Journal of Endodontics

Effect of heated sodium hypochlorite on the viscoelastic properties of dentin evaluated using Dynamic Mechanical Analysis --Manuscript Draft--

Manuscript Number:	JOE 19-128R2				
Article Type:	Basic Research - Technology				
Corresponding Author:	Yuan-Ling Ng, BDS, MSc, MRD RCS (Eng), PhD, FHEA UCL Eastman Dental Institute London, UNITED KINGDOM				
First Author:	Nikoletta Kafantari, BDS MSc				
Order of Authors:	Nikoletta Kafantari, BDS MSc				
	Kishor Gulabivala, BDS, MSc, FDSRCS, PhD, FHEA, FACD				
	George Georgiou, PhD				
	Jonathan C Knowles, PhD, FIMMM, CEng, FRSC, CSci				
	Yuan-Ling Ng, BDS, MSc, MRD RCS (Eng), PhD, FHEA				
Manuscript Region of Origin:	Europe & Central Asia				
Abstract:	Introduction: This study aimed to investigate the effect of immersion of standardised dentin bars in 5% sodium hypochlorite (NaOCI) solutions at 60°C and 80°C on their viscoelastic properties, using Dynamic Mechanical Analysis (DMA). Methods: Eighteen intact teeth were used to produce ninety-nine dentin bars of standard dimensions (12mm×1mm×2mm) and randomly allocated to 6 groups (n=15 each) for immersion in: (1) Saline at 26°C; (2) Saline at 60°C; (3) Saline at 80°C; (4) NaOCI at 26°C; (5) NaOCI at 60°C; (6) NaOCI at 80°C. The bars were individually tested using DMA at baseline and after every 10 minutes of immersion in the test medium, up to 40 minutes. The effects of media, temperature, duration of exposure, and aspect ratio of bars on storage modulus (SM) and tan delta (TD) were investigated using Generalised Estimating Equations. Results: There was significant interaction between test medium and duration of immersion (p<0.05). The SM of specimens immersed in NaOCI at 60°C or 80°C decreased significantly (p<0.0001) over time of exposure, but the changes in other groups were minimal and insignificant. The TD of specimens immersed in saline 80°C (p<0.05), NaOCI at 60°C (p<0.05) or 80°C (p<0.0001) increased significantly over time of exposure but the change in NaOCI at 26°C was minimal. Other groups displayed negligible changes. Conclusions: NaOCI at 60°C or 80°C significantly reduced the elastic behaviour but increased the hysteresis of dentin under cyclic loading.				

Effect of heated sodium hypochlorite on the viscoelastic properties of dentin evaluated using Dynamic Mechanical Analysis

Kafantari Nikoletta¹, Gulabivala Kishor¹, Georgiou George², Knowles Jonathan^{2,3,4,5}, Ng Yuan-Ling¹ 3

¹Unit of Endodontology, Division of Restorative Dental Science; ²Division of Biomaterials & Tissue 6 Engineering; UCL Eastman Dental Institute; ³The Discoveries Centre for Regenerative and Precision ⁷ Medicine, Uinversity College London, London: ⁴Department of Nanobiomedical Science and BK21 Plus 8 NBM, Global Research Center for Regenerative Medicine, Dankook University, Cheonan, Republic of Korea, 518-10 Anseo-dong, Dongnam-qu, Cheonan, Chungcheongnam-do, Republic of Korea; ⁵UCL Eastman-Korea Dental Medicine Innovation Centre, Dankook University, Cheonan 31114, Republic of Korea 12

15 Authors

4

13 14

16 17

21

25 26

30

41 42

45

46 47

53

54 55 56

57 58

61 62

63 64 65

Ms Kafantari Nikoletta MSc

18 Unit of Endodontology, Division of Restorative Dental Science, UCL Eastman Dental Insitute, 256 Grays 19 20 Inn Road, London WC1X 8LD

22 Gulabivala Kishor PhD

23 Unit of Endodontology, Division of Restorative Dental Science, UCL Eastman Dental Insitute, 256 Grays 24 Inn Road, London WC1X 8LD

George Georgiou PhD

27 Division of Biomaterials & Tissue Engineering, UCL Eastman Dental Insitute, 256 Grays Inn Road, London 28 WC1X 8LD 29

31 Knowles Jonathan PhD

32 Division of Biomaterials and Tissue Engineering, University College London Eastman Dental Institute, 256 33 Gravs Inn Road, London WC1X 8LD, UK

The Discoveries Centre for Regenerative and Precision Medicine, UCL Campus, London, UK 35

