Fossil biomass preserved as graphitic carbon in a late Paleoproterozoic banded

iron formation metamorphosed at more than 550°C DOMINIC PAPINEAU^{1,2,3}, BRADLEY T. DE GREGORIO⁴, JAMES SAGAR¹, RICHARD THOROGATE¹, JIANHUA WANG⁵, LARRY NITTLER⁵, DAVID A. KILCOYNE⁶, HUBERTUS MARBACH⁷, MARTIN DROST⁷, GEOFF THORNTON^{1,8} 1 London Centre for Nanotechnology, 17-19 Gordon Street, University College London, London, U.K. 2 Department of Earth Sciences, University College London, London, U.K. 3 Centre for Planetary Sciences, UCL/Birkbeck, London, U.K. 4 Materials Science and Technology Division, U.S. Naval Research Laboratory, Washington DC, U.S.A. 5 Department of Terrestrial Magnetism, Carnegie Institution for Science, Washington, DC, U.S.A. 6 Advanced Light Source, Lawrence Berkeley National Laboratory, Berkeley, CA, U.S.A. 7 Microscopy and Nanolithography Group, Lehrstuhl Für Physikalische Chemie II, Friedrich-Alexander-Universität Erlangen-Nürnberg, Erlangen, Germany. 8 Department of Chemistry, University College London, 20 Gordon Street, London, London U.K. Revised manuscript submitted for publication in the Journal of the Geological Society

Abstract

Metamorphism is thought to destroy microfossils, partly through devolatilization and graphitization of biogenic organic matter. However, the extent to which there is a loss of molecular, elemental, and isotope signatures from biomass during high-temperature metamorphism is not clearly established. We report on graphitic structures inside and coating apatite grains from the ca. 1,850 million years old Michigamme silicate banded iron formation from Michigan, metamorphosed above 550°C. Traces of N, S, O, H, Ca, and Fe are preserved in this graphitic carbon and X-ray spectra show traces of aliphatic groups. Graphitic carbon has an expanded lattice around 3.6Å, forms microscopic radiating rosettes, and has homogeneous δ¹³C values around -22‰, identical to bulk analyses. Graphitic carbon inside apatite is associated with nanometre-size ammoniated phyllosilicate. Precursors of these metamorphic minerals and graphitic carbon originated from ferruginous clay-rich sediments with biomass. We conclude that graphite coatings and inclusions in apatite grains indicate fluid remobilization during amphibolite facies metamorphism of precursor biomass. This new evidence fills-in observational gaps of metamorphosed biomass into graphite and supports the existence of biosignatures in the highly metamorphosed iron formation from the Eoarchean Akilia Association, which dates from the beginning of the sedimentary rock record.

Keywords: graphite, metasedimentary, carbon isotopes, biosignature, correlated microscopy.

1. The preservation of biosignatures in graphitic carbon

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Understanding the impact of metamorphism on biological organic matter (OM) is a fundamental problem to resolve in order to determine the origin of carbon in the oldest graphite on Earth and in rocks from other ancient planetary surfaces. The metamorphic alteration of OM is known to lead to the loss of heteroatoms and molecular functional groups (Vandenbroucke and Largeau, 2007), as well as to enrichments in heavy carbon and nitrogen isotopes (Desmarais, 2001; Boyd, 2001a). The composition of OM associated with fossils in the rock record can be altered first through diagenesis, when biomass is oxidized by microbes and when sediments are altered by non-equilibrium reactions, dehydration, and eventually by metamorphism. The latter has the greatest effect on the resulting composition and crystallinity because it leads to the graphitization of carbon, in which heteroatoms and light isotopes are lost from biological OM (Bernard and Papineau, 2014). Organic microfossils in Proterozoic rocks metamorphosed at less than 300°C preserve pristine carbon isotope compositions (House et al., 2000; Lepot et al., 2013; Williford et al., 2013; 2015). In fact, carbon isotope compositions are relatively unaffected by metamorphism at conditions less than the greenschist facies (P ca. 3-6 kbar and T ca. 250-450°C), with isotopic fractionations* only about 2 to 4‰ from precursor OM (Desmarais, 2001). Heteroatoms in OM such as H, N, O, and S can also be preserved in OM from microfossils in sedimentary rocks metamorphosed at the greenschist facies (Oehler et al., 2009; 2010) and if they can be quantified, heteroatom concentrations can serve to distinguish types of kerogen (Vandenbroucke et Largeau, 2007). Under high-pressure and temperature below 400°C, organic microfossil structures can still be well-preserved and can contain carboxylic, alcoholic, phenolic, ketonic, and olefinic functional groups (Bernard et al., 2007). Some molecular functional groups may also be preserved through

* Expressed as
$$\delta^{13}C = \left(\frac{\left(\frac{^{13}C}{^{12}C}\right)_{sample}}{\left(\frac{^{13}C}{^{12}C}\right)_{PDB}} - 1\right) \times 1000\%_{0}$$

metamorphism up to the amphibolite facies (P ca. 4-9 kbar and T ca. 450-700°C), however alkanes tend to be shorter and less abundant due to thermal cracking (Schwab et al., 2005). It is unknown however if sedimentary rocks metamorphosed at the high-pressure and high-temperature conditions of the amphibolite facies can preserve graphitic carbons with clear biosignatures that indicate a carbon source from biomass. In other words, it is unclear whether combined mineral, molecular, elemental, and isotope signatures can be preserved through such metamorphic regimes and retain diagnostic characteristics of original biomass. This is important, however, because documentation of biological signatures in graphitic carbon from highly metamorphosed rocks serves as a basis to interpret the geobiological record of the oldest sedimentary rocks and to search for biosignatures in extraterrestrial environments.

Because animal fossils have never been debated as non-biological in origin and are generally preserved as carbonate-apatite mixtures with OM, clays, and/or pyrite (Olempska and Wacey, 2016), such mineral assemblages in metamorphosed sedimentary rocks can also be indicative of mineralized decayed biomass. Sedimentary rocks from late Palaeoproterozoic age in the Lake Superior area preserve distinct diagenetic structures such as rosettes and granules (LaBerge, 1973; Carrigan and Cameron, 1991; Heaney and Veblen, 1992; Winter and Knauth, 1992; Papineau et al., 2017), microfossils (Barghoorn and Tyler, 1965; Lougheed, 1983; Wacey et al., 2013; Shapiro and Konhauser, 2015), and stromatolites, which demonstrate that microbial ecosystems were widespread in the studied environment. The samples in the current study came from the ca. 1.85 Ga Bijiki member of the Michigamme Formation in the Baraga Group, Michigan, United States, and were metamorphosed at the amphibolite facies. The objective is to provide a comprehensive sub-micron scale documentation of the composition and petrography of graphitic carbon in order to constrain the preservation of biological OM under amphibolite facies metamorphic conditions.

The composition of carbon in the amphibolite facies Michigamme banded iron formation (BIF) is also of particular relevance to the controversial origin of carbon in graphite from the

3.83 Ga Akilia quartz-pyroxene (Akilia *Qp*) rock, which has been debated as evidence for the earliest life on Earth (Papineau et al., 2010a; 2010b; McKeegan et al., 2007; Nutman and Friend, 2006; Mojzsis et al., 1996) and fluid-deposited graphite of possible non-biological origin (Papineau et al., 2010a; Lepland et al., 2002; 2010). The Akilia Op rock is an Fe-silicate BIF metamorphosed at the granulite facies at 3.65 Ga (T > 650°C and P > 5 kBar), retrograded at the amphibolite facies at 2.7 Ga (Manning et al., 2006), and finally around the upper greenschist facies at around 1.5 Ga. Like the Michigamme samples in this study, coarse quartz crystals in the Akilia *Qp* rock are associated with fields of fluid inclusions that commonly contain apatite grains between about 10 and 60 µm, which frequently have graphite coatings (Papineau et al., 2010a). These sizes of apatite crystals are also consistent with analogous apatite occurrences in metapelites from the granulite facies metamorphism in the Lachlan Fold Belt in Australia (Nutman, 2007). The Bijiki graphite is ¹³C-depleted and contains trace amounts of biologicallyimportant heteroatoms (Papineau et al., 2010b). However, in rocks from such metamorphic grades, graphite can also be associated with calcite, magnetite, multiphase sulphides, and typically enveloped in amphibole, which complicates interpretations of graphite-apatite associations (Papineau et al., 2010a). Thus, the possible biological origin of graphite in the Akilia *Op* rock remains unclear and debated. Here we make progress on this problem and report new results from the petrologically similar and younger Bijiki BIF that shed some light on the possible biological origin of carbon in graphite from the Akilia *Qp* rock.

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2. Geology and samples

The Michigamme BIF is just one of several BIF units deposited between 1.85 and 1.83 Ga in the Lake Superior region of North America due to hydrothermal and geotectonic activity (Schulz and Cannon, 2007). During the accretion of supercontinent Nuna (Laurentia) in the SW Superior Craton, the Penokean Orogeny resulted in the closure of the Baraga Basin in the Marquette range. These events resulted in the closing of many coeval basins with hydrothermal

activity that delivered vast quantities of Fe on the seafloor, now preserved as BIF of the Cuyuna, Mesabi, Iron River, Marquette, Gogebic, and Gunflint ranges. The Penokean Orogen is thought to have ended by 1.84 Ga (Schneider et al., 2002) and to have resulted in the suture of island arcs and the Wisconsin Magmatic Terrain south of the Superior Craton (Van Wyck et al., 1997). Regional volcanism at 1.88 Ga (Rasmussen et al., 2012) was swiftly followed by widespread hydrothermal activity and the deposition of late Paleoproterozoic BIF. Notably, if the Lake Superior BIF are stitched back together, the time-correlative Cuyuna and Mesabi ranges in Minnesota, the Iron River, Gogebic, and Marquette ranges in Michigan, and the Gunflint range of west Ontario would form a continuous, mostly linear belt more than 600 km long of late Paleoproterozoic BIF-pelite-chert successions (Schulz and Cannon, 2007). Collectively, the BIF of the Animikie and Baraga Groups includes various types of mineralogies, including stromatolitic and granular jasper, siderite BIF, grey-red silicate BIF, and grey magnetite cherty BIF and have collectively been metamorphosed at low to high grades, up to the amphibolite and granulite facies in areas of contact metamorphism (French, 1964). Age constraints for the Baraga Group are between 1.874 and 1.833 Ga (Fralick et al., 2002; Rasmussen et al., 2012; Schneider et al., 2002). The location of TVA-294 drill hole is in the area of the Great Lakes tectonic zone in Marquette County in the Upper Peninsula of Michigan, where pre-Penokean rocks have been metamorphosed up to the amphibolite facies during the Penokean Orogen (Holm et al., 2007). Drill core *TVA-294* was drilled by the Ford Motor Company into the Dead River Basin in

Drill core *TVA-294* was drilled by the Ford Motor Company into the Dead River Basin in the Upper Peninsula of Michigan and the studied samples come from a section of this core located between 190 and 200 m depth (621 to 659 feet) (Fig. 1a). The Michigamme Fe-silicate BIF† contains graphite-rich layers (Fig. 1b) as well as pyroxene, grunerite, and magnetite bands with subordinate quartz, biotite, stilpnomelane, chlorite, carbonate, apatite, graphite, pyrite,

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[†] Because of their varied mineralogy and metamorphic grade, Fe-silicate BIF have also been variably called by other workers 'banded silicate formation', 'silicate facies BIF', 'quartz-pyroxene rock' and 'slaty taconite' in the literature.

and pyrrhotite (Fig. 2; Table 1). Associated sedimentary rocks from this drill core contain garnet and inosilicate minerals that confirm metamorphism at the amphibolite facies. Dolomitic carbonate veins in this rock cross-cut layers and contain apatite grains between 5 to 50 μ m in size associated with inclusions and coatings of graphitic carbon (Papineau et al., 2010b). In the same rock, apatite grains are concentrated in layers and, based on trace elements of co-occurring graphite, they have been argued to have sourced biomass (Papineau et al., 2010b).

