



Development of a rapid, reliable and quantitative method – “SPOTi” for testing antifungal efficacy



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ABSTRACT

A reference method for the antimicrobial susceptibility testing of common fungal pathogens such as dermatophytes, is currently lacking. In this study, we report the successful adaptation of solid agar-based spot culture growth inhibition assay (SPOTi) for dermatophytes, currently being used as a gold-standard in the anti-tubercular drug discovery field. The fungal-SPOTi assay correlated with the disc-diffusion method, and is validated using mycelial plugs. We propose the fungal-SPOTi as a high-throughput alternative to the disc-diffusion and broth micro-dilution anti-fungal assays to screen novel anti-fungals.

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1. Introduction

Dermatophytes, a group of fungi, are responsible for causing infection of the hair, skin and nails in one-fifth of the world's population (Marques et al., 2000). Treatment of fungal infections is not always straight forward, for example, in onychomycosis, up to 50% of patients in clinical practice are estimated to be treatment failures (Arrese and Pierard, 2003) and 47% suffer a relapse, possibly as a result of incorrect treatment and strain heterogeneity (Bradley et al., 1999). An immediate solution to improve treatment outcomes is a rapid, straightforward and accurate means to profile the drug susceptibility of these fungal pathogens. This will allow clinicians to make an informed choice when prescribing a therapy. While several drug susceptibility screening methods for bacteria (Wiegand et al., 2008) and fungi such as yeasts and filamentous fungi have existed for many years (Rex et al., 1993), dermatophytes still lack an undisputable quantitative assay. An increase in research activity related to anti-fungal susceptibility testing of dermatophytes has led to the development of a number of broth micro- and macro-dilution methods (Araujo et al., 2009), agar-based methods using disc diffusion (Nweze et al., 2010; Macura, 1993); and E-Test (Mendez et al., 2008), colorimetric modifications (Castro et al., 2007), bioluminescence assays to enhance MIC determinations (Yoshida et al., 1997). A limitation of many of these reported methods is their

long incubation times (up to 15 days) (Macura, 1993; Karaca and Koc, 2004). In addition, solid agar-based methods for dermatophytes often report the minimum inhibitory concentration (MIC) of a drug as the concentration at which the fungal growth showed a relative growth inhibition in percent, which can give rise to discrepancies owing to operator-subjectivity (Ghannoum et al., 2006).

A rapid, but reliable method that is universally reproducible is needed in both the clinical setting, where determination of antibiotic resistance profile is the goal, and the academic and/or industrial drug-discovery wet-laboratories where irrefutable identification of novel lead molecules takes precedence. This calls for a stewardship engaged in the development of assay techniques that are objective, gold-standard and feasible.

We originally developed and previously reported a high-throughput, solid agar-based assay conducted in multi-well (six-well to ninety-six well) plates to test inhibitors of bacterial growth and viability, known as the SPOTi assay (Evangelopoulos and Bhakta, 2010; Gupta and Bhakta, 2012; Guzman et al., 2013). The latter has been independently used in a clinical setting for antibacterial drug susceptibility testing (Hall et al., 2012).

In this paper, we report the successful adaptation and application of this methodology, which can be used to test anti-fungal drug susceptibility as well as screening novel inhibitor libraries against fungal pathogens. At the first instance, we tested four antifungal agents; amorolfine, terbinafine, ciclopirox olamine and nystatin against four dermatophyte strains; *Trichophyton rubrum*, *Trichophyton tonsurans*, *Trichophyton*

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Table 1
The average conidial count and CFU of the dermatophytes.

Fungi	Count (spores/ml)	CFU (spores/ml)
<i>T. rubrum</i>	50×10^4	46×10^4
<i>T. equinum</i>	61×10^4	43×10^4
<i>T. tonsurans</i>	47×10^4	26×10^4
<i>T. mentagrophytes</i>	38×10^4	19×10^4

equinum and *Trichophyton mentagrophytes*, and validated the assay using disc diffusion and mycelial plug methods.

2. Materials and methods

2.1. Materials

Four antifungal agents; amorolfine HCl (Ranbaxy Research Laboratories, India), terbinafine HCl (AK Scientific, USA) and ciclopirox olamine (Insight Biotechnology Ltd, UK) and nystatin (Sigma Aldrich Ltd, UK) were used. Sabouraud's dextrose agar (SDA), Sabouraud's dextrose broth (SDB) and dimethyl sulfoxide DMSO were purchased from Sigma-Aldrich, UK.

