A new dimeric imidazole alkaloid plasmid conjugation inhibitor from Lepidium sativum

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Abstract

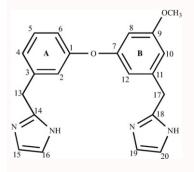
Phytochemical investigation of the methanolic extract of *Lepidium sativum* seeds led to the isolation of a new compound, named 2-(3-(3-((1*H*-imidazol-2-yl)methyl)-5-methoxyphenoxy)benzyl)-1*H*-imidazole and given the trivial name Lepidine AK (1), along with three known compounds; Lepidine E (2), Lepidine B (3) and 2-(3-(2-((1*H*-imidazol-2-yl)methyl)-6-methoxyphenoxy)benzyl)-1*H*-imidazole (4). The structures were elucidated based on NMR spectroscopy, UV, IR and high-resolution electrospray ionization mass spectrometry. The isolated compounds were tested for bacterial conjugation inhibition. Lepidine AK (1, 100 µg/mL) reduced the conjugal transfer of the Incl₂ plasmid TP114 to 44.7 ± 3.5% but interestingly promoted the conjugation of the IncN plasmid pKM101 to greater than 120%.

Keywords: Dimeric; Imidazole; Lepidine; Plasmids; Conjugation

Introduction

Lepidium sativum (garden cress) is an annual herb of the family Brassicaceae. It is a native of Egypt and West Asia but is widely farmed in temperate countries globally for its culinary and medicinal uses.^{1,2} In China, garden cress seeds are used for the treatment of abdominal colic, asthma, pleurisy and dropsy.³ In Africa, Ethiopians use the seeds primarily for treatment of throat diseases, asthma, bronchitis, headache and for baking and as a condiment.^{4,5} The Mauritians use the seeds for the treatment of hiccough and stomachache.⁵ Both seeds and leaves of *L. sativum* are used for the treatment of inflammation, bronchitis, rheumatism and muscular pain in the Unani system of medicine.^{3,6} It is also reported to be useful in the treatment of diabetes, hypertension, cough and bleeding piles.^{7–9} Phytochemical studies on *L. sativum* have shown the presence of essential oil, glucosinolates, alkaloids, sterols, cyanogenic glycosides (traces), flavonoids, tannins, saponins and triterpenes.^{3,6} Bahroun and Damak first reported dimeric imidazole alkaloids for *L. sativum* seeds after which Meinhart Zenk's group isolated five additional analogues.¹⁰

In this paper, we report the isolation and structural characterization by spectroscopic methods of a new dimeric imidazole alkaloid, 2-(3-(3-((1*H*-imidazol-2-yl)methyl)-5-methoxyphenoxy)benzyl)-1*H*-imidazole (1) (Fig. 1) along with three known compounds. The antibacterial and antifungal activities and inhibition of bacterial conjugal transfer of plasmids of the isolated compounds are reported herein for the first time.



Results and discussion

L. sativum seeds (245 g) were obtained from Seed Parade, UK, and defatted with petroleum ether and subsequently extracted with methanol. The MeOH extract was mixed with water, acidified with concentrated HCl and it was then liquid-liquid extracted with EtOAc. The EtOAc fraction was chromatographed over silica gel 60 (gravity column, 70–230 mesh, column size: 50×4 cm). Compound **1** (3.6 mg, *Rf* value: 0.39) was obtained by purification of the column fractions using preparative TLC (silica gel, CHCl₃-MeOH-NH₃, 90:9:1, $^{V}_{V_{V}}$).

Compound **1** was isolated as a pale yellow solid and HRTOFESIMS gave an *m/z* at 361.1659 [M+H]^{*}, which indicated that its molecular formula was $C_{21}H_{20}N_4O_2$. The ¹H and COSY NMR spectra revealed two aromatic rings; di-substituted (A) and tri-substituted (B) aromatic rings. The ¹H data was similar to the known compound **4** (2-(3-(2-((1/H-imidazol-2-yl))methyl)-6-methoxyphenoxy)benzyl)-1*H*-imidazole, only varying in the positioning of the methoxyl and imidazolyl methyl group on the B-ring. The di-substituted aromatic ring was identified by proton resonances at $\delta_{th} 6.70$ (dd, J = 2.0, 1.5 Hz, H-2), 6.86 (dd, J = 8.0, 0.5 Hz, H-4), 7.15 (t, J = 8.0 Hz, H-5), 6.64 (dd, J = 8.0, 2.0 Hz, H-6). The couplings between the aromatic hydrogens, and the meta-coupling of H-2 by H-6 (⁴J_{HH} = 2.0 Hz) confirmed their arrangement on the di-substituted ring. On the B-ring a meta-coupling of a hydrogen signal at δ_{th} 7.01 (d, J = 1.5 Hz, H-10) to δ_{th} 6.85 (s, J = 1.5 Hz, H-12) in the COSY spectrum also indicated the presence of an aromatic spin system, which was 1,3,5-tri-substituted (ring B). Both spin systems were confirmed by the HMQC and HMBC correlations (Table 1, Fig. 2). The aromatic rings were linked by oxygen, which was confirmed by a strong IR absorption band at 1053 cm⁻¹ (the ether functional group). A ³J_{H-C} correlation of the methoxyl hydrogens (3H, s, δ_{h} 3.70) to the aromatic quaternary carbon (δ_{c} 151.9, C-9) revealed the position of the methoxyl group on the aromatic ring B. It was meta-positioned to the imidazolylmethyl moiety and this was supported by the splitting pattern of the aromatic hydrogens signal at δ_{h} 3.97 (2H, s, H_{2} -13), δ_{c} 34.3 (C-13) and δ_{h} 3.95 (2H, s, H_{2} -17), δ_{c} 35.1 (C-17) were identified as the two methylene groups connecting the imidazole rings. The IR spectrum confirmed this with the presence of secondary amine functional group absorbance at 2920 cm⁻¹. The combined NMR, IR and HRTOFESIMS spectra data led t