Here we consider the potential contamination by more recent biological activity and fluids. Because Holocene glacial till is preserved as scattered pockets of soils around the Upper Peninsula of Michigan, soils are thin and sparse, and the underlying bedrocks are Late Palaeoproterozoic metasediments. While samples from outcrops are more susceptible to contamination by modern growth, e.g. from lichens and fungus, the studied samples were all from a drill core of Fe ore for the benefit of prospectors for the Ford Motor Company, well separated from the reach of these modern organisms. While phylogenetic analyses suggest that modern microorganisms can colonize and contaminate drill core material (Gérard et al., 2009), these microorganisms are likely only in trace abundance and the timing of their colonization of drill cores remains unknown. Considering the paucity of other sources of biomass younger than the Late Palaeoproterozoic depositional age, we do not expect that young organic molecules could be contaminants in the drill core specimens that we studied. However, contamination by drilling fluids and by late, but not modern, infiltration of OM remains possible and needs to be evaluated in light of the expected crystallinity of graphitic carbons in metamorphosed rocks.

3. Analytical methods

3.1. Micro-Raman spectroscopy

Raman spectroscopy was performed on a separate polished slab, about 1cm diameter by about 0.5 cm in thickness, using a WITec $\alpha 300$ Confocal Raman imaging system at the University College London. A 532 nm laser was used and focused at 1000X magnification for a single acquisition of 100 integrations of 1 second spectra. An optical fibre of 50 microns in diameter was used to collect a Raman spectrum at a confocal depth of 2 μ m below the polished surface of a thin section and inside transparent minerals, which minimizes the chances of spectral acquisition with artificially disordered graphite caused by polishing. The resulting average spectrum was corrected with a background subtraction of a polynomial fit of order 4. Raman D1, D2, and G peaks in the spectra were then modelled with Lorentz-fitted curves. The hyperspectral image was created from filters at 965 cm⁻¹ width of 20cm⁻¹) for apatite, 1090 cm⁻¹ (width 20cm⁻¹) for carbonate, 1575cm⁻¹ (width 60cm⁻¹) for graphite, and 1347cm⁻¹ (width 20cm⁻¹) for PCG.

3.2. Focused Ion Beam nanofabrication

Focused ion beam (FIB) milling and lift-out were performed using a FEI Nova 600 dual beam FIB-SEM at the U.S. Naval Research Laboratory. For FIB nanofabrication, a rock slab was shaped into a disk and was polished with 250 nm Al₂O₃ and DI water, rinsed with DI water, rinsed with isopropyl alcohol, dried with a dry N₂ blower, and covered with ca. 10Å Au coat. The polished rock slab, which was never in contact with epoxy, was inserted into the FIB-SEM, and a Pt shield was then deposited on the surface of the target prior to milling. A focused 30 keV Ga⁺ primary beam was then used to mill away material from both sides of the deposited Pt shield to expose the lamella. Following lift-out and welding onto an Omniprobe TEM half-grid made of Cu, the lamella was thinned down to approximately 100 nm with progressively smaller beam currents (Wirth, 2009) (down to 20 pA at 30 keV).

3.3. Scanning Electron Microscopy with Energy Dispersive Spectroscopy (SEM-EDS) and Electron Probe Micro-Analysis with Wavelength Dispersive Spectroscopy (EPMA-WDS)

These analyses were performed at the Department of Earth Sciences at University College London using a JEOL JSM-6480L SEM and at the Carnegie Institution of Washington with the JEOL 8900 Superprobe, respectively. In the SEM, the electron beam was set at 0.5 nA and 15 keV, whereas in the EPMA the beam was 5 nA at 30 keV, for spot sizes around 1 μm . The polished rock slab was C-coated prior to EPMA analysis. Abundances were determined in the EPMA using five WDS spectrometers and standards of Durango apatite, and in-house sulphide, garnet, and other silicates, which yield an error of around 0.1 wt% oxide. Other elemental abundances were measured by SEM-EDS at the University College London on Au-coated samples and were obtained from spectra

subtracted of Au, corrected with a XPP matrix correction routine in the Oxford Instruments Inca© software, and normalized to 100.0%, which yields an error around 5%.

3.4. Transmission Electron Microscopy (TEM)

High and low resolution TEM analysis of the lamella was performed with a 200 keV JEOL JEM 2500SE at the Astromaterials Research and Exploration Science e-beam facility at NASA Johnson Space Center (JSC) and with a 200 keV JEOL JEM 2100 in the Department of Chemistry at University College London. The TEM at JSC uses a field emission electron source, while the TEM at UCL uses a LaB₆ electron source. Detailed mineralogy of the FIB lamella was characterized in both conventional bright-field imaging mode and in scanning (STEM) mode using both bright-field (BF, unscattered electrons) and dark field (DF, scattered electrons) detectors. Elemental maps and selected "spot" compositional analyses were acquired at JSC using a Noran energy dispersive X-ray spectrometer (EDS) in STEM mode and at UCL using a liquid nitrogen cooled EDS detector.

3.5. Synchrotron-based Scanning Transmission X-Ray Microscopy (STXM)

Samples were analysed with the polymer STXM beamline 5.3.2.2 at the Advanced Light Source (ALS), Lawrence Berkeley National Laboratory (Kilcoyne et al., 2003). During analysis, the electron current in the storage ring was held constant in "topoff mode" at 500 mA at energy of 1.9 GeV, providing a nearly constant flux of photons at the STXM end-station. The dispersive and non-dispersive exit slits were set at 25 μ m. Focusing of the photon beam is produced by a Fresnel zone plate with a spot size of around 30 nm. STXM data were acquired as spectral image stacks (i.e. a series of X-ray absorption images at sequential energies), from which XANES spectra of regions of interest were extracted. The highest spectral resolution (0.1 eV step between subsequent images) was in the 282-292 eV range, where the near-edge spectral features for electronic transitions from core shell states to antibonding σ^* and π^* -orbitals are located. XANES spectra are presented as the ratio of transmission spectra from the region of interest, I, relative to background transmission spectra, I0, calculated as $A = -\ln(I/I_0)$. Modelling of the CNO spectrum was performed by fitting the Henke mass absorption coefficient curves for those elements to the entire XANES spectrum (Alléon et al. 2015).

3.6. Secondary Ion Mass Spectrometry and NanoSIMS

Secondary Ion Mass Spectrometry (SIMS) analyses were performed on the Gold-coated polished slab, after FIB, but before EPMA analyses and were to measure the δ^{13} C value of graphite with a Cameca ims 6F ion probe at the Carnegie Institution of Washington. Analytical conditions include extreme energy filtering conditions

(350 eV offset from -5 keV) and the use of the first field aperture and second energy slit. Each analysis consisted of 15 blocks of 10 cycles with a 3σ data rejection of cycles for an analysis time of 25 minutes, including 3 minutes of pre-sputtering. The primary Cs⁺ beam current was continuously monitored and kept at 3.0 ± 0.2 nA, to ensure at least 5×10^8 counts for the 12 C signal, which yielded an internal error better than 0.05%. Spot size was set at 30 μ m. The external error was calculated for a bracketing series of spot analyses on an in-house graphite (*BH*-graphite from Sri Lanka (Farquhar et al., 1999); n = 34) and diamond (standard *N198*; n = 15) (Table 2). Reproducibility was calculated as the square root of the sum of the internal error square plus the external error square, and averages $\pm 2.0 \%$ (2σ).

NanoSIMS analyses were performed on the FIB lamella prior to final thinning by Ne. The TEM half-grid with the FIB lamella was mounted on a clean polished Al stub using colloidal silver paint before insertion in the NanoSIMS holder, following the STXM analyses. Analyses were performed as before (Papineau et al., 2010b), but an electron gun was used for charge compensation due to loss of electrons in addition to extracted negative secondary ions, because the FIB lamella is essentially an uncoated insulator. Although a graphite standard was unavailable during these analyses, atomic ratios of C/N and C/O were calibrated using quantitative atomic ratios estimated from STXM measurements on the same areas in the FIB lamella, using the method described by Alléon (2015), and here tentatively applied to also include O/C estimates. We note that there are a number of possible model curves and chose the one that best showed the overall trend of the selected line spectrum. Unfortunately, the NanoSIMS map for O/C was deemed to unreliably show this atomic ratio in graphite due to signal contamination by the phyllosilicate, apatite, and dolomite surrounding the graphitic carbon, a spatial limitation of the technique, which did not enable quantification of the O/C atomic ratio from the CNO-XANES spectrum.

273 3.7. Helium-Neon Focused Ion Beam milling and microscopy

Surface analyses of the FIB lamella by He-Ne microscopy were performed using a Zeiss Orion NanoFAB FIB instrument located at the London Centre for Nanotechnology at the University College London. Final thinning of the lamella was performed with a 1.6 to 0.8 pA focused beam of Ne⁺ and secondary electron images were acquired using a 0.4 pA focused beam of He⁺. The He⁺ and Ne⁺ ions were accelerated to 25 KeV and 15 KeV respectively. Although the decreased Ne acceleration voltage typically results in a larger probe size it has previously been shown that lower landing energies for ion result in increased fidelity of milling (Tan et al., 2010). Organic contamination of the surface of the FIB lamella was minimized by cleaning the Orion NanoFab sample chamber using an oxygen plasma generated by an Evactron® system.

3.8. Atomic Force Microscopy

Atomic Force Microscopy was then used to measure the surface topography and evaluate the roughness of the lamella using a Dimension 3100 system at the London Centre for Nanotechnology at the University College London with the following analytical configuration parameters: Tapping mode AFM (TM-AFM) was carried out on the Dimension 3100 with a Nanoscope IV controller (Bruker Ltd., UK) equipped with a closed loop scanner. PPP-NCH cantilevers (NanosensorsTM) with an average spring constant of 42 N/m and a nominal tip radius of curvature <10nm were used for ambient air TM-AFM. The cantilevers were oscillated near their resonant frequency with a free amplitude at resonance of 2V. The amplitude setpoint was set to 80% of the value of the free amplitude. Typical scan rate was 1 Hz.

3.9. Auger Electron Spectroscopy

The local Auger Electron Spectroscopy (AES) measurements were performed at room temperature in a UHV chamber (instrument base pressure < $2 \cdot 10^{-10}$ mbar) equipped with a Leo Gemini electron column (resolution < 3 nm) at the Friedrich-Alexander-Universität Erlangen-Nürnberg, Germany. The electron beam of the SEM was used as ionization source, with a beam energy of 15 keV and a nominal probe current of 3 nA. Spectra were recorded with a hemispherical electron energy analyser (EA125, Omicron Nanotechnology) and Matrix 3.1 electronics and software (Omicron Nanotechnology). Data processing was performed with Igor Pro 6.22A (Wavemetrics). The depicted spectra were normalized such that both spectra have the same intensity at $E_{\rm Kin}$ = 950 eV (i.e. the background level). The quantitative atomic % values given were estimated based on the relevant peak areas (after linear background subtraction) and the corresponding element-specific sensitivity factors of 1.07, 0.38, and 0.35 for C, N, and O respectively as theoretically calculated (Mroczkowski and Lichtman, 1985). We note that some re-deposition of milled material onto the FIB lamellae, even after careful Ne thinning, could be included in the spectra; hence we choose to include these results regardless of their validity, as they suggest new analytical possibilities.