2.2. Isolates

Four dermatophytes strains *T. rubrum* (CBS 118892), *T. tonsurans* (CBS 112818), *T. equinum* (CBS 127.97) and *T. mentagrophytes* (MUCL 9823) were used.

2.3. Methods

2.3.1. Preparation of inocula

Stock inoculum suspensions of the dermatophytes were prepared from 14-day old cultures grown on Sabouraud's dextrose agar (SDA) slants at 25 °C. The fungal colonies were covered with 2 mL of Sabouraud's dextrose broth (SDB), and suspensions were obtained by gently probing the surface of the agar with the tip of a sterile Pasteur pipette, generating a mixture of conidial and hyphal fragments. The obtained suspensions were then filtered through four layers of sterile muslin, which retains hyphal fragments but permits the passage of dermatophyte micro-conidia. A filtration process was used as this has been shown to provide a greater reproducibility and reliability of susceptibility testing.

The number of conidia per milli-litre (mL) of the inocula were determined by: 1) counting the number of conidia using a haemocytometer

Table 2
MICs as determined from the SPOTi assay. (n = 3. Sd = 0 for all).

Fungi	MIC ($\mu\text{g/ml}$)			
	Amorolfine	Terbinafine	Ciclopirox	Nystatin
<i>T. rubrum</i>	0.01	0.0006	2.5	1.2
<i>T. equinum</i>	0.005	0.0012	2.5	1.2
<i>T. tonsurans</i>	0.01	0.0012	1.2	0.6
<i>T. mentagrophytes</i>	0.02	0.0006	2.5	1.2

and 2) determining colony forming units per mL (CFU mL^{-1}) with plate dilution assays and are shown in Table 1. For each isolate the numbers of conidia were determined in five viewing fields and averaged. With the CFU determination, the results shown are the average of three plates from three independent experiments in which strains were harvested and counted.

2.3.2. High throughput SPOTi assay

The methodology, with required modification from previously published method (Guzman et al., 2013; Evangelopoulos and Bhakta, 2010; Gupta and Bhakta, 2012), is schematically shown in Fig. 1. Serial two fold dilutions of drug were prepared from a stock solution in DMSO. Two μL of each dilution were dispensed into the individual wells of a 96-well plate followed by the addition of 200 μL of SDA media using a Multidrop™ Combi multi-dispenser (Thermo Scientific, UK). The final drug concentration range in the agar ranged between 10 and 0.005 $\mu\text{g/ml}$ for ciclopirox olamine and nystatin and 0.15 and 0.00007 $\mu\text{g/ml}$ for terbinafine HCl and amorolfine HCl. The wells were then spotted using a multichannel pipette with 5 μL of dermatophyte spore suspension. This was equivalent to approximately 2000 dermatophyte spores per well.

SDA media was used as it was found to be one of the most favoured for both soil and clinical dermatophytes isolates (Sharma and Sharma, 2011). An incubation temperature of 32 °C was used as these dermatophytes often infect keratinous tissue (Weitzman and Summerbell, 1995) such as skin, whose temperature is 32 °C. Furthermore, temperatures of 30 to 35 °C have been shown to result in optimal fungal growth and sporulation for *T. rubrum* and *T. mentagrophytes* (Sharma et al., 2012). Dermatophyte growth was assessed visually over a period of 6 days (Fig. 1).

2.3.3. Disc diffusion susceptibility testing

Discs loaded with ciclopirox olamine, nystatin, amorolfine HCl and terbinafine HCl were prepared according to the potency in the Neo-Sensitabs Susceptibility Testing Catalogue (Rosco Diagnostica Ltd,

