

Neurosurgery Focus Issue: – Traumatic Spinal Cord Injury Repair and Regeneration

Manuscript word count (not including table/references): 4244

References: 130

Figures: 4

Tables: 2

Authors: Christopher S. Ahuja^{1,3,5}, Satoshi Nori⁵, Lindsay Tetreault³, Jefferson Wilson^{1,3,4}, Brian Kwon^{6,7}, James Harrop⁸, David Choi⁹, Michael G. Fehlings¹⁻⁵

Toronto Western Hospital, West Wing 4th floor

399 Bathurst Street, Toronto, ON, M5T 2S8

Tel: 416-603-5675

Fax: 416-603-5298

Email: Michael.Fehlings@uhn.ca

Author Affiliations:

1. Division of Neurosurgery, Department of Surgery, University of Toronto, Toronto, Canada
2. Institute of Medical Science, University of Toronto, Toronto, Canada
3. Department of Surgery, University of Toronto, Toronto, Canada
4. Spine Program, University of Toronto, Toronto, Canada
5. Department of Genetics and Development, University of Toronto, Toronto, Canada
6. Vancouver Spine Institute, Vancouver General Hospital, Vancouver, Canada
7. Department of Surgery, University of British Columbia, Vancouver, Canada
8. Thomas Jefferson University Hospital, Philadelphia, PA
9. National Hospital for Neurology and Neurosurgery, University College London, London, England

Disclosures: Funded by AOSpine North America

KEYWORDS

Spinal cord injury, trauma, regenerative medicine, stem cells, neuroprotection, management, clinical trial

Introduction

Traumatic spinal cord injuries (SCI) have devastating physical, psychosocial, and vocational implications for patients and caregivers. Direct lifetime costs can reach a staggering \$1.1-4.6 million per patient with over 1 million people affected in North America alone (Figure 1)¹⁻³. For treating physicians, a working knowledge of current and emerging therapies in SCI is critical to expediently deliver care and improve long-term functional outcomes for patients^{4, 5}. This article summarizes the evidence-based management of a patient with acute SCI and discusses upcoming neuroprotective and neuroregenerative strategies on the cusp of translation. A primer on the unique pathophysiology of SCI is provided to aid in understanding the rationale behind the diverse range of therapeutic approaches discussed below.

Pathophysiology

Primary and secondary spinal cord injury

SCI can be divided into primary and secondary phases.^{6, 7} The primary SCI is caused by the physical forces of the initial traumatic event and is often the most important determinant of injury severity; physical forces involved can include compression, shearing, laceration and acute stretch/distraction.⁸ After the primary injury event a cascade of secondary injury events is initiated which serves to expand the zone of neural tissue injury and exacerbate neurological deficits.^{9, 10} Secondary SCI is a delayed and progressive tissue injury following the primary SCI. During this secondary injury cascade, inflammatory cells such as macrophages, microglia, T-cells and neutrophils infiltrate the injury site as a result of disruption of the blood-spinal cord barrier. These cells trigger the release of inflammatory cytokines such as tumor necrosis factor (TNF) α , interleukin (IL)-1 α , IL-1 β and IL-6, with levels of these cytokines peaking 6-12 hours after injury and remaining elevated up to 4 days

after injury¹¹. In addition, a loss of ionic homeostasis after SCI causes increased intracellular calcium, which activates calcium-dependent proteases and causes mitochondrial dysfunction, ultimately leading to cell death.¹² Notably, oligodendrocytes are susceptible to apoptotic loss. This apoptotic loss has been observed distant from the epicenter of SCI as well as at the lesion epicenter and leads to demyelination of preserved axons.¹³⁻¹⁵ Moreover, phagocytic inflammatory cells release reactive oxygen species (ROS) which causes DNA oxidative damage, protein oxidation and lipid peroxidation. Delayed necrosis and apoptosis are induced by this process.¹⁶⁻¹⁸ After SCI, upregulated release of excitatory amino acids, such as glutamate and aspartate, is observed due to release from disrupted cells.^{19, 20} The excessive activation of excitatory amino acid receptors produces excitotoxicity and further propagation of loss of neurons and glia by both necrotic and apoptotic cell death.²¹

Barriers to regeneration

It is widely recognized that regeneration in the adult mammalian central nervous system (CNS), which includes the spinal cord, is difficult due to limited plasticity and inhibitory factors produced from myelin degradation.²² Although recent progress in the field of SCI research has demonstrated that the CNS has more inherent regenerative capacity than that was once thought,^{23, 24} it does not have the same regenerative capacity that is observed in the peripheral nervous system (PNS). Compared with the PNS, not only is the regenerative capacity of CNS axons lower but it also decreases over time.²⁵

The inhibitory nature of CNS myelin, which is in contrast to PNS myelin, was first recognized in 1985.²⁶ Myelin-associated proteins, including neurite outgrowth inhibitor A (Nogo A),^{27, 28} myelin-associated glycoprotein (MAG)^{29, 30} and oligodendrocyte-myelin glycoprotein (OMgp)³¹, bind Nogo receptors (NgR) that form co-receptor complexes with TNF receptor family proteins such as p75, TROY, and LIGO-1 to activate the GTPase Rho A. Rho-associated protein kinase (ROCK) is the effector of Rho A, which regulates further

downstream effectors, and leads to growth cone collapse of regenerating axons and to neurite retraction.

Hypertrophied astrocytes form a physical barrier called the glial scar, which walls off injured tissue from the healthy tissue.³² The astrocytes also form a chemical barrier by secreting a number of growth inhibitory chondroitin sulfate proteoglycans (CSPGs) including neurocan, versican, brevican, phosphacan and NG2.³³ Fibroblasts also infiltrate the perilesional region and replace the extracellular matrix (ECM) with fibrous connective tissue. This is associated with the deposition of inhibitory ECM molecules which function as chemical barriers to axonal regeneration similar to myelin associated inhibitors (Figure 2).³⁴

Current Management

The current management of SCI largely follows the American Association of Neurological Surgeons (AANS) and Congress of Neurological Surgeons (CNS) joint section guideline series as well as an upcoming AOSpine 2016 guideline (Table 1)³⁵. Initial care in the field prioritizes securing the airway, breathing, and circulation (ABCs) followed by early recognition of SCI and rapid referral to specialized centers in order to expedite delivery of time-sensitive interventions³⁵. To limit further insult to the highly-vulnerable cord, spinal immobilization should be performed for all patients with suspected or confirmed injuries³⁵. This typically involves a rigid cervical collar, backboard for transport, and spinal precautions for patient transfers (e.g. logroll maneuver with inline manual cervical stabilization and a transfer board). Systemic hypotension (SBP <90 mmHg), even for brief periods, should be avoided as it is associated with worse long-term neurological outcomes³⁵. This can be particularly challenging as hypovolemia is common in polytrauma and interruption of spinal cord sympathetic fibers can induce a profound loss of vascular tone and bradycardia (neurogenic shock). Resuscitation with large-volume crystalloids is typical, however, alpha-

agonists (e.g. phenylephrine) or mixed alpha/beta-agonists (e.g. dopamine, norepinephrine) may also be required as adjuncts. Once resuscitated, an American Spinal Injury Association (ASIA) International Standards for Neurological Classification of SCI (ISNCSCI) examination should be documented to establish baseline function and the level of neurological injury (Table 1; Figure 3)³⁵.

Early localization and classification of osseoligamentous and neurological injuries is critical to expediently provide the outcome-altering therapies discussed below^{4, 5, 36, 37}. The management of particular fracture patterns is discussed in detail in ‘Spine Trauma’. CT imaging is recommended for all patients with suspected SCI as x-rays can miss up to 6% of injuries³⁸. When evaluating patients with high-energy mechanisms and confirmed cervical injuries, thoracolumbar imaging is recommended to rule out concomitant injuries that may not be clinically apparent³⁹. The role of MRI in the initial workup of patients remains unclear, however, urgent MRI is strongly recommended by the senior authors in all cases with unexplained neurological deficits to rule out ongoing spinal cord compression due to occult ligamentous injuries, epidural hematomas, or critical disc herniations. The utility of MRI in prognostication is also becoming more apparent as validated prediction scores continue to be published⁴⁰.

Concurrent with the diagnostic workup, patients should be transferred to a critical care unit providing continuous respiratory, cardiac, and hemodynamic monitoring³⁵. Immediate life- or limb-threatening injuries should be managed by the appropriate teams while maintaining strict spinal immobilization. Delivering effective care in SCI requires a collaborative multidisciplinary approach including fiberoptic intubation by anesthesia/critical care, modified intraoperative positions for general surgery/orthopedic procedures and early recognition of therapeutic windows which can positively alter long-term outcomes.

Early Surgical Decompression

After SCI, ongoing mechanical compression of the spinal cord can impair blood flow causing ischemia and an expanded zone of neural tissue injury. The goal of early surgical decompression after SCI is to relieve this compression thereby improving the vascular supply to the injured area and limiting the zone of secondary injury expansion. A sizable body of preclinical literature supports the positive effects of early surgical decompression on behavioral and pathological outcomes in animal SCI models⁴¹.