36 Department of Nanobiomedical Science & BK21 PLUS NBM Global Research Center for Regenerative

Medicine, Dankook University, Cheonan 31114, Republic of Korea

³⁸ UCL Eastman-Korea Dental Medicine Innovation Centre, Dankook University, Cheonan 31114, Republic of 39 40 Korea

43 Ng Yuan-Ling PhD

⁴⁴ Unit of Endodontology, Division of Restorative Dental Science, UCL Eastman Dental Insitute, 256 Grays Inn Road, London WC1X 8LD

48 Acknowledgments

The authors affirm that there was and is no financial affiliation (e.g., employment, direct payment, stock 50 holdings, retainers, consultantships, patent licensing arrangements or honoraria), or involvement with any 51 commercial organization with direct financial interest in the subject or materials discussed in this 52 manuscript, nor have any such arrangements existed. The authors deny any other conflicts of interest related to this study.

To whom correspondence should be addressed:

₅₉ Dr Y-L Ng, Unit of Endodontology, UCL Eastman Dental Institute, 256 Grays Inn Road, London WC1X 8LD 60 United Kingdom. Tel: 020 34561233; Fax: 020 34562371; E-mail: y.ng@ucl.ac.uk

Statement of clinical relevance

Exposure of dentin to 5% NaOCI solutions heated to 60°C-80°C, over 40 minutes, reduced its elasticity and increased its <u>viscous behaviour or hysteresis</u>. These changes could potentially increase fracture susceptibility, particularly on sustained <u>loading</u>.

Highlights

- Non-destructive dynamic mechanical analysis allows investigation of the viscoelastic properties of dentin specimens within their elastic limit <u>by cyclic loading</u>.
- Immersion of dentin in 5% NaOCI solutions heated to 60°C or 80°C, over a 40-minute period, altered its viscoelastic properties by reducing elasticity (storage modulus) and increasing viscous behaviour (tan δ) with time.

Effect of heated sodium hypochlorite on the viscoelastic properties of dentin evaluated using Dynamic Mechanical **Analysis**

Abstract

Introduction: This study aimed to investigate the effect of immersion of standardised dentin bars in 5% sodium hypochlorite (NaOCI) solutions at 60°C and 80°C on their viscoelastic

properties, using Dynamic Mechanical Analysis (DMA).

Methods: Eighteen intact teeth were used to produce ninety-nine dentin bars of standard

dimensions (12mm×1mm×2mm) and randomly allocated to 6 groups (n=15 each) for

immersion in: (1) Saline at 26°C; (2) Saline at 60°C; (3) Saline at 80°C; (4) NaOCl at 26°C;

(5) NaOCI at 60°C; (6) NaOCI at 80°C. The bars were individually tested using DMA at

baseline and after every 10 minutes of immersion in the test medium, up to 40 minutes. The

effects of media, temperature, duration of exposure, and aspect ratio of bars on storage

modulus (SM) and tan delta (TD) were investigated using Generalised Estimating Equations.

Results: There was significant interaction between test medium and duration of immersion

(p<0.05). The SM of specimens immersed in NaOCI at 60°C or 80°C decreased significantly

(p<0.0001) over time of exposure, but the changes in other groups were minimal and

insignificant. The TD of specimens immersed in saline 80°C (p<0.05), NaOCl at 60°C

(p<0.05) or 80°C (p<0.0001) increased significantly over time of exposure but the change in

NaOCI at 26°C was minimal. Other groups displayed negligible changes.

Conclusions: NaOCI at 60°C or 80°C significantly reduced the elastic behaviour but

increased the hysteresis of dentin under cyclic loading.

Key words: Heated NaOCI, storage modulus, tan delta.

Introduction

Sodium hypochlorite (NaOCI) is the preferred root canal irrigant because of its antimicrobial (1) and tissue-dissolving (vital and necrotic) properties (2, 3). However, NaOCI also unfavourably alters flexural strength and elastic modulus of dentine (4, 5) and surface strain in "whole teeth" (6). The altered strain behaviour of teeth is worse when 5% NaOCI and 17% EDTA are used in alternation (6). This "whole tooth" strain behaviour is not entirely explained by quasi-static properties of dentin bars (4, 7). Non-destructive dynamic loading of dentin bars may better explain whole-tooth strain behaviour.

The changes in mechanical properties of dentin are attributed to depletion of dentinal collagen by NaOCI and mineral by EDTA (8, 9). Type I collagen accounts for 22% organic material by weight and contributes considerably to the mechanical properties of dentin (10).