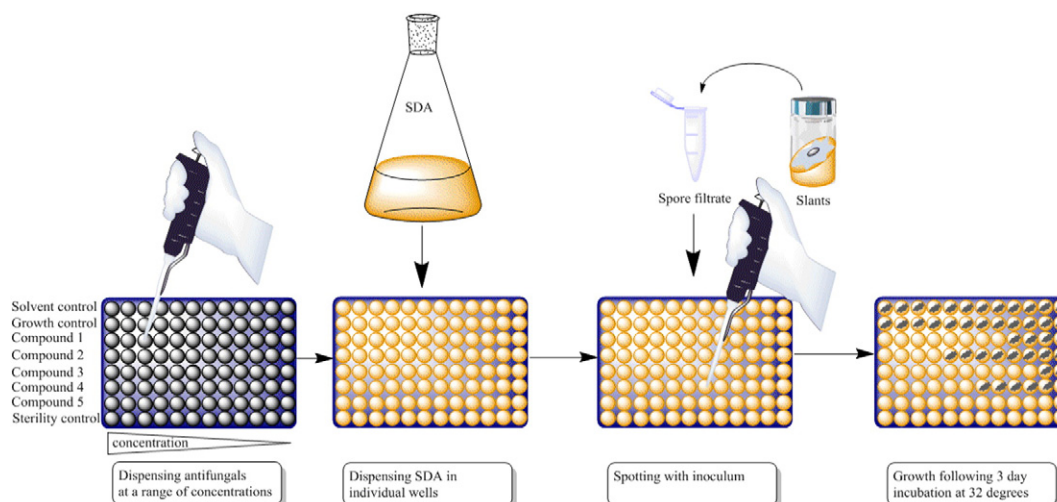


Fig. 1. Schematic explanation of the spot culture growth inhibition (SPOTi) assay.

