

Effect of geometry on dissolution profiles of 3D printed tablets

Alvaro Goyanes¹, Pamela Robles Martinez¹, Abdul Basit^{1,2}, Simon Gaisford^{1,2}

¹UCL School of Pharmacy, University College London, 29-39 Brunswick Square, London,
WC1N 1AX, UK

²FabRx Ltd., 3 Romney Road, Ashford, Kent TN24 0RW, UK

Key words

3D printing; controlled-release; fused deposition modeling; PVA; paracetamol; hot melt
extrusion

25 **Abstract**

26

27

28 **1. Introduction**

29 The future of medicine design and manufacture is likely to move away from mass production
30 of tablets/capsules of limited dose range towards extemporaneous fabrication of unit dosage
31 forms of any dose, personalised to the patient. The factors driving this change include the
32 development of low dose drugs with narrow therapeutic indices (for instance
33 immunosuppressants and/or blood thinners), the increasing awareness and importance of
34 pharmacogenomics (for instance in the drug sensitivity of cancer sufferers, Kim et al, 2012)
35 and the need to formulate drug combinations. To face this challenge, the pharmaceutical
36 industry needs to evaluate and embrace novel manufacturing technologies. One technology
37 with such potential is 3D printing (3DP).

38

39 Of the many types of 3D printer commercially available, fused-deposition modelling (FDM)
40 offers perhaps the most immediate potential to unit dose fabrication. In FDM 3DP an
41 extruded polymer filament is passed through a heated tip. The heat softens the polymer and
42 it is then deposited on a build plate. The temperature of the build plate can be controlled and
43 is set so that the polymer hardens. The print head deposits polymer on the build plate in the
44 x-y dimensions, creating one layer of the object to be printed. The build plate then lowers
45 and the next layer is deposited. In this fashion, an object can be fabricated in three
46 dimensions, and in a matter of minutes. The particular benefit of FDM 3DP to pharmaceuticals
47 is that the polymer filament can be loaded with a drug prior to printing, and so drug-loaded
48 unit dosage forms can be manufactured. This principle has been demonstrated, for example,
49 to tablets containing fluorescein (Goyanes et al, 2014), 4-aminosalicylate and 5-
50 aminosalicylate (Goyanes et al, 2015) and (Skowrya et al, 2015).

51

52 A further potential benefit of FDM 3DP, currently unexplored, is that the printer can be used
53 to fabricate tablets of any geometry, including shapes that would be impossible to create by
54 powder compaction. Since in principle the shape of a tablet could control its dissolution
55 profile, FDM 3DP would seem to offer a new route to design controlled-release or modified-

56 release dosage forms. The aim of this work, therefore, was to print tablets in a diverse range
57 of geometries, many not attainable by powder compaction, and to correlate their geometric
58 parameters with dissolution behaviour.

59

60 **2. Materials and methods**

61 Polyvinyl alcohol (PVA, a water-soluble synthetic polymer of molecular formula $(C_2H_4O)_n$)
62 was purchased as an extruded filament (1.75mm diameter, print temperature 190-220°C,
63 batch No: 2013-10-18, Makerbot Inc., USA). Paracetamol USP grade was obtained from
64 (Sigma-Aldrich, UK). Salts for preparing buffer dissolution media were purchased from VWR
65 International Ltd., Poole, UK.

66

67 *2.1 Preparation of PVA filament loaded with drug*

68 The commercial PVA filament (38g) was cut into small pieces (~1 mm) using a Pharma 11
69 Varicut Pelletizer (Thermo Fisher Scientific, UK) and mixed with paracetamol (2g, 5% drug
70 w/w) for 10 minutes in a Turbula[®] T2F shaker-mixer (Glen Mills Inc., USA). The mixture was
71 extruded using a single-screw FilaBot[®] hot melt extruder (Filabot, USA) at 180 °C through a
72 1.75 mm diameter nozzle (screw speed 35 rpm). The extruded filaments obtained were
73 protected from light and kept in a vacuum desiccator until printing. The drug-loading of the
74 filaments was determined by HPLC analysis.