With respect to clinical evidence on this topic, a number of comparative cohort studies have been published investigating the clinical impact of performing decompressive surgery prior to several thresholds. Notably, to investigate the efficacy of early decompression prior to a 24 hour threshold, The Surgical Treatment of Acute Spinal Cord Injury Study (STASCIS) prospectively enrolled 313 cervical SCI patients.³⁷ Patients receiving early decompression (<24 hours after SCI) experienced 2.8 times greater odds of experiencing an AIS improvement of at least 2-grades at 6 months as compared with patients who underwent late decompression (\geq 24 hours after SCI). Although not statistically significant, there was a trend to a reduced incidence of acute in-hospital complications after early decompression. A prospective Canadian cohort study (including cervical, thoracic and lumbar SCI, n=84) confirmed the findings observed in STASCIS, reporting that in the adjusted analysis early decompression was associated with a statistically greater improvement in ASIA Motor Score recovery at the time of rehabilitation facility discharge.⁴² Moreover, an observational Canadian cohort study showed that AIS A (complete injury) and AIS B (complete motor injury with incomplete sensory injury) patients who received early decompression experienced shorter length of hospital stay, while AIS B, C and D incomplete injury patients decompressed in an early fashion demonstrated an additional 6.3 points in motor recovery as compared to those decompressed late.⁴ Taken together, these findings support the concept of 'Time is Spine', emphasizing the importance of early diagnosis and

intervention to enhance long-term outcomes.

Central cord injury is the most common form of incomplete SCI. Historically, early decompression by durotomy and sectioning of the dentate ligaments has been avoided in cases of central cord injury due findings of poor outcomes after surgery.⁴³ However, an analysis of prospective data performed by the Spine Trauma Study Group associated early decompression (<24 hours after SCI) with an additional 6.3 points of ASIA motor score recovery and 2.8 times odds of ASIA Impairment Scale grade improvement at 12-month follow-up as compared to late decompression (\geq 24 hours after SCI).⁴⁴ In 2013, a randomized control trial Comparing Surgical Decompression Versus Conservative Treatment in Incomplete Spinal Cord Injury (COSMIC; NCT01367405; n=72) trial was initiated by Raboud University. The study will compare surgical decompression within 24 hours to normal conservative treatment without surgery and is currently recruiting participants.

Steroids for SCI

Methylprednisolone (MPSS) is a potent synthetic glucocorticoid which upregulates anti-inflammatory cytokine release and reduces oxidative stress to enhance neural cell survival in preclinical models of traumatic SCI. The National Acute Spinal Cord Injury Study (NASCIS) trial series (1990⁴⁵, 1997⁴⁶) found an increase in the number of infection-related complication (e.g. severe sepsis, severe pneumonia) with the high-dose 48-hour protocol which outweighed the potential neurological benefits. However, a shorter 24-hour course of IV MPSS (30mg/kg bolus + 5.4mg/kg/hr x 23hrs) had a substantially lower complication rate and, when administered to a subgroup of patients within 8 hours of injury, was still found to improve neurological outcomes. These subgroup analyses and the purported methodology have been a source of controversy for the last three decades. To definitively address the debate, a 2012 Cochrane Review meta-analysis pooling 6 key randomized trials and observational studies was completed. The study found that patients receiving MPSS within 8 hours of injury had a 4-

point greater ASIA motor score improvement⁴⁷. This modest benefit can have tremendous functional implications for patients when those motor points are recovered in key myotomes such as grip and deltoid function. As a result, an upcoming AOSpine 2016 guideline developed by an international expert panel will suggest 24 hours of IV MPSS be administered within 8 hours of injury to patients without significant medical contraindication.

Blood Pressure Augmentation

Vascular injury and localized edema contribute to ongoing ischemia in the perilesional region. Blood pressure augmentation has emerged as a viable strategy to neuroprotect at-risk tissue by enhancing perfusion. Current AANS/CNS guidelines recommend maintenance of mean arterial pressure (MAP) \geq 85-90mmHg for 7 days post-injury as this has been found to enhance long-term AIS grade outcomes for patients⁵. In application, this most often necessitates invasive blood pressure monitoring, IV fluid therapy and central venous access for continuous infusion of vasopressors. These substantial requirements have prompted a non-inferiority trial entitled Mean Arterial blood Pressure Treatment for Acute Spinal Cord Injury (MAPS; NCT02232165) comparing MAP \geq 85mmHg vs MAP \geq 65mmHg with results expected by 2017³.

These requirements can also be a significant hindrance to early mobilization, an important component of cardiorespiratory and dermatologic complication prevention. A collaborative interdisciplinary approach utilizing adjunctive measures such as prophylactic vasopressors, abdominal binding, and assistive devices is often required to safely elevate patients. The precise timing of mobilization is dictated by the patient's hemodynamic status and the expertise of the treating team.

Key Trials in Neuroprotection

In addition to early decompression, MAP augmentation and IV MPSS; several other

neuroprotective strategies targeting key components of the secondary injury cascade have emerged in preclinical research. The most promising therapies currently being translated are discussed in this section.

Pharmacological Therapies

Riluzole

Riluzole is a benzothiazole sodium channel blocker currently approved by the FDA, EMA, and Health Canada for the treatment of amyotrophic lateral sclerosis (ALS)^{48,49}. It protects against excitotoxic cell death by blocking sodium influx in injured neurons and restricting the pre-synaptic release of glutamate⁵⁰. Animal studies in SCI have demonstrated its ability to reduce neuronal loss and cavity size while improving sensorimotor and electrophysiological outcomes⁵¹⁻⁵⁴. A collaborative effort to study Riluzole for SCI is being led by the senior author (MGF) and includes the North American Clinical Trials Network (NACTN), AOSpine, the Ontario Neurotrauma Foundation (ONF) and the Rick Hansen Institute. This Phase II/III RCT (N=351) entitled “Riluzole in Spinal Cord Injury Study” (RISCIS; NCT01597518) is currently recruiting patients with acute C4-8 ASIA grade A/B/C injuries and will assess multiple outcomes including the ASIA Impairment Scale (AIS), Brief Pain Inventory (BPI), and Spinal Cord Independence Measure (SCIM)³. The study is expected to conclude in 2018.

Magnesium

Magnesium can act as an NMDA receptor antagonist to decrease excitotoxicity and also functions as an anti-inflammatory agent. Stable CSF levels can be generated by delivering magnesium with an excipient such as polyethylene glycol (PEG)⁵⁵⁻⁵⁷. In animal models, the Mg-PEG combination has been shown to enhance tissue sparing and lead to

behavioral recovery^{58, 59}. A Phase I trial (N=15; NCT01750684) of a Mg-PEG combination (AC105) led by Acorda Therapeutics Inc. concluded in February 2015 with results pending report³.

Minocycline

Minocycline is a second-generation bacteriostatic tetracycline antibiotic that has demonstrated neuroprotective properties in preclinical models of CNS disorders including Huntington's disease and multiple sclerosis^{60, 61}. This stems in part from its significant anti-inflammatory effect mediated by inhibition of microglial activation, interleukin-1 β (IL-1 β), tumour-necrosis factor- α (TNF- α), cyclooxygenase-2 (COX-2), and matrix metalloproteinases⁶²⁻⁶⁵. In animal studies, minocycline treatment after acute SCI has been shown to reduce lesion size and promote tissue sparing^{66, 67}. A Phase II trial demonstrated that patients with acute incomplete cervical SCI (N=25) may benefit from early minocycline administration as they found a 14-point ASIA motor score improvement compared to placebo (p=0.05)⁶⁸. This exciting result led to the development of a Phase III trial (N=248) entitled 'Minocycline in Acute Spinal Cord Injury' (MASC, NCT01828203) which will assess IV minocycline for 7 days versus placebo and is expected to report by 2018³.

GM-1 Ganglioside

Monosialotetrahexosylganglioside (GM-1) is a glycosphingolipid found in cell membranes with the ability to activate receptor tyrosine kinases to enhance neural plasticity and regeneration. It has been successfully used for neuroprotection in animal models of SCI where it enhanced tissue sparing.⁶⁹ A successful Phase II trial (N=37) found improved 1-year ASIA motor scores for those receiving daily GM-1 for 18 to 32 days post-injury⁷⁰. Unfortunately, a follow-up Phase III RCT (N=797) found no statistically significant

improvement with treatment⁷¹. No further studies have been registered.

Fibroblast Growth Factor

Fibroblast growth factor (FGF) is a heparin-binding protein found to be neuroprotective against excitotoxicity while also reducing oxygen free radical generation⁷². In animal models, it has been shown to reduce motor neuron loss and improve respiratory deficits^{73, 74}. A Phase I/II trial (N=62; NCT01502631) of the FGF-analogue, SUN13837 (Asubio Pharmaceuticals Inc.), completed in 2015 with results pending publication³.

G-CSF

Granulocyte colony-stimulating factor (G-CSF; CSF 3) is a cytokine glycoprotein found in numerous tissues throughout the body. It is capable of promoting cell proliferation, survival, and mobilization. In the CNS, it has been shown to facilitate survival of ischemic cells and reduce inflammatory cytokine expression (e.g. TNF- α , IL-1 β)⁷⁵⁻⁷⁷. A recent pair of non-randomized Phase I/IIa trials showed no increase in serious adverse events with G-CSF administration while also demonstrating improvement in AIS outcomes^{78, 79}. Additional well-designed RCTs will be required to establish the efficacy of G-CSF for SCI.

Hepatocyte Growth Factor

Hepatocyte growth factor (HGF) is a pro-survival, pro-motility c-Met receptor ligand. In small animal SCI models, HGF increases neuron survival and decreases oligodendrocyte apoptosis resulting in improved behavioral outcomes⁸⁰⁻⁸². More recently, HGF has been shown to promote angiogenesis and enhance upper limb recovery in a primate model of cervical SCI⁸¹. A Phase I/II randomized trial (N=48; NCT02193334) of KP-100IT (HGF; Kringle Pharma Inc.) is now underway with results expected in 2017³.