3.10. Isotope ratio mass spectrometry (IRMS)

Powder from drillates were obtained using a dental drill bur of SiC and 2 mm in diameter and about 2 mm deep, after discarding the first powder generated, which yields about 1-5 mg of rock powder. Samples accurately weighed to μ g precision were decarbonated in Ag boats, pre-muffled at 600°C for two hours, using 60-100 microliters of sequanal grade 10% HCl (Pierce ©). The decarbonated powders were then dried in a laminar airflow hood for more than 4 days. The samples were then combusted using a Carlo Erba elemental analyser coupled to a

Delta XL IRMS linked with a Conflo III gas injection system at the Carnegie Institution of Washington. Standards of Peru Mud and acetanilide were used to determine the accuracy and, combined with the precision (around 0.05 %), yield an error better than 0.3 %0 consistent with long-term reproducibility of δ^{13} C values on these phases.

4. Results

4.1. Petrology of graphite and apatite in an amphibolite facies silicate BIF

In thin section, the amphibolite facies Michigamme silicate BIF contains ferrous silicate bands interlayered with magnetite, quartz, and carbonate-rich layers, which can also be rich in OM (Fig. 2a-c; Fig. S1a-b). Occurrences of ferric-ferrous silicates include manganese-bearing proto-ferro-anthophyllite, grunerite, garnet, biotite, stilpnomelane, and green-brown chlorite (Table 1 and Fig. 2c, S1a-S1c). Graphitic carbon in these rocks occurs as massive layers free of clays or sulphides up to about 1 mm in thickness (Fig. 1b), and also as wavy, curvy, and circular filamentous structures in the silicate matrix (Fig. 2d, S2b-S2c) (Papineau et al., 2010b), as coatings and inclusions in about 78% of apatite grains that form in layers parallel to bedding (Papineau et al., 2010b), and as disseminated filamentous fragments in Mn-Fe-dolomite veins and layers (Fig. 2e-f, S2e-S2f). Magnetite grains are less than one mm in size (Fig. 1) and sometimes contain sulphides as micron-size spheroidal inclusions (Fig. S1c). Collectively, the mineral assemblage is consistent with recrystallization under the amphibolite facies conditions.

Raman spectra of both poorly crystalline graphite and well-ordered graphite associated with apatite from elsewhere in this dolomite vein has G-peaks located at 1575 cm⁻¹ and D-peaks located around 1347 cm⁻¹ (Fig. 3), typical position for graphite and identical to a previous report (Papineau et al., 2010b). The spectrum of well-ordered graphite was used to calculate a maximum metamorphic crystallization temperature (Beyssac et al., 2002a) of 567±50°C, consistent with amphibolite facies metamorphism, and matches graphite with a slightly expanded lattice structure from similar mineral assemblages, previously published from this

rock (Papineau et al., 2010b). Some Raman spectra display a more intense and broad D1 peak than graphitic carbon, but it is unclear whether this is due to polishing artefact (Pasteris et al., 1989), variable structures of turbostratic carbon (Beyssac et al., 2002b), variably crystallized graphitic carbon (Dodd et al., 2019), or possibly also a younger, remobilized, more disordered kind of organic matter. The Raman spectrum shows that this disordered graphitic carbon, or poorly crystalline graphite - PCG from hereon, has the same peak positions for its D1, D2, G, and 2D peaks as the well-ordered graphite (Fig. 3a-b). High-resolution Raman imaging shows the two are finely mixed (inset in Fig. 3a). The calculated crystallization temperature for the PCG was calculated to be 369°C, which may not be accurately estimated, since the Raman thermometer used is based on young organic matter in metapelites (Beyssac et al., 2002a). The PCG is also similar in its Raman spectrum to graphitic carbons found in similarly-aged greenschist to upper greenschist facies rocks elsewhere in the Michigamme Formation and in the Ihamarkotra Formation (Fig. 3c-d).

An apatite grain with graphitic carbon that intersects dolomite from the embedding matrix was targeted for nanofabrication into a lamella with a FIB-SEM (Fig. 2f, S1d-S1f) and subsequently thinned with a Ne beam (Fig. S1g-S1i). Secondary electron images from He ion microscopy, following *in situ* thinning by Ne, show contrast (Fig. 4c) given by the electrical conductivity of the different phases; graphitic carbon appears light grey, except when surrounded by an electrically insulating phase, such as dolomite or apatite. Bright-field TEM images show sub-micron size spherulites of graphite (Fig. 4d) as well as nanoscopic graphite "books" (Fig. 4e). Both of these types of graphitic carbons occur with phyllosilicate (Fig. 4e, S2c). Concentric graphite books have a euhedral habit and intimately co-occur with PCG with a more irregular and patchy texture (Fig. S2k-I). The full range of measured *d*-spacings reveals expanded graphite lattices between 3.55 and 3.72 Å (Fig. 2f; S2i-j). The C-XANES spectra of graphite show well-resolved 285.3 eV and 291.7 eV peaks, and a depressed region in between without strong peaks (Fig. 5c), consistent with highly-ordered graphitic carbon. The presence

of these peaks is also akin to graphitic carbons associated with apatite from greenschist to upper greenschist facies in the Michigamme and Jhamarkotra formations (Fig. 5c-d; Papineau et al., 2016; 2017). Hence, both forms of graphitic carbons found by micro-Raman are represented in the FIB lamella, where they are intimately mixed and have a range of *d*-spacings (Fig. 4f).

Graphitic carbon is systematically in direct contact with a clay-size, ferrous, crystalline silicate phase (Fig. 4c) that forms a complex and delicate network in apatite and dolomite (Fig. 4g). This Fe-silicate phase has 2.08 Å lattice fringes (Fig. S2h) and a composition similar to a calcium end-member of stilpnomelane (Table 1b, Fig. S2e), a ferric-ferrous phyllosilicate with several crystal orientations showing lattice spacings around 2.1 Å (Gruner, 1937; Blake, 1965). We note however that there is only less than 1 wt% Al in this phase, while stilpnomelane is expected to have around 4.3 wt% Al, and hence this phyllosilicate mineral could also be minnesotaite or greenalite. This phyllosilicate is best seen in the contrast offered by He secondary electrons (Fig. 4c) and in X-ray images near the Ca2p-edge (Fig. 5b). Although dolomite, apatite, and silicate minerals are electrical insulators, we suggest that Fe in the phyllosilicate raises its conductivity at the nanometer scale, which yields higher contrast than apatite or dolomite in the secondary electron image from He-microscopy.

4.2. Compositions of graphitic carbon and associated phases

The carbon isotope composition of graphite was measured independently by three techniques, all in good agreement. Bulk powder (about 10g) has δ^{13} C values (Papineau et al., 2010b) around -24.0% similar to three 2mm-diameter drilled spots (about 1 mg) that have δ^{13} C values between -23.4 and -21.4% (Fig. 1a), which shows relatively homogeneous compositions over cm scales. A pair of four 30 μ m diameter spot analyses *in situ* by SIMS, have δ^{13} C between -21.3 and -22.9% (Fig. 2a-b). This graphite layer analysed by SIMS is thus homogeneous with an average δ^{13} C_{gra} value (n = 10) of -21.7 ± 0.5% (1 σ) (Table 2). It thus

shows micron-scale homogeneous isotopic compositions over sub-millimetre layers of graphite. This rock is also known to have a $\delta^{13}C_{carb}$ value of -14.9 ‰ (Papineau et al., 2010b).

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X-ray images acquired at energies above the photoabsorption energy for C show strong contrast from the graphitic carbon (Fig. 5a), but variable contrast is present within graphitic carbon as well, as shown by X-ray images acquired near the aromatic carbon $(1s \rightarrow \pi^*)$ photoabsorption (~285 eV), especially for graphite completely embedded in apatite (Fig. S4bc). Since the intensity of this π^* peak is strongly dependent on the orientation of graphite lattice sheets (Rosenberg et al. 1986), this variable contrast at ~285 eV indicates multiple oriented sub-domains of graphitic carbon, also referred to as highly aromatic carbon. Carbon-XANES spectra from the lamella show no molecular functional group (spectrum 1 in Fig. 5c), whereas some trace residual groups were detected on bulk acid-insoluble OM from the same sample with resolvable peaks at 287.6 and possibly at 288.5 eV (spectra 2 in Fig. 5c). This observation can hardly be attributed to younger contamination, which would be expected to have higher absorption, and they are rather similar to the contemporary occurrences elsewhere in the Michigamme Formation and in the Jhamarkotra Formation. The weak peak at 287.6 eV could represent trace levels of aliphatic C-C bonds (Cody et al., 1996; De Gregorio et al., 2011; Bernard et al., 2012), whereas the peak at 288.5 eV represents the $1s\rightarrow\pi^*$ transition of carboxyl functional groups (Cody et al., 1996; Boyce et al., 2002; Bernard et al., 2012; Ishii and Hitchcock, 1988) and/or interlayer states in the graphite structure (Brandes et al., 2008; Fischer et al., 1991). These two peaks are not unexpected for graphitic carbon metamorphosed at the amphibolite facies, as they are well-resolved in acid insoluble residues from the phosphatic Michigamme chert and Jhamarkotra stromatolite (Fig. 5c), both metamorphosed at the greenschist to upper greenschist facies. It is possible that the PCG could host most of the trace molecular functional groups detected in acid-insoluble residues (Fig. 5c).

The relatively low intensity of these peaks however is consistent with a trace residual abundance due to metamorphism at the amphibolite facies. All samples analysed by STXM also

contain N with a peak centred at 404.0 eV, which could point to trace nitro-bearing groups (Alléon et al., 2016; Cody et al., 2011; Papineau et al., 2016), although the feature is broad enough that it could include contributions from many N-bearing organic functional groups (Fig. 5d). Other well-resolved peaks are absent from the Bijiki graphitic carbons N-XANES spectra, as well as from the greenschist to upper greenschist facies graphitic carbons of the Jhamarkotra and Michigamme formations (Fig. 5d; *cf.* Papineau et al., 2016). Peaks at 531.7 and 538.9 eV in the O-XANES spectrum may be consistent with C=O functional groups (Hitchcock and Brion, 1980) (Fig. S4g). The low intensity of the O-edge relative to that for the phyllosilicate and apatite (Fig. S4g) is consistent with minor 0 in graphitic carbon. Molecular functional groups commonly found in graphitic carbons of biological origin from the lower greenschist facies Gunflint Formation (De Gregorio et al., 2009; Alléon et al., 2016) and the greenschist to upper greenschist facies Jhamarkotra and Michigamme formations (Papineau et al., 2016; 2017) are therefore also preserved in the amphibolite facies graphitic carbon of the Bijiki BIF, although at significantly lower levels and sometimes undetectable.

NanoSIMS secondary ion images reveal the presence of H, N, O and S within graphitic carbons (Fig. 6a-f). The H and N distributions, as seen in the ¹²C¹H and ¹²C¹⁴N secondary ion images, match the graphitic carbon distribution shown by the ¹²C secondary ion image (Fig. 6a-c). However, N appears more enriched in graphitic carbon embedded in dolomite than that in apatite (Fig. 6c). Low levels of N also occur in the phyllosilicate and therefore presumably occurs as ammonium (NH₄⁺) (Boyd, 2001a; 2001b; Gruner, 1937). Oxygen was detected in all phases including in apatite, dolomite, and phyllosilicate, although it was not clearly resolved in the graphitic carbon by NanoSIMS due to the relative high flux of secondary ¹⁶O⁻ ions from associated phases (Fig. 6d; S3a). While no P was detected in dolomite (Fig. 6e), S was detected in relatively high levels in the dolomite, and in trace level in apatite, phyllosilicate, and graphitic carbons where it is non-uniform and tends to form 'hot spots' (Fig. 6f). Structural and chemical heterogeneities are documented in nanoscopic spatial resolutions for all the microscopy

techniques used and clearly shows that well-ordered graphite and PCG occur in the FIB lamella and that they contain H, N, O, and S (Fig. 6 and 7).

