

1 **Hominoid arcade shape: pattern and magnitude of covariation**

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25 **Abstract**

26           The shape of the dental arcade and canine size distinguish extant humans from all apes.  
27 Humans are characterized by a parabolic arcade with short postcanine tooth rows and small canines,  
28 whereas apes have long, U-shaped arcades with large canines. The evolutionary and biomechanical  
29 mechanisms underlying arcade shape differences between and within groups are not well understood. It  
30 is unclear, for example, whether evolutionary changes in the covariation among modules comprising the  
31 upper and lower jaws are the cause and/or consequence of different arcade shapes. Here we use 3D  
32 geometric morphometric methods to explore to what extent the morphological differences in arcade  
33 shape between living hominoids are related to differences in covariation of upper and lower jaws, and  
34 the premaxilla and the maxilla. We show that all extant hominoids follow a very similar covariation  
35 pattern between upper and lower dental arcades, as well as between the premaxilla and the maxilla. We  
36 find comparably high magnitudes of covariation between the premaxilla and the maxilla in all groups.  
37 Between the upper and lower jaws, levels of covariation are similar in apes (*Pan*, *Gorilla*, *Pongo*, and  
38 *Hylobates*), but overall lower in extant humans. Our results demonstrate an independence of the  
39 pattern of arcade shape covariation from dental spatial arrangements. Importantly, we show that a  
40 shared hominoid pattern of covariation between premaxilla and maxilla together with the covariation of  
41 upper and lower jaw is consistent with major evolutionary arcade shape changes in hominoids. We  
42 suggest that with the reduction of canine and diastema size in hominins, the incisors move posteriorly  
43 and the tooth row becomes more parabolic. Our study provides a framework for addressing questions  
44 about fossil hominin dentognathic diversity, including inter- and intraspecific variation and associations  
45 of upper and lower jaw morphology.

46

## 47 **Introduction**

48 Dentognathic morphology of extant humans and apes differs notably in canine size and arcade  
49 shape. In apes, the arcades are U-shaped with narrow and near-parallel postcanine tooth rows.  
50 Furthermore, dental arcade shape varies within and between ape species: posteriorly diverging or  
51 converging postcanine tooth rows result in a range of arcade contours (Hellman, 1919). In some  
52 individuals the arcade tapers in the premolar region to diverge again more posteriorly, giving it a saddle  
53 shape (Hellman, 1919; Remane, 1921). The large lower canines are accommodated in a diastema  
54 between the upper lateral incisor and the enlarged upper canine; in the mandible, the upper canine is  
55 accommodated between the lower canine and the lower premolars (Angle, 1899; Hellman, 1942), where  
56 canine size can vary strongly inter- and intraspecifically (Almquist, 1974; Leutenegger and Kelly, 1977;  
57 Oxnard et al., 1985; Leutenegger and Shell, 1987; Plavcan and van Schaik, 1992; Plavcan, 1993; Kelley,  
58 1995).

59 Modern human dental arcades are usually referred to as being parabolic (Angle, 1899; Broomell,  
60 1902; Le Gros Clark, 1950; Genet-Varcin, 1969), elliptic (Black, 1902; Garn, 1968; Currier, 1969; Brader,  
61 1972) or catenary (MacConaill and Scher, 1949; Scott, 1957; Engel, 1979), with upper and lower arcades  
62 having slightly different shapes (Hellman, 1919; Engel, 1979). Incisors and canines are of almost equal  
63 size (Schwartz, 1995) and their occlusal surfaces are on the same level as the postcanine dentition.

64 Intra- and interspecific differences in cranial and mandibular architecture among primates have  
65 been related to differences in masticatory behavior, including varying mechanical stress (Hrdlicka, 1940;  
66 Hylander, 1972; Carlson and Vangerven, 1977; Hylander, 1979; Weijs and Hillen, 1984; Bouvier, 1986;  
67 Weijs and Hillen, 1986; Armelagos et al., 1989; Ravosa, 1990; Corruccini, 1991; Herring, 1993; Anapol  
68 and Lee, 1994; Larsen, 1995; Cassidy et al., 1998; Sardi et al., 2004; Lieberman, 2008; von Cramon-  
69 Taubadel, 2011; Prasad et al., 2013). These biomechanical arguments emphasize that relative position

70 and length of the dental arcade influence the stress distribution (Witzel and Preuschoft, 2002). In this  
71 view, phenotypic variation of the upper and lower jaw is seen as a consequence rather than a  
72 precondition for different force distributions. In apes relatively long tooth rows and prognathic dental  
73 arcades seem to be biomechanically advantageous, whereas in humans the parabolic arcade is seen as a  
74 side effect of reduction in bite- and chewing forces and the reorganization of the face (Preuschoft, 1989;  
75 Witzel and Preuschoft, 1999) with little biomechanical significance (Preuschoft and Witzel, 2004). Recent  
76 studies in capuchin monkeys (Makedonska et al., 2012) and modern humans (Noback and Harvati, 2015)  
77 showed that, at least within groups, dental arcade shape (in contrast to size and position) is largely  
78 independent from masticatory forces. These authors found correlations between diet and shape of the  
79 temporalis muscle and the cranium, but none between subsistence and maxillary arch shape.

80         The evolutionary and biomechanical mechanisms underlying the differences in arcade shape  
81 between extant humans and apes, as well as those underlying the within-group variability, are not well  
82 understood. The evolutionary trajectory of our lineage is interesting in this regard, as arcade shape  
83 varies considerably among fossil hominins (e.g., Weidenreich, 1936; Tobias, 1967; Johanson et al., 1978;  
84 Johanson and White, 1979; Greenfield, 1992). Besides a general reduction in absolute canine size, early  
85 hominins like *Australopithecus anamensis* and *A. afarensis* possess primitive features such as a small  
86 diastema between the upper incisor and the canine, as well as parallel to slightly diverging tooth rows  
87 (Schwartz, 1995; White et al., 2000; Ward et al., 2001; Kimbel and Delezene, 2009). Some early *Homo*  
88 specimens retain long and almost parallel tooth rows, whereas others have shorter postcanine rows and  
89 non-projecting frontal tooth rows (Tobias, 1991; Wood, 1991; Rightmire, 1993; Kimbel et al., 1997;  
90 Clarke, 2012; Leakey et al., 2012; Spoor et al., 2015). Large-scale differences in jaw shapes have been  
91 used to support arguments about species diversity in early *Homo* (Spoor et al., 2015). Moreover, it has  
92 been argued that high variation in the anterior maxillary region, the premaxilla, indicates that it might  
93 have been subject to selection in hominin evolution (Villmoare et al., 2014). Understanding the

94 mechanisms contributing to the variation and covariation of jaw shapes in extant hominoids is thus  
95 informative for interpreting the hominin fossil record.

96 Morphological covariation is the statistical interrelationship of morphometric variables  
97 (Mitteroecker and Bookstein, 2007). It is the consequence of developmental, evolutionary, genetic, and  
98 functional processes (Cheverud, 1996a) that in combination lead to integration of structures. Integrated  
99 traits must covary, however, traits that covary are not necessarily integrated (Villmoare et al., 2014).  
100 Although not equivalent (compare Porto et al., 2009; Young et al., 2010; Grabowski et al., 2011),  
101 integration and covariation are mutually dependent in that covarying structures influence the way an  
102 organism can react to evolutionary forces. On the other hand, evolution can also act on the covariation  
103 of structures. Using 3D geometric morphometric methods we explore the pattern and magnitude of the  
104 covariation between and within upper and lower arcades in extant hominoids. We seek to understand if  
105 and how the morphological differences in arcade shape between living hominoids are related to  
106 differences in covariation of upper and lower jaws.