Non-Pharmacologic Therapies

Therapeutic Hypothermia

Therapeutic hypothermia (TH; 32-34°C) significantly reduces the basal metabolic rate of the CNS and decreases inflammatory cell activation⁸³. It has been successfully applied in neonatal hypoxic-ischemic encephalopathy and after in-hospital cardiac arrest⁸⁴⁻⁸⁶. In preclinical SCI models, it has been shown to enhance tissue sparing and promote behavioral recovery prompting a pilot study (N=14) of early systemic TH for patient with AIS A injuries which found no increase in complication rates and a trend towards increased neurologic recovery (43% vs 21%)^{87, 88}. A Phase II/III trial entitled Acute Rapid Cooling Therapy for Injuries of the Spinal Cord (ARCTIC) has been planned to definitively assess efficacy.

CSF Drainage

CSF drainage attempts to prevent cord hypoperfusion in the critical post-injury period by relieving pressure analogous to EVD drainage for raised ICP. A Phase I/II trial (N=22) completed in 2009 found no significant improvement outcomes with drainage, however, the study was not sufficiently powered to demonstrate efficacy⁸⁹. Recent large-animal trials have found CSF drainage acts synergistically with MAP augmentation to improve cord blood flow⁹⁰. Based on these key results, a Phase IIB trial (N=60; NCT02495545) of MAP elevation with CSF drainage has been launched with results expected by December 2017³.

Key Trials in Neuroregeneration

While timely neuroprotective interventions can have tremendous benefits in the acute injury period, the majority of our patients are in the chronic phase of their injuries where further

recovery is limited. This section discusses emerging neuroregenerative therapies in clinical trial or on the cusp of translation (illustrated in Figure 4).

Pharmacological Therapies

Rho-Rock Inhibitor

Components of the injured adult CNS including CSPGs, myelin-associated glycoproteins (MAG), and neurite outgrowth inhibitor (NOGO) potently inhibit axon outgrowth and attempts at regeneration via the Rho-ROCK signaling pathway. Cethrin/VX-210 (Vertex Pharmaceuticals) is a direct Rho inhibitor applied intraoperatively within a fibrin glue sealant to the epidural space⁹¹. A mixed open-label Phase I/IIa trial (N=48; NCT00500812) of patients with cervical or thoracic injuries found no increase in serious adverse events and a significant improvement in long-term motor scores (18.5 ASIA points) for cervical patients⁹². These very exciting results have led to a Phase III trial in patients with acute cervical SCI planned to begin in 2016.

Anti-NOGO antibody

Anti-NOGO is a monoclonal antibody against NOGO-A, a major inhibitor component of adult CNS myelin. Anti-NOGO treatment delivered by intrathecal injection has been shown to promote axonal sprouting and functional recovery in animal models by clearing the source of this inhibitory signaling⁹³. A Phase I trial (N=51; NCT004060160) of humanized anti-NOGO antibody (ATI-355; Novartis) has been completed in Europe with results pending dissemination³.

Cell Therapies

Cell-based regenerative therapies are an exciting field as transplanted cells are capable

of filling many roles including providing trophic support, modulating the inflammatory response, regenerating lost neural circuits, and remyelinating denuded axons⁹⁴⁻⁹⁶. Early research utilized embryonic stem cells (ESCs), however, ethical concerns and limited supplies have driven the field towards induced pluripotent stem cell (iPSCs) which can be derived from any somatic cell, including autologous sources⁹⁷. While unanticipated challenges have arisen, including early senescence and retained epigenetic memory, iPSCs remain a key therapeutic approach moving forward. Numerous animal studies over the last three decades have demonstrated the beneficial effects of a range of transplanted cell types. The most clinically-relevant approaches are discussed here.

Neural stem/precursor cells

Neural precursor cells are capable of differentiating to CNS-specific neurons, oligodendrocytes and astrocytes making them a particularly promising strategy. In animal studies, they are capable of integrating with host circuits to enhance behavioral recovery over several weeks^{98, 99}. A pair of phase II trials by Stem Cells Inc. of human CNS stem cell transplants for cervical (N=31; NCT02163876) and thoracic (N=12; NCT01321333) injury were terminated in 2016 prior to completion. While results regarding sensorimotor outcomes are pending dissemination, preliminary data suggest no increase in complications rates related to the treatments³. This provides confirmation of existing safety data that intraparenchymal stem cell transplants are feasible and suggests further optimization of the cells and/or their environment is required to produce meaningful changes in functional recovery.

Mesenchymal stem cells

Mesenchymal stem cells (MSCs) are multipotent cells capable of repairing connective tissues by differentiating to myocytes, osteoblasts, chondrocytes and adipocytes¹⁰⁰. They can

also modulate local and systemic inflammation which has been exploited in animal models of SCI where MSC treatment led to a decrease in peripheral inflammatory cell infiltration and an increase in parenchymal tissue volume¹⁰¹⁻¹⁰⁵. A Phase II/III RCT (N=32; NCT01676441) by Pharmicell Co. studying intraparenchymal and intrathecal MSC treatment for patients with acute AIS B injuries is ongoing with results expected in 2016³.

Schwann cells

The robust regeneration seen in the PNS is thought to be mediated in large part by Schwann cells (SCs). In animal models of SCI, peripheral SCs transplanted into the CNS were found to remyelinate axons, reduce cystic cavitation and enhance recovery¹⁰⁶. An open-label Phase I trial (N=10; NCT02354625) by the Miami Project to Cure Paralysis is now investigating SCs in the treatment of patients with chronic AIS A, B, and C cervical or thoracic injuries with results expected by 2018³. The group is also conducting a Phase I trial (N=10; NCT01739023) of autologous SCs for the treatment of subacute thoracic AIS A injuries with results expected by 2016³.

Olfactory ensheathing cells

Olfactory ensheathing cells (OECs) protect olfactory neurons exposed to the harsh conditions of the nasal mucosa. They rapidly phagocytose debris and microbes while also providing trophic support through growth factor signaling, and physical axonal guidance through guidance cues¹⁰⁷⁻¹¹⁰. OECs harvested from the nasal mucosa and transplanted into the cord have been shown to improve neurite outgrowth and endogenous remyelination resulting in impressive behavioral recovery in animal models¹¹¹. Numerous clinical trials of OECs for chronic SCI have been completed worldwide and analyzed in a meta-analysis (cumulative N=1193) which found no significant increase in complication rates related to the transplant¹¹².

However, human OECs have not been well characterized, and transplants invariably consist of mixtures of cells from the olfactory mucosa or bulb. More data is required regarding human cells, which are not as easy to culture in large numbers from patients compared to animal models. Moving forward, higher quality studies will be required to definitively establish the efficacy of OEC therapy.

Biomaterials

Regeneration is often hindered by the presence of a substantial post-injury cystic cavity which lacks the substrate to support cell migration and axon growth. Biomaterials have emerged as an exciting strategy to fill cavitation defects and reproduce the complex structural architecture of the extracellular matrix¹¹³⁻¹¹⁷. Many of these materials can be engineered to biodegrade over time, release growth factors, and can even be seeded with stem cells to enhance engraftment^{97,98}. Several biomaterials have been shown to be effective in animal models of SCI from the acute to chronic phases (e.g. HAMC¹¹⁴, QL6^{118, 119}, fibrinogen¹²⁰, etc.) As the technology evolves, more niche-specific biomaterials are expected to emerge with extended drug-release and cell support capabilities. Currently, a Phase III trial (N=20; NCT02138110) of InVivo Therapeutics' Neuro-Spinal Scaffold is now recruiting patients with acute AIS A thoracic injuries³. Results are expected in 2017.

Future Directions

The next substantial changes in the management of patients with SCI are likely to be translated from research which adapts to the heterogeneity of SCI. Modified trial designs which specifically target SCI subpopulations are likely to have the greatest impacts on long-term functional recovery. Stratifying patients in this way will require a combination of existing metrics (e.g. clinical exam, radiography) and novel assessment techniques (e.g. advanced imaging, biochemical biomarkers).

While many novel treatments show promise in animal models of SCI, these experimental paradigms typically involve very controlled injury and recovery conditions after biomechanically precise injuries in animals matched for age, weight, gender, species, and, in some cases, genetic background. This obviously pales in comparison to the natural variability that occurs in the acute human SCI setting. The appreciation of the heterogeneity of human SCI is partly the result of the challenges that have been experienced in the execution of clinical trials of novel therapies, particularly in the acute setting. Variability in neurologic recovery requires that many patients be recruited to complete such acute clinical trials in order to be sufficiently powered. Difficulties in achieving such recruitment has plagued the conduct of virtually all acute clinical studies, and the failure to enroll sufficiently large patient cohorts within realistic time frames has resulted in the premature cessation of numerous clinical trial programs. New approaches to overcome this will be needed in the future to facilitate the conduct of such clinical trials and enhance the speed with which novel treatments for SCI can be validated. Such approaches include narrowing the inclusion window to be more specific in the types and severities of cord injuries being studied, and establishing objective biomarkers for the stratification of injury severity and more precise prediction of neurologic outcome.

Seminal large-scale clinical trials for SCI have typically used broad inclusion criteria to bolster recruitment across participating centers. However, post hoc subgroup analyses have now demonstrated that patient characteristics, presentations, and the underlying pathophysiology in SCI can be highly heterogeneous which can influence the relationship between treatments and outcomes^{45-47, 121, 122}. As a result, more recent studies are recruiting carefully defined populations which we feel is key to success. The upcoming Riluzole in Acute Spinal Cord Injury (RISCIS; NCT01597518) trial is an example where recruitment is limited to patients with C4-8 injuries and ASIA grades A, B, or C^{3, 54}. Other clinical initiatives have similarly restricted inclusion both with regards to the level of injury (cervical

vs thoracic), severity of injury (AIS grade A, B, or C), and timing of intervention. While logistically demanding, this careful selection will allow a more valid assessment of the drug's efficacy.

