

1  
2 1 **Demodicosis in a captive African straw-coloured fruit bat (*Eidolon***  
3  
4 2 ***helvum*)**  
5

6 3  
7  
8 4 **Running title:** Demodicosis in an African fruit bat  
9

10 5  
11  
12 6 **Carlo Bianco<sup>1,2</sup>, Kate S. Baker<sup>3</sup>, Luca Pazzini<sup>4</sup>, Alessandra Cafiso<sup>5</sup>, Richard D. Suu-Ire<sup>6,7</sup>,**  
13  
14 7 **Andrew A. Cunningham<sup>8</sup>, James L. N. Wood<sup>9</sup>, Alejandro Nuñez<sup>1</sup>**  
15 8

16  
17 9 <sup>1</sup>Pathology Department, Animal Plant and Health Agency, Woodham Ln, Addlestone, Surrey,  
18  
19 10 KT15 3NB, UK; <sup>2</sup>Diagnostic & Consultant Avian Pathology, Pathology Department, Animal and  
20  
21 11 Plant Health Agency (APHA-Lasswade) Pentlands Science Park, Bush Loan, Penicuik, Midlothian,  
22  
23 12 EH26 0PZ, UK; <sup>3</sup>Institute of Integrative Biology, Biosciences Building University of Liverpool  
24  
25 13 Crown Street University of Liverpool, Liverpool, L69 7ZB, UK; <sup>4</sup>Veterinary Clinic 'Centro storico'  
26  
27 14 Via dei Vanga, 51, 39100 Bolzano (BZ), Italy; <sup>5</sup>Department of Veterinary Medicine, University of  
28  
29 15 Milan, Via Celoria 10, 20133 Milan, Italy; <sup>6</sup>Wildlife Division, Forestry Commission, Box M239  
30  
31 16 Accra, Ghana; <sup>7</sup>Veterinary Services, Ministry of Food and Agriculture, Box M161, Accra, Ghana;  
32  
33 17 <sup>8</sup>Institute of Zoology, Zoological Society of London, Regent's Park, NW1 4RY, UK; <sup>9</sup>Department  
34  
35 18 of Veterinary Medicine, University of Cambridge, Madingley Road, Cambridge CB3 0ES, UK  
36  
37 19

38 20 **<sup>1</sup>Corresponding author:** Diagnostic & Consultant Avian Pathology, Pathology Department,  
39  
40 21 Animal and Plant Health Agency (APHA-Lasswade) Pentlands Science Park, Bush Loan, Penicuik,  
41  
42 22 Midlothian, EH26 0PZ, UK. +44 020 8225 7611 E-mail: carlo.bianco@apha.gov.uk  
43  
44  
45 23  
46  
47 24

48  
49 25 **ABSTRACT**  
50  
51 26

52 27 Demodicosis is most frequently observed in the domestic dog (*Canis familiaris*), but it has rarely  
53  
54 28 been reported in bats (Chiroptera). The overpopulation of *Demodex* spp. that causes dermatological  
55  
56 29 changes is generally associated with a compromised immune system. We describe the gross and  
57  
58 30 histological features of generalized demodicosis in an adult female African straw-coloured fruit bat  
59  
60 31 (*Eidolon helvum*) drawn from a captive research colony. The histology of the lesions revealed  
61  
62 32 comedones and follicular infundibular cysts harbouring numerous *Demodex* spp. mites, eliciting a  
63  
64 33 minimal inflammatory response in the adjacent dermis. The histological examination of a full set of  
65  
66 34 tissues did not reveal clear evidence of immunosuppression, although a clinical history of recent



1  
2 1 abortion and possible stressors due to captivity could be considered risk factors for the demodicosis.  
3 2 Attempts to determine the *Demodex* species using PCR on DNA extracted from the formalin fixed  
4 3 paraffin embedded tissue failed. This is the first clinical and histological description of demodicosis  
5 4 in *Eidolon helvum*.  
6  
7  
8  
9 5