The dissolution effect of NaOCI on pulp collagen increases with temperature because of an increased rate of reaction, prompting the adoption of heated solutions in dental practice (11). Collagen consisting of 3 helical polypeptide chains bound together by hydrogen and covalent bonds into a super-helix (12), undergoes various temperature-dependent and time-dependent changes (13). Collagen structure is altered to different degrees at different temperatures (20-200°C) and is influenced by levels of hydration and physical confinement in mineralized tissues (12-16).

Given the avocation of using heated NaOCI to 60°C (https://www.endoruddle.com/blogs/show/15/heating-sodium-hypochlorite-peer-reviewed-evidence-supporting-the-heating-of-naocl-, accessed 13/03/2019) and its testing at such temperatures (11) and beyond (17, 18), it would be logical to investigate the effect of heated sodium hypochlorite on the viscoelastic properties of dentin under dynamic loading (19). We hypothesized that exposure of dentin bars to heated NaOCI would significantly change their *viscoelastic* properties. The aim of this study was to investigate the effect of immersion of standardized dentin bars in 5%

NaOCI solutions at 60°C or 80°C over a period of 40 minutes, on their viscoelastic properties.

Materials and Methods

Preparation of dentin bars

Approval was granted for the use of extracted teeth from the UCL Eastman Biobank (Study number: 1301). Forty-one recently extracted wisdom teeth were stored in 4% formal-saline for 2 to 12 weeks before commencement of experiment. Eighteen intact, disease- and crack-free teeth (from patients aged 18-35 years) with fused roots of at least 13 mm length, were selected. Adherent soft or hard tissues were removed gently from the teeth with a scalpel.

The teeth were sectioned longitudinally in the mesio-distal plane and slices were produced in the bucco-lingual plane using a revolving diamond-coated saw (Leica Instruments Gmbtt Mod. 1.600, D-6907 Nussloch, Germany). Dentin bars, of standard dimensions (12 mm × 1 mm × 2 mm) were produced under copious water irrigation and their length and width measured with a micrometer (Moore and Wright, Bradford, UK) and recorded (Pyris software, Perkin Elmer instruments, Beaconsfield, UK). Each tooth produced two to six bars, mainly from the root, yielding a total of ninety-nine bars; all specimens were marked with an indelible pen to identify the surface facing the canal lumen and stored in 0.9% saline solution (Seven Biotech Ltd., Kidderminster, UK). The dentinal tubules ran across the bar so that loading was parallel to the dentinal tubules.

Allocation of dentin bars to test groups

Ninety dentin bars were randomly assigned to six groups (n=15), three were used for a pilot study (Table 1) and six were randomly pre-assigned as 'spares' in case of need for replacement. All dentin bars were stored in saline just prior to the experiment.

Preparation of sodium hypochlorite (5%) test solution

Stock sodium hypochlorite solution (6-14% active chlorine) (Merck KGaA, Darmstadt, Germany) was diluted with distilled water to produce 5% NaOCI, confirmed by iodometric titration, just before use.

The test solutions (40 mL) were decanted into individual 120 mL glass jars and heated to the designated temperatures (60°C and 80°C) over a hot-plate (Jenway 1000, Chelmsford, Essex, UK). The temperature was monitored with a thermometer (Fisherbrand, Thermo Fisher Scientific Inc., Loughborough, UK) throughout the experiment. The glass jar remained on the hot plate, covered with a lid during the immersion period, as the solution was constantly stirred. Fresh solution was prepared and heated for each bar.

Dynamic mechanical analysis (DMA) test conditions

The viscoelastic properties of dentin were evaluated at baseline and after every 10 minutes of immersion in the test medium, up to 40 minutes, using the 3-point bend test in a DMA-7e (Perkin Elmer instruments, Beaconsfield, UK) with Pyris software (Perkin Elmer instruments, Beaconsfield, UK). The loss and storage moduli and tan δ were recorded. The specimens were subjected to oscillatory and static deformation. A static tension control of 120% (i.e. dynamic stress x 1.2 = static stress) and a constant dynamic strain control of 0.02% (deformation over length of specimen) were maintained. The initial loads applied were 200 mN and 240 mN for dynamic and static force, respectively; these parameters adjust accordingly, but at the same ratio with respect to each other, so as to satisfy the dynamic strain control conditions throughout testing. The frequency of loading was held at 1 Hz. All tests were performed at room temperature (26°C).