107 Previous studies of cranial integration and covariation have consistently demonstrated  
108 conserved patterns among primates and even mammals (Cheverud, 1996b; Ackermann and Cheverud,  
109 2000; Lieberman et al., 2000; Marroig and Cheverud, 2001; Ackermann, 2002; Gonzalez-Jose et al.,  
110 2004; Marroig et al., 2004; Ackermann, 2005; Goswami, 2006; Gunz and Harvati, 2007; Mitteroecker and  
111 Bookstein, 2008; Porto et al., 2009; Makedonska et al., 2012; Villmoare et al., 2014). In this study we  
112 assess (1) whether the same applies to hominoid upper and lower dental arcades, or (2) whether the  
113 variation in extant arcade shapes is associated with a variation of the underlying patterns of covariation.

114 The magnitude of covariation has been related to material properties of food, in that taxa  
115 relying on a mechanically challenging diet have stronger integrated upper and lower jaws (Marroig and  
116 Cheverud, 2001; Makedonska et al., 2012). *Pan*, *Pongo*, *Gorilla*, and *Hylobates* consume different

117 proportions of leaves, fruit, other plant parts, or insects, and their dietary preferences are dependent on  
118 season, food availability, habitat and sex. In general, however, *Pan*, *Pongo*, and *Hylobates* are more  
119 frugivorous, whereas *Gorilla* is more reliant on leaves (MacKinnon, 1974; Rijksen, 1978; Gittins and  
120 Raemaekers, 1980; Watts, 1984; Galdikas, 1988; Tutin and Fernandez, 1993; Knott, 1998; McConkey et  
121 al., 2003; Rogers et al., 2004; Boesch et al., 2006; Doran-Sheehy et al., 2009). Morphologically, this  
122 difference is reflected in relative dental sizes. Frugivores have relatively larger incisors, while foliovores  
123 have relatively larger molars (Hylander, 1975; Kay and Hylander, 1978). In comparison to apes, modern  
124 humans have a more generalist diet, where extra-oral food processing via tool use or cooking is thought  
125 to have reduced the masticatory effort since the emergence of our species or even earlier (e.g. Brace et  
126 al., 1987; Wrangham et al., 1999; Richards et al., 2001; Teaford et al., 2002; Henry, 2010). While a  
127 correlation with diet seems likely, some authors have suggested that the lower covariation magnitudes  
128 in the human cranium (Marroig et al., 2009; Porto et al., 2009) and pelvis (Grabowski et al., 2011) may  
129 indicate a general relaxation of integration in the hominin lineage. These authors argued that the  
130 change in magnitude of integration might have paved the way for the development of the  
131 morphological changes characterizing our lineage. In addition to the two aforementioned aims, we  
132 therefore also assess (3) whether there are differences in the magnitude of covariation between the  
133 groups, and (4) whether there are differences between females and males within groups.

134         It has been argued that within the maxilla, the premaxilla represents its own module that has  
135 been a target of selection in the hominin lineage leading to higher variation in the anterior region in  
136 hominins compared to other anthropoids (Villmoare et al., 2014). The reduction of canine and thereby  
137 diastema size is associated with an earlier closure of the premaxillary sutures (McCollum and Ward,  
138 1997; Braga, 1998). We explore whether those differences are associated with a change in the pattern  
139 or the magnitudes of covariation. We therefore also assess (5) the pattern and magnitudes of  
140 covariation between the premaxilla and the maxilla.

141

## 142 **Materials and methods**

### 143 *Samples*

144           Our samples are summarized in Table 1 and in the Supplementary Online Material (SOM) Table  
145 S1. The modern human sample ( $n=53$ ) comprises geographically diverse pre-industrial populations, and  
146 was obtained from the Institute of Anatomy at Leipzig University (ULAC), Germany; the Natural History  
147 Museum London, UK; the National Historical Museum, Buenos Aires, Argentina; the American Museum  
148 of Natural History, New York; and the Smithsonian Institution, Washington, D.C., USA. The *Pan*  
149 *trogodytes* sample ( $n=44$ ) is comprised of specimens from Côte d'Ivoire, Liberia, Gabon, Cameroon and  
150 Equatorial Guinea, housed at the Senckenberg Museum, Frankfurt, Germany; at the Max Planck Institute  
151 for Evolutionary Anthropology in Leipzig (MPI-EVA), Germany; the Phyletisches Museum, Jena,  
152 Germany; and the Smithsonian Institution; plus one zoo specimen housed in the Phyletisches Museum  
153 Jena. The *Gorilla* sample ( $n=53$ ) includes *Gorilla gorilla* specimens from Cameroon, Gabon, and the  
154 Congo, and *Gorilla beringei* from Rwanda, housed at the Museum für Naturkunde Berlin, Germany; the  
155 Phyletisches Museum, Jena and the Smithsonian Institution. The *Pongo* sample ( $n=52$ ) comprises *P.*  
156 *pygmaeus* and *P. abelii* specimens from Borneo and Sumatra, respectively, and one specimen with  
157 unknown provenance. The *Pongo* specimens are housed at the Museum für Naturkunde Berlin and the  
158 Smithsonian Institution. The *Hylobates* specimens ( $n=63$ ) include the species *H. agilis* (Sumatra), *H.*  
159 *albibarbis* (Borneo), *H. klossii* (Sumatra), *H. lar* (Thailand, Myanmar), and *H. muelleri* (Borneo). The  
160 specimens are housed at the Museum für Naturkunde Berlin and the Smithsonian Institution. Sex  
161 attributions were taken from museum records when available. Sex of the remaining specimens of *Pan*,  
162 *Gorilla* and *Pongo* was determined by several observers based on the formation of cranial  
163 superstructures and canine size.

164 We used computed tomography (CT) of all specimens, including both industrial CT (BIR ACTIS  
165 225/300 at the Max Planck Institute for Evolutionary Anthropology, Leipzig, Germany; isotropic voxel  
166 sizes 0.03 – 0.093 mm) and medical CT (Vivantes Klinikum Berlin, Germany; CIMED, La Plata, Argentina;  
167 the Senckenberg Museum, and the Smithsonian Institution, Washington, U.S.A. (modern human CT  
168 scans obtained as part of Copes (2012)); pixel sizes 0.13 –0.47 mm, slice intervals 0.33 - 0.50 mm). Our  
169 sample comprises scans of adult individuals (defined as third molars in occlusion) that preserved both  
170 cranium and mandible and did not display malocclusions or other severe pathologies. So as to achieve  
171 comparably large sample sizes for all groups that meet these strict criteria, we pooled the respective  
172 species of *Gorilla*, *Pongo*, and *Hylobates* in our analyses. Species specific morphologies within genera are  
173 known (e.g. Groves, 1970, 1972; Rörer-Ertl, 1984; Uchida, 1998; Guy et al., 2003; Schmittbuhl et al.,  
174 2007, but see Groves et al. 1992; Courtenay et al. 1998 for a different view on *Pongo*), however, the  
175 differences in dental arcade shape seem to be negligible, when compared to the shape differences  
176 between genera, as can be seen in SOM Figures S1-S3, in which the species of each individual are color  
177 coded to illustrate that they are randomly scattered within the convex hulls of their respective genera.