The next generation of trials will also need to further define subpopulations based on quantifiable imaging and biochemical biomarkers. MRI is a key imaging modality for most CNS pathologies, however, its adoption in SCI has been slow. This is likely because the most common sequences (T1- and T2-weighted) rely on gross measurements of hemorrhage and compression providing only modest utility in predicting outcomes. Future MR imaging will need to quantify the cord microstructure to better estimate damage and recovery potential. Emerging techniques for this include diffusion tensor imaging (DTI; axon integrity), myelin water fraction (MWF; myelination), MR spectroscopy (MRS; gliosis or ischemia), and functional MRI (fMRI; connectivity)^{123, 124}.

Biochemical biomarkers are also being extensively explored. The Canadian Multicentre CSF Monitoring and Biomarker Study (CAMPER; NCT01279811) is testing CSF over 5 days for inflammatory cell proteins, interleukins, and other cytokines³. Specific proteins such as IL-6, S100 β , and tau within the cerebrospinal fluid of acute SCI patients have been shown to be able to objectively stratify injury severity and predict AIS grade and motor score improvement^{125, 126}. An additional class of biomarkers currently under study through the Rick Hansen Institute is micro RNAs (miRNA) which are short non-coding RNA segments that can regulate post-transcriptional gene expression. miRNAs are specifically up or down regulated with varying grades of SCI and may hold important prognostic information as they are further understood¹²⁷. Together these biomarkers will yield important data to help identify subgroups within the heterogeneous SCI population, and when combined with clinical examination, will allow patients to be stratified by their specific pathophysiologic niche into targeted trials.

The breadth of therapeutic approaches discussed within this review and the rapidly-evolving management of a patient with SCI highlight the excitement and progress continuing to be made in the field by thousands of collaborating physicians, scientists, and allied health workers worldwide.

Acknowledgements

Thank you to AOSpine for funding this work, Madeleine O'Higgins for copyediting and Chi Lam for organizational support.

References

1. Center NSCIS. Spinal Cord Injury Facts and Figures at a Glance. *The Journal of Spinal Cord Medicine*. 2014/01 2014;37(1):117-118.
2. Foundation CaDR. One degree of separation: paralysis and spinal cord injury in the United States 2010:
http://www.christopherreeve.org/site/c.ddJFKRNoFiG/b.5091685/k.58BD/One_Degree_of_Separation.htm.
3. Clinical Trials.gov. 2016; <https://clinicaltrials.gov/>. Accessed August 4, 2016.
4. Dvorak MF NV, Fallah N, Fisher CG, Finkelstein J, Kwon BK, Rivers CS, Ahn H, Paquet J, Tsai EC, Townson A, Attabib N, Bailey CS, Christie SD, Drew B, Fournery DR, Fox R, Hurlbert RJ, Johnson MG, Linassi AG, Parent S, Fehlings MG. The influence of time from injury to surgery on motor recovery and length of hospital stay in acute traumatic spinal cord injury: an observational Canadian cohort study. *J Neurotrauma*. 2015;32(9):645.
5. Wilson JR, Forgiione N, Fehlings MG. Emerging therapies for acute traumatic spinal cord injury. *CMAJ*. Apr 2 2013;185(6):485-492.
6. Tator CH. Update on the pathophysiology and pathology of acute spinal cord injury. *Brain Pathol*. Oct 1995;5(4):407-413.
7. McDonald JW, Sadowsky C. Spinal-cord injury. *Lancet*. Feb 2 2002;359(9304):417-425.
8. Ackery A, Tator C, Krassioukov A. A global perspective on spinal cord injury epidemiology. *J Neurotrauma*. Oct 2004;21(10):1355-1370.
9. Yip PK, Malaspina A. Spinal cord trauma and the molecular point of no return. *Molecular neurodegeneration*. 2012;7:6.
10. Norenberg MD, Smith J, Marcillo A. The pathology of human spinal cord injury: defining the problems. *J Neurotrauma*. Apr 2004;21(4):429-440.
11. Nakamura M, Houghtling RA, MacArthur L, Bayer BM, Bregman BS. Differences in cytokine gene expression profile between acute and secondary injury in adult rat spinal cord. *Exp Neurol*. Nov 2003;184(1):313-325.
12. Schanne FA, Kane AB, Young EE, Farber JL. Calcium dependence of toxic cell death: a final common pathway. *Science*. Nov 9 1979;206(4419):700-702.
13. Beattie MS, Farooqui AA, Bresnahan JC. Review of current evidence for apoptosis after spinal cord injury. *J Neurotrauma*. Oct 2000;17(10):915-925.
14. Crowe MJ, Bresnahan JC, Shuman SL, Masters JN, Beattie MS. Apoptosis and delayed degeneration after spinal cord injury in rats and monkeys. *Nat. Med*. Jan 1997;3(1):73-76.
15. Dong H, Fazzaro A, Xiang C, Korsmeyer SJ, Jacquin MF, McDonald JW. Enhanced oligodendrocyte survival after spinal cord injury in Bax-deficient mice and mice with delayed Wallerian degeneration. *J Neurosci*. Sep 24 2003;23(25):8682-8691.
16. Waxman SG. Demyelination in spinal cord injury. *Journal of the Neurological Sciences*.

- 1989/06 1989;91(1-2):1-14.
17. Dizdaroglu M, Jaruga P, Birincioglu M, Rodriguez H. Free radical-induced damage to DNA: mechanisms and measurement. *Free Radic. Biol. Med.* Jun 1 2002;32(11):1102-1115.
 18. Hausmann ON. Post-traumatic inflammation following spinal cord injury. *Spinal Cord.* 2003/07 2003;41(7):369-378.
 19. Liu M, Wu W, Li H, et al. Necroptosis, a novel type of programmed cell death, contributes to early neural cells damage after spinal cord injury in adult mice. *J. Spinal Cord Med.* Nov 2015;38(6):745-753.
 20. Wang Y, Wang H, Tao Y, Zhang S, Wang J, Feng X. Necroptosis inhibitor necrostatin-1 promotes cell protection and physiological function in traumatic spinal cord injury. *Neuroscience.* 2014/04 2014;266:91-101.
 21. Li S, Stys PK. Mechanisms of ionotropic glutamate receptor-mediated excitotoxicity in isolated spinal cord white matter. *J Neurosci.* Feb 1 2000;20(3):1190-1198.
 22. Cajal Ry. *Degeneration and Regeneration of the Nervous System.* London: Oxford University Press; 1928.
 23. Barnabe-Heider F, Frisen J. Stem cells for spinal cord repair. *Cell Stem Cell.* Jul 3 2008;3(1):16-24.
 24. Okano H, Yamanaka S. iPS cell technologies: significance and applications to CNS regeneration and disease. *Mol Brain.* 2014;7:22.
 25. Kwon BK, Liu J, Messerer C, et al. Survival and regeneration of rubrospinal neurons 1 year after spinal cord injury. *Proc. Natl. Acad. Sci. U. S. A.* Mar 5 2002;99(5):3246-3251.
 26. Schwab ME, Thoenen H. Dissociated neurons regenerate into sciatic but not optic nerve explants in culture irrespective of neurotrophic factors. *J. Neurosci.* Sep 1985;5(9):2415-2423.
 27. Chen MS, Huber AB, van der Haar ME, et al. Nogo-A is a myelin-associated neurite outgrowth inhibitor and an antigen for monoclonal antibody IN-1. *Nature.* Jan 27 2000;403(6768):434-439.
 28. Freund P, Schmidlin E, Wannier T, et al. Nogo-A-specific antibody treatment enhances sprouting and functional recovery after cervical lesion in adult primates. *Nat Med.* Jul 2006;12(7):790-792.
 29. Cafferty WB, Duffy P, Huebner E, Strittmatter SM. MAG and OMgp synergize with Nogo-A to restrict axonal growth and neurological recovery after spinal cord trauma. *J Neurosci.* May 19 2010;30(20):6825-6837.
 30. DeBellard ME, Tang S, Mukhopadhyay G, Shen YJ, Filbin MT. Myelin-associated glycoprotein inhibits axonal regeneration from a variety of neurons via interaction with a sialoglycoprotein. *Mol. Cell. Neurosci.* Feb 1996;7(2):89-101.
 31. Barton WA, Liu BP, Tzvetkova D, et al. Structure and axon outgrowth inhibitor binding of the Nogo-66 receptor and related proteins. *EMBO J.* Jul 1 2003;22(13):3291-3302.