10  
11 6 **Keyword:** bat, *Demodex*, dermatology, veterinary pathology, wildlife diseases.  
12  
13 7  
14 8  
15

## 16 9 INTRODUCTION

17  
18 10 *Demodex* mites (Acarina: Prostigmata: Demodicidae) are obligate host-specific mammalian ecto-  
19 11 parasites inhabiting the pilo-sebaceous units or the adjacent *stratum corneum* of the skin. Over 140  
20 12 *Demodex* species have been identified in at least 11 orders of domestic and wild mammals,  
21 13 reflecting the constancy of the sheltered ecologic niche (Sastre et al. 2016). The mites' primary food  
22 14 source are follicular keratinocytes and the secretions of sebaceous glands (Gross et al. 2005;  
23 15 Mauldin et al. 2015). Low population densities of mites are considered commensals as part of the  
24 16 normal skin fauna (Gross et al. 2005; Mauldin et al. 2015). The host's immune system inhibits their  
25 17 proliferation, keeping mite numbers low in the absence of an inflammatory response (Gross et al.  
26 18 2005; Mauldin et al. 2015). *Demodex* mite infestations usually remain asymptomatic, but these  
27 19 mites can be important causative agents for clinical dermatological disease, called demodicosis or  
28 20 demodectic mange.  
29  
30

31 21 Demodicosis is defined as a non-contagious parasitic inflammatory skin disease  
32 22 characterized by the presence of an overpopulation of *Demodex* spp. mites. The disease is often  
33 23 associated with immunodepression (a defect in the immune system), underlying  
34 24 immunosuppressive disease or debilitating conditions such as poor nutrition, stress, concurrent  
35 25 diseases or genetic predisposition. Demodicosis is distributed worldwide and is reported most  
36 26 frequently in domestic dogs and cats. In veterinary dermatology, canine and feline demodicosis may  
37 27 be divided into two forms based on the clinical manifestation: a generalized form, a potentially life-  
38 28 threatening disease, and a localized form, a mild self-limiting disease. The gross clinical features of  
39 29 canine demodicosis are typically small, well-circumscribed, erythematous, scaly, multifocal areas of  
40 30 alopecia that can coalesce, and sometimes form nodular lesions (Gross et al. 2005; Mauldin et al.  
41 31 2015). The three histopathological patterns of inflammation are folliculitis and furunculosis,  
42 32 parafollicular granulomas, and lymphocytic mural interface folliculitis (Gross et al. 2005; Mauldin  
43 33 et al. 2015). All patterns may coexist, or individual patterns may predominate, and follicular  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65



1 melanosis with pigmentary incontinence frequently occurs. In very rare cases, mites may greatly  
2 distend superficial hair follicles to form comedone-like lesions (Gross et al. 2005).

3  
4  
5 In other domestic species, apart from the goat and cattle in Africa, demodicosis is  
6 considered rare (Mauldin et al., 2015). The clinical appearance of demodicosis in goats is papulo-  
7 nodular, and histologically the nodules are follicular cysts lined by flattened squamous epithelium,  
8 filled with keratin scales and large numbers of demodicid mites. A mild inflammatory infiltrate may  
9 occur around the epithelial lining of affected hair follicles (Mauldin et al. 2015). Demodicosis has  
10 rarely been reported in bats (Chiroptera). Here we describe the gross and histological features of  
11 generalized demodicosis in an adult female African straw-coloured fruit bat (*Eidolon helvum*)  
12 drawn from a captive research colony.  
13  
14  
15  
16  
17  
18  
19  
20

## 21 MATERIAL AND METHODS

### 22 Clinical history of the animal and post mortem investigations

23  
24  
25 An adult female African straw-coloured fruit bat maintained within a group of 111 conspecifics for  
26 about 9 months, in a large flight cage (closed to public view) in the grounds of Accra Zoological  
27 Gardens in Achimota Forest Reserve, Accra, Ghana, was humanely euthanised on welfare grounds  
28 due to the presence of numerous cutaneous nodular lesions across the face and ventrum. The subject  
29 had been caught as part of a viral antibody dynamic study and tagged for individual identification  
30 following a protocol approved by the Zoological Society of London's Ethic Committee (Baker et al.  
31 2014). Although pregnant when placed in the colony, the bat suffered an abortion after 5 months.  
32 Two months later a laceration to the leading-edge of the *patagium* was observed. The cutaneous  
33 nodular lesions were observed after a further 2 months, and the animal was euthanised  
34 subsequently.  
35  
36  
37  
38  
39  
40  
41  
42  
43