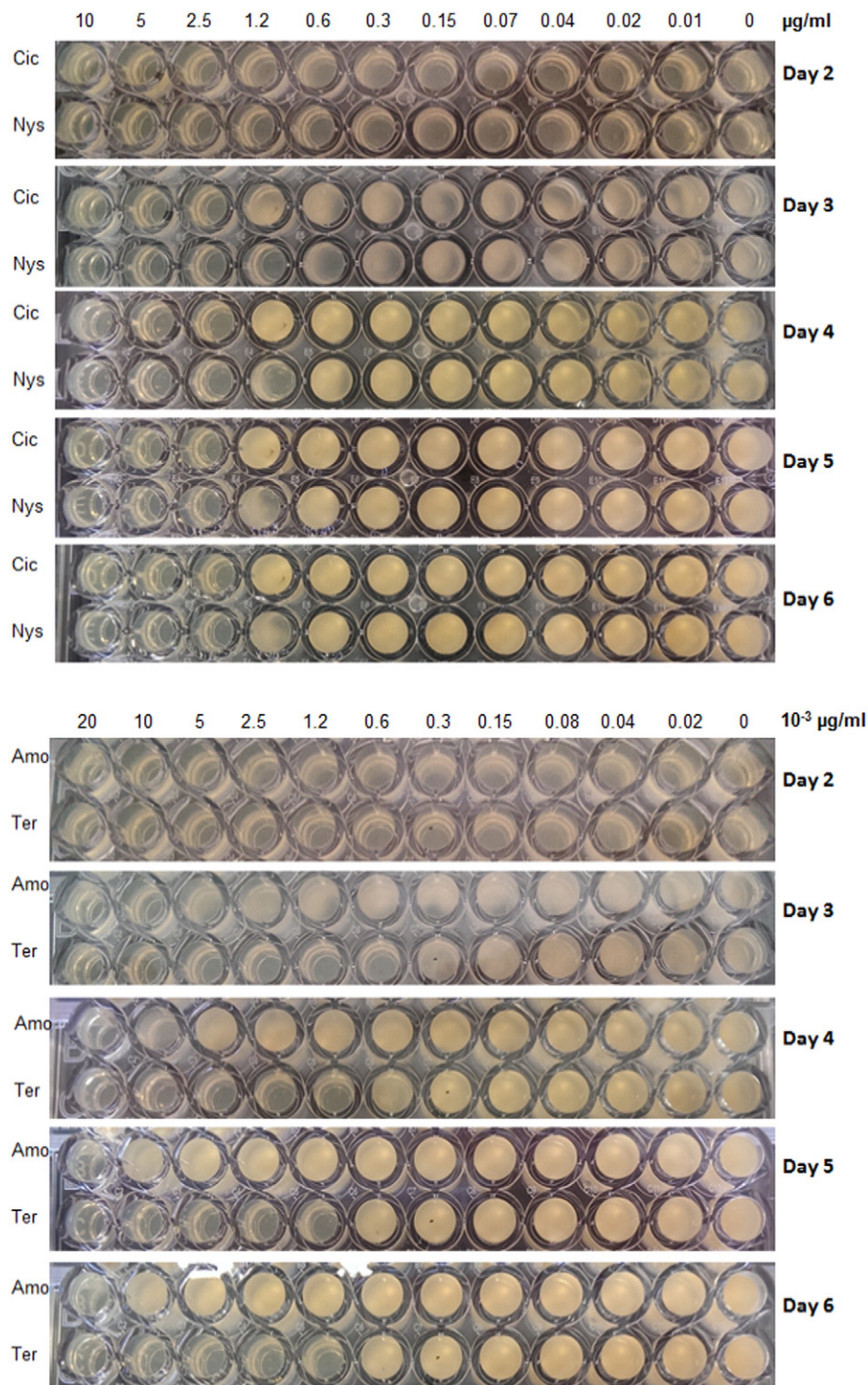


Fig. 2. Susceptibility of *T. rubrum* to antifungal drugs using the 96-well plate SPOTi assay. The assay was performed in triplicates. Cic, ciclopirox; Nys, nystatin; Amo, amorolfine; Ter, terbinafine.

Table 3

ZOIs of ciclopirox, nystatin, amorolfine and terbinafine against *T. rubrum*. Mean \pm standard deviation are shown ($n = 3$).

Antifungal agents	Zone of inhibition (cm)
Ciclopirox	5.39 ± 0.31
Nystatin	2.65 ± 0.37
Amorolfine	9.00 ± 0.00
Terbinafine	7.95 ± 0.32

2007/2008). The drug load per disc, hence the amounts available for drug diffusion into the agar gel were 50 μg for ciclopirox olamine and nystatin, and 5 μg for terbinafine and amorolfine. 100 μl of *T. rubrum* inocula was spread on the surface of SDA plates then the diffusion discs were placed in the centre of the plates. The plates were then incubated at 32 $^{\circ}\text{C}$ for 6 days. The diameters of ZOI were measured, using Image J software. Results were averaged for 3 plates for each antifungal agent.

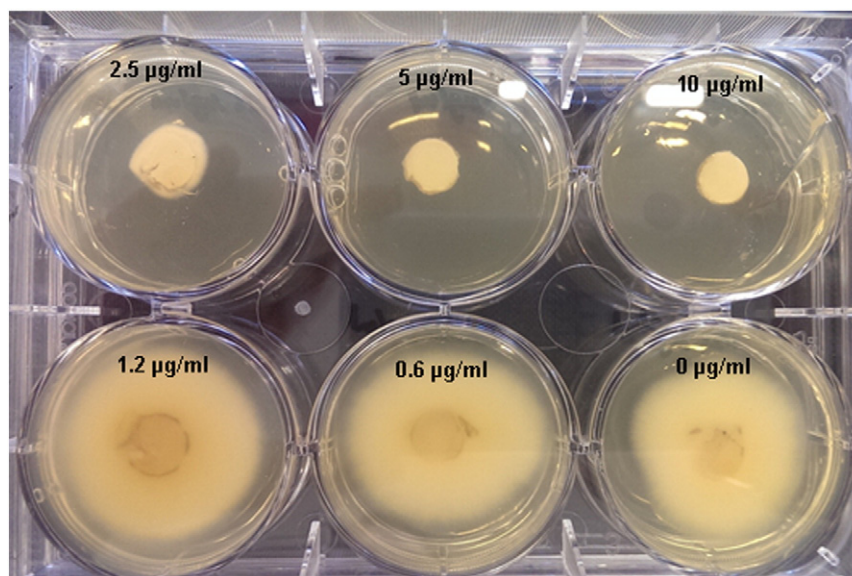


Fig. 3. Susceptibility of *T. rubrum* to ciclopirox olamine using mycelial plugs in 6-well plates.