32. Guha A, Tator CH, Rochon J. Spinal cord blood flow and systemic blood pressure after experimental spinal cord injury in rats. *Stroke*. 1989/03/01 1989;20(3):372-377.
33. McKeon RJ, Schreiber RC, Rudge JS, Silver J. Reduction of neurite outgrowth in a model of glial scarring following CNS injury is correlated with the expression of inhibitory molecules on reactive astrocytes. *J. Neurosci*. Nov 1991;11(11):3398-3411.
34. Brazda N, Muller HW. Pharmacological modification of the extracellular matrix to promote regeneration of the injured brain and spinal cord. *Prog. Brain Res*. 2009;175:269-281.
35. Resnick DK. Updated Guidelines for the Management of Acute Cervical Spine and Spinal Cord Injury. *Neurosurgery*. Mar 2013;72 Suppl 2:1.
36. Wilson JS, A; Craven, C; Verrier, M; Drew, B; Ahn, H; Ford, M; Fehlings, MG. Early versus late surgery for traumatic spinal cord injury: the results of a prospective Canadian cohort study. *Spinal Cord*. 2012;50(11):840.
37. Fehlings MG, Vaccaro A, Wilson JR, et al. Early versus delayed decompression for traumatic cervical spinal cord injury: results of the Surgical Timing in Acute Spinal Cord Injury Study (STASCIS). *PLoS One*. 2012;7(2):e32037.
38. Ryken TC, Hadley MN, Walters BC, et al. Radiographic assessment. *Neurosurgery*. Mar 2013;72 Suppl 2:54-72.
39. Sixta S, Moore FO, Ditillo MF, et al. Screening for thoracolumbar spinal injuries in blunt trauma: an Eastern Association for the Surgery of Trauma practice management guideline. *J Trauma Acute Care Surg*. Nov 2012;73(5 Suppl 4):S326-332.
40. Wilson JR, Grossman RG, Frankowski RF, et al. A clinical prediction model for long-term functional outcome after traumatic spinal cord injury based on acute clinical and imaging factors. *J Neurotrauma*. Sep 2012;29(13):2263-2271.
41. Batchelor PE, Wills TE, Skeers P, et al. Meta-analysis of pre-clinical studies of early decompression in acute spinal cord injury: a battle of time and pressure. *PLoS One*. 2013;8(8):e72659.
42. Wilson JR, Singh A, Craven C, et al. Early versus late surgery for traumatic spinal cord injury: the results of a prospective Canadian cohort study. *Spinal Cord*. Nov 2012;50(11):840-843.
43. Schneider RC, Cherry G, Pantek H. The syndrome of acute central cervical spinal cord injury; with special reference to the mechanisms involved in hyperextension injuries of cervical spine. *J. Neurosurg*. Nov 1954;11(6):546-577.
44. Lenehan B, Fisher CG, Vaccaro A, Fehlings M, Aarabi B, Dvorak MF. The urgency of surgical decompression in acute central cord injuries with spondylosis and without instability. *Spine (Phila Pa 1976)*. Oct 1 2010;35(21 Suppl):S180-186.
45. Bracken MB, Shepard MJ, Collins WF, et al. A randomized, controlled trial of methylprednisolone or naloxone in the treatment of acute spinal-cord injury. Results of the Second National Acute Spinal Cord Injury Study. *N Engl J Med*. May 17

1990;322(20):1405-1411.

46. Bracken MB, Shepard MJ, Holford TR, et al. Administration of methylprednisolone for 24 or 48 hours or tirilazad mesylate for 48 hours in the treatment of acute spinal cord injury. Results of the Third National Acute Spinal Cord Injury Randomized Controlled Trial. National Acute Spinal Cord Injury Study. *JAMA*. May 28 1997;277(20):1597-1604.
47. Bracken M. Steroids for acute spinal cord injury. *Cochrane Database of Systematic Reviews*: John Wiley & Sons, Ltd; 2012.
48. Bhatt JM, Gordon PH. Current clinical trials in amyotrophic lateral sclerosis. *Expert Opinion on Investigational Drugs*. 2007/08 2007;16(8):1197-1207.
49. Bensimon G, Lacomblez L, Delumeau JC, Bejuit R, Truffinet P, Meininger V. A study of riluzole in the treatment of advanced stage or elderly patients with amyotrophic lateral sclerosis. *J. Neurol*. May 2002;249(5):609-615.
50. Azbill RD, Mu X, Springer JE. Riluzole increases high-affinity glutamate uptake in rat spinal cord synaptosomes. *Brain Research*. 2000/07 2000;871(2):175-180.
51. Nogradi A, Szabo A, Pinter S, Vrbova G. Delayed riluzole treatment is able to rescue injured rat spinal motoneurons. *Neuroscience*. Jan 19 2007;144(2):431-438.
52. Stutzmann JM, Pratt J, Boraud T, Gross C. The effect of riluzole on post-traumatic spinal cord injury in the rat. *Neuroreport*. Jan 31 1996;7(2):387-392.
53. Simard JM, Tsybalyuk O, Keledjian K, Ivanov A, Ivanova S, Gerzanich V. Comparative effects of glibenclamide and riluzole in a rat model of severe cervical spinal cord injury. *Exp Neurol*. Jan 2012;233(1):566-574.
54. Schwartz G, Fehlings MG. Evaluation of the neuroprotective effects of sodium channel blockers after spinal cord injury: improved behavioral and neuroanatomical recovery with riluzole. *J Neurosurg*. Apr 2001;94(2 Suppl):245-256.
55. Kwon BK, Roy J, Lee JH, et al. Magnesium chloride in a polyethylene glycol formulation as a neuroprotective therapy for acute spinal cord injury: preclinical refinement and optimization. *J Neurotrauma*. Aug 2009;26(8):1379-1393.
56. Luo J, Borgens R, Shi R. Polyethylene glycol immediately repairs neuronal membranes and inhibits free radical production after acute spinal cord injury. *J Neurochem*. Oct 2002;83(2):471-480.
57. Kaptanoglu E, Beskonakli E, Solaroglu I, Kilinc A, Taskin Y. Magnesium sulfate treatment in experimental spinal cord injury: emphasis on vascular changes and early clinical results. *Neurosurgical review*. Oct 2003;26:283-287.
58. Kaptanoglu E, Beskonakli E, Okutan O, Selcuk Surucu H, Taskin Y. Effect of magnesium sulphate in experimental spinal cord injury: evaluation with ultrastructural findings and early clinical results. *Journal of clinical neuroscience : official journal of the Neurosurgical Society of Australasia*. May 2003;10(3):329-334.
59. Kaptanoglu E, Beskonakli E, Solaroglu I, Kilinc A, Taskin Y. Magnesium sulfate treatment

in experimental spinal cord injury: emphasis on vascular changes and early clinical results. *Neurosurg. Rev.* Oct 2003;26(4):283-287.

60. Chen M, Ona VO, Li M, et al. Minocycline inhibits caspase-1 and caspase-3 expression and delays mortality in a transgenic mouse model of Huntington disease. *Nat Med.* Jul 2000;6(7):797-801.
61. Giuliani F, Fu SA, Metz LM, Yong VW. Effective combination of minocycline and interferon-beta in a model of multiple sclerosis. *J Neuroimmunol.* Aug 2005;165(1-2):83-91.
62. Seabrook TJ, Jiang L, Maier M, Lemere CA. Minocycline affects microglia activation, Abeta deposition, and behavior in APP-tg mice. *Glia.* May 2006;53(7):776-782.
63. Zanjani TM, Sabetkasaei M, Mosaffa N, Manaheji H, Labibi F, Farokhi B. Suppression of interleukin-6 by minocycline in a rat model of neuropathic pain. *Eur J Pharmacol.* May 24 2006;538(1-3):66-72.
64. Lee SM, Yune TY, Kim SJ, et al. Minocycline inhibits apoptotic cell death via attenuation of TNF-alpha expression following iNOS/NO induction by lipopolysaccharide in neuron/glia co-cultures. *J Neurochem.* Nov 2004;91(3):568-578.
65. Drabek T, Janata A, Wilson CD, et al. Minocycline attenuates brain tissue levels of TNF-alpha produced by neurons after prolonged hypothermic cardiac arrest in rats. *Resuscitation.* Feb 2014;85(2):284-291.
66. Wells JE, Hurlbert RJ, Fehlings MG, Yong VW. Neuroprotection by minocycline facilitates significant recovery from spinal cord injury in mice. *Brain.* Jul 2003;126(Pt 7):1628-1637.
67. Festoff BW, Ameenuddin S, Arnold PM, Wong A, Santacruz KS, Citron BA. Minocycline neuroprotects, reduces microgliosis, and inhibits caspase protease expression early after spinal cord injury. *J Neurochem.* Jun 2006;97(5):1314-1326.
68. Casha S, Zygun D, McGowan MD, Bains I, Yong VW, Hurlbert RJ. Results of a phase II placebo-controlled randomized trial of minocycline in acute spinal cord injury. *Brain.* Apr 2012;135:1224-1236.
69. Imanaka T, Hukuda S, Maeda T. The role of GM1-ganglioside in the injured spinal cord of rats: an immunohistochemical study using GM1-antisera. *J. Neurotrauma.* Mar 1996;13(3):163-170.
70. Geisler FD, Frank; Coleman, William. Recovery of Motor Function after Spinal-Cord Injury — A Randomized, Placebo-Controlled Trial with GM-1 Ganglioside — NEJM. *N Engl J Med.* 1991;324:1829.
71. Geisler FC, William; Grieco, Giacinto; Poonian, Devinder. The Sygen multicenter acute spinal cord injury study. *Spine.* 2001;26(24 Suppl):S87.
72. Siddiqui AM, Khazaei M, Fehlings MG. Translating mechanisms of neuroprotection, regeneration, and repair to treatment of spinal cord injury. *Prog Brain Res.* 2015;218:15-54.
73. Teng YD, Mocchetti I, Taveira-DaSilva AM, Gillis RA, Wrathall JR. Basic fibroblast growth