44 A representative area of cutaneous lesions and samples from internal organs was fixed in  
45 formalin and paraffin embedded for routine histopathology (haematoxylin and eosin stained slides).  
46 Formalin-fixed, paraffin embedded sample of skin was subjected to xylene treatment. Total DNA  
47 isolation was performed using the QIAamp DNA Investigator kit (Qiagen). DNA was eluted in 30  
48 µl of sterile water, quantified (Nanodrop 1000 Spectrophotometer) and stored at -80 °C until  
49 molecular analyses. Quality of the extraction was assessed by PCR amplification of two gene  
50 fragments of the mammal host. 12S rRNA and mitochondrial 16S rRNA gene fragments (expected  
51 fragments size: 200-250 bp) were amplified using universal PCR primers designed for the  
52 identification of vertebrate species (Kitano et al. 2006). The presence of *E. helvum* DNA was  
53 assessed in both PCR assays performed with universal vertebrate primers, the result was confirmed  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65



1 by Sanger sequencing.

2 For the molecular identification of the mite, the PCR amplification was performed using  
3 both primers for species belonging to the genus *Demodex* and broad-range primers for the suborder  
4 Prostigmata. The PCR amplifications performed specifically for *Demodex* spp. were targeted on: i)  
5 a 340 bp 16S rRNA gene fragment (Zhao and Wu 2012); ii) a 429 bp *COXI* gene fragment (Zhao et  
6 al. 2013); a 230 bp 18S rDNA gene fragment (Zewe et al. 2017). The broad-range PCR  
7 amplification for the suborder Prostigmata targeted on a 500 bp 18S rDNA gene fragment (Otto and  
8 Wilson 2001). PCR amplifications were performed on undiluted and serially diluted DNA (from  
9 1:10 to 1:1000) and possible inhibition of the amplification was also assessed.

## 10 RESULTS

11 The post-mortem examination revealed multiple nodular cutaneous lesions with asymmetrical  
12 distribution on the facial and ventral regions of the body. The dome-shaped nodules, 3-12 mm in  
13 diameter, were raised above the skin surface and were fluctuant, often with a central ostium (Figure  
14 1). The other organs were grossly unremarkable. On histological examination, the cutaneous  
15 nodular lesions were found to comprise distended hair follicles (up to 4 mm in diameter) filled with  
16 numerous cigar-shaped arthropod parasites admixed with a scant amount of eosinophilic lamellar  
17 material consistent with flaky keratin (cystic comedonal appearance). These lesions elevated the  
18 epidermis, displacing and compressing adjacent dermis and adnexa. The arthropods were  
19 approximately 40  $\mu$ m in diameter and 200  $\mu$ m in length, with a body divided into three segments (a  
20 gnathosoma with mouthparts, a podosoma with short jointed appendages and a worm-shaped  
21 opisthosoma), and were characterized by an eosinophilic, chitinous, exoskeleton, a haemocoel,  
22 striated muscle, and digestive and reproductive tracts (Figure 2A,B). These histological features  
23 were consistent with those of a demodicid (*Demodex* sp.) mite.