75

76 *2.2. Printing of paracetamol dosage forms*

77 Tablets were fabricated with the drug-loaded filaments using a standard fused-deposition
78 modelling 3D printer, MakerBot Replicator 2X Desktop 3D printer (MakerBot Inc, USA). The
79 templates used to print the tablets were designed with AutoCAD 2014[®] (Autodesk Inc., USA)
80 and exported as a stereolithography (.stl) file into MakerWare v. 2.4.1 (MakerBot Inc., USA).
81 The .stl format encodes only the surface data of the object to be printed and requires the
82 thickness of the surface to be defined in order to print the desired object. The printer settings
83 were as follows: standard resolution with the raft option deactivated and an extrusion

84 temperature of 180 °C, speed while extruding (90mm/s), speed while traveling (150mm/s),
85 number of shells (2) and layer height (0.20mm). The infill percentage was 100% in order to
86 produce tablets of high density. The selected 3D geometries were cube, pyramid, cylinder,
87 sphere and torus (Figure 1). The sizes of the shapes were varied using the scale function of
88 the software to fabricate tablets of constant surface area (275 mm²), surface area/volume
89 ratio (1:1) or weight (500 mg), Tables 1-3. In all cases, however, the ratio of the length, width
90 and height of each shape was kept constant.

91

92 *2.3 Scanning electron microscopy (SEM)*

93 Surface and cross-section images of the filaments were taken with an SEM (JSM-840A
94 Scanning Microscope, JEOL GmbH, Eching, Germany). All samples for SEM testing were
95 coated with carbon (~30–40 nm).

96

97 *2.4 Thermal analysis*

98 Filaments were characterised with differential scanning calorimetry (DSC) and
99 thermogravimetric analysis (TGA). DSC measurements were performed with a Q2000 DSC
100 (TA instruments, Waters, LLC, USA) at a heating rate of 10°C/min. Calibration for cell
101 constant and enthalpy was performed with indium (T_m = 156.6°C, ΔH_f = 28.71 J/g) according
102 to the manufacturer instructions. Nitrogen was used as a purge gas with a flow rate of 50
103 mL/min for all the experiments. Data were collected with TA Advantage software for Q series
104 (version 2.8.394), and analysed using TA Instruments Universal analysis 2000. All melting
105 temperatures are reported as extrapolated onset unless otherwise stated. TA aluminium
106 pans and lids (Tzero) were used with an average sample mass of 8-10mg.

107

108 For TGA analysis, samples were heated at 10°C/min in open aluminium pans with a
109 Discovery TGA (TA instruments, Waters, LLC, USA). Nitrogen was used as a purge gas with
110 a flow rate of 25 mL/min. Data collection and analysis were performed using TA Instruments
111 Trios software and % mass loss and/or onset temperature were calculated.

112 *2.5 Characterisation of tablet morphology*

113 The physical dimensions of the tablets were measured using a digital calliper. Pictures of the
114 tablets were taken with a Nikon CoolpixS6150 with the macro option of the menu. The
115 surface areas and volumes of the tablets were calculated based on these dimensions. The
116 surface area to volume ratio was obtained dividing these values.

117

118 *2.6 Determination of drug loading*

119 A tablet or a section of drug-loaded strand (approx. 0.3g) was placed in a volumetric flask
120 with deionized water (1L) with magnetic stirring until complete dissolution. Samples of
121 solution were then filtered through 0.45 µm filters (Millipore Ltd, Ireland) and the
122 concentration of drug determined with HPLC (Hewlett Packard 1050 Series HPLC system,
123 Agilent Technologies, UK). The validated high performance liquid chromatographic assay
124 entailed injecting 20 µL samples for analysis using a mobile phase, consisting of methanol
125 (15%) and water (85%), through a Luna 5µm C8 column, 25 x 4.6 cm (Phenomenex, UK)
126 maintained at 40 °C. The mobile phase was pumped at a flow rate of 1 mL/min and the
127 eluent was screened at a wavelength of 247 nm. All measurements were made in duplicate.