2.3.4. Validation of the SPOTi assay using mycelial plugs

Using ciclopirox as a model drug, the SPOTi assay described above in 96-well plates was validated using a 6-well plate and mycelial plugs of 5 mm diameter. Drug solutions were diluted in 5 ml SDA in each well to produce drug concentrations of: 10, 5, 2.5, 1.2 and 0.6 $\mu\text{g}/\text{mL}$. In the last well, the pure solvent (DMSO) was used to ensure that it does not negatively affect fungal growth. *T. rubrum* mycelial plugs of 0.6 mm diameter were then placed in the centre of each well and incubated at 32 °C. This experiment was performed in triplicates. An additional 6-well plate was used as a growth control where no antifungals or solvents were mixed with SDA.

3. Results and discussion

3.1. MICs determined by SPOTi, ZOI and mycelial plugs

With the modified SPOTi assay, dermatophyte growth was assessed over a 6 day period after inoculation the MICs (Table 2.) of the antifungal drugs could be reliably determined at day 3, as shown by photographic images shown in Fig. 2 for *T. rubrum*. Compared to the disc-diffusion and broth micro-dilution methods, where a 7 day incubation period is recommended (Norris et al., 1999), SPOTi offers a significant advantage and ease of performance.

Encouragingly, the results from the SPOTi, quantitative in nature, agreed with the qualitative ones obtained from the disc-diffusion tests (a qualitative analysis) that were performed alongside to validate the former approach, i.e. a higher potency of amorolfine and terbinafine which gave larger ZOI at lower concentrations, compared to ciclopirox and nystatin (Table 3).

Taking ciclopirox as a model drug, the SPOTi assay was compared with the traditional method of inoculation using mycelial plugs (Wright et al., 1983) (Fig. 3), where the MIC of ciclopirox was found to be 2.5 $\mu\text{g}/\text{mL}$ (Fig. 3). This is the same as that determined with SPOTi (Table 2) and establishes the fact that, in spite of using only the conidia of the fungi in the SPOTi assay, the MIC determination by SPOTi is representative of all forms of the pathogen-hyphae and spores, thereby displaying the robustness and reproducibility of the assay.

In the absence of a dermatophytes' susceptibility assessment standard, disc-diffusion method is still widely used primarily due to its convenience. The major critique of this method arises from the fact that when a disc containing a given amount of an antifungal is placed on

an agar plate, there follows a slow release of the drug out of the disc, into the agar, followed by its diffusion within the agar over time. This is likely to form a concentration gradient with higher drug concentrations found near the drug-loaded disc. Thus dermatophytes further away from the disc will be exposed to a lower drug concentration. Furthermore, the interpretation of zone of inhibition (ZOI) can often be subjective, with definitions of measured areas ranging from "area of no growth" to "areas up to normal growth". Finally, ZOI does not allow the direct comparison of various agents in terms of MIC; and is only qualitative even when areas of inhibitions are reported, as these values cannot be used to determine MIC. For example, an area of more than 20 mm is indicative of susceptibility when testing terbinafine but only an area of 15 mm is indicative of susceptibility with nystatin (Rosco Diagnostica Ltd, 2007/2008). Nonetheless, comparison of this method with the broth micro-dilution assay has resulted in favourable correlation with some antifungal agents, where an increase of MIC is accompanied by a decrease of inhibition zone diameters (National Committee for Clinical Laboratory Standards, 1997; National Committee for Clinical Laboratory Standards, 2002), as is also seen through this work (Tables 2 and 3).

With the SPOTi assay, more quantitative results are obtained through direct observation. Drug concentrations resulting in 'no fungal growth' are identified as the MIC. The main advantage of this assay, compared to the disc diffusion method, is that the concentration of the drugs that fungal colonies are exposed to is uniform in each well as the drug is mixed thoroughly in the agar. Hence, SPOTi allows for quantitative measurement of MICs.

4. Conclusions

We have shown SPOTi to be a rapid, reliable and quantitative susceptibility assay for dermatophytes. The encouraging results attest to the use of this assay on other fungi, for antifungal susceptibility testing as well as for the screening of novel antifungal agents and it may prove to be a gold-standard susceptibility assay for fungi. SPOTi has already proven to be a success story in the identification of antimycobacterials from varied sources such as natural, synthetic and has recently been instrumental in repurposing drugs (Guzman et al., 2013). We expect this success to translate effortlessly when using fungi, and endeavour to adapt this convenient, effective technique for other types of fungal pathogens.

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Competing financial interests

The authors declare no competing financial interests.

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