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Correlated microscopy can be achieved using semi-transparent and aligned overlays of different images, which can be useful to visualize correlations between nanoscopic chemical heterogeneities and specific mineral phases. This approach is taken here to show the following observations: 1) well-ordered graphite occurs as spherulites and linear books (Fig. 4d, 4e), whereas the associated PCG exhibits variable to random orientations (Fig. 7a), 2) the PCG is mixed with well-ordered graphite and a phyllosilicate resembling stilpnomelane (Fig. 7b), 3) highly aromatic domains are preferentially associated with PCG (Fig 7c, 7l), 4) the Fe-Mg phyllosilicate contains N (Fig. 7d, 7e, 7k), probably as structural ammonium, and 5) PCG contains preferential enrichments of H (Fig. 7g, 7n), N (Fig. 7h, 7k), and S (Fig. 7m). The picture that emerges from this complex and poly-crystalline mixture of graphitic carbons is one where the PCG contains the most compositional heterogeneities in the form of nanoscopic domains with elemental enrichments and where well-ordered graphite has more homogeneous compositions (Fig. 7o). Similar observations of heterogeneous S in graphitic carbon were previously reported in the Akilia Qp rock, in which nanoscopic sulphide inclusions occur (Papineau et al., 2010a), and are consistent with partial sulphidization of microorganisms during diagenesis, and the condensation and crystallization of S during metamorphism. We also note the presence of Mn in the apatite with around 0.2 wt% (Table 1a), in phyllosilicates with 0.2 to 0.4 wt% (Table 1b), in amphiboles with 1.8 to 2.7 wt% (Table 1c), and in dolomite with 4.5 to 6.5 wt% (Table 1d).

By fitting a combined CNO-XANES line spectrum collected along the edge of the FIB lamella to a linear combination of tabulated X-ray mass-absorption coefficients (Alléon et al., 2016; Cody et al., 2008) (Fig. S4e), we obtained atomic N/C and O/C ratios of 0.033 ± 0.019 and 0.051 ± 0.021 , respectively. This method has been used before for the calibration of NanoSIMS elemental ratios in graphitic carbons (Alléon et al., 2015). An artefact in the intensity of

transmitted X-rays between the C-EXAFS region and the N-edge caused a small shift in the spectrum and yielded a poor error of around ±60% for the estimated N/C of graphitic carbon. The STXM line-scan used for the calibration of NanoSIMS secondary ion ratios also had variable O intensities across the graphite (Fig. S4g), which suggests that more accurate calibrations could be achieved for smaller regions of the sample. Using the same line of pixels as that used for the XANES line-scan, we performed a calibration of the atomic N/C ratio measured by NanoSIMS to obtain a quantitative atomic N/C map from secondary ion images (Fig. 6g), which represents an improvement over previous semi-quantitative secondary ion images (Wacey et al., 2013; 2012; Oelher et al., 2010; 2009). In the lamella, the atomic N/C ratio is usually less than 0.04 (Fig. 6g), although there is a highly localized area with N/C ratio up to 0.4, which occurs on the edge of a nanoscopic apatite grain in the lamella (Fig. 6h, 6i). These compositional ratios are higher than previously published measurements by NanoSIMS up to 0.002 in three other graphite occurrences associated with apatite in this same rock and calibrated using a Nbearing graphite standard (Papineau et al., 2010b). These results show that N/C is heterogeneous over three orders of magnitude at sub-micron scales, although most commonly less than 0.04.

Auger Electron Spectroscopy (AES) measures the chemical composition of the first few atomic layers of the sample, hence atomically-flattened surfaces are ideal for chemical analysis with this method (Fig. 8a). Auger electron spectra were acquired from two areas of the lamella for which TEM, STXM, and NanoSIMS data were also acquired (Fig. 8b-c), but after re-thinning the surfaces by Neon. Simultaneous SEM monitoring during the AES measurement ensured that the beam position did not shift significantly during acquisition (Schirmer et al., 2011). Both AES spectra contain electron peaks of C (\sim 270 eV), O (\sim 510 eV), Fe (\sim 654 eV) and Ca (\sim 290 eV), while the area marked in red also exhibits a clearly resolved N peak (\sim 380 eV – Fig. 8c). Besides the occurrence of N in both graphitic carbon and phyllosilicate, Fe and Ca were also detected in both phases by AES. The spectrum for the phyllosilicate (blue curve in Fig. 8c) has a carbon

peak area more than twice that for graphitic carbon (i.e. ~ 63 at.% vs ~ 25 at.%, respectively), which is interpreted as contamination of the surface by carbon re-deposition and therefore, this Auger data can only be used qualitatively for interpretations. Nevertheless, there are disordered sub-micron domains of graphite and phyllosilicate rich in Ca and N as detected by AES, which independently confirm some TEM, NanoSIMS and STXM observations (Fig. 5b, 6c, 6g, S2f, S4e), and point to co-crystallization with phyllosilicate.

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5. Discussion

5.1. Fluid-deposition of apatite with graphitic carbons from carbonic fluids

Metamorphic recrystallization at the amphibolite facies in the Michigamme BIF yielded graphitic carbon associated with stilpnomelane included in and coating manganiferous apatite grains (Table 1a) remobilized in a Mn-dolomite vein cross-cutting sedimentary layering (Fig. 1b). This petrographic context suggests that apatite was fluid-deposited earlier than dolomite and that it was assimilated along with OM from the sedimentary layers and transported in carbonic fluids. Since bulk carbonate has a δ^{13} C value of -14.9% (Papineau et al., 2010b), much of the carbonate in dolomite can be interpreted to be the mineralized product of organic oxidation, as in other BIF (e.g. Heimann et al., 2010). NanoSIMS data also show that trace sulphate is present in dolomite (Fig. 6f), which suggests that it was present in the diagenetic environment and available for possible microbial or thermochemical sulphate reduction. The presence of Mn in dolomite, ankerite, apatite, stilpnomelane, and grunerite from this rock (Table 1) is consistent with dolomite precipitation during decreased alkalinity in the presence of Ca²⁺, Mg²⁺, Mn²⁺, and Fe²⁺ during the remobilization of the carbonic fluids. This probably took place during peak metamorphism. The graphitic carbon inside the apatite and dolomite shows a slightly distorted structure, with expanded lattice spacings between 3.55-3.72 Å (n = 6), which explains the presence of a small D1-peak in the Raman spectra. Using the Raman spectrum acquired from this well-ordered graphite, the peak metamorphic temperature is estimated to

be 567±50°C (Fig. 3a) (Beyssac et al., 2002a), consistent with an indigenous source of OM. The presence of H, O, N, and S heteroatoms in both well-ordered graphite and PCG is seen in NanoSIMS images, although it appears that the PCG hosts more trace heteroatoms, and possibly also carboxyl and aliphatic groups.

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Based on the observation that the Raman spectra of the Michigamme graphite and PCG have the same D1, D2, G, and 2D peak positions, we suggest that both these graphitic carbons have gone through the same metamorphic history. Comparable occurrences of distinct types of graphitic carbons associated with apatite occur in several other Precambrian BIF, and they are most common in highly metamorphosed rocks where fluid-deposition played an important role (Dodd et al., 2019). It is thus possible that both types of graphitic carbons were fluid-deposited and/or directly graphitized from biomass. Hence the PCG is unlikely to represent younger migrated organic matter. This also unlikely because there are no suitable Phanerozoic source rocks in the Michigan Upper Peninsula, and arguments for syngenetic oil generation and migration in the late Palaeoproterozoic sedimentary rocks of the Michigamme Formation have been based on vitrinite reflectance, heteroatom contents, and carbon isotope ratios of "anthraxolite" pyrobitumen (Mancuso et al., 1989). Hence, we find it more likely that the PCG and well-ordered graphite in the Raman spectra differ most likely by having variable levels of functional groups and heteroatoms, and variable ordering possibly due to metamorphic crystallization. Z, whereas graphite likely represents both fluid-deposited and directly graphitized sedimentary organic matter formed during peak metamorphism.

Oxidized products of biomass include carbonate and phosphate, which could have initially precipitated as carbonate-apatite layers between silicate- and magnetite-rich BIF layers. Thermal metamorphism during the Penokean Orogen then caused the remobilization of Mn-dolomite into veins, the fluid-deposition of manganiferous apatite (Table 1a), the crystallization of graphitic carbon containing aliphatic groups, the fluid-deposition of graphite spherulites, and the crystallization of stilpnomelane, grunerite, and anthophyllite from ferric-

ferrous and manganiferous clay precursors (Tables 1b, 1c). The presence of Mn in the Michigamme BIF is consistent with a syn-depositional hydrothermal-sedimentary source, the same source as Fe, and akin to other manganiferous Precambrian BIF occurrences (Roy, 2006).

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5.2. Assessing the possible non-biological origins of graphitic carbon and associated minerals

In the context of the search for evidence of early life in metamorphosed sedimentary rocks, the null hypothesis should be evaluated, which means that all possible known nonbiological processes should be considered before a biological conclusion can be established. Microscopic spherulites of graphitic carbon preserved in the Bijiki BIF (Fig. 4d) likely formed in part due to the non-biological fluid-deposition of graphite, from the spontaneous reaction of CO₂ and CH₄ either from non-biological mantle sources, or from decomposed microbial biomass (next section). They have a pseudo-hexagonal habit different from that of millimetric graphite spherulites from the Pogranichnoe carbonatite formed from non-biological carbon (Doroshkevitch et al., 2007) and from alkaline pegmatite in the Kola Peninsula with a hydrothermal origin (Jaszczak et al., 2007). Graphite spherulites in both of these Russian occurrences are formed by radially-aligned graphite needles and tubes, whereas the Michigamme graphite forms distinct concentrically-aligned books. In addition, the nonbiological graphite from Pogranichnoe and Kola also co-occurs with apatite, showing that these mineral associations can also be non-biological in origin (Doroshkevitch et al., 2007; Jaszczak et al., 2007). Other mantle-fluid processes can also produce non-biological OM associated with apatite as in the case of the Chassigny dunite from Mars where these associated phases have been reported (Papineau, 2012). Instead, the Bijiki graphitic carbon appears unique to sedimentary lithologies and they are similar to graphite filaments from the Jingangku Fm in the ca. 2.5 Ga Wutai Complex of China (Schiffbauer et al., 2007). However, the association of apatite with graphitic carbon can occur through non-biological processes and hence the null

hypothesis cannot be rejected for this mineral association nor the morphology of the Michigamme graphitic carbon.

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To further test the null hypothesis, possible non-biological sources of complex OM could be remobilized to form mantle carbon with an expected δ^{13} C value around -5\%, characteristic of the upper mantle (Horita, 2005). Graphite can be spontaneously fluid-deposited during the reduction of CO₂ and the oxidation of CH₄ in aqueous fluids (Rumble, 2014), which can lead to the deposition of non-biological graphitic carbon in basalt and pyroxenite on Mars (Steele et al., 2012). Fisher-Tropsch Type (FTT) synthesis can also lead to the formation of short-chain alkanes through catalysis by Cr and Fe-bearing spinels, although Ni-sulphides tend to favour the production of CH₄ over short alkanes (Foustoukos and Seyfried, 2004). Experiments of FTT synthesis show that organic carbon can be significantly depleted in ¹³C by up to 36‰ (McCollom and Seewald, 2007), although in modern hydrothermal systems FTT carbon is not usually depleted by more than 16% (Proskorowski et al, 2008). In addition, FTT experiments with variably complex reactants show that various organic compounds containing C, H, O, N, and S can be synthesized non-biologically under hydrothermal conditions (Rushdi and Simoneit, 2004). Because the hydrothermally-influenced environment of the Michigamme BIF is a plausible site for hydrocarbons generated by FTT reactions, the null hypothesis for the isotopic and elemental composition of the graphitic carbon cannot be entirely rejected.