128

129 *2.7 Dissolution testing*

130 Dissolution profiles were obtained using a USP-II apparatus (Model PTWS, Pharmatest,
131 Germany). In each assay, the tablets were placed at the bottom of the vessel in phosphate
132 buffer (pH= 6.8, 900 mL) under constant paddle stirring (50 rpm) at 37°C. During the
133 dissolution test, samples of paracetamol were automatically removed and filtered through
134 10µm filters and drug concentration was determined using an in-line UV spectrophotometer
135 (Cecil 2020, Cecil Instruments Ltd., Cambridge, UK) operated at the wavelength of
136 maximum absorbance of the drug in phosphate buffer (243 nm). Data were processed using
137 Icalis software (Icalis Data Systems Ltd, Berkshire, UK). Tests were conducted in triplicate
138 under sink conditions.

139

140 3. Results and discussion

141 The first important result was that it was possible to fabricate all of the shapes with 3DP,
142 Figure 2. Manufacture of such complex and intricate shapes by powder compaction would
143 be extremely challenging and so the study immediately suggests that 3DP offers a route of
144 manufacture of dosage forms of novel geometries not previously possible. As found in our
145 previous studies (Goyanes et al, 2014, 2015), 3DP tablets were not friable and so easy to
146 handle.

147
148 DSC and TGA analyses of the pure substances and extruded filament were performed in
149 order to understand how the drug was incorporated in the polymer, Figures 3-5. It is
150 apparent that paracetamol raw material melts around 168 °C, indicative of form I while PVA
151 shows a glass transition around 135 °C and melting between 175-200 °C. Significant
152 degradation of PVA is seen above 260 °C, but the printhead temperature used during tablet
153 fabrication is 230 °C. The DSC data of the paracetamol-loaded PVA filament shows no
154 evidence of melting around 168 °C, indicating that the drug is molecularly dispersed within
155 the polymer matrix as a solid solution. A glass transition is seen in both experiments
156 involving PVA (highlighted by the arrows in Figure 4), the temperature of which rises when
157 paracetamol is present, suggesting the drug is acting as an anti-plasticiser. TGA data
158 suggests that PVA is stable until 260 °C, consistent with the DSC data, while paracetamol
159 degrades significantly above 200 °C. When paracetamol is incorporated into the PVA, no
160 appreciable mass loss is seen, suggesting the polymer is stabilising the drug.

161
162 Percentage drug loadings were measured for the filament ($3.95\% \pm 0.01$) and printed tablets
163 ($3.78\% \pm 0.01$), indicating little degradation of the drug during printing, consistent with the
164 thermal stability of paracetamol above its melting point seen by DSC. Electron micrographs
165 show little evidence of paracetamol crystallites within the polymer filaments, Figure 7,
166 consistent with the DSC data. Images of the cross-section of the printed torus clearly show
167 the individual strands deposited by the printer. The strands are ca. 100 μm in diameter,

168 consistent with the nozzle diameter of the printhead, and there is some evidence of fusion of
169 strands, leading to the strength of the tablet noted earlier.

170

171 Dissolution tests reveal that the geometry plays an important role in defining drug release
172 profiles, Figure 7. When the surface area of the tablets was kept constant, drug release rates
173 were in the following order (fastest first); pyramid > torus > cube > sphere and cylinder. The
174 time to 90% release (t_{90}) varied from just under 2h (pyramid) to nearly 12 h (sphere and
175 cylinder). When tablets were prepared with a constant surface area/volume ratio, the order
176 of release rates was (fastest first); sphere and cube > torus > cylinder > pyramid. Less
177 differentiation was seen in the t_{90} values, with most shapes having a value between 2-3h.
178 Only the pyramid gave noticeably slower release (9 h). Interestingly, when tablets were
179 prepared to constant weight, dissolution behavior was similar for all geometries. Since PVA
180 releases drug via erosion, it seems that the controlling factor is simply the mass of polymer
181 present. The data do show, however, that it is possible to design a tablet with a controlled-
182 release profile (varying over 10h) by careful selection of shape and/or size.