Position	$\delta_{ m c}$, type	$\delta_{ extsf{H}}$, (<i>J</i> in Hz)	HMBC	
1	159.9, C			
2	117.6, CH	6.70, dd (2.0, 1.5)		
3	141.0, C			
4	123.3, CH	6.86, dd (8.0, 0.5)	C-5	
5	130.6, CH	7.15, t	C-1, C-3	
6	115.6, CH	6.64, dd (8.0, 2.0)		
7	145.6, C			
8	114.5, CH	7.01, s	C-9	
9	151.9, C			
10	126.4, CH	7.01, d (1.5)	C-11	
11	132.5, C			
12	123.0, CH	6.85, d (1.5)		
13	34.3, CH ₂	3.97, s	C-2, C-3, C-4, C-14	
14	148.1, C			
15	123.3, CH	6.91, s	C-14	
16	123.3, CH	6.91, s	C-14	

Table 1 ¹H (500 MHz) and ¹³C NMR (125 MHz) NMR data and HMBC correlations of 1 recorded in methanol-d₄.

17	35.1, CH ₂	3.95, s	C-10, C-11, C-12, C-18
18	148.4, C		
19	123.0, CH	6.91, s	C-18
20	123.0, CH	6.91, s	C-18
O-Me	56.5, CH ₃	3.70, s	C-9

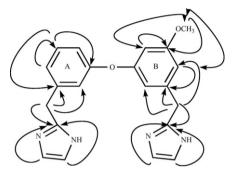


Fig. 2 Key HMBC (single-headed arrows) and NOESY (double-headed arrows) correlations of compound 1.

The compounds were tested for their ability to inhibit bacterial plasmid conjugation¹¹ (Table 2). The essence of this assay is to identify compounds that can inhibit the spread of antibiotic-resistance genes via the bacterial type-IV secretion systems (conjugation machinery). The inhibition of this process is important because of the clinical significance of conjugation machinery in the transfer of toxins and effector proteins directly into eukaryotic target cells, and its involvement in biofilm formation as well as the aforementioned transfer of antibiotic-resistance genes among microorganisms.^{12,13}

Table 2 The effect of the isolated dimeric imidazoles on conjugal transfer of plasmids pKM101 and TP114.

Conjugation pair	Compounds (100 µg/mL)				Controls		
Donor	Recipient	1	2	3	4	Novobiocin (10 µg/mL)	No drug
pKM101ª	ER1793°	120 ± 0.0	31.0 ± 7.0	120 ± 0.0	28.0 ± 5.0	-	100.0 ± 0.0
TP114 ^b		44.0 ± 3.5	26.0 ± 4.0	120 ± 0.0	23.5 ± 4.0	17.0 ± 4.2	100.0 ± 0.0

The values represent the mean transfer frequency (%) ± standard deviation of at least three independent experiments.

^a E. coli strain WP2 bearing plasmid pKM101 (IncN; ampicillin-resistant).

^b E. coli strain K12 J53-2 bearing plasmid TP114 (Incl₂; kanamycin-resistant).

^c E. coli strain ER1793, streptomycin resistant.

It was interesting to note that some of the dimeric imidazoles (compounds 2 and 4) at sub-inhibitory concentration (100 µg/mL) exhibited anti-conjugal activity in *Escherichia coli* while compound 3 at the same concentration enhanced conjugation activity (conjugal transfer frequency, greater than 120%). With compound 1, the activity varied.

