	11.07	-0.385	N
HEZATM	22	O2A	FFP	B	670	-8.105	31.185	31.990	1.00	26.35	-0.624	OA
HEZATM	23	O2B	FFP	B	670	-11.189	33.334	29.876	1.00	14.28	-0.616	OA
HEZATM	24	C5*	FFP	B	670	-8.419	26.941	38.715	1.00	13.90	0.019	A
HEZATM	25	O3A	FFP	B	670	-10.518	31.603	31.476	1.00	18.42	-0.173	OA
HEZATM	26	O3B	FFP	B	670	-12.271	31.028	29.842	1.00	18.52	-0.616	OA
HEZATM	27	C6*	FFP	B	670	-7.550	27.494	39.598	1.00	12.25	0.102	A
HEZATM	28	C7*	FFP	B	670	-9.885	27.369	38.723	1.00	13.72	0.172	C
HEZATM	29	PA	FFP	B	670	-9.198	32.182	32.091	1.00	19.03	0.418	P
HEZATM	30	PB	FFP	B	670	-11.770	32.192	30.618	1.00	18.84	0.431	P
ATCOM	34	H1*	FFP	B	670	14.558	24.087	43.661	1.00	0.00	0.166	HD
ATCOM	45	H3*	FFP	B	670	15.022	26.390	47.008	1.00	0.00	0.168	HD
ATCOM	50	1H4*	FFP	B	670	18.336	25.661	47.594	1.00	0.00	0.157	HD
ATCOM	51	2H4*	FFP	B	670	16.948	26.552	48.201	1.00	0.00	0.157	HD
TER	52	FFP	B	670								
HEZATM	31	CM2	FFP	B	670	13.263	25.676	45.343	1.00	12.39	0.089	C
HEZATM	32	CM4	FFP	B	670	19.449	24.862	42.708	1.00	16.57	0.069	C
HEZATM	33	N1*	FFP	B	670	15.159	24.422	44.414	1.00	11.86	-0.355	N
HEZATM	35	S1	FFP	B	670	20.992	27.390	45.462	1.00	30.40	-0.087	SA
HEZATM	36	C2	FFP	B	670	20.126	26.072	46.130	1.00	37.83	0.141	A
HEZATM	37	N3	FFP	B	670	19.619	25.216	45.225	1.00	15.93	-0.349	N
HEZATM	38	C4	FFP	B	670	19.904	25.620	43.919	1.00	15.83	0.021	A