Upon removal from the storage solution, a test run was performed before immersion in its respective test solution. The duration of each run was one minute and the readings for storage modulus, loss modulus and δ were taken at 30 seconds into each run. The total

time to complete each test, including the time to equilibrate the apparatus (for each test) and establish a stable dynamic strain was approximately 7 minutes.

The specimens were then immersed in their respective test solutions for 10 minutes and then quenched by rinsing in saline for three minutes before the test. The bars were left damp during the test and their orientation maintained constant with the marked side under compression so that the dentinal tubules had a consistent orientation to the probe. The cycle of immersion, wash and test-run was repeated 4 times until a total of 40 minutes exposure to the test medium was completed for each sample.

Data analysis

The data for storage modulus, loss modulus and Tan δ were recorded and stored on Pyris Data Analysis software and analyzed using the SPSS statistical software package (SPSS Inc., Chicago, IL, USA). The means and 95% confidence intervals of storage modulus (SM) and tan δ were plotted against the duration of immersion by each group. Generalized Estimating Equations (GEE) were used to investigate the effects of test medium and duration of immersion on SM and Tan δ , accounting for the effects of specimen dimensions.

Results

It was observed that the bars in groups with heated NaOCI at 60°C and 80°C effervesced during immersion in the test solution, gaining a chalky appearance; those in the latter group being more obvious.

The mean values and 95% confidence intervals for SM and tan δ at baseline and following each 10-minute immersion for all test and control groups are presented in figure 1. The mean SM at baseline (t = 0 minute) varied between 14.35 GPa (95% confidence interval [CI]: 12.41 – 16.29 GPa) and 19.92 GPa (95% CI: 17.25 – 22.59), while the mean tan δ at

baseline (t = 0 minute) varied between 0.077 (95% CI: 0.069 - 0.085) and 0.098 (95% CI: 0.089 - 0.108) amongst the test groups, respectively.

In groups with NaOCl 60°C and 80°C, a clear downward trend was evident in the storage modulus values (reduced by 5.6 [95% Cl: 4.3, 6.8] GPa, 7.4 [95% Cl: 5.1, 9.7] GPa, respectively) with time but an upward trend in the tan δ values (increased by 0.03 [95% Cl: 0.02, 0.05] and 0.07 [95% Cl: 0.03, 0.10] units, respectively).

The GEE models revealed test medium and duration of immersion had significant (p<0.0001) interactive effects on SM and $\tan \delta$ of the dentin bars, respectively (Tables 2&3). This indicated that the duration of immersion had different effects among different test media. Every minute of immersion in NaOCI at 60°C or 80°C resulted in 0.014 Pa (p<0.0001) or 0.019 Pa (p<0.0001) reduction in SM, respectively. Concomitantly, there was an increase in $\tan \delta$ by 0.001 or 0.002 units for every minute of immersion in NaOCI at 60°C (p<0.01) or 80°C (p<0.0001), respectively. Specimens immersed in saline at 80°C (p<0.05) also displayed significant increase in $\tan \delta$, albeit small in magnitude.

The reduction of SM (p<0.0001) and increase in tan δ (p<0.05) amongst samples in the heated NaOCI groups was significantly greater than those in the heated Saline groups, regardless of temperature.

Discussion

The aspect ratio of the dentin bars can influence outcomes and thus every effort was made to minimize sample deviation. The potential effect of aspect ratios (ratio of the span between the supports for the specimen and the height) was accounted for in the regression models and found to be statistically insignificant (p>0.05).

The potentiating effect of heat on the antibacterial and tissue-dissolving properties of sodium hypochlorite has been clinically applied using solution temperatures of up to 60°C

(https://www.endoruddle.com/blogs/show/15/heating-sodium-hypochlorite-peer-reviewed-evidence-supporting-the-heating-of-naod-, accessed 13/03/2019). Previous studies had tested NaOCI at 21°C to 60°C (11); the upper limit temperature in this study (80°C) was used as a positive control because the tri-helical structure of collagen denatures between 65°C – 80°C in a specific temperature-dependent manner (12, 13, 20).

Storage of teeth in formal-saline (4%) was found to have no effect on the viscoelastic properties of dentin harvested in a pilot study, even though it is known that it can induce collagen cross-links that may affect the denaturation temperature of extracted native collagen (21). Other forms of storage, such as freezing, may also induce conformational and cross-linking changes in collagen but may affect the thermodynamic properties of collagen to a greater degree (22).