178

#### 179 *Data*

180 The occlusal surfaces of teeth are modified by wear, which makes placing homologous  
181 landmarks problematic. Hence, we took landmarks on the alveolar margin of every tooth and on the  
182 cervix of the postcanine dentition in order to capture arcade shape, spatial arrangement of the teeth,  
183 and dental size proportions. We recorded 224 homologous 3D landmarks on the mandibular and the  
184 maxillary dental arcades (112 landmarks each) (Fig. 1). All measurements were taken in Avizo 7.1  
185 (Visualization Sciences Group).



186 To capture overall length and breadth patterns, landmarks were placed on the distal, buccal,  
187 mesial and lingual surfaces of the incisor, canine and premolar alveolus. On the molars, besides a distal  
188 and a mesial landmark, we set two landmarks buccally and two lingually at the position of each root. On  
189 the third molar we took one landmark buccally and lingually at the position of the mesial root. The  
190 cervical landmarks were positioned distally, buccally, mesially, and lingually.

191 - Table 1 -

192 - Figure 1 -

### 193 *Analysis*

194 Landmarks on the upper and the lower jaw were superimposed separately applying Generalized  
195 Procrustes Analysis (GPA) to extract shape information independent from rotation, translation and  
196 scaling (Rohlf and Slice, 1990; Bookstein, 1991). We performed a principal component analysis (PCA) on  
197 the Procrustes shape coordinates for mandibles and maxillae separately to assess taxonomic shape  
198 differences.

199 Landmarks on the premaxilla (incisors) and the maxilla (canines, premolars, and molars) were  
200 superimposed together to retain information on the exact position of the premaxilla relative to the  
201 maxilla. In the analysis of the covariation between premaxilla and maxilla, only the alveolar, not the  
202 cervical, landmarks were used to capture the morphology of the postcanine dentition.

203 Pattern of covariation A two-block partial least squares (PLS) analysis was used to quantify the  
204 covariation of the mandibular and the maxillary arcade shape, or the premaxilla and the maxilla,  
205 respectively (Rohlf and Corti, 2000; Bookstein et al., 2003). We used the mean of the original data and  
206 the reflected relabeled landmark configuration in order to symmetrize the data and therewith remove  
207 asymmetric shape variation that also arises from asymmetric measurement error (Klingenberg and  
208 McIntyre, 1998; Mardia et al., 2000; Mitteroecker and Gunz, 2009). The PLS analysis computes

209 correlated pairs of vectors, so-called singular warps (SW), for the maxilla and for the mandible, or the  
210 premaxilla and the maxilla, respectively, that account for the maximum covariance between the two  
211 sets of landmarks (Wold, 1966; Rohlf and Corti, 2000; Bookstein et al., 2003).

212 We performed two PLS analyses per data set: one includes all groups with the data mean  
213 centered for each group. This explores the overall pattern of covariation while accounting for large-scale  
214 differences between the groups. The second PLS analysis was performed for every group separately,  
215 mean centered by sex. If all groups follow a similar pattern of covariation, shape variation associated  
216 with PLS axes of the pooled sample and within groups should be similar.

217 Magnitude of covariation We used two metrics to quantify the magnitude of covariation; the  
218 covariance ratio (CR, Adams, 2016) and the correlation coefficient between the singular warp scores  
219 (Bookstein et al., 2003). The CR has been proposed recently by Adams (2016) as an alternative to the RV  
220 coefficient that overcomes some of the fundamental methodological issues associated with the RV  
221 coefficient (Smilde et al., 2009; Fruciano et al., 2013; Bookstein, 2016). The CR quantifies the overall  
222 pattern of covariation between the blocks. Therefore one does not know which aspect of covariation the  
223 CR is quantifying. In contrast, the correlations of the singular warp scores pertain to the pattern  
224 visualized in the respective singular warp plots, and are easier to interpret. As our sample sizes are not  
225 equal across all groups, we used a resampling approach for computing the values for the CR. To get a  
226 representative distribution, the CR was calculated for 30 randomly selected individuals in 1000 iterations  
227 per group.

228 A permutation test (Good, 2000) was used to determine the significance of the mean  
229 differences between the permuted CR values (Bonferroni-corrected for multiple testing, considered  
230 significant at  $\alpha < 0.05$ ).

231           Sexual dimorphism A permutation test (Good, 2000) was used to determine the significance of  
232 the mean shape differences between male and female mandibles and maxillae. To do so, we randomly  
233 selected individuals and assigned them to male or female and computed a mean shape and shape  
234 difference between the permuted sexes 10 000 times (considered significant at  $\alpha < 0.05$ ).

235           Allometry To assess the amount of the total shape variance that can be explained by jaw size we  
236 separately computed multivariate regressions of upper and lower arcade shapes on the natural  
237 logarithms of their centroid sizes. We evaluated the statistical significance of these regressions using a  
238 permutation test based on the explained variance (Mitteroecker et al., 2013).

239           Intra-observer error All data were measured by one of the authors (S.S). Intra-observer error  
240 was assessed by an analysis of repeated measurements: one specimen (*Gorilla gorilla*, ZMB 14645,  
241 medical CT scan, voxel size 0.227x0.227x0.335 mm) was measured fifteen times. The largest Procrustes  
242 distance between repeated measurements of this individual was considerably smaller than the smallest  
243 Procrustes distance between different specimens of the same species. Specimen affinity is therefore not  
244 affected by intra-observer error.

245 All analyses and visualizations were performed in Mathematica 8.0 (Wolfram Research Inc., 2010).

246

## 247 **Results**

### 248 *1. PCA*

249           Table 2 shows the results of the principal components (PC) analysis. The first PC (PC1, 66.9% of  
250 total shape variance in maxillae, 70.8% in mandibles) separates humans from the other groups. Both,  
251 maxillae (Fig. 2a) and mandibles (Fig. 2c), have short parabolic arcades and small canines on the positive  
252 end (humans) compared to a U-shaped arch, long parallel tooth rows with large canines on the negative  
253 side (apes).

254 For the maxillae, PC2 (11.0% of the total shape variance) describes the relation between a  
255 relatively large postcanine dentition with relatively small incisors in the positive direction (*Gorilla*); and  
256 relatively large incisors with a relatively small postcanine dentition in the negative direction (*Pan*) (Fig.  
257 2a). For the mandibles (10.9% of the total shape variance), PC2 combines a long postcanine tooth row  
258 with small incisors on the positive end (*Gorilla*), whereas the negative extreme of PC2 combines a short  
259 postcanine tooth row with a large anterior dentition (*Pan*) (Fig. 2c).

260 PC3 (5.8% of total shape variance in maxillae, 6.3% for mandibles) separates *Hylobates* from the  
261 other hominoids. For the maxilla in the negative direction, relatively small postcanine teeth are  
262 accompanied by relatively large canines and small incisors; the arcade is slightly V-shaped (*Hylobates*)  
263 (Fig. 2b). For the mandibles, moderately sized canines are associated with relatively large postcanine  
264 teeth and incisors in the positive direction of PC3 (other groups). The lower scores of PC3 describe  
265 relatively small postcanine dentition associated with relatively small incisors and slightly enlarged  
266 canines (*Hylobates*).

267 - Figure 2 –

268 - Table 2 –

## 269 2. Pattern and magnitudes of covariation between the upper and the lower arcades

270 Pattern of covariation (all groups, group-mean centered) Table 3 shows the percentages of the  
271 explained covariance for the first five PLS components. On the positive side of PLS1 (61.4% of the total  
272 covariance,  $r = 0.73$ ), parabolic mandibles are associated with parabolic maxillae. On the negative side,  
273 maxillary and mandibular arcades are U-shaped (Fig. 3a). Humans have positive PLS1 scores and are  
274 separated from the apes that have decreasing scores along a diagonal line, starting at chimpanzees, to  
275 gibbons, orang-utans and then gorillas.