It has also been suggested that Mn, Mg, Ca, and Fe-bearing carbonic fluids reacted to form non-biological graphite associated with apatite grains in metasomatically-altered carbonate rocks from the Eoarchean Isua Supracrustal Belt (Lepland et al., 2002). The decarbonation of siderite occurs at temperatures above 450°C and can produce graphite along with magnetite and CO₂ (French, 1971; McCollom, 2003), which could conceivably have reprecipitated as carbonate. Dolomitic carbonate has been proposed as a biosignature (Nutman et al., 2010), although recent carbonation experiments show that clay minerals such as illite and montmorillonite can stimulate the non-biological precipitation of proto-dolomite under

ambient conditions (Liu et al., 2019). Similarly, NH₄+ in phyllosilicates has also been proposed to be a biosignature (Boyd, 2001b), but it could have been sourced from hydrothermal fluids, possibly from non-biological sources such as N₂ reduction with H₂S (Schoonen and Xu, 2001) or the reduction of NO₃- or NO₂- with Fe-Ni metals (Summers and Chang, 1993; Brandes et al., 1998; Smirnov et al., 2008). Hence there are known non-biological processes in hydrothermal and metamorphic systems that can result in the precipitation of dolomite with graphitic carbon as well as ammoniated phyllosilicates and therefore the null hypothesis for these biosignatures cannot be rejected.

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In summary, there are processes under hydrothermal conditions that may produce purely non-biological OM with ¹³C-depletion, trace elements of H, N, O, and S, and molecular functionalities, or associations with apatite or ammoniated phyllosilicates. However, these nonbiological scenarios can only explain some characteristics of the Michigamme graphite and associated minerals; the key fact remains that all observations combined point to a biological origin. A number of non-biological processes can produce the features reported here, but those mechanisms have to converge to coherently explain all the combined key lines of evidence: (1) the geological context in a BIF unit, known to have contemporary versions with prolific stromatolites and microfossils, (2) the presence of syngenetic graphitic carbon with an expanded crystalline structure consistent with metamorphic history, (3) the observed carbon isotope homogeneity, heteroatom heterogeneity, and trace molecular compositions, and (4) the association of graphitic carbon with NH₄+-bearing phyllosilicate inside apatite, and with sulphate-bearing ¹³C-depleted dolomite. Indeed, non-biological processes are unlikely to produce all these observed elemental, isotopic, molecular, structural, and mineralogical features documented here together for the metamorphosed Michigamme BIF. Hence, because life had also been present throughout the late Palaeoproterozoic and since at least the Eoarchean, we conclude that a microbial origin is the simplest and most likely explanation for the observed compositions of graphitic carbons and their associated minerals.

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5.3. The preservation of metamorphosed biological signatures in graphitic carbon

In the Michigamme Formation, the age of peak metamorphism was around 1.833 Ga with the intrusion of various plutons and 1.830 Ga with regional metamorphism (Schulz and Cannon, 2007). The origin of the carbon in the Michigamme graphitic carbon must therefore be indigenous and syngenetic, as is the case for graphite in the Biwabik Formation in northern Minnesota (French, 1964). The degree of crystallinity of graphitic carbon associated with apatite in the Michigamme silicate BIF was significantly modified by thermal metamorphism as seen in the Raman spectrum with a low G-peak position and a weak D1-peak (Fig. 3a), commonly observed in metamorphosed graphitic carbon (Beyssac et al., 2002a; 2002b).

We now consider that the Michigamme graphite represents the metamorphosed remains of microorganism and that the most graphitized micron-size domains (e.g. Fig. 4d, 4e) represent microbial biomass that has been partly fluid-deposited (Rumble, 2014). This would have occurred during thermal metamorphism, possibly from dehydration reactions involving phyllosilicate precipitation, CH₄ cracked from alkanes, and CO₂ from oxidized and decarboxylated kerogen. No fluid inclusions have been observed so far in the specimens. Direct graphitization of biomass is proposed for the graphitic carbon associated with phyllosilicate (Fig. 7a, 7b). The graphitic carbon seen in BF-TEM images typically has nanoscopic inclusions of highly aromatic carbon (as seen from the most opaque domains at 285.3 eV; Fig. 7c, 7j, 7l), and contains heterogeneities of N, S, and H (Fig. 7g, 7h, 7m). We observed no nanoscopic inclusions of sulphide inside the graphite (Fig. S2k-S2o). Hence, these observations are consistent with nanoscopic-size chemical heterogeneities observed in biological carbon from chert metamorphosed at sub-greenschist facies in the Gunflint Fm (Wacey et al., 2013). Organic matter from the Gunflint chert also contains heteroatoms of N, O, P, and S as well as aromatic alcohol, phenol, and carboxylic acid functional groups (Alléon et al., 2016; De Gregorio et al., 2009). Residual heteroatoms are largely preserved at the greenschist facies, where H/C is

typically between 0.1 and 0.5 (Watanabe et al., 1997; Hayes et al., 1983). The above observations are also consistent with trace element compositions in metamorphosed graphitic carbons from stromatolitic phosphorite and pelite including black shale in the contemporary Aravalli Group, India, which have Raman spectra with significant D1-peaks (Fig. 7 in Papineau et al., 2009) and C-XANES spectra often with resolved peaks for carboxyl, aliphatic, nitrile, and nitro functional groups (Fig. 11 in Papineau et al., 2016).

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Hence, all characteristics of graphitic carbon and associated minerals in the Michigamme Formation are independently consistent with a biological source of carbon from the original depositional environment, and they include: 1) Raman peaks for graphitic carbon consistent with the metamorphic history and silicate mineral assemblages (Fig. 3), 2) the common filamentous graphite morphology (Fig. 2d) never observed in igneous rocks, where graphite usually forms geometric crystal habits (Jaszczak, et al., 2007; 2003), 3) the slightly expanded graphite lattice spacings (Fig. 4f) consistent with crystallization of an organic precursor with heteroatoms, 4) the trace levels of molecular functional groups in graphitic carbon, which is consistent with the independently detected presence of H, N, O, and Ca (Fig. 5c, 5d, 6a-6f, 8c, S4e), and similar to what was observed in kerogen from the Gunflint Fm (Alléon et al., 2016), 5) the heterogeneous distribution of N and S that are concentrated over areas less than 500 nm in size (Fig. 7k, 7m), similar to heterogeneous N distributions in OM from the Gunflint Fm (Wacey et al., 2013), 6) the ¹³C-depleted Mn-Fe dolomite (down to -14.9‰) in veins that most likely sourced carbonate from diagenetically-oxidized biomass (Papineau et al., 2010b), similar to other BIF (Heimann et al., 2010), 7) the systematic association of graphitic carbon with C- and N-bearing phyllosilicate (Fig. 7h, 7k), which may contain NH₄+ of biological origin (Boyd, 2001b) such as in co-eval BIF from Minnesota (Blake, 1965), 8) the co-occurrence with manganiferous dolomite, apatite, stilpnomelane, and grunerite (Tables 1) that indicate deposition from fluids with hydrothermally-sourced metals (Roy, 2006), and 9) fractionated δ^{13} C values consistent with a metamorphic overprint on the signature of photoautotrophic biomass (Table 2) (Desmarais, 2001). These indications point to the Michigamme graphitic carbons having a biological origin, likely dominated by oxygenic photosynthesis and having formed oil, kerogen, and pyrobitumen during metamorphism (Mancuso et al., 1989). Fluid-deposition during the retrograde phase of peak metamorphism can explain why there is PCG in this rock, which could have formed from devolatilized biomass and reaction products such as CO₂ and CH₄.

The compositions of the Michigamme graphitic carbon are those expected for microbial biomass metamorphosed at the amphibolite facies and thus include trace levels of biological element heterogeneities and carbon isotope distributions that are homogeneous from centimetre to micrometre scales. Heating experiments of acritarch microfossils composed of OM have shown that residual ketone and aldehyde functional groups can be preserved after several days of exposure to temperatures at 500°C, and also that recognizable biological structures are best preserved under anoxic metamorphic conditions, but also under oxic conditions (Schiffbauer et al., 2012). These details are important because they demonstrate key characteristics of kerogen metamorphosed at low metamorphic grades and lead to the conclusion that some elemental and molecular characteristics of degraded biomass can be preserved at high metamorphic grades (Bernard et al., 2009; 2007; Boyce et al., 2002).

In summary for the Michigamme BIF, the oxidative and thermal degradation of photoautotrophic microbial biomass with silicate-rich ferruginous sediments produced kerogen, apatite, dolomite with sulphate, sulphide, and ferric-ferrous oxides like hydromagnetite and phyllosilicates like greenalite and stilpnomelane. These diagenetic minerals were then crystallized graphite during prograde metamorphism, whereas PCG is interpreted to have formed from thermally cracked products that were solubilized, remobilized, and fluid-deposited during amphibolite facies metamorphism, but after peak metamorphism. These processes would have begun with precursor biomass mixed with silica and ferruginous clays, and are now preserved as a complex mixture of fluid-deposited graphite

and PCG from residual biomass, mixed with apatite, Mn-Fe-dolomite with sulphate, phyllosilicate with NH₄+, and associated with mineral reaction products from diagenesis and metamorphism including magnetite, pyrrhotite, biotite, chlorite, and grunerite. However, examples from other highly metamorphosed Fe-silicate BIF and graphitic schists should also be rigorously tested by studying graphite petrology and geochemistry.

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5.5. Implications for the Akilia Qp rock

Comparisons with graphite from the granulite facies Akilia *Qp* rock should reveal a more extreme effect of metamorphism above 640°C on OM with similar original characteristics. While graphite from the Akilia *Qp* rock has characteristics consistent with a biological origin, the geochemical pathways of fluid-deposited graphite with apatite are unknown for such hightemperature metamorphism, and thus uncertainties persist for possible contributions from non-biological sources of carbon in the Akilia graphite. The key observations of Akilia graphite are listed in Table 3 (Papineau et al., 2010a; 2010b) and include 1) its common association as coatings on apatite in linear fields of fluid inclusions in coarse quartz, suggesting precipitation from fluids, 2) its slightly expanded graphite lattice spacings, consistent with the presence of heteroatoms, 3) its heterogeneous distribution of N and S consistent with similar observations in younger metamorphosed graphitic carbons, 4) its co-occurrence with ¹³C-depleted calcite, consistent with some oxidized organic matter, 5) its ¹³C-depleted composition not inconsistent with isotopic fractionation of biomass metamorphosed at the granulite facies, 6) the presence of curled graphite structures consistent with granulite facies thermal metamorphism, and 7) the absence of molecular functional groups in C-XANES spectra that confirm the graphitic nature of the carbon (Papineau et al., 2010a; 2010b), and consistent with a complete removal of residual functional groups at the granulite facies. The PCG shares spectral and compositional similarity with younger graphitic carbons of biological origin, e.g. from the Aravalli Supergroup (Papineau et al., 2009; 2016), and C-XANES spectra for bulk acid-insoluble organic matter in the Bijiki BIF suggest that it could be the host of trace residual functional groups and perhaps also heteroatoms.

Fluid-deposited graphitic carbon can form from microbial biomass, possibly from thermally-cracked alkanes and decarboxylated kerogen, and could be associated with ¹³C-depleted carbonate, hydrated minerals, as well as fluid inclusions in quartz that contain CH₄ and CO₂ (Lepland et al., 2010). Collectively, the new observations demonstrate the preservation of molecular, elemental, and isotopic biosignatures in graphitic carbon metamorphosed at the amphibolite facies, and strengthen the case for the biological origin of carbon in graphite from the Akilia *Qp* rock. While some workers have suggested that the protracted metamorphic history of the *Akilia Qp* rock precludes any biogenic interpretation and makes it 'impossible' to prove a biological origin (Lepland et al., 2011), progress can still be achieved as demonstrated in this work. The new observations of graphitic carbons associated with fluid-deposited apatite from the Michigamme Formation show highly similar compositions (Table 3), consistent with slightly lower metamorphic temperatures than in the Akilia *Qp* rock. A continuum of the key characteristics of graphitic carbon observed in biomass metamorphosed at the amphibolite and granulite facies is thus established for the first time.

