Dear Sirs,

We read with great interest the paper by Strassburger-Krogias and colleagues showing a significant effect of dimethyl fumarate (DMF) on disability progression in progressive multiple sclerosis (MS) patients.(1) They enrolled 26 patients, 12 affected by primary progressive MS and 14 with secondary progressive MS of whom 18 were treated with Fumaderm (Biogen Idec GmbH, Ismaning, Germany) and 8 on pharmacy prepared DMF. During a total mean follow-up period of 13.2 ± 7.5 months, 57.7% of patients did not experience any disability progression and 19.2% of them showed an improvement of their disability measured with Expanded Disability Status Score (EDSS), with a overall favorable safety profile. There was no significant difference between the Fumaderm and DMF.

DMF is one of the new oral drugs available for the treatment of relapsing MS. The exact mechanism of action of DMF is still under research and seems to involve the modulation of the immune response, the interference with the intracellular redox balance and possibly also an important effect on mitochondria, by inhibition of the mitochondrial respiratory complex II succinate dehydrogenase whose reaction product is precisely fumarate, and by activating the human mitochondrial NAD(P)+- dependent malic enzyme.(2,3)

Following the encouraging results on RRMS patients (4,5), and considering the multifaceted effects of DMF, a phase 3 study was planned and to be commenced in secondary progressive MS (SPMS) patients (Biogen INSPIRE study NCT02430532). However, despite the recent negative results from the natalizumab phase 3 study in SPMS, [ASCEND, (NCT01416181)], the decision made by Biogen to close the INSPIRE study seems at least disappointing, especially as the mechanisms of action of natalizumab and DMF are very different.

Although we agree that neurodegeneration, rather than inflammation, should be the primary therapeutic target in progressive MS, DMF has important effects on the immune response that may be relevant in progressive MS. Longbrake and colleagues (6) recently analyzed and characterized circulating blood leukocytes in 41 stable MS patients treated with DMF and found that circulating CD8+ and CD4+ T cells, CD56dim natural killer (NK) cells, CD19+ B cells and plasmacytoid dendritic cells were lower in the lymphopenic MS patients compared to either control, with no changes of CD56hi NK cells, monocytes or myeloid dendritic cells. Whether lymphopenic or not, within the CD4+ and CD8+ subsets, they found a reduction of memory cells and a relative expansion of naïve cells.

Relevant to DMF, there are two potential mechanisms pertinent to progressive MS. Firstly, the percentage of circulating CD56dim NK lymphocytes expressing perforin, a pore forming cytolytic protein found in the granules of cytotoxic cells, has been demonstrated to be increased in both PPMS and SPMS.(7) CD56dim cytotoxic NK cells have been hypothesized to have a role in the progression of the disease and, interestingly, also CD8+ T cells expressing perforin, representing another cytotoxic lymphocyte subset, have been proposed to play a role in disability progression both in Theiler's murine encephalomyelitis virus model of MS (8,9) and in human studies.(10,11) The reduction in CD56dim cytotoxic NK cells, along with CD4+ and CD8+ memory T cells, crucial in the MS pathophysiology,(12–14) in patients on DMF (6) is therefore relevant.

Secondly, DMF may facilitate mitochondrial function, that pathological studies show to be altered in progressive MS,(15) with alterations of ATP synthesis, permeability transition pore opening, release of proapoptotic factors, electron transport chain and ionic homeostasis dysfunction.(15) For all of these reasons, we would support on-going testing of DMF in SPMS.

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References

- Strassburger-Krogias K, Ellrichmann G, Krogias C, Altmeyer P, Chan a., Gold R. Fumarate treatment in progressive forms of multiple sclerosis: first results of a single-center observational study. Ther Adv Neurol Disord [Internet].
 2014;232–8. Available from: http://tan.sagepub.com/cgi/doi/10.1177/1756285614544466
- Ruggieri S, Tortorella C, Gasperini C. Pharmacology and clinical efficacy of dimethyl fumarate (BG-12) for treatment of relapsing-remitting multiple sclerosis. Ther Clin Risk Manag [Internet]. 2014;10:229–39. Available from: http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3972027&tool=pmce ntrez&rendertype=abstract
- Scannevin RH, Chollate S, Jung M -y., Shackett M, Patel H, Bista P, et al. Fumarates Promote Cytoprotection of Central Nervous System Cells against Oxidative Stress via the Nuclear Factor (Erythroid-Derived 2)-Like 2 Pathway. J Pharmacol Exp Ther [Internet]. 2012;341(1):274–84. Available from: http://www.ncbi.nlm.nih.gov/pubmed/22267202\nhttp://jpet.aspetjournals.org/co ntent/341/1/274.full.pdf\nhttp://jpet.aspetjournals.org/cgi/doi/10.1124/jpet.111.1 90132
- Fox RJ, Miller DH, Phillips JT, Hutchinson M, Havrdova E, Kita M, et al. Placebo-Controlled Phase 3 Study of Oral BG-12 or Glatiramer in Multiple Sclerosis. N Engl J Med. 2012;367(12):1087–97.