The overall duration of test was based on the average root canal treatment procedure duration plus pilot studies, which showed that viscoelastic properties of dentin reached a plateau between 40-60 minutes. Every effort was made to reposition the bars accurately on the DMA platform for each sequential test and to ensure the orientation of the tubules remained constant in relation to the probe.

This study found no significant change in $\tan \delta$ and SM values with time (over 40 minutes) for dentin bars immersed in saline at 26°C, as anticipated from thermo-mechanical studies (12). Those immersed in saline at 60°C also showed no significant change in either $\tan \delta$ or SM over time, however, the saline 80°C group, although showing no significant change in SM, indicated a significant albeit small increase in $\tan \delta$ with time. The minor change in viscous behaviour was clinically insignificant in respect of the absolute $\tan \delta$ values.

Temperature-mediated collagen denaturation is dependent on hydration, neutral salt concentration and confinement of the collagen molecules (14, 23). Unconfined collagen may begin denaturing at around 60°C (23) but is substantially affected by hydration (15). In the

present study, dehydration was not a risk because of constant water saturation but the molecules are confined within mineralised structures, which is said to confer a stabilising effect on thermal denaturation by a so-called "polymer-in-a-box" mechanism (15).

Static mechanical properties may undergo a linear decrease with increasing temperature from 0-80°C (24), although flexural strength and microtensile strength increased with increasing temperature from 50°C to 200°C (25). These changes are reversed upon rehydration and have been attributed to dehydration and the formation of cross-linkages that are reversible upon rehydration (26). Yamashita *et al.* (27) assessed the viscoelasticity of bone heated to 100°C or 200°C with DMA and concluded that heat was not the main contributor to the increased viscoelasticity of bone. The observed increase in viscoelastic behaviour may reflect increasing compliance of the collagen network with increase in temperature and greater absorption of water beginning at 60°C to around 80°C, possibly enhanced by changes in collagen conformation (28, 29).

Dentin bars immersed in NaOCI at 60° C exhibited a significant decrease in SM and an appreciable increase in $\tan \delta$, while those in the NaOCI 80° C showed significant decreases in SM coupled with significant increases in $\tan \delta$. Notwithstanding, the individual specimen variations, the change in the viscoelastic behaviour of dentin in these two groups can clearly be attributed to the heated NaOCI. The detectable changes in the viscoelastic properties of dentin in the heated NaOCI groups may be explained by NaOCI-mediated dentin surface depletion of collagen and deeper denaturation of collagen within dentin (8). These would be manifested as alterations in chemistry and structure of collagen (8, 12).

Collagen denaturation by heat is mediated by unfolding of its triple helices into individual helices due to disruption of hydrogen bonds between the polypeptide chains; the individual helices may then unfold into random coils due to disruption of intra-helical hydrogen bonds (12). Such unfolding may enhance water flow and hydration of the exposed collagen matrix, increasing viscoelastic behaviour (28).

The decrease in SM also signals some effect on the mineral matrix or mineral-collagen interaction (30). Whilst it is known that mineral may be lost from dentine under saline (31) or NaOCI (32) immersion, this is unlikely to be the mechanism at play. It is more likely that the heated NaOCI has a direct effect on the interface between mineral and collagen. This is consistent with the effect of saline at 80°C, where there may be a mild effect on the collagen-mineral bonding interface through increase in the mean length of vibrating chain segments (29). Collagen shrinks on heating at 70°C in saline solution but falls apart at temperatures higher than 80°C (16). In the presence of NaOCI, the effect is likely enhanced by deeper penetration of NaOCI into dentin and thus probable damage to protein interface with the mineral structure (12). This may cause anelastic behaviour in the mineral matrix (29), coupled with greater damping effect. The potential clinical relevance of these two phenomenon is that the increased viscosity would cause greater strain, whilst the loss of elasticity or anelastic behaviour may delay recovery. This enhanced hysteresis, may upon sustained loading cause or support microcracking and subsequent structure failure (33).

The above observed effects on viscoelastic properties are likely to be a function of dependent rather than independent effects between the medium and temperature given the negligible and insignificant effects of heated saline and unheated NaOCl on SM or $\tan \delta$.

In the present experimental model, the temperature was effectively maintained at the designated level but when heated NaOCI is delivered into the root canal system, the temperature may drop in the syringe within 4 minutes (17). Although, heat transfer into dentin may be limited (34), temperature changes to 60°C can thermally induce deformation of human dentin (35). Therefore, further investigation of the depth and nature of chemical changes *in situ* within roots, associated with heated NaOCI irrigation is merited, together with their effects on tooth surface strain (6).