276 At the negative extreme of PLS2 (21.6% of the total covariance,  $r = 0.81$ ) large maxillary canines  
277 are associated with large mandibular canines. The postcanine dentition is relatively small, short and  
278 parallel. At the positive extreme, small mandibular canines and V-shaped arcades are associated with  
279 small maxillary canines and arcades that are more rounded than the mandibles (Fig. 3b). Humans have  
280 positive PLS2 scores and are separated from the apes along a diagonal line.

281 At the negative end of PLS3 (7.9% of the total covariance,  $r = 0.73$ ), both arcades are broad and  
282 converge distally (more pronounced in the maxilla). In the maxillary arcade, the front teeth lie on the  
283 bicanine line. At the positive end of PLS3, both upper and lower arcades are V-shaped. The anterior  
284 region is narrow and projecting anteriorly (more pronounced in the maxilla) (Fig. 3c). Separation among  
285 groups along the PLS3 axis is less clear.

286 In all PLS dimensions the specimens of all five groups scatter around a diagonal line, indicating  
287 that the patterns of covariation of all groups are similar along these PLS axes. In PLS2 *Hylobates*  
288 specimens are shifted away from the hominid trajectory, but their trajectory has the same slope.

289 - Figure 3 -

290 - Table 3 -

291 Pattern of covariation (groups separate, sex-mean centered) Table 3 shows the percentages of  
292 the explained covariance for the first five PLS components for each group. At the positive end of PLS1, in  
293 all groups, rounded maxillae are associated with broad and rounded mandibles (Fig. 4 a-e). In the apes,  
294 the maxillary arcade is anteriorly wider than the mandibular arcade. At the negative end of PLS1, long  
295 and straight postcanine maxillary arcades are associated with straight mandibular postcanine arcades  
296 that taper slightly in the premolar region. In *Hylobates*, PLS1 and PLS2 are interchanged, so that the  
297 aforementioned shape changes are described by PLS2 (SOM Fig. S4). In addition, the maxillary arcade in  
298 *Hylobates* also tapers in the premolar region. The anterior regions in both upper and lower arcades are

299 rounded and extend anteriorly. PLS scores of PLS2 and PLS3 and wireframes of the first three PLS  
300 components are shown in SOM Figs. S4 and S5.

301 At the positive end of PLS2 in all groups, straight diverging posterior maxillary arcades are  
302 associated with likewise diverging mandibular arcades. At the negative end of PLS2, straight converging  
303 posterior maxillary tooth rows are associated with mandibular tooth rows that behave similarly. These  
304 shape changes are described by PLS1 in *Hylobates*. In addition, on the positive side of this PLS  
305 component, *Hylobates* is more straight than converging. In *Homo*, PLS2 and PLS3 are interchanged. The  
306 incisal part of the upper and lower arcades in all groups are flat, with the exception of *Homo* and  
307 *Hylobates*, where the mandibular arcades are more rounded anteriorly (SOM Figs. S4 and S5).

308 At the negative end of PLS3 in all groups, straight posterior tooth rows and flat anterior teeth in  
309 the maxilla are associated with likewise shaped mandibular arcades. In *Pan*, *Gorilla* and *Pongo*, the  
310 postcanine tooth rows are parallel, in *Homo* and *Hylobates* the arcades diverge in both upper and lower  
311 jaws. At the positive end of PLS3, anteriorly extending incisors and large diastemata in the maxilla are  
312 associated with long straight mandibular arcades, where the incisors extend anteriorly as well. In *Homo*  
313 both arcades are more rounded in the postcanine row and less pointed in the anterior region (SOM Figs.  
314 S4 and S5).

315 Overall, all groups show similar covariation patterns.

316

317 Morphological comparison between the variation within blocks and the covariation between  
318 blocks The associated shape (co)variation between the maxillary arcades and the mandibular arcades  
319 along PLS1 and PLS2 is comparable to the shape variation within the maxillae and within the mandibles  
320 along PC1 (i.e., parabolic arcades with small canines versus long U-shaped arcades with large canines).

321 Also the shape changes along PLS3 are partly represented by the variation described by PC3 (broad  
322 versus V-shaped arcades).

323

324 Magnitude of covariation (all groups combined) Results for the covariance ratio (CR) are given  
325 in Table 4. The distribution of values for the permuted CRs from varied sample composition is shown  
326 in Fig. 5. The apes overlap completely with CRs varying from 0.71 to 0.89. *Homo* shows values between  
327 0.64 and 0.89. Median values are 0.81 (*Pongo*), 0.81 (*Pan*), 0.80 (*Gorilla*), 0.81 (*Hylobates*), and 0.76  
328 (*Homo*). *Homo* is significantly different from every other group ( $p < 0.02$ , Bonferroni corrected). *Pan*,  
329 *Gorilla*, *Pongo*, and *Hylobates* are not significantly different from each other ( $p > 0.12$  at least) (Table 4).

330 The correlation coefficient of PLS1 is lowest in *Gorilla*, followed by *Homo*, *Pongo*, and *Pan*.  
331 *Hylobates* show the highest value (Table 5). The correlation coefficient of PLS2 is lowest in *Hylobates*,  
332 followed by *Homo*. *Pan*, *Pongo*, and *Gorilla* show higher values. In PLS3, the correlation coefficient is  
333 lowest in *Hylobates* and *Homo*. *Gorilla* falls between the latter two and *Pongo* and *Pan*.

334 - Figure 5 -

335 - Table 4 -

336 - Table 5 -

337 Magnitude of covariation (groups separate) We used a subsample of 15, i.e., the smallest  
338 number of subsamples (female *Homo*) minus one, and considered the bootstrapping distribution  
339 resulting from 1000 iterations for the calculation of the CR (Table 6). Correlation coefficients are given in  
340 Table 7.

341 Overall, female *Pan*, *Gorilla*, *Pongo* and *Homo* show higher mean values in the covariance ratio  
342 than their male conspecifics (SOM Fig. S6) in the CR. In *Hylobates*, however, the males show higher  
343 values.

344 In PLS1 female *Gorilla*, *Pongo*, and *Homo* show higher correlation coefficients than in their male  
345 counterparts. *Hylobates* show the opposite pattern, so do male *Pan* even though the difference from  
346 the females is small. In PLS2 male *Pan*, *Gorilla*, and *Homo* show higher values than the females. *Pongo*,  
347 *Homo*, and *Hylobates* show the opposite pattern. In PLS3 males show higher values than females in all  
348 groups.

349 - Table 6 -

350 - Table 7 -

### 351 3. Allometry

352 For the maxillary arcades, size explains 14.3% of the total variance in *Gorilla*, 12.6% in *Pongo*,  
353 6.8% in *Pan*, 2.5% in *Homo*, and 2.4% in *Hylobates* ( $p < 0.01$ ). For the mandibular arcades size explains  
354 8.5% of the total variance in *Gorilla*, 9.7% in *Pongo*, 4.8% in *Pan*, 2.4% in *Homo*, and 3.6% in *Hylobates*  
355 ( $p < 0.01$ ) (Table 8).