- Gold R, Kappos L, Arnold DL, Bar-Or A, Giovannoni G, Selmaj K, et al. Placebo-controlled phase 3 study of oral BG-12 for relapsing multiple sclerosis. N Engl J Med [Internet]. 2012;367(12):1098–107. Available from: http://www.ncbi.nlm.nih.gov/pubmed/22992073
- Longbrake EE, Ramsbottom MJ, Cantoni C, Ghezzi L, Cross AH, Piccio L. Dimethyl fumarate selectively reduces memory T cells in multiple sclerosis patients. Mult Scler J [Internet]. 2015;1–10. Available from: http://msj.sagepub.com/cgi/doi/10.1177/1352458515608961
- Plantone D, Marti A, Frisullo G, Iorio R, Damato V, Nociti V, et al. Circulating CD56dim NK cells expressing perforin are increased in progressive multiple sclerosis. J Neuroimmunol [Internet]. Elsevier B.V.; 2013;265(1-2):124–7. Available from: http://dx.doi.org/10.1016/j.jneuroim.2013.10.004
- Deb C, LaFrance-Corey RG, Schmalstieg WF, Sauer BM, Wang H, German CL, et al. CD8+ T Cells Cause Disability and Axon Loss in a Mouse Model of Multiple Sclerosis. Kleinschnitz C, editor. PLoS One [Internet]. 2010 Aug 30 [cited 2015 Oct 22];5(8):e12478. Available from: http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=2930011&tool=pmce ntrez&rendertype=abstract
- 9. Murray PD, McGavern DB, Lin X, Njenga MK, Leibowitz J, Pease LR, et al.

Perforin-dependent neurologic injury in a viral model of multiple sclerosis. J Neurosci [Internet]. 1998;18(18):7306–14. Available from: http://www.ncbi.nlm.nih.gov/pubmed/9736651

- Giovanni F, Domenico P, Alessandro M, Raffaele I, Viviana N, Katia PA, et al. Circulating CD8+CD56-perforin+ T cells are increased in multiple sclerosis patients. J Neuroimmunol [Internet]. Elsevier B.V.; 2011;240-241:137–41. Available from: http://dx.doi.org/10.1016/j.jneuroim.2011.09.002
- Denic A, Wootla B, Rodriguez M. CD8+ T cells in Multiple Sclerosis. Expert Opin Ther Targets. 2013;17(9):1053–66.
- Bielekova B, Sung M-H, Kadom N, Simon R, McFarland H, Martin R.
 Expansion and Functional Relevance of High-Avidity Myelin-Specific CD4+ T
 Cells in Multiple Sclerosis. J Immunol [Internet]. 2004;172(6):3893–904.
 Available from: http://www.jimmunol.org/cgi/doi/10.4049/jimmunol.172.6.3893
- Crawford MP, Yan SX, Ortega SB, Mehta RS, Hewitt RE, Price D a, et al. High prevalence of autoreactive, neuroantigen-specific CD8+ T cells in multiple sclerosis revealed by novel flow cytometric assay. Blood [Internet].
 2004;103(11):4222–31. Available from: http://www.ncbi.nlm.nih.gov/pubmed/14976054
- 14. Okuda Y, Okuda M, Apatoff BR, Posnett DN. The activation of memory CD4(+)

T cells and CD8(+) T cells in patients with multiple sclerosis. J Neurol Sci [Internet]. 2005;235(1-2):11–7. Available from: http://www.ncbi.nlm.nih.gov/pubmed/15972217

 Su KG, Banker G, Bourdette D, Forte M. Axonal degeneration in multiple sclerosis: The mitochondrial hypothesis. Curr Neurol Neurosci Rep. 2009;9(5):411–7.