- factor increases long-term survival of spinal motor neurons and improves respiratory function after experimental spinal cord injury. *J. Neurosci.* Aug 15 1999;19(16):7037-7047.
74. Teng YD, Mocchetti I, Wrathall JR. Basic and acidic fibroblast growth factors protect spinal motor neurones in vivo after experimental spinal cord injury. *Eur. J. Neurosci.* Feb 1998;10(2):798-802.
 75. Wallner S, Peters S, Pitzer C, Resch H, Bogdahn U, Schneider A. The Granulocyte-colony stimulating factor has a dual role in neuronal and vascular plasticity. *Front Cell Dev Biol.* 2015;3:48.
 76. Nishio Y, Koda M, Kamada T, et al. Granulocyte colony-stimulating factor attenuates neuronal death and promotes functional recovery after spinal cord injury in mice. *J Neuropathol Exp Neurol.* Aug 2007;66(8):724-731.
 77. Koda M, Nishio Y, Kamada T, et al. Granulocyte colony-stimulating factor (G-CSF) mobilizes bone marrow-derived cells into injured spinal cord and promotes functional recovery after compression-induced spinal cord injury in mice. *Brain Res.* May 29 2007;1149:223-231.
 78. Takahashi H, Yamazaki M, Okawa A, et al. Neuroprotective therapy using granulocyte colony-stimulating factor for acute spinal cord injury: a phase I/IIa clinical trial. *Eur Spine J.* 2012;21(12):2580-2587.
 79. Kamiya K, Koda M, Furuya T, et al. Neuroprotective therapy with granulocyte colony-stimulating factor in acute spinal cord injury: a comparison with high-dose methylprednisolone as a historical control. *Eur Spine J.* May 2015;24(5):963-967.
 80. Kitamura K, Iwanami A, Fujiyoshi K, et al. Recombinant Human Hepatocyte Growth Factor Promotes Functional Recovery After Spinal Cord Injury. In: Uchida KN, M; Ozawa, H; Katoh, S; Toyama, Y, ed. *Neuroprotection and Regeneration of the Spinal Cord.* Japan: Springer Japan; 2014:147-167.
 81. Kitamura K, Fujiyoshi K, Yamane J, et al. Human hepatocyte growth factor promotes functional recovery in primates after spinal cord injury. *PLoS One.* 2011;6:e27706.
 82. Kitamura K, Iwanami A, Nakamura M, et al. Hepatocyte growth factor promotes endogenous repair and functional recovery after spinal cord injury. *J. Neurosci. Res.* Aug 15 2007;85(11):2332-2342.
 83. Kwon BK, Mann C, Sohn HM, et al. Hypothermia for spinal cord injury. *The Spine Journal.* 2008/11 2008;8(6):859-874.
 84. Dehaes M, Aggarwal A, Lin PY, et al. Cerebral oxygen metabolism in neonatal hypoxic ischemic encephalopathy during and after therapeutic hypothermia. *J Cereb Blood Flow Metab.* Jan 2014;34(1):87-94.
 85. Dingley J, Tooley J, Liu X, et al. Xenon ventilation during therapeutic hypothermia in neonatal encephalopathy: a feasibility study. *Pediatrics.* May 2014;133(5):809-818.
 86. Hypothermia after Cardiac Arrest Study G. Mild therapeutic hypothermia to improve the

- neurologic outcome after cardiac arrest. *N Engl J Med*. Feb 21 2002;346(8):549-556.
87. Lo TP, Jr., Cho KS, Garg MS, et al. Systemic hypothermia improves histological and functional outcome after cervical spinal cord contusion in rats. *J Comp Neurol*. Jun 10 2009;514(5):433-448.
 88. Levi AD, Green BA, Wang MY, et al. Clinical application of modest hypothermia after spinal cord injury. *J Neurotrauma*. Mar 2009;26:407-415.
 89. Kwon BK, Curt A, Belanger LM, et al. Intrathecal pressure monitoring and cerebrospinal fluid drainage in acute spinal cord injury: a prospective randomized trial. *J Neurosurg Spine*. Mar 2009;10:181-193.
 90. Martirosyan NL, Kalani MY, Bichard WD, et al. Cerebrospinal fluid drainage and induced hypertension improve spinal cord perfusion after acute spinal cord injury in pigs. *Neurosurgery*. Apr 2015;76:461-468; discussion 468-469.
 91. Forgiione N, Fehlings MG. Rho-ROCK inhibition in the treatment of spinal cord injury. *World Neurosurg*. Sep-Oct 2014;82(3-4):e535-539.
 92. Fehlings MG, Theodore N, Harrop J, et al. A phase I/IIa clinical trial of a recombinant Rho protein antagonist in acute spinal cord injury. *J Neurotrauma*. May 2011;28:787-796.
 93. Bregman BS, Kunkel-Bagden E, Schnell L, Dai HN, Gao D, Schwab ME. Recovery from spinal cord injury mediated by antibodies to neurite growth inhibitors. *Nature*. Nov 30 1995;378(6556):498-501.
 94. Arriola A, Kiel ME, Shi Y, McKinnon RD. Adjunctive MSCs enhance myelin formation by xenogenic oligodendrocyte precursors transplanted in the retina. *Cell Res*. 2010/05/04 2010;20(6):728-731.
 95. Wang L, Shi J, van Ginkel FW, et al. Neural stem/progenitor cells modulate immune responses by suppressing T lymphocytes with nitric oxide and prostaglandin E2. *Experimental Neurology*. 2009/03 2009;216(1):177-183.
 96. Okamura RM, Lebkowski J, Au M, Priest CA, Denham J, Majumdar AS. Immunological properties of human embryonic stem cell-derived oligodendrocyte progenitor cells. *Journal of Neuroimmunology*. 2007/12 2007;192(1-2):134-144.
 97. Shi Y, Desponts C, Do JT, Hahm HS, Scholer HR, Ding S. Induction of pluripotent stem cells from mouse embryonic fibroblasts by Oct4 and Klf4 with small-molecule compounds. *Cell Stem Cell*. Nov 6 2008;3(5):568-574.
 98. Salewski RP, Mitchell RA, Li L, et al. Transplantation of Induced Pluripotent Stem Cell-Derived Neural Stem Cells Mediate Functional Recovery Following Thoracic Spinal Cord Injury Through Remyelination of Axons. *Stem Cells Transl Med*. Jul 2015;4(7):743-754.
 99. Ahuja C, Fehlings M. Bridging the gap: novel neuroregenerative and neuroprotective strategies in spinal cord injury. *Stem Cells Translational Medicine*. 2016;in press.
 100. Dasari VR, Veeravalli KK, Dinh DH. Mesenchymal stem cells in the treatment of spinal cord injuries: A review. *World J Stem Cells*. Apr 26 2014;6(2):120-133.

101. Quertainmont R, Cantinieaux D, Botman O, Sid S, Schoenen J, Franzen R. Mesenchymal stem cell graft improves recovery after spinal cord injury in adult rats through neurotrophic and pro-angiogenic actions. *PLoS One*. 2012;7(6):e39500.
102. Kim JW, Ha KY, Molon JN, Kim YH. Bone marrow-derived mesenchymal stem cell transplantation for chronic spinal cord injury in rats: comparative study between intralesional and intravenous transplantation. *Spine (Phila Pa 1976)*. Aug 1 2013;38(17):E1065-1074.
103. Swartzlander MD, Blakney AK, Amer LD, Hankenson KD, Kyriakides TR, Bryant SJ. Immunomodulation by mesenchymal stem cells combats the foreign body response to cell-laden synthetic hydrogels. *Biomaterials*. Feb 2015;41:79-88.
104. Bessout R, Semont A, Demarquay C, Charcosset A, Benderitter M, Mathieu N. Mesenchymal stem cell therapy induces glucocorticoid synthesis in colonic mucosa and suppresses radiation-activated T cells: new insights into MSC immunomodulation. *Mucosal Immunol*. May 2014;7(3):656-669.
105. Lim JH, Kim JS, Yoon IH, et al. Immunomodulation of delayed-type hypersensitivity responses by mesenchymal stem cells is associated with bystander T cell apoptosis in the draining lymph node. *J Immunol*. Oct 1 2010;185(7):4022-4029.
106. Williams RR, Bunge MB. Schwann cell transplantation: a repair strategy for spinal cord injury? *Prog Brain Res*. 2012;201:295-312.
107. Windus LC, Lineburg KE, Scott SE, et al. Lamellipodia mediate the heterogeneity of central olfactory ensheathing cell interactions. *Cell Mol Life Sci*. May 2010;67(10):1735-1750.
108. Silva NA, Cooke MJ, Tam RY, et al. The effects of peptide modified gellan gum and olfactory ensheathing glia cells on neural stem/progenitor cell fate. *Biomaterials*. Sep 2012;33(27):6345-6354.
109. Zhang J, Chen H, Duan Z, et al. The Effects of Co-transplantation of Olfactory Ensheathing Cells and Schwann Cells on Local Inflammation Environment in the Contused Spinal Cord of Rats. *Mol Neurobiol*. Jan 20 2016.
110. Ekberg JA, St John JA. Olfactory ensheathing cells for spinal cord repair: crucial differences between subpopulations of the glia. *Neural Regen Res*. Sep 2015;10(9):1395-1396.
111. Liu J, Chen P, Wang Q, et al. Meta analysis of olfactory ensheathing cell transplantation promoting functional recovery of motor nerves in rats with complete spinal cord transection. *Neural Regen Res*. Oct 15 2014;9(20):1850-1858.
112. Li L, Adnan H, Xu B, et al. Effects of transplantation of olfactory ensheathing cells in chronic spinal cord injury: a systematic review and meta-analysis. *Eur Spine J*. May 2015;24(5):919-930.
113. Caicco MJ, Zahir T, Mothe AJ, et al. Characterization of hyaluronan-methylcellulose

