## 404 words

# Authors' response to Letters from Gunst et al (20-3415) and Lew et al (20-3583), CHEST

Angela S. McNelly, PhD1,2,3, Danielle E. Bear, MRes4,5,6, Bronwen A. Connolly, PhD<sup>6,7</sup>, Gill Arbane, BSc<sup>7</sup>, Laura Allum, BSc<sup>7</sup>, Azhar Tarbhai BSc<sup>2</sup>, Jackie A. Cooper MSc<sup>2</sup>, Philip A. Hopkins, PhD<sup>8</sup>, Matthew P. Wise, MBBS<sup>9</sup>, David Brealey, PhD<sup>3</sup>, Kieron Rooney, MBBS<sup>10</sup>, Jason Cupitt, MBBS<sup>11</sup>, Bryan Carr, MBBS<sup>12</sup>, Kiran Koelfat, MD<sup>13</sup>, Steven Olde Damink, PhD<sup>13,14</sup>, Philip J. Atherton, PhD<sup>15</sup>, \*Nicholas Hart, PhD<sup>7</sup>, \*Hugh E. Montgomery, MD<sup>2,3</sup> and \*Zudin A. Puthucheary, PhD<sup>1,16</sup> <sup>1</sup>William Harvey Research Institute, Barts and The London School of Medicine & Dentistry, Queen Mary University of London; <sup>2</sup>University College London (UCL); <sup>3</sup>UCL Hospitals NHS Foundation Trust (UCLH), National Institute for Health Research (NIHR) Biomedical Research Centre (BRC), London; <sup>4</sup>Department of Nutrition and Dietetics St Thomas' NHS Foundation Trust; <sup>5</sup>Department of Critical Care, Guy's and St. Thomas' NHS Foundation Trust; <sup>6</sup>King's College London (KCL) NIHR BRC, London; <sup>7</sup>Lane Fox Clinical Respiratory Physiology Research Centre Guy's and St. Thomas' NHS Foundation Trust; <sup>8</sup>Kings College Hospital, London; <sup>9</sup>University Hospital of Wales, Cardiff; <sup>10</sup>Bristol Royal Infirmary; <sup>11</sup>Blackpool Victoria Hospital; <sup>12</sup>University Hospitals of North Midlands, Stoke-on-Trent; <sup>13</sup>Department of Surgery and School of Nutrition and Translational Research in Metabolism (NUTRIM), University of Maastricht, Maastricht, The Netherlands; 14Department of General, Visceral and Transplantation Surgery, RWTH University Hospital Aachen, Germany; <sup>15</sup>Medical Research Council/Arthritis Research UK Centre for Musculoskeletal Aging, University of Nottingham; 16Adult Critical Care Unit, Royal London Hospital, London, UK.

\*Joint senior authors

### **CORRESPONDENCE TO:**

Dr Angela McNelly, Critical Care and Perioperative Medicine Research Group,

Adult Critical Care Unit, Royal London Hospital, London, E1 1BB, United Kingdom

Email: <a href="mailto:angela.mcnelly@qmul.ac.uk">angela.mcnelly@qmul.ac.uk</a>

#### **FUNDING:**

JP Moulton Charitable Foundation, (JM29/04/14; JM02/06/15); NIHR UCL/UCLH BRC Cardiometabolic research grant (BRC202 rev/CM/AM/101320; RCF236/AMcN/2015); Intensive Care Foundation (New Investigator Award A.S.McN); London South Local Clinical Research Network (LCRN) (D. E. B.: November 2014 – May 2015); North Thames LCRN (A. S. McN,: January 2017-March 2017); ASPEN Rhoads Research Foundation (Z. A. P. January 2018-January 2020).

## **SUMMARY CONFLICT OF INTEREST STATEMENTS:**

DEB has received speaker fees, conference attendance support or advisory board fees from Nutricia, Baxter, BBraun, Nestle Nutrition, Fresenius Kabi, Abbott Nutrition, Cardinal Health and Avanos. ZP has received honoraria for consultancy from GlaxoSmithKline, Lyric Pharmaceuticals, Faraday Pharmaceuticals and Fresenius-Kabi and speaker fees from Orion and Nestle. HM holds patents relating to intravenous hydration and to regulation of metabolic efficiency using reninangiotensin system antagonists and consults for Google Health. MW has accepted accommodation and attendance at educational meeting organised by Orion. Other authors have no conflicts of interest to declare.