6. Conclusions

In the amphibolite facies Michigamme Fe-silicate BIF, the diagenetic oxidation of biomass along with metamorphism at temperatures in excess of 550°C produced the precursors to the mineralization of Mn-Fe dolomite, graphitic carbons, apatite, and sulphides as well as various ferric-ferrous phases including magnetite, anthophyllite, grunerite, and phyllosilicates. Apatite associated with graphitic carbon in this silicate BIF was fluid-deposited from remobilized carbonic fluids that also contained highly polyaromatic hydrocarbons from oxidized and decarboxylated kerogen. The preservation of graphitic carbon as micron-size coatings and inclusions in apatite grains is likely the result of co-precipitation of apatite,

carbonate, and graphitic carbon from phosphatic and carbonic fluids under amphibolite facies temperatures. The graphitic carbon contains trace levels of heterogeneous H, N, O, and S as well as trace amounts of aliphatic and possibly carboxyl groups. Carbon isotope ratios of graphite layers are homogeneous with a δ^{13} C around -22‰ in centimetre to micrometre scales, which shows that carbon isotopes were fractionated during prograde metamorphism. It also co-occurs with fine-grained ammoniated phyllosilicate and is embedded in manganiferous apatite and sulphate-bearing dolomite. Because all the expected elemental, isotopic, molecular, mineralogical, and morphological biosignatures of degraded microorganisms occur in the Bijiki member of the Michigamme BIF, the reported data is most consistent with a biological origin. There are also no known non-biological graphitic carbons with comparable petrographic distribution, mineral associations, and geochemical composition, such that the null hypothesis can be reasonably rejected for our combined observations. However, none of the null hypotheses for each independent line of evidence evaluated on its own can be rejected.

These observations shorten the observation gap of graphitic carbon from biological origin between the amphibolite and granulite facies. In the Eoarchean Akilia *Qp* rock, graphite occurs in several forms such as apatite coatings (Papineau et al., 2010a), as hydrothermal mineral associations with calcite (Papineau et al., 2010a), and as fluid inclusions that also contain CO₂ and CH₄ (Lepland et al., 2010). Graphite coatings on apatite grains from the Akilia *Qp* rock could have formed from recrystallized biomass (Papineau et al., 2010b; McKeegan et al., 2007; Nutman and Friend, 2006; Mojzsis et al., 1996), and our new data further supports this interpretation by filling the observational gap between well-characterized kerogen from greenschist facies metamorphism (Alléon et al., 2016; Papineau et al., 2017; 2016; Wacey et al., 2013; De Gregorio et al., 2009), and more highly metamorphosed graphitic carbons at amphibolite facies (Schiffbauer et al., 2007; Beyssac et al., 2002; French, 1964). Graphitic carbon associated with apatite, dolomite, and ferric-ferrous oxides and silicates can constitute a solid biosignature in metamorphosed sedimentary rocks if 1) the geological context is

permissive both from the metamorphic history and the inferred depositional environment and 2) the mineralogical, elemental, isotopic, and molecular signatures of life are preserved with or in graphitic carbon and their abundances and distributions are consistent with the metamorphic grade and analogous occurrences. This robust evaluation of the geological transformation of biomass through thermal metamorphism further paves the way to search for biosignatures in other ancient environments, including ancient planetary surfaces such as Mars.

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1065	
1066	
1067	Author contributions

D.P. designed and led the research and participated in all analyses; D.P., B.T.D., and D.A.K.

performed STXM analyses; D.P. and B.T.D. performed FIB nanofabrication and STEM-EDS

analyses; J.S. performed He-Ne FIB nano-fabrication and microscopy; D.P. performed sample

preparation and CF-EA-IRMS analyses; R.T. performed the AFM analyses; H.M. and M.D.

performed the AES and analysed the data along with G.T.; D.P. and J.W. performed SIMS analyses,

L.R.N. contributed to the analysis of NanoSIMS data collected by D.P. and J.W.; D.P. wrote the paper

with significant input from all co-authors who also variably contributed to the analysis of the data.

1076	Figures – see attached
1077	
1078	<u>Tables</u>
1079	- Table 1: X-ray-based compositional analyses of Michigamme minerals using different
1080	electron beam instruments.
1081	- Table 2: In situ carbon isotope analyses by SIMS on graphite band from TVA294-659.6 and
1082	standards.
1083	- Table 3: Comparative table of mineralogical and geochemical data between the
1084	Michigamme BIF and the Akilia <i>Qp</i> rock.
1085	
1086	Supplementary information
1087	- Figure S1: Additional petrographic context.
1088	- Figure S2: Compositional and structural analysis of graphite and associated
1089	minerals.
1090	- Figure S3: Additional contextual images and analysis details of the analysed area
1091	used to calibrate NanoSIMS 12C14N-/12C- map with STXM.
1092	- Figure S4: Additional STXM data of the analyzed target.
1093	- Figure S5: Selected line profiles measured by AFM

Table 1: Elemental compositions of apatites, carbonates, and silicates obtained from EDS and WDS analyses.

a) Apatites analyzed by WDS in the EPMA

Targets	TVA294-659.6	TVA294-659.6	TVA294-659.6	TVA294-659.6
element Wt.(%) / spot	3	12	14	4
CaO	55.1	54.5	55.0	55.1
P ₂ O ₅	41.2	41.5	40.6	40.9
F	3.2	3.2	3.3	3.2
CI	0.0	0.0	0.0	0.0
FeO	0.5	0.6	1.0	0.4
MnO	0.2	0.3	0.2	0.2
Na ₂ O	0.0	0.0	0.0	0.0
MgO	0.0	0.0	0.0	0.0
SiO ₂	0.0	0.0	0.0	0.0
Ce ₂ O ₃	0.1	0.1	0.0	0.1
Y ₂ O ₃	0.0	0.0	0.0	0.0
Total	99.1	98.8	98.8	98.7
	fluoranatite	fluoranatite	fluoranatite	fluoranatite

b) Fe-silicates analyzed by EDS in the TEM (on FIB lamellae)

Targets	TVA294-656.9	TVA294-656.9	
element Wt.(%) / spot	01b	01a	
ок	48.6	49.9	
Mg K	0.4	0.4	
AI K	0.8	0.5	
Si K	36.9	39.6	
Ca K	0.7	0.8	
Mn K	0.4	0.2	
Fe K	12.2	8.6	
Total	100.0	100.0	

Ca-Mn-stilpnomelane Ca-Mn-stilpnomelane

c) Ferric-ferrous silicates analysed by EDS in the SEM

Targets	TVA294-622.1	TVA294-622.1	TVA294-622.1	TVA294-622.1	TVA294-622.1	TVA294-622.1	TVA294-622.1
element Wt.(%) / spot	1	2	5	8	7	4	6
C K			14.3				13.0
оĸ	36.2	36.1	36.3	37.1	39.9	39.2	37.8
/lg K	5.9	5.2	4.1	5.6	5.5	5.1	4.0
AI K					11.2	8.9	8.2
Si K	26.9	26.2	19.5	25.6	12.6	13.9	10.1
KK						1.4	
In K	2.3	1.8	1.9	2.7			
e K	28.8	30.7	24.0	29.1	30.8	31.5	26.9
otals	100.0	100.0	100.0	100.0	100.0	100.0	100.0
	proto-Mn,Fe-anthophylite	Mn,Fe-anthophylite	Mn,Fe-anthophyllite + graphite	grunerite	chlorite	chlorite	graphite with chlor

d) Sulphides and oxides with graphitic carbon analysed by EDS in the SEM

Targets	TVA294-622.1	TVA294-622.1	TVA294-622.1	TVA294-622.1
element Wt.(%) / spot	1	3	9	2
CK		22.4		8.8
ок			11.5	22.9
SK	34.7	26.7	22.3	
Fe K	65.3	50.9	66.2	68.3
Totals	100.0	100.0	100.0	100.0

e) carbonates analysed by EDS in the SEM

Targets	TVA294-656.9	TVA294-656.9	TVA294-622.1	TVA294-622.1	TVA294-622.1	TVA294-622.1
element Wt.(%) / spot	1	2	4	5	6	3
CK	15.5	13.6	13.6	13.6	11.7	63.2
ок	43.0	34.4	43.2	43.2	45.1	20.9
Mg K	9.8	9.8	6.1	6.1	6.7	2.2
CIK					1.4	
Ca K	26.0	24.8	21.4	21.4	20.7	7.9
Si K	1.1	1.1				0.6
Mn K	4.5	4.9	6.5	6.5	5.4	2.5
Fe K		11.0	9.2	9.2	9.2	2.8
Totals	99.9	99.6	100.0	100.0	100.0	100.0
	Mn-dolomite	Mn-Fe-dolomite	Mn-Fe-dolomite	Mn-Fe-dolomite	Mn-Fe-dolomite	graphite in dolomit

Table 2: In situ carbon isotope analyses on graphite band from TVA294-659.6 and stan

	In situ carbon isotope a				
Analysis	#Analysis spot name ^l	³ C/ ¹² C (uncorr	err.	¹² C count rate	(c/s) ¹² C counts
					_
1	N198a@2.ais	1.04659E-02	4.12E-06	8.67E+05	1.30E+08
2	N198a@3.ais	1.04666E-02	4.53E-06	8.72E+05	1.31E+08
3	N198a@4.ais	1.04786E-02	4.18E-06	8.68E+05	1.30E+08
4	N198a@5.ais	1.04638E-02	4.08E-06	8.69E+05	1.30E+08
5	N198a@6.ais	1.04796E-02	3.87E-06	8.64E+05	1.30E+08
6	N198a@7.ais	1.04731E-02	4.14E-06	8.22E+05	1.23E+08
7	N198a@8.ais	1.04715E-02	3.39E-06	8.49E+05	1.27E+08
8	N198a@9.ais	1.04735E-02	4.04E-06	8.56E+05	1.28E+08
9	N198a@10.ais	1.04856E-02	3.97E-06	8.62E+05	1.29E+08
10	N198a@11.ais	1.04895E-02	4.34E-06	8.65E+05	1.30E+08
11	N198a@12.ais	1.04829E-02	4.40E-06	8.57E+05	1.29E+08
12	N198a@13.ais	1.04928E-02	4.27E-06	8.56E+05	1.28E+08
13	N198a@14.ais	1.04770E-02	4.50E-06	8.43E+05	1.26E+08
14	N198a@15.ais	1.04857E-02	4.09E-06	8.44E+05	1.27E+08
15	N198a@16.ais	1.04869E-02	4.44E-06	8.50E+05	1.27E+08
16	BH_graphite@1.ais	1.04838E-02	4.73E-06	7.24E+05	1.09E+08
17	BH_graphite@2.ais	1.04950E-02	4.54E-06	7.05E+05	1.06E+08
18	BH_graphite@3.ais	1.04853E-02	4.11E-06	7.20E+05	1.08E+08
21	BH_graphite@6.ais	1.04610E-02	4.90E-06	6.03E+05	9.04E+07
22	BH_graphite@7.ais	1.04700E-02	5.30E-06	6.16E+05	9.23E+07
23	BH_graphite@8.ais	1.04810E-02	4.66E-06	6.06E+05	9.09E+07
24	BH_graphite@9.ais	1.04799E-02	4.79E-06	6.23E+05	9.35E+07
25	BH_graphite@10.ais	1.04781E-02	4.72E-06	6.21E+05	9.32E+07
26	BH_graphite@11.ais	1.04712E-02	4.40E-06	6.21E+05	9.31E+07
34	BH_graphite@12.ais	1.04775E-02	4.88E-06	5.89E+05	8.84E+07
35	BH_graphite@13.ais	1.04899E-02	4.83E-06	5.74E+05	8.60E+07
36	BH_graphite@14.ais	1.04869E-02	4.91E-06	5.61E+05	8.41E+07
37	BH_graphite@15.ais	1.04739E-02	5.13E-06	5.16E+05	7.73E+07
38	BH_graphite@16.ais	1.04632E-02	5.79E-06	5.39E+05	8.09E+07
39	BH_graphite@17.ais	1.04860E-02	5.27E-06	5.12E+05	7.68E+07
46	BH_graphite@18.ais	1.04743E-02	5.12E-06	5.24E+05	7.86E+07
47	BH_graphite@19.ais	1.04901E-02	5.30E-06	5.04E+05	7.56E+07
48	BH_graphite@20.ais	1.04813E-02	5.32E-06	5.09E+05	7.63E+07
54	BH_graphite2@1.ais	1.04712E-02	4.83E-06	6.97E+05	1.05E+08
55	BH_graphite2@2.ais	1.04774E-02	4.45E-06	7.17E+05	1.08E+08
56	BH_graphite2@3.ais	1.04744E-02	4.59E-06	7.31E+05	1.10E+08
67	BH_graphite2@4.ais	1.04792E-02	4.71E-06	7.28E+05	1.09E+08
68	BH_graphite2@5.ais	1.04674E-02	4.59E-06	7.35E+05	1.10E+08
69	BH_graphite2@6.ais	1.04747E-02	4.38E-06	7.43E+05	1.11E+08
70	BH_graphite2@7.ais	1.04716E-02	4.10E-06	7.46E+05	1.12E+08
71	BH_graphite2@8.ais	1.04647E-02	4.49E-06	7.33E+05	1.10E+08
72	BH_graphite2@9.ais	1.04865E-02	4.72E-06	7.12E+05	1.07E+08
73	BH_graphite2@10.ais	1.04899E-02	4.55E-06	7.10E+05	1.07E+08
74	BH_graphite2@11.ais	1.04971E-02	4.76E-06	6.75E+05	1.01E+08
79	BH_graphite2@13.ais	1.04540E-02	4.95E-06	5.81E+05	8.71E+07
80	BH_graphite2@14.ais	1.04561E-02	5.28E-06	5.81E+05	8.72E+07