Conclusions

Immersion of standardized dentin bars in saline solution at 80° C resulted in small but significant increase in their tan δ with time. Immersion in NaOCI solutions heated to 60° C or 80° C, significantly reduced storage modulus and increased tan δ with time.

Figure caption

Figure 1: Means (solid lines) and upper and lower limits of 95% confidence intervals (dotted lines) of storage modulus (GPa) or tan delta plotted against time (minutes), by test medium and temperature

Table captions

- **Table 1:** Test and control group composition.
- **Table 2:** Multivariable GEE model incorporating Storage modulus as the dependent variable, and test medium and duration of immersion as the independent variables.
- **Table 3:** Multivariable GEE model incorporating Tan δ as the dependent variable, and test medium and duration of immersion as the independent variables.

References

- 1. Bystrom A, Sundqvist G. Bacteriologic evaluation of the effect of 0.5 percent sodium hypochlorite in endodontic therapy. Oral Surg Oral Med Oral Pathol 1983;55(3):307-312.
- 2. Naenni N, Thoma K, Zehnder M. Soft tissue dissolution capacity of currently used and potential endodontic irrigants. J Endod 2004;30(11):785-787.
- 3. Rosenfeld EF, James GA, Burch BS. Vital pulp tissue response to sodium hypochlorite. J Endod 1978;4(5):140-146.
- 4. Sim TP, Knowles JC, Ng YL, Shelton J, Gulabivala K. Effect of sodium hypochlorite on mechanical properties of dentine and tooth surface strain. Int Endod J 2001;34(2):120-132.
- 5. Zhang K, Tay FR, Kim YK, Mitchell JK, Kim JR, Carrilho M, et al. The effect of initial irrigation with two different sodium hypochlorite concentrations on the erosion of instrumented radicular dentin. Dent Mater 2010;26(6):514-523.
- 6. Rajasingham R, Ng YL, Knowles JC, Gulabivala K. The effect of sodium hypochlorite and ethylenediaminetetraacetic acid irrigation, individually and in alternation, on tooth surface strain. Int Endod J 2010;43(1):31-40.
- 7. Grigoratos D, Knowles J, Ng YL, Gulabivala K. Effect of exposing dentine to sodium hypochlorite and calcium hydroxide on its flexural strength and elastic modulus. Int Endod J 2001;34(2):113-119.
- 8. Ramirez-Bommer C, Gulabivala K, Ng YL, Young A. Estimated depth of apatite and collagen degradation in human dentine by sequential exposure to sodium hypochlorite and EDTA: a quantitative FTIR study. Int Endod J 2018;51(4):469-478.
- 9. Ryou H, Amin N, Ross A, Eidelman N, Wang DH, Romberg E, et al. Contributions of microstructure and chemical composition to the mechanical properties of dentin. J Mater Sci Mater Med 2011;22(5):1127-1135.
- 10. Miguez PA, Pereira PN, Atsawasuwan P, Yamauchi M. Collagen cross-linking and ultimate tensile strength in dentin. J Dent Res 2004;83(10):807-810.
- 11. Sirtes G, Waltimo T, Schaetzle M, Zehnder M. The effects of temperature on sodium hypochlorite short-term stability, pulp dissolution capacity, and antimicrobial efficacy. J Endod 2005;31(9):669-671.
- 12. Wang X, Li X, Bank RA, Agrawal CM. Effects of collagen unwinding and cleavage on the mechanical integrity of the collagen network in bone. Calcif Tissue Int 2002;71(2):186-192.
- 13. Suwa Y, Nam K, Ozeki K, Kimura T, Kishida A, Masuzawa T. Thermal denaturation behavior of collagen fibrils in wet and dry environment. J Biomed Mater Res B Appl Biomater 2016;104(3):538-545.
- 14. Hayashi M, Furuya Y, Minoshima K, Saito M, Marumo K, Nakashima S, et al. Effects of heating on the mechanical and chemical properties of human dentin. Dent Mater 2012;28(4):385-391.
- 15. Miles CA, Ghelashvili M. Polymer-in-a-box mechanism for the thermal stabilization of collagen molecules in fibers. Biophys J 1999;76(6):3243-3252.
- 16. Vangsness CT, Jr., Mitchell W, 3rd, Nimni M, Erlich M, Saadat V, Schmotzer H. Collagen shortening. An experimental approach with heat. Clin Orthop Relat Res 1997(337):267-271.
- 17. Macedo RG, Verhaagen B, Versluis M, van der Sluis L. Temperature evolution of preheated irrigant injected into a root canal ex vivo. Clin Oral Investig 2017;21(9):2841-2850.
- 18. Iandolo A, Amato M, Dagna A, Poggio C, Abdellatif D, Franco V, et al. Intracanal heating of sodium hypochlorite: Scanning electron microscope evaluation of root canal walls. J Conserv Dent 2018;21(5):569-573.
- 19. Ryou H, Romberg E, Pashley DH, Tay FR, Arola D. Nanoscopic dynamic mechanical properties of intertubular and peritubular dentin. J Mech Behav Biomed Mater 2012;7:3-16.
- 20. Bozec L, Odlyha M. Thermal denaturation studies of collagen by microthermal analysis and atomic force microscopy. Biophys J 2011;101(1):228-236.