183

184

185

186

Shape	Surface area (mm ²)	Volume (mm ³)	SA/V ratio	Weight (mg)	Density (mg/mm ³)
Cube	287.9 ± 2.1	332.3 ± 3.6	0.866 ± 0.003	268.2 ± 15.7	0.81 ± 0.05
Pyramid	270.4 ± 0.4	231.3 ± 0.5	1.169 ± 0.001	187.5 ± 3.9	0.81 ± 0.02
Cylinder	268.5 ± 3.9	314.4 ± 6.5	0.854 ± 0.005	355.3 ± 23.7.5	1.13 ± 0.05
Sphere	280.8 ± 1.4	442.3 ± 3.2	0.634 ± 0.002	505.3 ± 36.0	1.14 ± 0.08
Torus	266.8 ± 1.0	266.4 ± 1.9	1.002 ± 0.004	276.0 ± 19.6	1.04 ± 0.08

187

188 **Table 1: Physical parameters for tablets with similar surface areas**

189

190

Shape	Surface area (mm ²)	Volume (mm ³)	SA/V ratio	Weight (mg)	Density (mg/mm ³)
Cube	212.1 ± 2.2	210.1 ± 3.3	1.009 ± 0.005	186.9 ± 19.3	0.89 ± 0.11
Pyramid	356.1 ± 2.7	353.7 ± 5.2	1.007 ± 0.008	451.2 ± 12.0	1.28 ± 0.05
Cylinder	200.8 ± 3.2	202.3 ± 4.0	0.992 ± 0.004	197.6 ± 4.0	0.97 ± 0.18
Sphere	111.5 ± 1.2	110.7 ± 1.9	1.007 ± 0.006	98.5 ± 0.9	0.89 ± 0.05
Torus	266.8 ± 1.0	266.4 ± 1.9	1.002 ± 0.004	276.0 ± 19.6	1.04 ± 0.08

191

192 **Table 2: Physical parameters for tablets with similar surface/volume ratios**

193

194

195

Shape	Surface area (mm ²)	Volume (mm ³)	SA/V ratio	Weight (mg)	Density (mg/mm ³)
Cube	341.4 ± 0.5	429.2 ± 1.0	0.795 ± 0.001	477.9 ± 11.9	1.11 ± 0.03
Pyramid	393.0 ± 15.2	406.2 ± 23.4	0.968 ± 0.019	494.8 ± 16.6	1.22 ± 0.03
Cylinder	335.0 ± 15.7	438.3 ± 29.3	0.765 ± 0.016	480.0 ± 11.0	1.10 ± 0.06
Sphere	280.8 ± 1.4	442.3 ± 3.2	0.634 ± 0.002	505.3 ± 36.0	1.14 ± 0.08
Torus	387.0 ± 3.0	479.5 ± 4.3	0.807 ± 0.001	509.9 ± 29.0	1.06 ± 0.05

196

197 **Table 3: Physical parameters for tablets with similar weights**

198

199

200

201

202

203

204

205

206

207

208

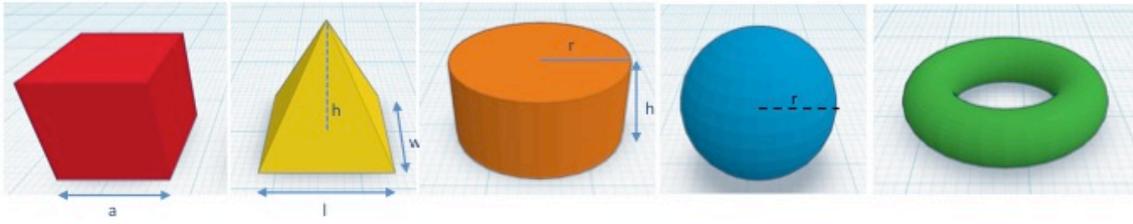
209

210

211

212

213



214

215 Figure 1. 3D representation of the 3D geometries printed (left to right; cube, pyramid,
216 cylinder, sphere and torus)