24 The affected follicles were distended and lined by an attenuated infundibular epithelium,  
25 displaying keratohyalin granules. Distended follicles were surrounded by a thin rim of dense  
26 collagen with the rare presence of mononuclear inflammatory cells (Figure 2A). The adjacent  
27 dermis did not show inflammatory changes. Single parasites were rarely observed in hair follicles or  
28 in areas of the skin without associated changes close to the comedonal and cystic follicular lesions.  
29 Moderate follicular melanosis and a minimal pigmentary incontinence was present. Other  
30 histopathological findings included mild multifocal splenic follicular vascular hyalinosis, mild  
31 random multifocal chronic granulomatous hepatitis with minimal diffuse hepatocellular  
32 degeneration. Kidney sections displayed mild multifocal tubular mineralization and multifocal  
33



1 minimal membrano-proliferative glomerular segmental changes. Other tissues (brain, adrenals,  
2 ovary, oviduct, urinary bladder, heart, lung, lymph nodes, abdominal fat) were unremarkable.

3 The amplification of DNA produced extremely weak bands at the expected molecular  
4 weight only for the undiluted sample, whereas the diluted samples did not show any positive  
5 amplification. No inhibition in the PCR reaction was highlighted. No amplification of *Demodex*  
6 DNA was achieved.

## 7 8 **DISCUSSION**

9 The growing awareness of bats as reservoir species for multiple infectious agents has increased  
10 interest in them as vectors of zoonotic pathogens (Mühldorfer et al. 2011; Baker et al. 2013, 2014).  
11 Old World fruit bats (family Pteropodidae) are a diverse group of non-echolocating bats that inhabit  
12 tropical regions in Africa, Asia, Australia, and many Pacific islands. In recent years bat species  
13 have been recognized as natural reservoir hosts for an increasing number of important zoonotic  
14 diseases, including those caused by filoviruses and paramyxoviruses (Baker et al. 2013). *Demodex*  
15 commensal parasitism in wild mammals is common, suggesting that this parasitic relationship is  
16 very ancient (Sastre et al. 2016). Fountain et al. (2017) investigated the epidemiology of skin  
17 lesions among captive bats, but without aetiological diagnosis. A small number of parasitological  
18 investigations have focused on the identification of commensal demodicid mites, including a report  
19 of infection in the New Zealand short-tailed bat (*Mystacina tuberculata*) in the absence of disease  
20 (Desch, 1989) and a description of demodicid mites from the eyelids of a Mexican bat (*Artibeus*  
21 *aztecus*) with blepharitis, but histological examination was not carried out (Vargas et al., 1995).

22 *Demodex* has been reported in fruit bats as a common commensal, but reports of  
23 demodicosis are rare. In the Egyptian fruit bat (*Rousettus aegyptiacus*), demodicosis has been  
24 reported, but has not been characterized clinically or described histologically (Childs-Sanford et al.  
25 2009). In the big brown bat (*Eptesicus fuscus*), preputial *Demodex* sp. infestation has been  
26 described in the absence of gross lesions but in three out of four cases, folliculitis, dermatitis or  
27 preputial adenitis was diagnosed on histological examination (Lankton et al. 2013). A study on zoo-  
28 kept Egyptian fruit bats (*Rousettus aegyptiacus*) found an association between demodicosis and  
29 concurrent disease, supporting the hypothesis that predisposing factors causing immunosuppression  
30 lead to an increase in the *Demodex* sp. population and disease (Childs-Sanford et al. 2009). Among  
31 possible exogenous factors resulting in *Demodex* overgrowth are social and hierarchical stress  
32 (Childs-Sanford et al. 2009).

33 In our subject, although histopathology did not reveal morphological evidence suggestive of  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65



1 impairment of the immune system, the clinical history of the animal revealed pregnancy, abortion  
2 and trauma, as well as capture from the wild and co-housing with new conspecifics, which are  
3 debilitating and predisposing factors that likely triggered changes in the immune status. In addition,  
4 the mild hepatic and nephropathic changes could have acted synergistically in debilitating the  
5 animal, making it more prone to the development of demodicosis. Parasitological characterization  
6 of the *Demodex* species present could not be conducted because no fresh tissue was available for  
7 parasitological examination (Mathison et al. 2014) and the weak positivity observed for the  
8 amplification of the host DNA suggested a high fragmentation of the extracted nucleic acid, usually  
9 due to overfixation (Rait et al. 2006; Dedhia et al. 2007). Considering the weak amplification of the  
10 vertebrate host DNA, the negative result for the amplification of the mite DNA could be thus  
11 attributed to the low quality of the available DNA template. The lack of amplification using PCR  
12 primer sets for both *Demodex* species and Prostigmata members occurred for short amplicons  
13 (around 230 bp) supported this hypothesis.  
14