- 21. Fathima NN, Madhan B, Rao JR, Nair BU, Ramasami T. Interaction of aldehydes with collagen: effect on thermal, enzymatic and conformational stability. Int J Biol Macromol 2004;34(4):241-247.
- 22. Badii F, Howell NK. Elucidation of the effect of formaldehyde and lipids on frozen stored cod collagen by FT-Raman spectroscopy and differential scanning calorimetry. J Agric Food Chem 2003;51(5):1440-1446.
- 23. Rochdi A, Foucat L, Renou JP. Effect of thermal denaturation on water-collagen interactions: NMR relaxation and differential scanning calorimetry analysis. Biopolymers 1999;50(7):690-696.
- 24. Watts DC, el Mowafy OM, Grant AA. Temperature-dependence of compressive properties of human dentin. J Dent Res 1987;66(1):29-32.
- 25. Hayashi M, Koychev EV, Okamura K, Sugeta A, Hongo C, Okuyama K, et al. Heat treatment strengthens human dentin. J Dent Res 2008;87(8):762-766.
- 26. Hayashi M, Okamura K, Koychev EV, Furuya Y, Sugeta A, Ota T, et al. Effects of rehydration on dentin strengthened by heating or UV irradiation. J Dent Res 2010;89(2):154-158.
- 27. Yamashita J, Li X, Furman BR, Rawls HR, Wang X, Agrawal CM. Collagen and bone viscoelasticity: a dynamic mechanical analysis. J Biomed Mater Res 2002;63(1):31-36.
- 28. McGee MP, Morykwas M, Shelton J, Argenta L. Collagen unfolding accelerates water influx, determining hydration in the interstitial matrix. Biophys J 2012;103(10):2157-2166.
- 29. Amadori S, Bonetti E, Campari EG, Cappelloni I, Montanari R. Anelastic phenomena associated to water loss and collagen degradation in human dentin. Mater Sci Eng C Mater Biol Appl 2013;33(3):1455-1459.
- 30. Forien J-B, Zizak I, Fleck C, Petersen A, Fratzl P, Zolotoyabko E, et al. Water-mediated collagen and mineral nanoparticle interactions guide functional deformation of human tooth dentin. Chemistry of Materials 2016;28(10):3416-3427.
- 31. Habelitz S, Balooch M, Marshall SJ, Balooch G, Marshall GW, Jr. In situ atomic force microscopy of partially demineralized human dentin collagen fibrils. J Struct Biol 2002;138(3):227-236.
- 32. Sakae T, Mishima H, Kozawa Y. Changes in bovine dentin mineral with sodium hypochlorite treatment. J Dent Res 1988;67(9):1229-1234.
- 33. Eltit F, Ebacher V, Wang R. Inelastic deformation and microcracking process in human dentin. J Struct Biol 2013;183(2):141-148.
- 34. Lin M, Xu F, Lu TJ, Bai BF. A review of heat transfer in human tooth--experimental characterization and mathematical modeling. Dent Mater 2010;26(6):501-513.
- 35. Kishen A, Murukeshan VM, Krishnakumar V, Asundi A. Analysis on the nature of thermally induced deformation in human dentine by electronic speckle pattern interferometry (ESPI). J Dent 2001;29(8):531-537.