88	BH_graphite2@17.ais	1.04880E-02	5.05E-06	5.36E+05	8.04E+07
106	BH_graphite2@20.ais	1.04763E-02	4.67E-06	5.49E+05	8.23E+07
107	BH_graphite2@21.ais	1.04660E-02	4.66E-06	5.65E+05	8.48E+07
108	BH_graphite2@22.ais	1.04711E-02	5.01E-06	5.63E+05	8.44E+07
57	TVA294-6596@1.ais	1.03354E-02	5.36E-06	5.73E+05	8.60E+07
58	TVA294-6596@2.ais	1.03385E-02	5.03E-06	5.89E+05	8.83E+07
59	TVA294-6596@3.ais	1.03297E-02	4.93E-06	5.78E+05	8.68E+07
60	TVA294-6596@4.ais	1.03323E-02	5.47E-06	5.71E+05	8.57E+07
61	TVA294-6596@5.ais	1.03359E-02	4.95E-06	5.98E+05	8.97E+07
62	TVA294-6596@6.ais	1.03383E-02	4.56E-06	5.92E+05	8.88E+07
63	TVA294-6596@7.ais	1.03362E-02	5.73E-06	5.56E+05	8.34E+07
64	TVA294-6596@8.ais	1.03347E-02	6.08E-06	5.29E+05	7.94E+07
65	TVA294-6596@9.ais	1.03220E-02	5.34E-06	5.31E+05	7.96E+07
66	TVA294-6596@10.ais	1.03383E-02	5.38E-06	4.94E+05	7.41E+07

^{*} The two in-house standards used were diamond N198 (δ^{13} C = -6.0‰, IMF correction fa

I primary (nA)	¹³ C/ ¹² C (IMF corr.	s ¹³ C /0/ \	int err (1 ~)	evt err (2 ~\
i primary (IIA)	CI C (IIVIF COFF.	υ ບ (‰)	mr en (10)	GAL GII (4 0)
2.07	4 44200E 02	0.2	0.4	2.0
3.27	1.11208E-02	-9.2	0.4	2.0
3.25	1.11215E-02 1.11343E-02	-9.2	0.4	2.0
3.21 3.18	1.11343E-02 1.11186E-02	-8.0 -9.4	0.4 0.4	2.0
3.16	1.11106E-02 1.11354E-02		0.4	2.0
3.08	1.11354E-02 1.11284E-02	-8.0	0.4	2.0
3.12	1.11264E-02 1.11267E-02	-8.6 -8.7	0.3	2.0 2.0
3.14	1.11287E-02 1.11289E-02	-6. <i>1</i> -8.5	0.4	2.0
3.09	1.11209E-02 1.11417E-02	-6.5 -7.4	0.4	2.0
3.12	1.11417E-02 1.11459E-02	-7.4 -7.0	0.4	2.0
3.12	1.11389E-02	-7.6	0.4	2.0
3.06	1.11494E-02	-6.7	0.4	2.0
3.08	1.11326E-02	-8.2	0.4	2.0
3.09	1.11418E-02	-0.2 -7.4	0.4	2.0
3.05	1.11431E-02	-7.4	0.4	2.0
3.03	1.114312-02	-7.5	0.4	2.0
3.04	1.11398E-02	-7.6	0.5	2.1
3.02	1.11517E-02	-6.5	0.4	2.0
2.99	1.11414E-02	-7.4	0.4	2.0
3.11	1.11156E-02	-9.7	0.5	2.1
3.07	1.11252E-02	-8.9	0.5	2.1
3.03	1.11368E-02	-7.8	0.4	2.1
2.84	1.11357E-02	-7.9	0.5	2.1
2.76	1.11338E-02	-8.1	0.5	2.1
2.92	1.11264E-02	-8.7	0.4	2.0
3.30	1.11331E-02	-8.1	0.5	2.1
3.28	1.11463E-02	-7.0	0.5	2.1
3.27	1.11431E-02	-7.3	0.5	2.1
3.04	1.11293E-02	-8.5	0.5	2.1
3.04	1.11179E-02	-9.5	0.6	2.2
3.03	1.11422E-02	-7.3	0.5	2.1
2.96	1.11297E-02	-8.5	0.5	2.1
2.94	1.11465E-02	-7.0	0.5	2.1
2.90	1.11372E-02	-7.8	0.5	2.1
3.49	1.11264E-02	-8.7	0.5	2.1
3.47	1.11330E-02	-8.2	0.4	2.0
3.44	1.11298E-02	-8.4	0.4	2.0
3.30	1.11349E-02	-8.0	0.4	2.1
3.27	1.11224E-02	-9.1	0.4	2.1
3.26	1.11301E-02	-8.4	0.4	2.0
3.23	1.11269E-02	-8.7	0.4	2.0
3.22	1.11195E-02	-9.4	0.4	2.0
3.20	1.11427E-02	-7.3	0.4	2.1
3.22	1.11463E-02	-7.0	0.4	2.0
3.21	1.11539E-02	-6.3	0.5	2.1
3.07	1.11081E-02	-10.4	0.5	2.1
3.09	1.11104E-02	-10.2	0.5	2.1

3.08	1.11443E-02	-7.2	0.5	2.1
2.83	1.11318E-02	-8.3	0.4	2.1
2.88	1.11209E-02	-9.2	0.4	2.1
2.82	1.11263E-02	-8.8	0.5	2.1
3.43	1.09821E-02	-21.6	0.5	2.1
3.39	1.09854E-02	-21.3	0.5	2.1
3.41	1.09761E-02	-22.1	0.5	2.1
3.38	1.09788E-02	-21.9	0.5	2.1
3.33	1.09827E-02	-21.6	0.5	2.1
3.34	1.09852E-02	-21.3	0.4	2.1
3.31	1.09830E-02	-21.5	0.6	2.2
3.34	1.09814E-02	-21.7	0.6	2.2
3.33	1.09679E-02	-22.9	0.5	2.1
3.32	1.09852E-02	-21.3	0.5	2.1

actor is -60.8‰) and BH-graphite (δ^{13} C = -8.2‰, IMF correction factor is -58.9‰)

Table 3: Comparative table of mineralogical and geochemical data between the Michigamme BIF and the Akilia Qp rock.

	Amphibolite facies Michigamme silicate BIF	Granulite facies Akilia Qp rock*
Age	ca. 1.85 Ga	>3.83 Ga
Metamorphic temperature	$T = 567 \pm 50^{\circ}C$	T > 635°C
Mineralogy	Grunerite, magnetite, quartz, pyroxene,	Quartz, pyroxene (hedenbergite-
	dolomite, stilpnomelane, chlorite, biotite,	ferrosilite), magnetite, grunerite-
	graphite, apatite, pyrrhotite, pyrite	hornblende, calcite, apatite, graphite, pyrrhotite, chalcopyrite, pentlandite,
Specific mineral associations	Apatite, dolomite, magnetite,	Apatite, calcite, magnetite,
with graphitic carbon	stilpnomelane, grunerite, chlorite, pyrrhotite	hornblende, chalcopyrite, pyrrhotite
δ^{13} Ccarb	-14.9%*	-3.3. to -5.5‰
$\delta^{13} C_{gra}$	-21.3 to -24.0‰	-4.1 to -49.0‰
Crystallinity of graphitic carbon	D-bands between 1346 and 1353 cm ⁻¹	D-bands between 1344 and 1358 cm
	(FWHM between 41 and 51 cm ⁻¹ and	(FWHM between 22 and 29 cm ⁻¹) and
	between 82 and 85 cm ⁻¹) and G-bands	G-bands between 1573 to 1581 cm ⁻¹
	between 1574 to 1581 cm ⁻¹ (FWHM between 41 and 77 cm ⁻¹) *	(FWHM between 10 and 46 cm ⁻¹); presence of curled graphite structures
d-spacing of graphitic carbon	3.55-3.72Å	3.41-3.64Å
Atomic N/C in graphitic carbon	10 ⁻⁴ to 10 ⁻¹	10 ⁻⁶ to 10 ⁻⁴
Heteroatoms in graphitic carbon	H, N, O, S, P, Ca, Fe	H, N, O, S, P, Fe
Molecular functional groups	Aliphatic and carboxyl (both minor)	None detected

^{*} Data from Papineau et al. (2010b) and Papineau et al. (2010a)

Figure 1 – Papine au et al. (2019) res_submitted-

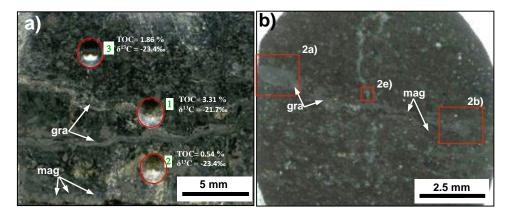


Figure 1: Petrography of graphite in the Michigamme silicate BIF. a) reflected light images of graphite layers in polished slabs where the C-isotope compositions were measured on micro-drilled powders (red circles), b) reflected light image of the round-shape polished slab used for SIMS spot analyses (red boxes 2a and 2b) and FIB-based nanofabrication (box 2e). Mineral abbreviations: gra = graphite, mag = magnetite.

Figure 2 – Papineau et al. (2019)

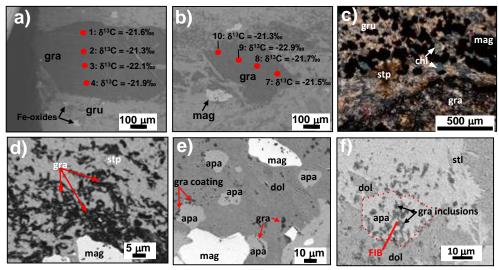


Figure 2: Petrography of apatite and graphite in the Michigamme silicate BIF. a-b) Secondary electron images of spot locations analyzed by SIMS showing the corrected δ^{13} C values in Table 2, c) cross-polar transmitted light image of the silicate matrix that embeds layers of coarse euhedral magnetite, d) BSE image of graphite filaments in silicate matrix, e) BSE image of apatite grains with graphite coatings in dolomite vein, and f) BSE image of apatite associated with graphite, dolomite, and phyllosilicate, targeted for FIB (lamella location is shown as a red line). Mineral abbreviations: gra = graphite, apa = apatite, cal = calcite, dol = dolomite (Mn and Fe-rich), gru = grunerite, bio = biotite, chl = chlorite, stl = stilpnomelane, mag = magnetite.