217

218

219

220

221

222

223

224

225

226

227

228

229

230



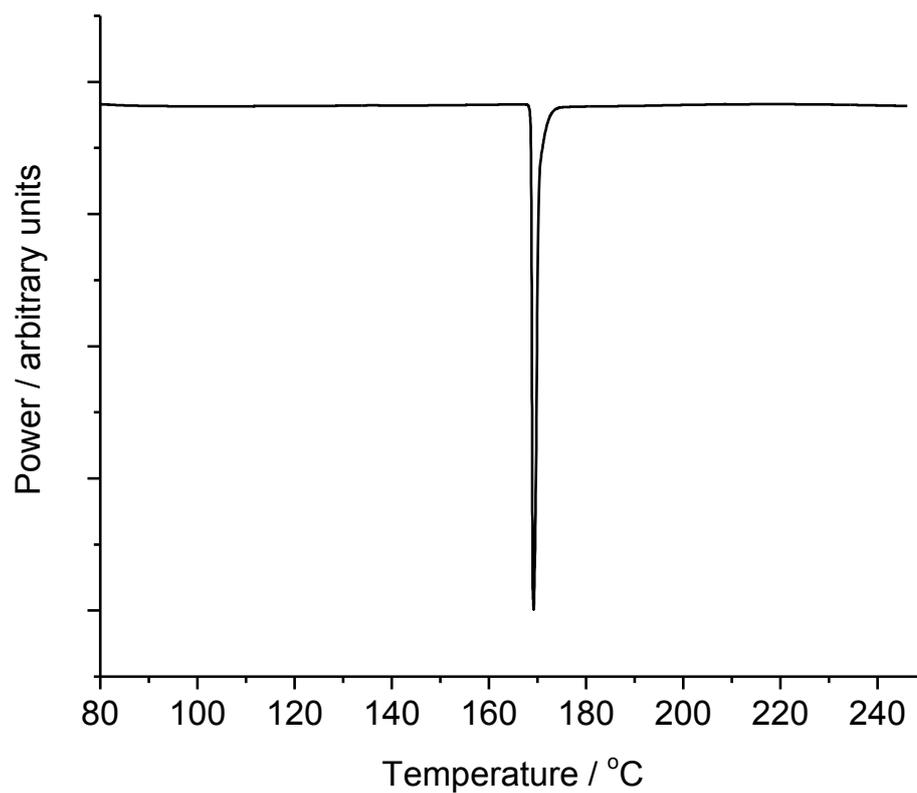
231

232

233

234

Figure 2: Images of the 3DP fabricated tablets at constant (A) Surface area, (B) surface area/volume ratio and (C) mass.