Compound **2** reduced the conjugal transfer frequency of the lncN plasmid pKM101 and lncl₂ plasmid TP114 to 31.0 ± 7.0% and 26.0 ± 4.0%, respectively. In comparison to compound **2**, **4** had a slightly better reduction in conjugal transfer frequency, a 28.0 ± 5.0% for lncN plasmid pKM101 and 23.5 ± 4.9% for lncl₂ plasmid TP114. Although the difference in anti-conjugal activity for compounds **2** and **4** is marginal, we suspect that the structural differences of substituent groups on the aromatic ring B of the compounds may have contributed to this. At position 8 of the aromatic ring B, compound **2** has a hydroxyl group attached while compound **4** has a methoxyl group. The imidazolylmethyl moiety is attached to position 11 for compound **2**, while for compound **4** it is at the position 12. A general observation made was that, the isolated compounds were similar to each other but only varied in the positioning of the substituent groups on the aromatic ring B (either an hydroxyl or a methoxyl group and an imidazolylmethyl moiety), and this may have influenced the varied outcome of activity against the conjugal transfer of plasmids in *E. coli*.

The only compound, which showed specificity in activity against the tested plasmids strains, was the new imidazole (1). It exhibited moderate anti-conjugal activity against the Incl₂ plasmid TP114 (transfer frequency 44.0±3.5%) and enhanced the conjugation activity of the IncN plasmid pKM101 to greater than 120%. We found this interesting, as specificity would limit its potential use but promotion may have utility in promoting plasmid transfer, which would be useful in many areas of molecular biology. In conclusion, this finding could serve as a good start point for structural modification for improved anti-conjugal activity with specificity. The development of an anti-conjugal molecule has potential druggability in reducing transfer and spread of resistance and reducing virulence.

Structure elucidation

2-(3-(3-((1H-imidazol-2-yl)methyl)-5-methoxyphenoxy)benzyl)-1H-imidazole (compound 1).

Pale yellow solid; λ_{max} (log ϵ) 218 (3.97), 228 (4.18) nm; IR (film) V_{max} : 2919.76, 1045.02, 971.15, 799.93, 610.64 cm⁻¹; ¹H NMR (500 MHz, methanol- d_4) and ¹³C NMR (125 MHz, methanol- d_4): see Table 1; positive HRTOFESIMS *m/z* 361.1659 [M+H]⁺ (calcd for C₂₁H₂₀N₄O₂, 361.1664).

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A Supplementary data

Supplementary data associated with this article can be found, in the online version, at https://doi.org/10.1016/j.tetlet.2018.04.028.

References

1. S.S. Gokavi, N.G. Malleshi and M. Guo, Plant Foods Hum Nutr 59, 2004, 105–111.

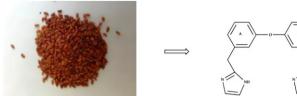
- 2. I.A. Nehdi, H. Sbihi, C.P. Tan and S.I. Al-Resayes, Bioresour Technol 126, 2012, 193–197.
- 3. S. Sharma, N. Agarwal and J. Indian, Nat Prod Resour 2, 2011, 292–297.
- 4. H.J. Kloos, Ethiopian Pharm Assoc 2, 1976, 18–28.
- 5. P.C.M. Jansen, In: G.J.H. Grubben and O.A. Denton, (Eds.), Plant Resources of Tropical African 2. Vegetables, 2004, PROTA Foundation/Backhuys Publishers; Wageningen, Netherlands, 365–367.
- 6. M.H. Ghante, S.L. Badole and S.L. Bodhankar, In: V.R. Preedy, R.R. Watson and V.B. Patel, (Eds.), Nuts and seeds in health and disease prevention, 2011, Academic Press; United States of America, 521–526.
- 7. M. Eddouks, M. Maghrani, N.A. Zeggwagh and J.B. Michel, J Ethnopharmacol 97, 2005, 391–395.
- 8. M. Maghrani, N.A. Zeggwagh, J.B. Michel and M. Eddouks, J Ethnopharmacol 100, 2005, 193–197.
- $\textbf{9.} \text{ R.G. Mali, S.G. Mahajan and A.A. Mehta, \textit{Orient Pharm Exp Med 7}, 2007, 331–335.}$
- 10. U.H. Maier, H. Gundlach and M.H. Zenk, Phytochemistry 49, 1998, 1791–1795.
- 11. L.B. Rice and R.A. Bonomo, In: V. Lorian, (Ed), Antibiotics in laboratory medicine, 2005, Lippincott Williams & Wilkins; USA, 483–484.
- 12. C.E. Alvarez-Martinez and P.J. Christie, Microbiol Mol Biol Rev 73, 2009, 775–808.
- 13. E.L. Zechner, S. Lang and J.F. Schildbach, Philos Trans R Soc Lond B Biol Sci 367, 2012, 1073–1087.

A Supplementary data

Multimedia Component 1

Supplementary data 1

Graphical abstract



Highlights

- A new dimeric imidazole alkaloid was isolated from the seeds of Lepidium sativum.
- · Structure elucidation was achieved by analysis of the NMR spectroscopic and mass spectral data.
- The new compound, named lepidine AK (1), reduced the conjugal transfer of the Incl₂ plasmid TP114 to 44.7%.
- Interestingly, compound 1 also promoted the conjugation of the IncN plasmid pKM101 to greater than 120%.
- Plasmid transfer inhibitors may have utility in reducing the spread of antibiotic resistance genes and bacterial virulence.

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