Dear Editor.

We thank the authors of the above letters for their interest in and thoughtful comments regarding our trial of intermittent versus continuous feeding in the critically ill.

Muscle ultrasound does indeed underestimate muscle loss when compared to protein:DNA ratio<sup>1</sup>. Whilst local tissue oedema might contribute to such discrepancy, fluid status per se does not appear to be a confounder<sup>2,3</sup>.

The respondents suggest that increased protein delivery in the intermittent feeding group may have worsened muscle loss, but the papers to which they refer relate to continuous feeding. Indeed, these observations (confirmed in our original paper) were what prompted us to test whether intermittent feeding may offer benefit over continuous amino acid provision. We agree that intermittent feeding may still be beneficial, but as stated, this is likely to be in the context of a multimodal intervention.

Both primary outcomes and adjusted analyses are appropriately powered, and details are provided in the manuscript and online supplement. No differences were seen between groups in withdrawal rates, and the risk of selection bias is low, given that our sensitivity analyses included use of imputation.

Daily median glucose and episodes of hyperglycaemia in the Per Protocol cohort are shown in Table 1. No differences were seen in baseline demographics between groups in this cohort (Table 2).

Both baseline and acute illness factors are likely to affect the nutrition/starvation response to autophagic flux<sup>4,5</sup>. We agree that the intervention may have been too short, in common with other critical care nutrition trials<sup>6,7</sup>. Future studies might extend beyond the ICU itself. As stated in the manuscript, we calculated feed delivery based on feeding days, which is appropriate for examination of the process of feed delivery<sup>8</sup>.

Lew et al offer insightful comments on the leucine peak data. A direct comparison of plasma leucine concentrations (or percentage change) is not possible, given the different physiological and metabolic states between groups<sup>9</sup>. We currently do not know the bounds of leucinaemia in young or old critically ill patients. The higher doses of protein needed to sustain muscle protein synthesis in the study quoted only appear to occur when exercise is overlaid<sup>10</sup>.

The fact that the two letters express diametrically opposed views on the role of nutritional protein in preserving muscle mass or provoking muscle wasting, demonstrates the equipoise on the role of current methods of nutritional support for the critically ill patient, and the need for trials such as the one we performed.

#### REFERENCES

- 1. Puthucheary ZA, Rawal J, McPhail M, et al. Acute skeletal muscle wasting in critical illness. *JAMA*. 2013;310(15):1591-1600.
- 2. Puthucheary ZA, Phadke R, Rawal J, et al. Qualitative Ultrasound in Acute Critical Illness Muscle Wasting. *Critical care medicine*. 2015;43(8):1603-1611.
- 3. Puthucheary ZA, McNelly AS, Rawal J, et al. Rectus Femoris Cross-Sectional Area and Muscle Layer Thickness: Comparative Markers of Muscle Wasting and Weakness. *American journal of respiratory and critical care medicine.* 2017;195(1):136-138.
- 4. Dethlefsen MM, Bertholdt L, Gudiksen A, et al. Training state and skeletal muscle autophagy in response to 36 h of fasting. *J Appl Physiol (1985)*. 2018;125(5):1609-1619.
- 5. Tardif N, Polia F, Tjader I, Gustafsson T, Rooyackers O. Autophagy flux in critical illness, a translational approach. *Scientific reports*. 2019;9(1):10762.
- 6. Bear DE, Puthucheary ZA. Designing nutrition-based interventional trials for the future: addressing the known knowns. *Critical care (London, England)*. 2019;23(1):53.
- 7. Casaer MP, Mesotten D, Hermans G, et al. Early versus Late Parenteral Nutrition in Critically Ill Adults. *New England Journal of Medicine*. 2011;365(6):506-517.
- 8. McNelly AS, Bear DE, Connolly BA, et al. Effect of Intermittent or Continuous Feed on Muscle Wasting in Critical Illness: A Phase 2 Clinical Trial. *Chest.* 2020.