235

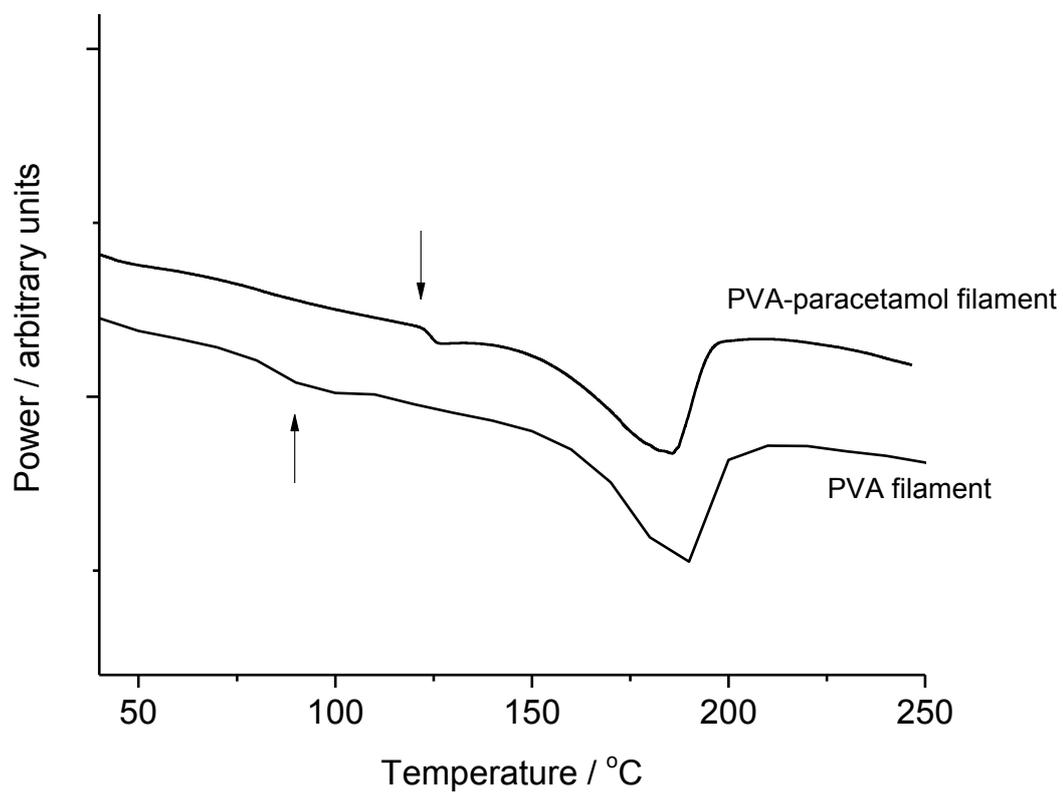
236

237 Figure 3. DSC thermal trace for paracetamol raw material, showing melt of the stable form I

238 at 168 °C.

239

240

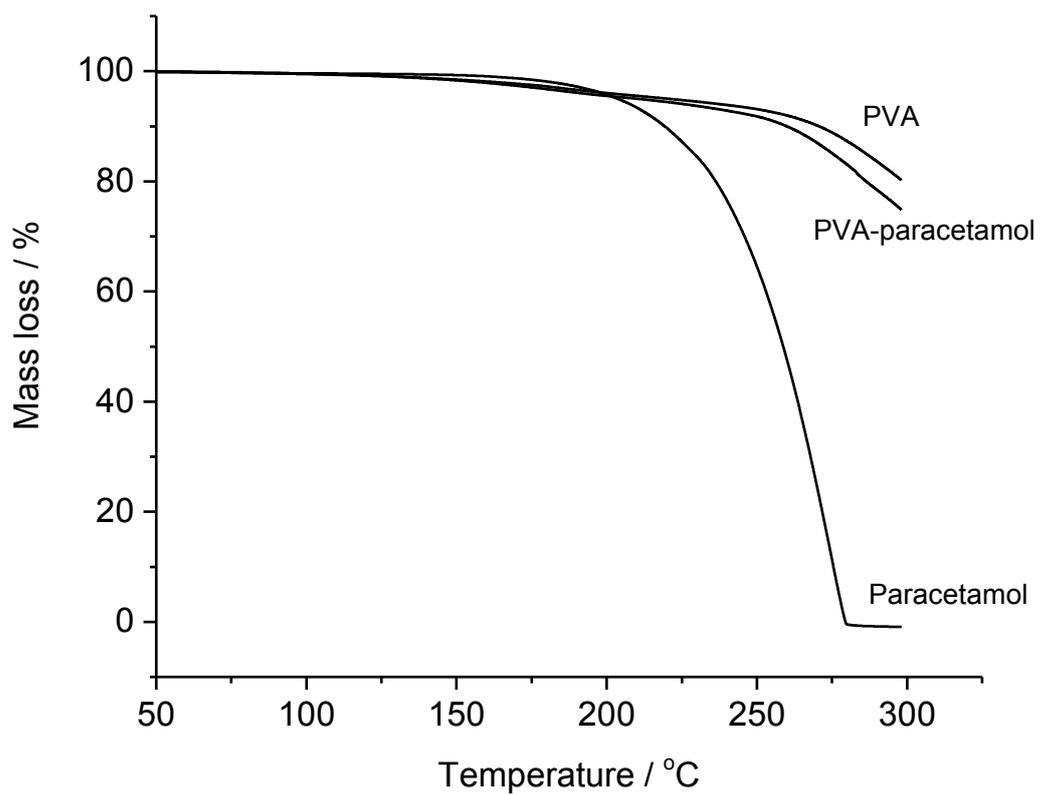


242

243 Figure 4. DSC thermal traces for PVA and PVA-paracetamol filaments. The PVA melts
244 between 175-200 °C and the arrows indicate glass transition temperatures.

