**Running title:** Primate fermented foods

1 2 3

4

## Fermented food consumption in wild non-human primates and its ecological drivers

5 Katherine R. Amato<sup>1\*</sup>, Óscar M. Chaves<sup>2</sup>, Elizabeth K. Mallott<sup>1</sup>, Timothy M. Eppley<sup>3,4</sup>,

- 6 Filipa Abreu<sup>5</sup>, Andrea L. Baden, <sup>6,7</sup> Adrian A. Barnett<sup>8</sup>, Julio Cesar Bicca-Marques<sup>9</sup>, Sarah
- A. Boyle<sup>10</sup>, Christina J. Campbell<sup>11</sup>, Colin A. Chapman<sup>12,13,14</sup>, María Fernanda De la
- 8 Fuente<sup>5</sup>, Pengfei Fan<sup>15</sup>, Peter J. Fashing<sup>16,17</sup>, Annika Felton<sup>18</sup>, Barbara Fruth<sup>19,20,21</sup>, Vanessa
- 9 B. Fortes<sup>22</sup>, Cyril C. Grueter<sup>23,24</sup>, Gottfried Hohmann<sup>25</sup>, Mitchell Irwin<sup>26</sup>, Jaya K.
- 10 Matthews<sup>24,27</sup>, Addisu Mekonnen<sup>17</sup>, Amanda D. Melin<sup>28</sup>, David B. Morgan<sup>29</sup>, Julia
- Ostner<sup>30,31</sup>, Nga Nguyen<sup>16,17</sup>, Alex Piel<sup>32</sup>, Braulio Pinacho-Guendulain<sup>33, 34</sup>, Erika Patricia
- Quintino Aredes<sup>9</sup>, Patrick Tojotanjona Razanaparany<sup>35,36</sup>, Nicola Schiel<sup>5</sup>, Crickette M.
- 13 Sanz<sup>37,38</sup>, Oliver Schülke<sup>30,31</sup>, Sam Shanee<sup>39</sup>, Antonio Souto<sup>40</sup>, João Pedro Souza-Alves<sup>40</sup>,
- Fiona Stewart<sup>20</sup>, Kathrine M. Stewart<sup>21</sup>, Anita Stone<sup>41</sup>, Binghua Sun<sup>42</sup>, Stacey Tecot<sup>43</sup>, Kim
- Valenta<sup>44</sup>, Erin R. Vogel<sup>45</sup>, Serge Wich<sup>20</sup>, Yan Zeng<sup>46</sup>

- <sup>1</sup>Department of Anthropology, Northwestern University, Evanston, USA
- <sup>2</sup>Escuela de Biología, Universidad de Costa Rica, UCR, San Pedro de Montes de Oca 2060,
- 19 San José, Costa Rica
- <sup>3</sup>Institute for Conservation Research, San Diego Zoo Global, San Diego, CA, USA
- <sup>4</sup>Department of Anthropology, Portland State University, Portland, OR, USA
- <sup>5</sup>Department of Biology, Federal Rural University of Pernambuco, Recife, Brazil
- <sup>6</sup>Department of Anthropology, Hunter College of the City University of New York, New
- 24 York, USA
- <sup>7</sup>The New York Consortium in Evolutionary Primatology (NYCEP)
- <sup>8</sup>Amazon Mammals Research Group, National Amazon Research Institute (INPA),
- 27 Manaus, AM, Brazil & Dept. Zoology, Federal University of Pernambuco, Recife, PE,
- 28 Brazil
- 29 <sup>9</sup>Laboratório de Primatologia, Escola de Ciências da Saúde e da Vida,
- 30 Pontifícia Universidade Católica do Rio Grande do Sul, PUCRS, Porto Alegre, RS, Brazil
- 31 <sup>10</sup>Department of Biology, Rhodes College, Memphis, USA
- 32 <sup>11</sup>Department of Anthropology, California State University Northridge, Northridge, USA
- 33 <sup>12</sup>Department of Anthropology, Center for the Advanced Study of Human Paleobiology,
- 34 George Washington University, Washington DC, USA
- 35 <sup>13</sup>School of Life Sciences, University of KwaZulu-Natal, Scottsville, Pietermaritzburg,
- 36 South Africa.
- 37 <sup>14</sup>Shaanxi Key Laboratory for Animal Conservation, Northwest University, Xi'an, China.
- 38 <sup>15</sup> School of Life Sciences, Sun Yat-Sen University, Guangzhou, China
- 39 <sup>16</sup>Department of Anthropology and Environmental Studies Program, California State
- 40 University Fullerton, Fullerton CA USA
- 41 <sup>17</sup>Centre for Ecological and Evolutionary Synthesis (CEES), University of Oslo, Oslo,
- 42 Norway
- 43 <sup>18</sup>Southern Swedish Forest Research Centre, Swedish University of Agricultural Sciences
- 44 (SLU), Alnarp, Sweden
- 45 <sup>19</sup> Department of Human Behavior, Ecology and Culture, Max-Planck-Institute for
- 46 Evolutionary Anthropology, Leipzig, Germany