356

### 357 4. Sexual dimorphism

358 Mean arcade shapes of female and males differ significantly (all  $p$ -values smaller than 0.05) in  
359 *Gorilla*, *Pan* and *Pongo*. While the mandible shapes of female and male *Homo sapiens* are not  
360 statistically different, the maxillae fall just below the significance level ( $p = 0.047$ ). There is no significant  
361 shape difference between female and male *Hylobates* (Fig. 6).



362 The main difference between male and female great apes is basal crown area of the canine. In  
363 *Pongo* and *Gorilla*, this is associated with small differences in the anterior region. In females, the incisors  
364 are shifted slightly posteriorly. In the postcanine dentition, the premolars are shifted anteriorly, whereas  
365 the molar region remains unaffected. In humans, the subtle difference is also canine crown area.

366 - Figure 6 -

#### 367 5. Pattern and magnitudes of covariation between the premaxilla and the maxilla

368 Pattern of covariation (all groups, group-mean centered) Table 9 shows the percentages of  
369 explained covariance for the first five PLS dimensions. In PLS1 (84.1% of the total covariance,  $r = 0.96$ ),  
370 all groups scatter along a diagonal (Fig. 7a). At the negative end of PLS1 (gorillas followed by the other  
371 non-human apes) large incisors are placed far anteriorly, creating a diastema between the maxilla and  
372 the premaxilla. The posterior dentition is parallel, and canines are large. At the positive end (humans),  
373 incisors and canines are small and incorporated in the dental arcade, and the posterior arcade is  
374 parabolic (Fig. 7a).

375 In PLS2 (10.2% of the total covariance,  $r = 0.59$ ), the groups still cluster along the diagonal, but  
376 are shifted parallel from each other with large overlap among groups (Fig. 7b). At the negative end of  
377 PLS2, large spaciouly arranged incisors are associated with rounded posterior arcades and there is no  
378 diastema. At the positive end of PLS2, smaller incisors are positioned in closer proximity and are placed  
379 anteriorly followed by a diastema and the posterior dentition is straight and parallel (Fig. 7b).

380 - Figure 7 -

381 - Table 9 -

382 Pattern of covariation (groups separate, sex-mean centered) Table 9 shows the percentages of  
383 explained covariance for the first five PLS components for each group separately. SOM Figure S7 shows  
384 the PLS scores for every group in the first three PLS dimensions.

385 At the negative end of PLS1 for every group, anteriorly expanding incisors are associated with  
386 straight, parallel posterior tooth rows. In *Pan*, *Gorilla*, and *Pongo*, diastemata are present. At the  
387 positive end there are no diastemata between the premaxillae and the maxillae. The incisors are  
388 incorporated in the dental arcade. The posterior row is more rounded (SOM Fig. S8, top row).

389 In the negative direction of PLS2, rather flat anterior regions are associated with wide, more  
390 rounded posterior arcades. At the positive end, the incisal region extends anteriorly, creating a diastema  
391 between the premaxilla and the maxilla. The posterior arcade is straight. *Gorilla* differs in showing  
392 straight posterior arcades in both directions; *Homo* shows no diastema and the posterior arcades are  
393 parabolic in both directions (SOM Fig. S8, middle row).

394 PLS3 describes the spacing of the incisors. In the negative direction, incisors are narrowly  
395 positioned. In the positive direction, incisors are widely spaced (SOM Fig. S8, bottom row).

396 The associated shape changes along all three PLS dimensions in the separate PLS analysis  
397 correspond to the shape changes observed in the combined analysis.

398

399 Magnitudes of covariation (all groups) Magnitudes of covariation (covariance ratio) are slightly  
400 smaller between the premaxilla and the maxilla than between the upper arcade and the lower arcade in  
401 the apes (Fig. 8, Table 10). In *Homo*, the values are comparable.

402 *Gorilla* is significantly different from *Homo* and *Hylobates* ( $p < 0.015$ , Bonferroni corrected), but  
403 not from *Pongo*. The difference between *Gorilla* and *Pan* is close to the significance level ( $p \leq 0.0559$ ).  
404 *Pan*, *Pongo*, *Homo*, and *Hylobates* are not significantly different from each other ( $p > 0.27$  at least) (Table  
405 10).

406 The correlation coefficient in PLS1 is lowest in *Pan* and *Hylobates*, followed by *Pongo*, *Homo* and  
407 *Gorilla* (Table 11). In PLS2, the correlation coefficient is lowest in *Gorilla*, followed by *Pongo*. *Homo* and

408 *Hylobates* show similar values. *Pan* shows the highest correlation coefficient. In PLS3, *Gorilla* shows the  
409 highest value, followed by *Pongo*, *Pan*, and *Hylobates*. *Homo* shows the lowest correlation coefficient.

410 - Figure 8 –

411 - Table 10 –

412 - Table 11 -

413 Magnitudes of covariation (groups separate) We used a subsample of 15, i.e., the smallest  
414 number of subsamples (female *Homo*) minus one, and considered the bootstrapping distribution  
415 resulting from 1000 iterations for the calculation of the CR (SOM Table S2). Correlation coefficients are  
416 given in SOM Table S3.

417 *Pan* and *Gorilla* show a marked sexual dimorphism with females showing lower values than  
418 males in the covariance ratio (SOM Table S2). In *Pongo*, *Homo*, and *Hylobates*, male and female  
419 distributions overlap. In all groups, magnitudes of covariation are in general slightly smaller between the  
420 premaxilla and the maxilla than between the upper and the lower arcades. Only male *Gorilla* show  
421 higher magnitudes (SOM Fig. S9).

422 In PLS1 male *Pan*, *Gorilla*, *Hylobates* show higher correlation coefficients than their female  
423 counterparts. *Homo* shows the opposite pattern, in *Pongo* values are similar (SOM Table S3). In PLS2  
424 male *Pan*, *Gorilla*, and *Pongo* show higher values than females. In *Homo* and *Hylobates* the opposite is  
425 true. In PLS3 correlation coefficients are low in all groups except for *Pan*, where males show higher  
426 values than females.

427

428 **Discussion**

429           In this study we sought to explore the pattern and magnitude of the covariation between the  
430 upper and the lower jaw, as well as between the premaxilla and the maxilla in extant hominoids. In the  
431 first three PLS components, that together explain more than 90% of the total covariance of the upper  
432 and the lower arcade, the data points scatter along the diagonal (Fig. 3). Such an arrangement is  
433 interpreted as the same pattern of covariation (e.g. Mitteroecker and Bookstein, 2008). While retaining  
434 the same slope, *Hylobates* are shifted from the hominid trajectory in PLS2 (Fig. 3b), probably owing to  
435 the presence of absolutely and relatively large canines in males and females in this group.

436           We also performed separate PLS analyses which showed that the related shape changes of the  
437 mandibular and the maxillary arcades are similar across species (Fig. 4 and SOM Figs. S4 and S5). In  
438 general, this result suggests a common hominoid pattern of covariation and that the evolutionary shape  
439 changes that led to hominoid arcade variation are not associated with modification of the underlying  
440 covariation of arcades. Importantly, our data demonstrate an independence of the pattern of  
441 covariation, which is similar in the analyzed species, from dental spatial arrangements, which are  
442 different in the analyzed species. This conclusion is in accordance with earlier studies that suggest  
443 conserved covariation patterns in primates and other mammals in cranial parts other than the jaws  
444 (Cheverud, 1996b; Ackermann and Cheverud, 2000; Lieberman et al., 2000; Marroig and Cheverud,  
445 2001; Ackermann, 2002; Gonzalez-Jose et al., 2004; Marroig et al., 2004; Ackermann, 2005; Goswami,  
446 2006; Gunz and Harvati, 2007; Mitteroecker and Bookstein, 2008; Porto et al., 2009; Makedonska et al.,  
447 2012; Villmoare et al., 2014).