245

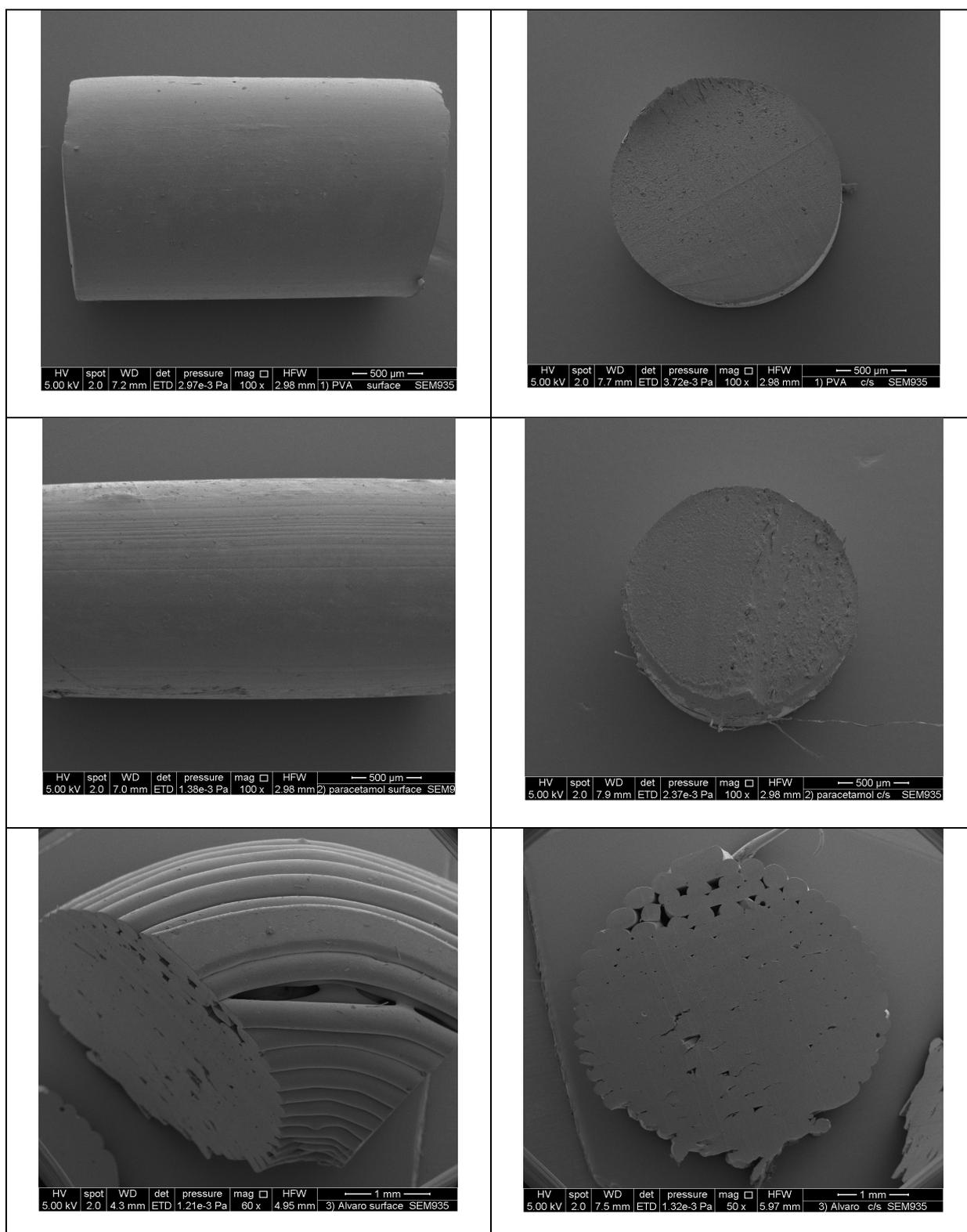
246



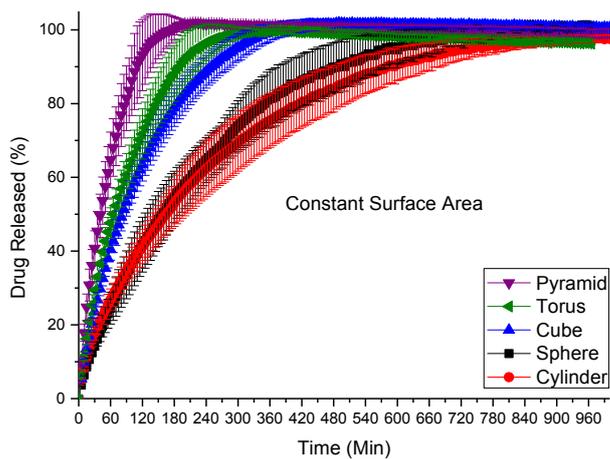
248

249 Figure 5. TGA thermal traces for paracetamol raw material and PVA and PVA-paracetamol
250 filaments.

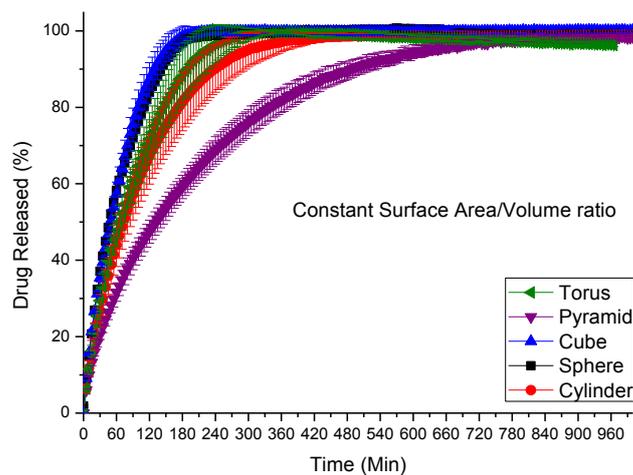
251



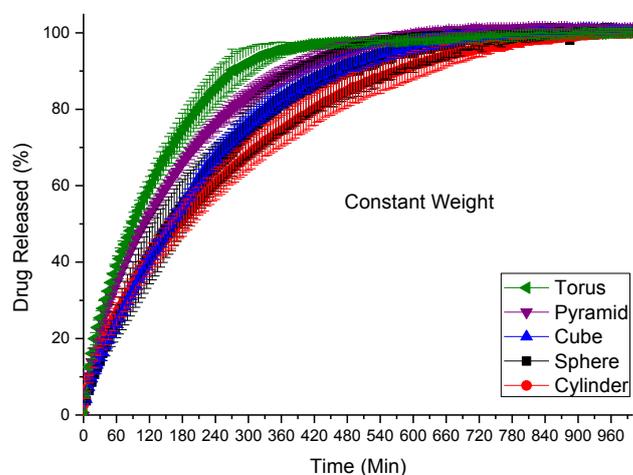
254 Figure 6. SEM images of the surface and cross-section of; Top) PVA filament, middle) PVA-
 255 paracetamol filament and bottom) a section of printed filament (in this case from the torus).



256



257



258

259 Figure 7: Paracetamol dissolution profiles from 3DP solid dosage with A) surface area 275
 260 mm², B) surface area/volume ratio 1 and C) 500 mg mass in phosphate buffer (pH 6.8)

261 **Conclusions**

262

263 **Acknowledgement**

264 Alvaro Goyanes would like to thank Fundación Alfonso Martín Escudero for the post-doctoral
265 fellowship.

266

267

268 **References**

269 Goyanes, A. Buanz, A.B.M., Basit, A.W., Gaisford, S., 2014. Fused-filament 3D printing
270 (3DP) for fabrication of tablets. *Int. J. Pharm.* 476, 88-92.

271 Goyanes, A. Buanz, A.B.M., Hatton, G.B., Gaisford, S., Basit, A.W., 2015. 3D printing of
272 modified-release aminosalicylate (4-ASA and 5-ASA) tablets. *Eur. J. Pharm. Biopharm.* 89,
273 157-162.

274 Skowrya, J., Pietrzak, K., Alhnan, M.A. 2015. Fabrication of extended-release patient-
275 tailored prednisolone tablets via fused deposition modelling (FDM) 3D printing. *Eur. J.*
276 *Pharm. Sci.* 68, 11-17.

277