- 9. Puthucheary ZA, Astin R, McPhail MJW, et al. Metabolic phenotype of skeletal muscle in early critical illness. *Thorax.* 2018;73(10):926-935.
- 10. Wilkinson DJ, Bukhari SSI, Phillips BE, et al. Effects of leucine-enriched essential amino acid and whey protein bolus dosing upon skeletal muscle protein synthesis at rest and after exercise in older women. *Clin Nutr.* 2018;37(6 Pt A):2011-2021.

Day	Intermittent arm	Hyperglycaemic episodes/day	Continuous arm	Hyperglycaemic episodes/day
0	8.5 (5.6-56.5)	0 (0-5)	8.4 (4.8-28)	0 (0-4)
1	9.5 (7.0-24.7)	2 (0-6)	8.4( 4.8-25.4)	0 (0-6)
2	8.7(6.5-18.0)	1 (0-6)	8.6 (5.5-9.2)	0 (0-6)
3	9.1 (5.5-18.3)	1 (0-6)	8.3 (5.4-11.3)	0 (0-4)
4	8.8 (4.4-16.3)	1 (0-6)	8.7 (5.6-13.5)	0 (0-6)
5	9.1 (4.6-13.7)	1 (0-6)	8.5 (6.5-14.3)	0 (0-6)
6	9.2 (5.9-12.2)	2 (0-5)	8.7 (6.1-14.4)	0 (0-6)
7	8.4 (6.8-14.5)	1 (0-4)	8.1 (5.4-13.1)	0 (0-5)
8	9.5 (5.5-16.2)	2 (0-6)	8.4 (4.4-11.9)	0 (0-4)
9	8.6 (5.4-14.7)	1 (0-5)	7.9 5.2-13.9)	0 (0-5)
10	8.7 (4.9-12.8)	0 (0-4)	7.4 (6.0-14.6)	0 (0-5)

**Table 1:** Median (95%CI) glucose concentrations per day and median (range) hyperglycemic episodes per day during the 10-day trial period in per protocol cohort (n=63)

	Intermittent	feeding	Continuous feeding	р
	(n=31)		(n=32)	
Age, y	55.1 (49.5-60.7)		61.3(55.6-66.9)	0.122
Male, No. (%) ¥	19 (61.3)		23 (71.8)	0.373
APACHE II score	23.2 (18.3-28.1)		19.2(16.3-22.2)	0.153
ICU LOS, d*	18 (9-84)		17.5 (6-52)	0.694
Hospital LOS, d*	32 (11-103)		34 (13-102)	0.416
SOFA score on admission	10.2 (9.1-11.3)		10.8 (9.7-11.9)	0.458
Comorbidities, No. (%)				
Hypertension	10 (32.3)		12 (37.5)	
<b>Chronic Respiratory Diseases</b>	11 (35.5)		8 (25.0)	
Diabetes Mellitus	10 (32.3)		7 (21.9)	
Ischemic heart disease	3 (9.7)		3 (9.4)	
Psychiatric diseases	9 (29.0)		3 (9.4)	

Renal impairment	2 (6.5)	3 (9.4)
Obesity	2 (6.5)	1 (3.1)
Liver cirrhosis	3 (9.7)	2 (6.3)
Haem-oncological disease	0 (0.0)	3 (9.4)
Thyroid disease	1 (3.2)	1 (3.1)
Crohns disease	0 (0.0)	1 (3.1)
Previous CVA	0 (0.0)	1 (3.1)

**Table 2:** Patient characteristics and demographics in per protocol cohort (n=63). ICU=Intensive Care Unit, APACHE II=Acute Physiology and Chronic Health Evaluation score, CVA=Cerebrovascular Accident, SOFA=Sequential Organ Failure Assessment, LOS=Length of Stay. Data are mean (95% confidence intervals) except for \* (median and range), Student's T-test was used except for ¥ (Chi-squared) and \* (Mann-Whitney U).