## 15 **Conclusions**

16 To the best of our knowledge, this report is the first to describe the gross and histological  
17 appearance of demodicosis in an African straw-coloured fruit bat (*E. helvum*). Demodicosis should  
18 be considered as a differential diagnosis in cases of cutaneous nodular and comedone-like lesions in  
19 this species and might be suggestive of non-specific stressors in affected individuals. More  
20 investigations are warranted to identify the species of *Demodex* spp. colonising *E. helvum* and  
21 possibly other species of fruit bats.  
22

## 23 **Declaration of conflicting interests**

24 The authors declared no potential conflicts of interest with respect to the research, authorship and/or  
25 publication of this article.  
26

## 27 **Funding**

28 This work was part-funded by a Wellcome Trust Research Training Fellowship (KSB) and KSB is  
29 supported by a Wellcome Trust Clinical Career Development Fellowship (106690/Z/14/Z).  
30

## 31 **Acknowledgements**

32 The authors thank Richard Irvine for his constructive review of the manuscript.  
33

## 34 **REFERENCES**

35 Baker KS, Leggett RM, Bexfield NH, Alston M, Daly G, Todd S, Tachedjian M, Holmes CE,  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65



- 1 Crameri S, Wang LF, Heeney JL, Suu-Ire R, Kellam P, Cunningham AA, Wood JL,  
2  
3 Caccamo M, Murcia PR (2013) Metagenomic study of the viruses of African straw-coloured  
4  
5 fruit bats: detection of a chiropteran poxvirus and isolation of a novel adenovirus. *Virology*  
6  
7 441: 95–106  
8
- 9 Baker KS, Suu-Ire R, Barr J, Hayman DT, Broder CC, Horton DL, Durrant C, Murcia PR,  
10  
11 Cunningham AA, Wood JL. (2014) Viral antibody dynamics in a chiropteran host. *J Anim*  
12  
13 *Ecol* 83: 415-28  
14
- 15 Childs-Sanford SE, Kollias GV, Abou-Madi N, McDonough PL, Garner MM, Mohammed HO  
16  
17 (2009) *Yersinia pseudotuberculosis* in a closed colony of Egyptian fruit bats (*Rousettus*  
18  
19 *aegyptiacus*). *J Zoo Wildl Med* 40: 8-14  
20
- 21 Dedhia P, Tarale S, Dhongde G, Khadapkar R, Das B (2007) Evaluation of DNA extraction  
22  
23 methods and real time PCR optimization on formalin-fixed paraffin-embedded tissues.  
24  
25 *Asian Pacific J of Cancer Prev* 8: 55-59  
26
- 27 Desch CE (1989) Two new species of *Demodex* (Acari: Demodicidae) from the New Zealand short-  
28  
29 tailed bat, *Mystacina tuberculata* Gray, 1843 (Chiroptera: Mystacinidae). *New Zeal J of*  
30  
31 *Zool* 16: 221-229  
32
- 33 Fountain KI, Stevens KB, Lloyd DH, Loeffler A (2017) Skin disease in captive bats: results of an  
34  
35 online survey of zoos and rehabilitators in Europe, North America and Australasia. *Vet*  
36  
37 *Dermatol* 28: 219-223  
38
- 39 Gross TL, Ihrke PJ, Walder EJ, Affolter VK (2005) Pustular and nodular disease with adnexal  
40  
41 destruction. In: Gross T.L, Ihrke P.J, Walder E.J, Affolter V.K, editors. *Skin Diseases of the*  
42  
43 *Dog and Cat: Clinical and Histopathologic Diagnosis. Second Edition. John Wiley & Sons,*  
44  
45 *pp. 420–459*  
46
- 47 Kitano T, Umetsu K, Tian W, Osawa M (2007) Two universal primer sets for species identification  
48  
49 among vertebrates. *Int. J Legal Med* 121: 423–427  
50
- 51 Lankton JS, Chapman A, Ramsay EC, Kania SA, Newkirk KM (2013) Preputial *Demodex* species  
52  
53 in big brown bats (*Eptesicus fuscus*) in eastern Tennessee. *J Zoo Wildl Med* 44: 124-129  
54
- 55 Mathison BA, Pritt BS (2014) Laboratory Identification of Arthropod Ectoparasites. *Clin Microbiol*  
56  
57 *Rev* 27: 48–67  
58
- 59 Mauldin EA, Peters-Kennedy J (2015) Integumentary System. In: Jubb, Kennedy, and Palmer's  
60  
61 *Pathology of Domestic Animals, 6th Edition, pp 509-736.*  
62
- 63 Mühldorfer K, Speck S, Kurth A, Lesnik R, Freuling C, Müller T, Kramer-Schadt S, Wibbelt G  
64  
65 (2011) Diseases and causes of death in European bats: dynamics in disease susceptibility