- hydrogels for cell delivery to the injured spinal cord. *J Biomed Mater Res A*. May 2013;101(5):1472-1477.
114. Mothe AJ, Tam RY, Zahir T, Tator CH, Shoichet MS. Repair of the injured spinal cord by transplantation of neural stem cells in a hyaluronan-based hydrogel. *Biomaterials*. May 2013;34(15):3775-3783.
 115. Tam RY, Cooke MJ, Shoichet MS. A covalently modified hydrogel blend of hyaluronan–methyl cellulose with peptides and growth factors influences neural stem/progenitor cell fate. *Journal of Materials Chemistry*. 2012;22(37):19402.
 116. Ansorena E, De Berdt P, Ucakar B, et al. Injectable alginate hydrogel loaded with GDNF promotes functional recovery in a hemisection model of spinal cord injury. *Int J Pharm*. Oct 15 2013;455(1-2):148-158.
 117. Itosaka H, Kuroda S, Shichinohe H, et al. Fibrin matrix provides a suitable scaffold for bone marrow stromal cells transplanted into injured spinal cord: A novel material for CNS tissue engineering. *Neuropathology*. 2009/06 2009;29(3):248-257.
 118. Zweckberger K, Ahuja CS, Liu Y, Wang J, Fehlings MG. Self-assembling peptides optimize the post-traumatic milieu and synergistically enhance the effects of neural stem cell therapy after cervical spinal cord injury. *Acta Biomater*. Jun 10 2016.
 119. Liu Y, Ye H, Satkunendrarajah K, Yao GS, Bayon Y, Fehlings MG. A self-assembling peptide reduces glial scarring, attenuates post-traumatic inflammation and promotes neurological recovery following spinal cord injury. *Acta Biomater*. Sep 2013;9(9):8075-8088.
 120. Lu P, Wang Y, Graham L, et al. Long-distance growth and connectivity of neural stem cells after severe spinal cord injury. *Cell*. Sep 14 2012;150(6):1264-1273.
 121. Bracken MB. Methylprednisolone in the management of acute spinal cord injuries. *Med J Aust*. Sep 17 1990;153(6):368.
 122. Bracken MB, Collins WF, Freeman DF, et al. Efficacy of methylprednisolone in acute spinal cord injury. *JAMA*. Jan 6 1984;251(1):45-52.
 123. Martin AR AI, Cohen-Adad J, Tarmohamed Z, Tetreault L, Smith N, Cadotte DW, Crawley A, Ginsberg H, Mikulis D, Fehlings MG. Translating state-of-the-art spinal cord mri techniques to clinical use: A systematic review of clinical studies utilizing DTI, MT, MWF, MRS, and fMRI. *Neuroimage: Clinical (in press; accepted 2015-Nov-27)*. 2015.
 124. Cadotte DW, Fehlings MG. Will imaging biomarkers transform spinal cord injury trials? *Lancet Neurol*. Sep 2013;12(9):843-844.
 125. Kwon BK, Streijger F, Fallah N, et al. Cerebrospinal Fluid Biomarkers to Stratify Injury Severity and Predict Outcome in Human Traumatic Spinal Cord Injury. *J Neurotrauma*. Jun 27 2016.
 126. Kwon BK, Stammers AM, Belanger LM, et al. Cerebrospinal fluid inflammatory cytokines and biomarkers of injury severity in acute human spinal cord injury. *J Neurotrauma*. Apr 2010;27(4):669-682.

127. Nieto-Diaz M, Esteban FJ, Reigada D, et al. MicroRNA dysregulation in spinal cord injury: causes, consequences and therapeutics. *Front Cell Neurosci.* 2014;8:53.
128. Martin AR AI, Fehlings MG. Diagnosis and acute management of spinal cord injury: Current best practices and emerging therapies. *Current Trauma Reports.* 2015;1(3):169-181.
129. Kirshblum SC, Waring W, Biering-Sorensen F, et al. Reference for the 2011 revision of the International Standards for Neurological Classification of Spinal Cord Injury. *J Spinal Cord Med.* Nov 2011;34(6):547-554.
130. Singh A, Tetreault L, Kalsi-Ryan S, Nouri A, Fehlings MG. Global prevalence and incidence of traumatic spinal cord injury. *Clin Epidemiol.* 2014;6:309-331.

Table 1: “Current best practices for the diagnosis and management of SCI. The table displays several key recommendations, many of which are from the 2013 updated guidelines from the Joint Section on Disorders of the Spine and Peripheral Nerves of the American Association of Neurological Surgeons and the Congress of Neurological Surgeons.”

Reprinted with permission from Martin AR, Aleksanderek I, Fehlings MG. Diagnosis and acute management of spinal cord injury: current best practices and emerging therapies.

Current Trauma Reports. 2015;1(3):169-181¹²⁸.

Topic	Level of AANS/CNS Recommendation	Guideline/Recommendation
Hypotension	Level III	Correction of hypotension to systolic blood pressure > 90 mm Hg) as soon as possible
	Level III	Maintenance of mean arterial blood pressure between 85 and 90 mm Hg for 7 days
Hypoxia	None	Hypoxia (PaO ₂ < 60 mm Hg or O ₂ saturation < 90%) should be avoided [3]
ICU Monitoring	Level III	SCI patients should be managed in an ICU setting with cardiac, hemodynamic, and respiratory monitoring to detect cardiovascular dysfunction and respiratory insufficiency
Immobilization	Level II	Patients with SCI or suspected SCI (except in penetrating injury) should be immobilized
	Level III	Spinal immobilization should be performed with rigid cervical collar and supportive blocks on a backboard with straps
Specialized Centers	Level III	SCI patients should be transferred expediently to specialized centers of SCI care
Examination	Level II	The ASIA ISNCSCI examination should be performed and documented
Imaging	Level I	No cervical imaging is required in awake trauma patients that have no neck pain/tenderness, normal neurological examination, normal range of motion, and no distracting injuries
	Level I	CT is recommended in favour of cervical x-rays
	Level I	CT angiography is recommended in patients that meet the modified Denver screening criteria [9]
Neuroprotection	Level I	Methylprednisolone is not recommended *

Spinal Cord Decompression	None	Surgical decompression prior to 24 hours after SCI can be performed safely and is associated with improved neurological outcome [10]
	Level III	Early closed reduction of fracture/dislocation in awake patients without a rostral injury is recommended, and pre-reduction MRI does not appear to influence outcome

* The authors do not agree with this guideline.

Table 2. Summary of International Standards for Neurological Classification of Spinal Cord Injury

Parameter	Definition
Motor Score	Score out of 100 points representing motor power in 5 key myotomes (each grade out of 5) in each limb
Sensory Score	Score out of 224 points representing light touch and pin prick sensation in 28 dermatomes bilaterally
AIS grade	Cumulative measure of injury severity ranging from AIS grade A (most severe motor sensory complete lesion) to AIS grade E (least severe no neurological deficit)
AIS grade A	No motor or sensory preservation below the neurological level of injury (including the distal sacral segments)
AIS grade B	Sensory, but no motor, preservation below the neurological level of injury (including the distal sacral segments)
AIS grade C	Motor preservation below the neurological level of injury (including the distal segments) with less than half of key muscles below the neurological level graded antigravity or better
AIS grade D	Motor preservation below the neurological level of injury (including the distal segments) with at least half of key muscles below the neurological level graded antigravity or better
AIS grade E	Neurological normal in a patients who previously had deficit
Neurological Level of Injury	The lowest segment where motor and sensory function is normal on both sides
Zone of Partial Preservation	In AIS grade A patient, lowest dermatome or myotome with partial innervation

Modified from Kirshblum et al. J Spinal Cord Med. 2011;34(6):535-546.¹²⁹

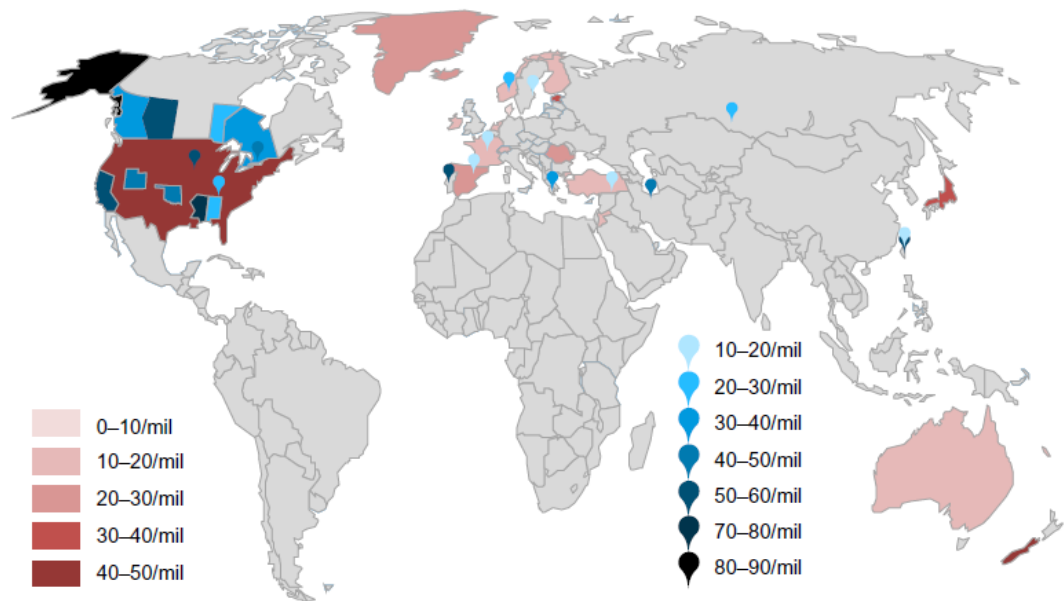


Figure 2 Relative annual incidences of countries, states/provinces, and regions.

Notes: The red color scheme illustrates incidences of countries. The blue color scheme highlights incidences of states/provinces and regions.

Abbreviation: mil, million.