448           To evaluate the magnitude of the covariation we used the covariance ratio (CR, Adams, 2016)  
449 and the correlations between upper and lower arcades, or the premaxilla and the maxilla, respectively.  
450 For the CR we considered the distributions from permutations. *Homo* stands out in showing lower  
451 overall CR values (Fig.5, Table 4). Covariation magnitudes of individual PLS axes are quantified by the  
452 correlation coefficient. When the first PLS is considered, which explains almost two thirds of the total

453 covariance (i.e., arcade shapes from U-shaped to parabolic), *Gorilla* shows the weakest covariation,  
454 followed by *Homo*, *Pongo*, and *Pan* which show similar covariation magnitudes. *Hylobates* has slightly  
455 higher values (Table 5). Thus the magnitude of covariation does not reflect the shape gradient from U-  
456 shaped to parabolic, and might be more taxon specific than arcade shape specific. In PLS2, which  
457 explains almost 22% of the total covariance (i.e., canine size), *Homo* and *Hylobates* display lower  
458 correlations than the other groups. Interestingly, while it seems as if absolute canine size itself is of  
459 minor importance for the magnitude of covariation, this result might indicate that sexual dimorphism in  
460 canine size contributes to the magnitude of covariation, in a way that less dimorphic taxa show lower  
461 magnitudes of covariation. In PLS3, that explains about 8% of the total covariance (i.e., spatial  
462 arrangement of anterior dentition and postcanine divergence), *Homo* and *Hylobates* also show lower  
463 values, followed by *Gorilla* which falls in-between the latter and *Pan* and *Pongo* (Fig. 5d). These  
464 differences in covariation magnitudes might be related to different strategies of incorporating large  
465 canines into the dental arcade. While in *Pan*, *Pongo* and *Gorilla* intercanine distance is enlarged, which  
466 results in a flat anterior region, in *Hylobates* the incisal region is extended anteriorly, resulting in a V-  
467 shaped arcade. The lower magnitudes in *Homo* might be the consequence of the generally reduced  
468 canines. In combination with the results for the overall measure for the magnitude of covariation these  
469 findings imply that the lower magnitude in modern humans is not a reflection of the large-scale  
470 differences between humans and apes.

471         While hominoids do show variation in dental morphology and dental size proportions, the  
472 pattern of the interplay between upper and lower arcades remained generally unchanged. This suggests  
473 that selection acted on maintaining a proper fit between the upper and the lower arcade. The lower  
474 magnitudes of maxillo-mandibular covariation in modern humans which we report here may be related  
475 to a systemic change in the hominin lineage. It was shown previously that magnitudes of covariation in  
476 the cranium and the pelvis are relaxed in modern humans in general compared to other primates and

477 mammals (Marroig et al., 2009; Porto et al., 2009; Grabowski et al., 2011). Following this argument, a  
478 relaxation initially allowed new body plans characteristic of hominins to emerge, potentially including  
479 the development of the parabolic arcade shape. If lower magnitudes can account for the development  
480 of hominin morphologies then they should be identifiable before new morphologies appear, i.e., at or  
481 shortly after the split from the chimpanzee lineage. While the resolution of the current fossil record  
482 might not be good enough to determine when this relaxation occurred in the hominin lineage, fossil  
483 evidence from about 2.1 to 1.5 million years ago suggests that bipedal adaptations, brain enlargement  
484 and short and rounded dental arcades are not necessarily associated with each other. While brain size of  
485 *H. habilis*, *H. rudolfensis* and *H. erectus* largely overlap in range, they show distinct facial morphologies  
486 including the primitive morphology of long and parallel post-canine tooth rows in *H. habilis*, the derived  
487 condition in *H. erectus*, and the distinct morphology of *H. rudolfensis* with short and parallel post-canine  
488 tooth rows plus a flat anterior region (Spoor et al., 2015). While these seemingly conflicting lines of  
489 evidence clearly need more data, another, mutually non-exclusive, explanation for a lower covariation in  
490 *Homo* compared to all apes could be diet. It has been found that those platyrrhine species consuming  
491 softer foods show lower magnitudes of covariation than species reliant on harder foods (Marroig and  
492 Cheverud, 2001; Makedonska et al., 2012). A similar relationship could contribute to the difference we  
493 found between apes and modern humans. This would be in accordance with the notion that at least in  
494 *H. sapiens* the masticatory effort was reduced due to extra-oral food processing via tool use and cooking  
495 (e.g. Brace et al., 1987; Wrangham et al., 1999; Richards et al., 2001; Teaford et al., 2002; Henry, 2010).  
496 This explanation, however, does not sufficiently explain the differences we observe between ape  
497 genera, which show similar magnitudes of covariation but at the same time different dietary habits.

498         We also analyzed the pattern and magnitudes of covariation between the premaxilla and the  
499 maxilla. Corroborating the results of Villmoare et al. (2014) the magnitudes of covariation between the  
500 premaxilla and the maxilla are in general slightly lower than between the upper and the lower jaw.

501 In the first PLS components, all groups follow a similar pattern of covariation between the  
502 premaxilla and the maxilla. When canines are large, incisors are placed anteriorly to create a diastema  
503 for the lower canine. The posterior dentition is long and parallel. With the reduction of canine and  
504 diastema size, incisors move posteriorly and the posterior arcades get more parabolic (PLS1). This  
505 finding of a shared pattern of covariation between the premaxilla and the maxilla as well as *Homo* falling  
506 within the apes' range of covariation magnitude is in accordance with Villmoare et al. (2014). Either  
507 result shows that the variation in hominin premaxilla is not associated with a change in the pattern of  
508 character covariation. Previous studies suggested that there is a relation between anterior tooth size  
509 and premaxilla size (Bromage, 1989; Mooney and Siegel, 1991; Lockwood, 1997; Braga, 1998). Our  
510 results show that canine and diastema size contributes substantially to maxillary arcade shape.

511 To assess differences related to sexual dimorphism, we evaluated magnitudes of covariation for  
512 females and males in each group between the upper and the lower jaw (SOM Fig. S6) and between the  
513 premaxilla and the maxilla (SOM Fig. S9). Magnitudes of the latter overlap almost completely in all  
514 groups except for *Pan* and *Gorilla*, where males show higher values. Mean magnitudes between the  
515 upper and the lower jaw are higher in females except for *Hylobates*, where males have higher values.  
516 Due to small sample sizes in the within-group comparisons these results should be treated with caution.  
517 Additionally, an uncertainty about modern human sex attribution must be considered, since their sexing  
518 is based on traditional morphological methods, rather than actual records (dissection, autopsy or  
519 graveyard). In general, it is probable that the mechanisms responsible for the results presented here  
520 might be multiple and different for each taxon as there are different patterns and magnitudes of  
521 dimorphism between species (Plavcan, 2002).

522 Our results suggest that variation in hominoid arcade shape is not the consequence of a change  
523 of the underlying covariation patterns. Our results provide a model for the evolutionary arcade shape

524 change in the hominin lineage: with the loss of interlocking canines and the decreased need for space in  
525 the antagonistic arcade in the hominin lineage, the tooth rows become more rounded and eventually  
526 parabolic. Driven by the necessity of the upper and the lower jaw to form an effective masticatory  
527 apparatus, the morphological covariation between the mandibular and the maxillary arcade has been  
528 maintained.