Figure 3 – Papineau et al. (2019)

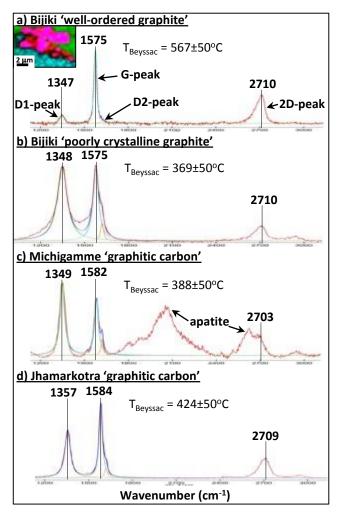


Figure 3: Raman spectra of mixed graphitic carbons in the immediate vicinity the nano-fabricated FIB lamella in Bijiki sample TVA294-659.6 (a-b) compared to other late Paleoproterozoic graphitic carbons (c-d). The inset is a hyperspectral Raman image of Bijiki graphitic carbons showing 'well-ordered graphite' in red (spectrum shown in (a)), mixed with PCG in purple (spectrum shown in (b)), and apatite and carbonate in turquoise and green, respectively. Similar spectra of graphitic carbons from contemporary phosphatic rocks metamorphosed at the greenschist to upper-greenschist facies are shown for comparison in c) for the Michigamme Formation (sample MA0708, Papineau et al., 2017) and d) for the Jhamarkotra Formation (sample UV0807, Papineau et al., 2016). The measured spectra of these graphitic carbons (red) were modelled using a linear combination of Lorentz-fitted peaks (blue) for each of the D1 (green), D2 (orange), and G (turquoise) peaks. Peak metamorphic temperatures were all estimated from the geothermometer of Beyssac et al. (2002), calibrated with minerals in metapelite.

Figure 4 – Papineau et al. (2019)

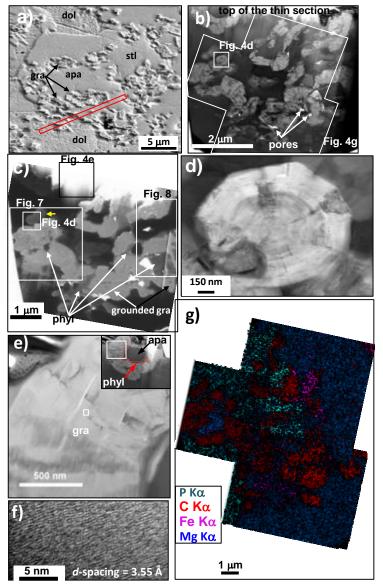


Figure 4: Apatite grain with graphite filaments from TVA294-659.6. a) secondary electron image of the targeted apatite grain, b) bright-field TEM image of graphite (light gray) in the apatite+dolomite matrix (dark gray) prior to Ne thinning — white spots are pores in the rock, c) He secondary electron image which best highlights the thin phyllosilicate (medium gray) coatings on graphite (light gray) inside apatite and dolomite (dark gray), d) cross-section view of concentrically-zoned and well-ordered graphite spherulite, e) longitudinal section of a well-ordered graphite filament associated with phyllosilicate (red area in inset) around and apatite core, f) HRTEM images showing lattice fringes for well-ordered graphite, g) EDS map of four major elements showing the close association of phyllosilicate (purple+blue) with graphite (red) with a legend for colours. Abbreviations: phyl = phyllosilicate, gra = graphite, dol = dolomite, apa = apatite.

Figure 5 - Papineau et al. (2019)

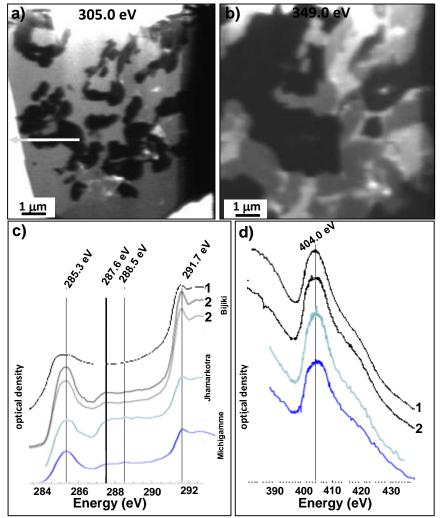


Figure 5: X-ray absorption images and spectra of graphitic carbons with apatite. a) 305.0 eV image, where graphitic carbon is the only phase strongly absorbing photons, hence appearing black. The white part of the arrow represents the line scan over the graphitic carbon (see also Fig. S4). b) 349.0 eV image, where Ca-bearing phases are the strongest absorbers, showing the distribution of Ca in apatite, dolomite (both black) and graphite in the FIB lamella. In this image, phyllosilicate is the brightest phase, closely associated with the distribution of graphitic carbon. XANES spectra at the c) C-edge and d) N-edge with major peaks identified. Graphitic carbons from Bijiki FIB lamella is labelled '1', acid-insoluble Bijiki graphitic carbons is labelled '2', the turquoise and blue spectra are from greenschist facies phosphorite from the Jhamarkotra Formation (turquoise; sample UV0807 from Papineau et al., 2016) and from a phosphatic chert at the Big Eric's crossing locality of the Michigamme Formation (blue; sample MA0708 from Papineau et al., 2017), respectively. Nitrogen was not detected by XANES in the phyllosilicate. See also Fig. S4b-S4e for broader energy context.

Figure 6 – Papineau et al. (2019)

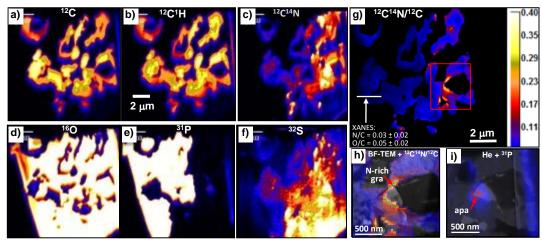


Figure 6: Geochemical context graphite associated with apatite and dolomite in Michigamme Fesilicate BIF. a-f) NanoSIMS secondary ion images. The distribution of graphite is visible in a) 12 C and b) 12 C 14 F images, and the co-location of organic N is shown in c). Dolomite-associated sulphate and the near absence of S in apatite can be seen in f). g) Map of the secondary 12 C 14 N/ 12 C ratio calibrated for atomic N/C ratios using XANES data. h) BF-TEM image superimposed by a semi-transparent 12 C 14 N/ 12 C map and i) He secondary electron image superimposed by semi-transparent 31 P, together showing N-rich graphitic carbon in contact with a nanoscopic grain of apatite. The location of these images is shown by the red box in (g).

Figure 7 - Papineau et al. (2019)

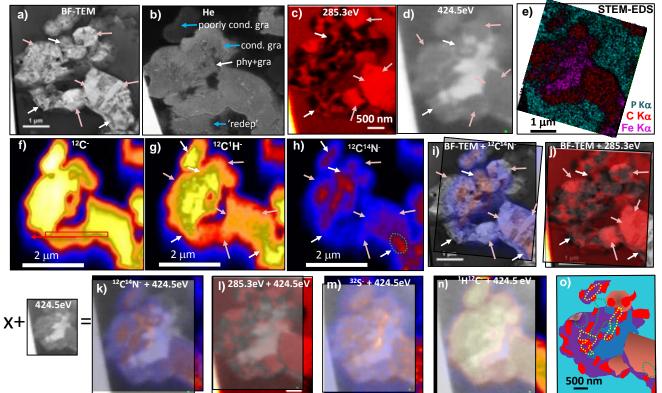


Figure 7: Correlated microscopy of filamentous graphitic carbon from Michigamme BIF that shows disordered graphitic carbon and the well-ordered graphite both correlate to compositionally-specific areas. a) BF-TEM image of the region of interest showing well-ordered graphite (pink arrows) and disordered graphitic carbon (white arrows) as distinguished by sub-domains with variable to random orientations. (b) secondary electron image from focused He beam showing phyllosilicate associated with graphitic carbon as well as highlighting the main contrast differences (blue arrows) in the He image attributed to the good electronic conductivity of re-deposition and parts of the graphitic carbon that are grounded (lightest gray) and poorly conductive graphite surrounded by apatite. X-ray images of graphite showing c) differences in absorption intensity at 285.3 eV are caused by the presence of variably-oriented nanoscopic aromatic domains (black) in disordered graphitic carbon (white arrows), whereas well-ordered graphite (pink arrows) is best highlighted in the brightest red contrast. d) At 424.5eV, the association of well-ordered graphite (pink arrows) and disordered graphitic carbon (white arrows) with less homogeneous gray contrast for the latter. Phyllosilicate is the brightest phase. e) STEM-EDS image of characteristic x-rays showing Fe in phyllosilicate, C in graphite, and P in apatite. Negative secondary ion images of f) ¹²C, showing homogeneous graphitic carbon (the red box shows the pixels selected for inter-instrumental calibration of atomic N/C, using the corresponding STXM line scan, Alléon et al., 2015), and g) ¹²C¹H⁻ and h) ¹²C¹⁴N⁻ showing nanoscopic heterogeneities within mostly the disordered graphitic carbon. Images superimposed on a correlated BF-TEM image are i) a secondary ion image of ¹²C¹⁴N⁻ and j) an X-ray image at 285.3 eV. Images superimposed over a correlated X-ray image at 424.5 eV, which shows phyllosilicate include: k) secondary ion image for ¹²C¹⁴N⁻ showing nanoscopic zones of disordered graphitic carbon and phyllosilicate enriched in N, I) X-ray image at 285.3 eV (in red) showing nanoscopic domains of aromatic carbon excluded from well-ordered graphite in the lower right, m) secondary ion image for ³²S⁻ showing nanoscopic Srich disseminations in the disordered graphitic carbon and a more homogeneous distribution in the well-ordered graphite, and n) secondary ion image for ¹²C¹H⁻ showing highest H levels in the disordered graphitic carbon. o) Interpretive summary diagram of phases, which shows chemical zones (green dotted line = N-rich and yellow line = Srich) preferentially located in the disordered graphitic carbon (turquoise = apatite; blue = phyllosilicate; purple = disordered graphitic carbon; pink shaded = concentric or aligned books of graphite; red = highly aromatic carbon).

Figure 8 – Papineau et al. (2019)

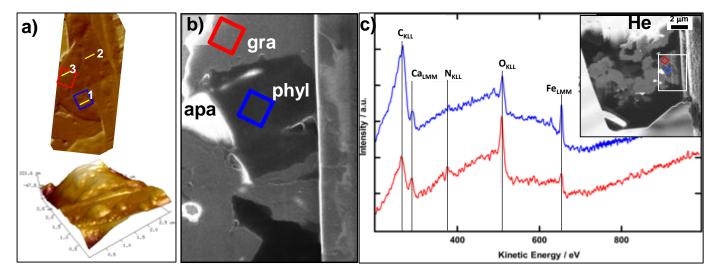


Figure 8: Nanoscale topography of lamella of Michigamme graphite polished using a 400 fA beam of Ne ions and analyzed by AFM and Auger spectroscopy. (a) AFM differential topographic height image of a section of the FIB lamella. Yellow lines indicate the locations of line profiles shown in Fig. S5. (b) He secondary electron image of the area where two spots were analyzed by Auger electron spectroscopy. (c) Two Auger spectra of the blue and red boxes in (b) with the localization in the lamella shown in inset. These show the presence of C, N, O, Ca and Fe in both graphite and the phyllosilicate, although the relative intensity of the peaks is affected by re-deposition on the lamella.

supplementary material

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