Figure 1. Annual incidence of spinal cord injury across reported countries, states/provinces, and regions. Reprinted with permission from Singh A, et al. Clin Epidemiol. 2014;6:309-331.¹³⁰

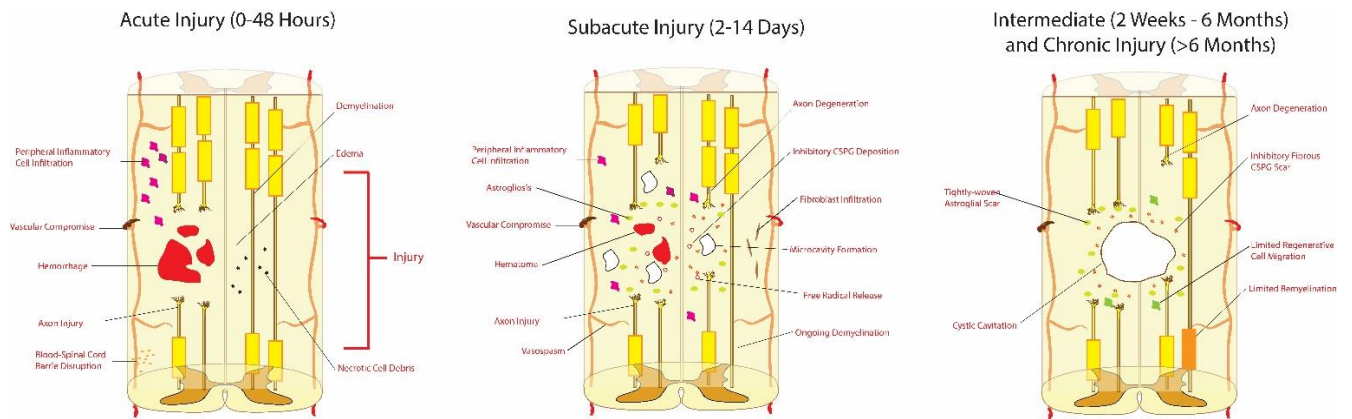


Figure 2. Pathophysiological evolution of spinal cord injury. In the acute injury period (0-48hrs) hemorrhage, edema, and pro-apoptotic factors (e.g. cytokines, K⁺, DNA, necrotic debris, etc.) contribute to ongoing cell death. Neurons and oligodendrocytes are injured resulting in further loss of function beyond the initial traumatic insult. Astrocytes rapidly activate, proliferate, and infiltrate the site of injury while depositing chondroitin sulfate proteoglycans (CSPGs) into the microenvironment and release additional pro-inflammatory factors which propagate the injury cascade. Demyelinated and injured axons begin to dieback from the inflamed and ischemic perilesional region. In the late subacute and intermediate phases, continued apoptotic and necrotic cell death leave microcystic cavities which eventually coalesce to form formidable barriers to regeneration in the chronic phase (>6 months). The final chronic phase scar is a dynamic entity consisting of a tightly-woven network of astrocytic processes with a dense fibrous deposit of CSPG acting as a physical and biochemical barrier to neurite outgrowth and regenerative cell migration.

ASIA INTERNATIONAL STANDARDS FOR NEUROLOGICAL CLASSIFICATION OF SPINAL CORD INJURY (ISNCSCI) **ISICOS**
AMERICAN SPINAL INJURY ASSOCIATION INTERNATIONAL SPINAL CORD SOCIETY

Patient Name _____ Date/Time of Exam _____
 Examiner Name _____ Signature _____

RIGHT

MOTOR KEY MUSCLES

Elbow flexors: **C5**

Wrist extensors: **C6**

Elbow extensors: **C7**

Finger flexors: **C8**

Finger abductor (little finger): **T1**

Hip flexors: **L2**

Knee extensors: **L3**

Ankle dorsiflexor: **L4**

Long toe extensor: **L5**

Ankle plantar flexor: **S1**

(VAC) Voluntary Anal Contraction (Yes/No)

RIGHT TOTALS (MAXIMUM) (50) (56) (56)

MOTOR SUBSCORES

UER + UEL = UEMS TOTAL (MAX(25) (25) (50))

LER + LEL = LEMS TOTAL (MAX(25) (25) (50))

SENSORY KEY SENSORY POINTS

Light Touch (LTR) Pin Prick (PPR)

C2

C3

C4

T2

T3

T4

T5

T6

T7

T8

T9

T10

T11

T12

L1

S2

S3

S4-5

SENSORY KEY SENSORY POINTS

Light Touch (LTL) Pin Prick (PPL)

C2

C3

C4

T2

T3

T4

T5

T6

T7

T8

T9

T10

T11

T12

L1

S2

S3

S4-5

LEFT TOTALS (MAXIMUM) (56) (56) (50)

SENSORY SUBSCORES

LTR + LTL = LT TOTAL (MAX(56) (56) (112))

PPR + PPL = PP TOTAL (MAX(56) (56) (112))

NEUROLOGICAL LEVELS Steps 1-5 for classification as on reverse

1. SENSORY R L

2. MOTOR R L

3. NEUROLOGICAL LEVEL OF INJURY (NLI)

4. COMPLETE OR INCOMPLETE? Incomplete = Any sensory or motor function in S4-5

5. ASIA IMPAIRMENT SCALE (AIS)

(In complete injuries only) ZONE OF PARTIAL PRESERVATION R L

MOTOR R L

This form may be copied freely but should not be altered without permission from the American Spinal Injury Association. REV 11/15

Figure 3. International Standards for Neurological Classification of Spinal Cord Injury clinical examination form. The standardized assessment and calculation of motor and sensory scores is demonstrated on this template.

Reprint of the 2015 American Spinal Injury Association and International Spinal Cord Society ISNCSCI assessment form retrieved from <http://www.asia-spinalinjury.org/elearning/International%20Stds%20Diagram%20Worksheet%2011.2015%20opt.pdf>.

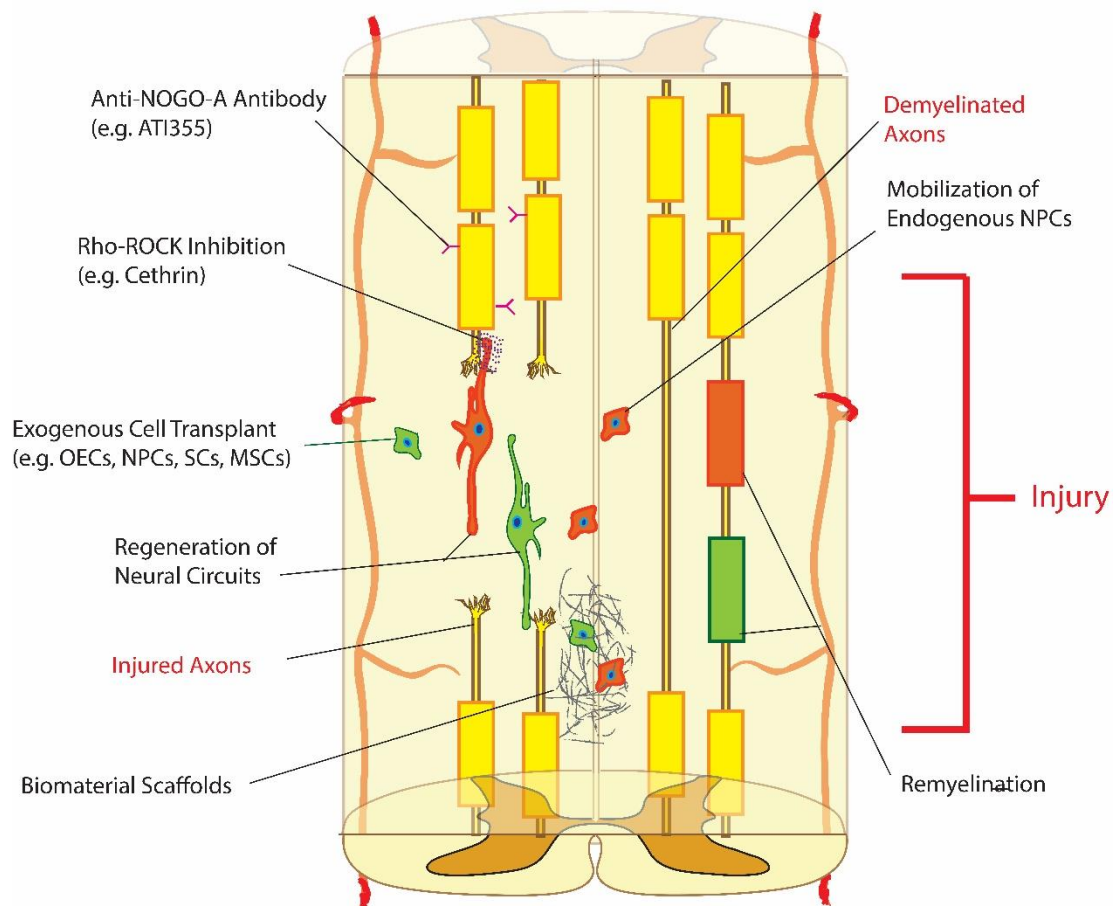


Figure 4 Neuroregenerative strategies for spinal cord injury. Schematic of a traumatic spinal cord injury with demyelination and loss of axons. Regenerative therapies actively being translated are shown including Anti-NOGO-A antibody treatment (e.g. ATI355), Rho-ROCK inhibition (e.g. Cethrin), cell transplants (e.g. iPSC-NPC, ES-NPC, OEC, SC, BMC, MSC), implantation of biomaterials, and mobilization of endogenous cell pools (e.g. Metformin).