529           The strong focus on dentognathic structures in palaeoanthropology is due to their good  
530 preservation and their taxonomic significance. Nonetheless, our limited understanding of the degree  
531 and pattern of intraspecific variation has resulted in different views regarding the number of species  
532 that can be recognized in the fossil record (e.g. Wood, 1992; Rightmire, 1993; Leakey et al., 2012).  
533 Another factor adding to differences in hypodigm composition is that there is a lack of consensus over  
534 which maxillae and mandibles can be accommodated within a single species (Spoor et al., 2015). The  
535 results of the current study will help address these issues, as they provide reference data which  
536 document the intraspecific and interspecific variation of the upper and lower dental arcades in extant  
537 taxa. Moreover, the degree and pattern of covariation between upper and lower jaws can help with  
538 identifying the probability that certain maxillae and mandibles in the fossil record are conspecific. Using  
539 a smaller landmark set, we have shown recently that this approach can indeed be used to recognize  
540 distinct morphs within early *Homo*, identifying those pairs of maxillae and mandibles which, based on  
541 extant hominid variation, can be excluded statistically from belonging to a single species (Spoor et al.,  
542 2015).

543

#### 544 **Conclusion**

545           We found that *Pan*, *Gorilla*, *Pongo*, *Hylobates*, and *Homo* share a very similar covariation pattern  
546 between the upper and the lower arcade, as well as between the premaxilla and the maxilla, that is



547 independent from dental spatial arrangements. We suggest that in the hominin lineage, with the  
548 reduction of canine and diastema size, the incisors move posteriorly and the whole arcade becomes  
549 parabolic. The upper and the lower arcades keep track of each other through high magnitudes of  
550 covariation. Modern humans show lower magnitudes of covariation between the upper and the lower  
551 jaw compared to apes, which might be related to a mechanically less challenging diet in living humans.  
552 Alternatively or additionally, lower magnitudes might follow from a general relaxation in covariation  
553 attributable to a systemic change in the hominin lineage that enabled the development of new shapes in  
554 the first place.

555

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570

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829 **Figure captions**

830 Figure 1. Data acquisition. Landmarks are placed on the alveolar margin (blue) and on the cervix  
831 of the postcanine dentition (orange) of the maxilla (a) and the mandible (c); b) and d) show the  
832 wireframe of the symmetrized data.

833 Figure 2. Arcade shape space of maxillae (a, b) and mandibles (c). *Homo* is shown in yellow, *Pan*  
834 in blue, *Gorilla* in black, *Pongo* in red, and *Hylobates* in green. The wireframes show the shape variation  
835 along PC1, PC2 and PC3. In PC3, *Hylobates* mandibles plot similarly to the maxillae and are therefore not  
836 shown.

837 Figure 3. Two-block partial least squares (PLS) analysis between the maxillary and the  
838 mandibular arcade. Group colors as in Figure 2. The wireframes show the shape changes along SW1 (a),  
839 SW2 (b), and SW3 (c). All groups scatter along the diagonal. *Hylobates* shows a shift along SW2, parallel  
840 with the hominid trajectory.

841 Figure 4. Two-block partial least squares (PLS) analysis between the maxillary and the  
842 mandibular arcade for each group separately: *Pan* (a), *Gorilla* (b), *Pongo* (c), *Homo* (d), and *Hylobates*  
843 (e). Males are shown in dark, females in light colors. The wireframes show the shape changes along  
844 SW1.

845 Figure 5. Distribution of the covariance ratio (CR) (sample size 30, 1000 iterations). *Homo* shows  
846 lower overall values than the apes

847 Figure 6. Female (red) and male (blue) mean shapes for the maxilla (a-e) and the mandible (f-j).  
848 Mean shape differences statistically significant at  $\alpha \leq 0.05$ ; *p*-values given below the wireframe. Female  
849 and male arcades are significantly different between *Pan*, *Gorilla*, *Pongo*, and *Homo* (maxillae). *Homo*  
850 mandibles and *Hylobates* are not significantly different. Smaller canines (females) are associated with a

851 more posteriorly placed anterior arcade. In the postcanine row, the premolars are placed more  
852 anteriorly, the molars remain unaffected.

853 Figure 7. PLS analysis between the premaxilla (anterior, black wireframe) and the maxilla  
854 (posterior, red wireframe). Group colors as in Figure 2.

855 Figure 8. Distributions of the CR (sample size 30, 1000 iterations) between the premaxilla and  
856 the maxilla.

857 SOM Figure S1. Species distribution within the genus *Gorilla* in arcade shape space of maxillae  
858 (a, b) and mandibles (c). *G. gorilla* is shown in black, *G. beringei* in gray. The wireframes show the shape  
859 variation along PC1, PC2 and PC3.

860 SOM Figure S2. Species distribution within the genus *Pongo* in arcade shape space of maxillae (a,  
861 b) and mandibles (c). *P. pygmaeus* is shown in light red, *P. abelii* in yellow, *Pongo* sp. in dark red. The  
862 wireframes show the shape variation along PC1, PC2 and PC3.

863 SOM Figure S3. Species distribution within the genus *Hylobates* in arcade shape space of  
864 maxillae (a, b) and mandibles (c). *H. muelleri* is shown in green, *H. lar* in pink, *H. klossii* in orange, *H.*  
865 *albibarbis* in blue, *H. agilis* in yellow. The wireframes show the shape variation along PC1, PC2 and PC3.

866 SOM Figure S4. Shape changes along SW1, SW2, SW3 between the maxillary arcade (black) and  
867 mandibular arcade (red) for each group separately. The left wireframe in each group is the negative  
868 extreme, the right wireframe is the positive extreme for each SW.

869 SOM Figure S5. PLS scores of the maxillary and the mandibular arcade for each group  
870 separately. Dark colors in each group are males, light colors females.

871 SOM Figure S6. Sexual dimorphism in the magnitude of covariation. For each taxon, distributions  
872 (sample size 15, 1000 iterations) and single values (horizontal lines) are given. Light group colors are

873 females, dark group colors are males. Female hominids show higher magnitudes (distributions and  
874 single values) than their male counterparts. *Hylobates* show the opposite pattern. In *Pan*, the single  
875 values are almost the same.

876 SOM Figure S7. PLS scores of the premaxilla and the maxilla for each group separately. Dark  
877 colors in each group are males, light colors females.

878 SOM Figure S8. Shape changes along SW1 (a), SW2 (b), and SW3 (c) between the premaxilla  
879 (black) and the maxilla (red). The left column is the negative extreme, the right column is the positive  
880 extreme.

881 SOM Figure S9. Sexual dimorphism in the covariance ratio (CR) between the premaxilla and the  
882 maxilla. For each taxon distributions subsample size is 15 (1000 iterations); light group colors are  
883 females, dark group colors are males.

884

Table 1. Sample composition

<b>Taxon</b>	♀	♂	<b>Unknown sex</b>
<i>Pan</i>	25	19	-
<i>Homo</i>	16	24	13
<i>Gorilla</i>	22	31	-
<i>Pongo</i>	25	27	-
<i>Hylobates</i>	29	30	4

Table 2. Results of the PC analyses.