1 and infection rates. PLoS One 6: e29773.

2 Otto J C, Wilson K (2001) Assessment of the usefulness of ribosomal 18S and mitochondrial COI  
3 sequences in Prostigmata phylogeny. In Acarology: Proceedings of the 10th International  
4 Congress- CSIRO PUBLISHING, pp. 100-109

5 Rait VK, Zhang Q, Fabris D, Mason JT, O'Leary TJ (2006) Conversions of formaldehyde-modified  
6 2'-deoxyadenosine 5'-monophosphate in conditions modeling formalin-fixed tissue  
7 dehydration. J Histochem Cytochem 54: 301-310

8 Sastre N, Francino O, Curti JN, Armenta TC, Fraser DL, Kelly RM, Hunt E, Silbermayr K, Zewe  
9 C, Sánchez A, Ferrer L (2016) Detection, Prevalence and Phylogenetic Relationships of  
10 *Demodex* spp and further Skin Prostigmata Mites (Acari, Arachnida) in Wild and Domestic  
11 Mammals. PLoS One 1: e0165765

12 Vargas M, Bassols IB, Desch CE, Quintero MT, Polaco OJ (1995) Description of two new species  
13 of the genus *Demodex* Owens, 1843 (Acari: Demodecidae) associated with mexican bats.  
14 Internat J Acarol 21: 75-82

15 Zewe CM, Altet L, Lam ATH, Ferrer L (2017) Afoxolaner and fluralaner treatment do not impact  
16 on cutaneous *Demodex* populations of healthy dogs. Vet Dermatol 28: 468

17 Zhao Y, Ma J, Hu L, Wu L, De Rojas M (2013) Discrimination between *Demodex folliculorum*  
18 (Acari: Demodicidae) isolates from China and Spain based on mitochondrial cox1  
19 sequences. J Zhejiang Univ Sci B 14: 829-836

20 Zhao YE, Wu LP (2012) Phylogenetic relationships in *Demodex* mites (Acari: Demodicidae) based  
21 on mitochondrial 16S rDNA partial sequences. Parasitol Res 111: 1113-1121

## 22 **Figure captions**

23 **Figure 1** Adult female African straw-coloured fruit bat (*Eidolon helvum*): post mortem gross  
24 appearance of demodicosis: multifocal dome-shaped nodules 3-12 mm in diameter, raised above  
25 skin surface and fluctuant with frequently evident central follicular ostium.

26 **Figure 2** Adult female African straw-coloured fruit bat (*Eidolon helvum*), haired skin. (A) Low  
27 magnification appearance of extremely dilated, cystic hair follicle comedones with numerous  
28 intralesional demodicid mites, evocating a minimal perifollicular inflammatory response.  
29 (haematoxylin and eosin stained slide; objective: 10×). (B) Detail of adult *Demodex* spp., elongated  
30 cigar-shaped parasites dwelling in the dilated hair follicle. (haematoxylin and eosin stained slide,  
31 objective: 40×).



FIG 1