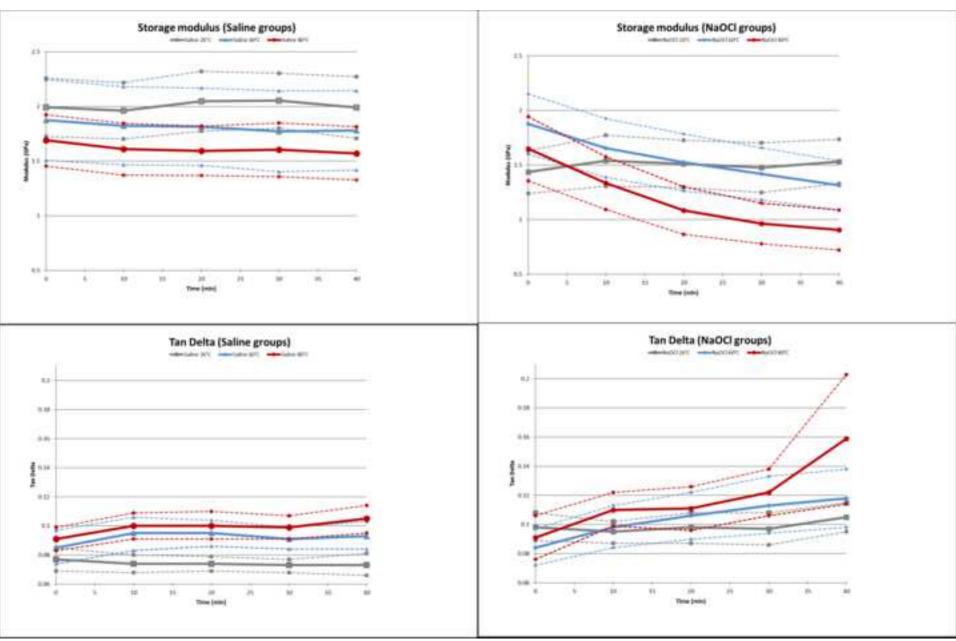


Table 1: Test and control group composition.

Groups	Test medium	Temperature	No. of specimens	Immersion time (mins)	Loading time (min)
1 (control)	Saline (0.9%)	26°C	15	40 (4×10)	1
2	Saline (0.9%)	60°C	15	40 (4×10)	1
3	Saline (0.9%)	80°C	15	40 (4×10)	1
4	NaOCI (5%)	26°C	15	40 (4×10)	1
5	NaOCI (5%)	60°C	15	40 (4×10)	1
6	NaOCI (5%)	80°C	15	40 (4×10)	1

Table 2: Multivariable GEE model incorporating Storage modulus as the dependent variable, and test medium and duration of immersion as the independent variables

Variable	Coefficient	95% Wald Confidence Interval		P value
		Lower	Upper	
Test medium				
Saline 26°C (Ref)	-	-	-	-
Saline 60°C	-0.130	-0.529	0.270	0.5
Saline 80°C	-0.329	-0.643	-0.014	0.04
NaOCI 26°C	-0.518	-0.820	-0.216	0.001
NaOCl 60°C	-0.162	-0.506	0.182	0.4
NaOCI 80°C	-0.432	-0.770	-0.093	0.01
Time (minute)				
Saline 26°C*Time	0.001	-0.002	0.004	0.6
Saline 60°C*Time	-0.003	-0.007	< 0.0001	0.09
Saline 80°C*Time	-0.003	-0.007	< 0.0001	0.08
NaOCI 26°C*Time	< 0.0001	-0.004	0.005	0.8
NaOCl 60°C*Time	-0.014	-0.019	-0.010	< 0.0001
NaOCI 80°C*Time	-0.019	-0.025	-0.014	< 0.0001

Time = duration of immersion; * = interaction of test solution with duration of exposure

Table 3: Multivariable GEE model incorporating Tan δ as the dependent variable, and test medium and duration of immersion as the independent variables.

Variable	Coefficient	95% Wald Confidence Interval		P value
		Lower	Upper	
Test medium				
Saline 26°C (Ref)	-	-	-	-
Saline 60°C	0.014	0.002	0.025	0.02
Saline 80°C	0.018	0.009	0.026	< 0.0001
NaOCI 26°C	0.019	0.011	0.028	< 0.0001
NaOCI 60°C	0.011	< 0.0001	0.023	0.06
NaOCI 80°C	0.013	0.003	0.024	0.02
Time (minute)				
Saline 26°C*Time	-<0.0001	< 0.0001	< 0.0001	0.3
Saline 60°C*Time	< 0.0001	< 0.0001	< 0.0001	0.4
Saline 80°C*Time	< 0.0001	< 0.0001	0.001	0.04
NaOCI 26°C*Time	< 0.0001	< 0.0001	< 0.0001	0.3
NaOCl 60°C*Time	0.001	0.001	0.001	0.001
NaOCI 80°C*Time	0.002	0.001	0.002	<0.0001

Time = duration of immersion; * = interaction of test solution with duration of exposure