	<b>Eigenvalues</b>	<b>% variance</b>	<b>Cumulative %</b>
<b><i>Maxillae</i></b>			
PC1	0.00789125	66.958	66.958
PC2	0.00129874	11.02	77.978
PC3	0.00067848	5.757	83.735
<b><i>Mandibles</i></b>			
PC1	0.01059464	70.795	70.795
PC2	0.00163547	10.928	81.723
PC3	0.00093735	6.264	87.987



Table 3. Percentage of the explained covariance between the upper and the lower arcade for the first five PLS components. Symmetrized data; combined PLS: group-mean centered; separate PLS: sex-mean centered.

% covariance combined PLS		% covariance separate PLS				
		<i>Pan</i>	<i>Gorilla</i>	<i>Pongo</i>	<i>Homo</i>	<i>Hylobates</i>
PLS1	61.39	68.44	51.79	57.80	65.53	79.40
PLS 2	21.55	11.74	19.31	15.00	13.27	8.78
PLS 3	7.92	6.26	13.06	10.85	6.55	4.44
PLS 4	3.46	3.50	3.83	6.38	3.41	1.36
PLS 5	1.38	2.60	2.65	2.39	2.09	1.27

Table 4. Results for the CR and the permutation tests for the upper and the lower jaw. Subsample size for the permutation is 30, 1000 resamples. Below, Bonferroni corrected  $p$ -values from permutation test of group mean differences (considered significant at  $\alpha < 0.05$ ).

	<i>Pan</i>	<i>Gorilla</i>	<i>Pongo</i>	<i>Homo</i>	<i>Hylobates</i>
<b>CR</b>	0.777	0.771	0.786	0.726	0.789
Min	0.712	0.715	0.739	0.635	0.721
Max	0.894	0.867	0.886	0.892	0.891
Median	0.805	0.804	0.812	0.755	0.814
Mean	0.805	0.802	0.813	0.757	0.814
SD	0.031	0.025	0.024	0.044	0.030
<i>Pan</i>	15	14.580	0.165	0.015	0.150
<i>Gorilla</i>	14.536	15	0.270	0.015	0.165
<i>Pongo</i>	0.120	0.240	15	0.015	11.613
<i>Homo</i>	0.015	0.015	0.015	15	0.015
<i>Hylobates</i>	0.210	0.150	10.984	0.015	15

Table 5. Results for the correlation coefficient ( $r$ ) between the upper and the lower arcade.

	PLS1	PLS2	PLS3
<b><i>Pan</i></b>	0.757	0.841	0.858
<b><i>Gorilla</i></b>	0.645	0.853	0.757
<b><i>Pongo</i></b>	0.728	0.851	0.828
<b><i>Homo</i></b>	0.695	0.664	0.668
<b><i>Hylobates</i></b>	0.831	0.569	0.625

**Table 6**

Table 6. Results for the CR and the permutation tests grouped by species and sex (subsample size 15, 1000 resamples) for the upper and the lower arcade. Below, Bonferroni corrected  $p$ -values from permutation test of sex mean differences (considered significant at  $\alpha < 0.05$ ).

	<i>Pan</i> M	<i>Pan</i> F	<i>Gorilla</i> M	<i>Gorilla</i> F	<i>Pongo</i> M	<i>Pongo</i> F	<i>Homo</i> M	<i>Homo</i> F	<i>Hylo</i> M	<i>Hylo</i> F
<b>CR</b>	0.837	0.831	0.720	0.809	0.771	0.834	0.756	0.859	0.836	0.795
Min	0.786	0.797	0.682	0.749	0.737	0.778	0.715	0.843	0.771	0.719
Max	0.926	0.921	0.912	0.910	0.893	0.933	0.904	0.872	0.940	0.935
Median	0.847	0.872	0.804	0.840	0.809	0.858	0.793	0.864	0.862	0.844
Mean	0.853	0.869	0.803	0.839	0.810	0.857	0.796	0.862	0.861	0.842
SD	0.030	0.023	0.038	0.029	0.023	0.031	0.033	0.007	0.032	0.040
$p$	0.950		0.002		0.002		0.002		0.214	

Table 7. Correlation coefficients ( $r$ ) grouped by species and sex for the upper and the lower arcade.

	PLS1	PLS2	PLS3
<i>Pan</i> M	0.924	0.810	0.879
<i>Pan</i> F	0.910	0.780	0.827
<i>Gorilla</i> M	0.660	0.906	0.876
<i>Gorilla</i> F	0.764	0.780	0.800
<i>Pongo</i> M	0.764	0.703	0.797
<i>Pongo</i> F	0.852	0.784	0.705
<i>Homo</i> M	0.745	0.858	0.860
<i>Homo</i> F	0.907	0.741	0.806
<i>Hylobates</i> M	0.865	0.779	0.802
<i>Hylobates</i> F	0.787	0.837	0.728

Table 8. Percentage of the total variance explained by size.

Taxon	Maxillae		Mandibles	
	%	<i>p</i>	%	<i>p</i>
<i>Pan</i>	6.80	<0.001	4.84	<0.001
<i>Gorilla</i>	14.25	<0.001	8.53	<0.001
<i>Pongo</i>	12.58	<0.001	9.68	<0.001
<i>Homo</i>	2.49	<0.001	2.38	<0.004
<i>Hylobates</i>	2.36	<0.001	3.55	<0.001
Total	5.38	<0.001	8.39	<0.001

Table 9. Percentage of the explained covariance between the premaxilla and the maxilla for the first five PLS components. Symmetrized data; combined PLS: group-mean centered; separate PLS: sex-mean centered.

% covariance combined PLS		% covariance separate PLS				
		<i>Pan</i>	<i>Gorilla</i>	<i>Pongo</i>	<i>Homo</i>	<i>Hylobates</i>
PLS1	84.14	44.28	85.99	67.19	79.75	74.83
PLS2	10.18	39.06	7.37	17.07	7.72	13.70
PLS3	2.20	6.44	2.21	6.47	5.41	3.95
PLS4	1.16	3.55	1.42	3.87	2.50	3.11
PLS5	0.74	1.96	1.06	1.85	1.42	1.27

Table 10. Results for the CR and the permutation tests between the premaxilla and the maxilla. Subsample size for the permutation is 30. 1000 resamples. Below, Bonferroni corrected  $p$ -values from permutation test of group mean differences (considered significant at  $\alpha < 0.05$ ).

	<i>Pan</i>	<i>Gorilla</i>	<i>Pongo</i>	<i>Homo</i>	<i>Hylobates</i>
<b>CR</b>	0.678	0.731	0.702	0.694	0.646
Min	0.611	0.621	0.620	0.596	0.576
Max	0.803	0.882	0.844	0.834	0.809
Median	0.707	0.751	0.727	0.719	0.688
Mean	0.707	0.749	0.726	0.719	0.687
SD	0.031	0.044	0.037	0.038	0.042
<i>Pan</i>	15	0.030	8.571	5.784	0.300
<i>Gorilla</i>	0.060	15	0.135	0.015	0.015
<i>Pongo</i>	8.796	0.120	15	3.267	0.270
<i>Homo</i>	5.619	0.015	3.282	15	2.742
<i>Hylobates</i>	0.554	0.015	0.360	2.892	15



Table 11. Results of the correlation coefficient ( $r$ ) between the premaxilla and the maxilla.

	PLS1	PLS2	PLS3
<b><i>Pan</i></b>	0.752	0.885	0.395
<b><i>Gorilla</i></b>	0.815	0.707	0.668
<b><i>Pongo</i></b>	0.798	0.752	0.492
<b><i>Homo</i></b>	0.814	0.794	0.060
<b><i>Hylobates</i></b>	0.758	0.796	0.207