- 47 <sup>20</sup>School of Biological and Environmental Sciences, Liverpool John Moores University,
- 48 Liverpool, UK
- 49 <sup>21</sup> Centre for Research and Conservation, Royal Zoological Society of Antwerp, B-2018
- 50 Antwerp, Belgium
- 51 <sup>22</sup> Laboratório de Primatologia, Departamento de Zootecnia e Ciências Biológicas,
- 52 Universidade Federal de Santa Maria, Palmeira das Missões, RS, Brasil
- 53 <sup>23</sup>School of Human Sciences, The University of Western Australia, Perth, Australia
- 54 <sup>24</sup>Centre for Evolutionary Biology, School of Biological Sciences, The University of
- 55 Western Australia, Perth, Australia
- 56 <sup>25</sup> Department of Human Evolution, Max Planck Institute for Evolutionary Anthropology,
- 57 Leipzig, Germany
- 58 <sup>26</sup> Dept. of Anthropology, Northern Illinois University, DeKalb, IL, USA
- 59 <sup>27</sup> Africa Research & Engagement Centre, The University of Western Australia, Crawley,
- 60 Western Australia, Australia
- 61 <sup>28</sup>Department of Anthropology and Archaeology, University of Calgary, Calgary, Canada
- 62 <sup>29</sup>Lester E. Fisher Center for the Study and Conservation of Apes, Lincoln Park Zoo,
- 63 Chicago, IL, USA
- 64 <sup>30</sup>Department of Behavioral Ecology, University of Goettingen, Goettingen, Germany
- 65 <sup>31</sup>Research Group Primate Social Evolution, German Primate Center, Leibniz Institute for
- 66 Primate Research, Goettingen, Germany
- 67 <sup>32</sup>Department of Anthropology, University College London, London, UK
- 68 <sup>33</sup>Departamento de Ciencias de la Salud, Universidad Autónoma Metropolitana (UAM),
- 69 Lerma, México.
- 70 <sup>34</sup>Centro Interdisciplinario de Investigación para el Desarrollo Integral Regional (CIIDIR),
- 71 Unidad Oaxaca, Instituto Politécnico Nacional, México.
- 72 <sup>35</sup>Graduate School of Asian and African Area Studies, Kyoto University, Japan
- 73 <sup>36</sup>Department of Zoology and Animal Biodiversity, University of Antananarivo,
- 74 Madagascar
- 75 <sup>37</sup>Department of Anthropology, Washington University in St. Louis, St. Louis, MO, USA
- 76 <sup>38</sup>Congo Program, Wildlife Conservation Society, Brazzaville, Republic of Congo
- 77 <sup>39</sup>Neotropical Primate Conservation, Cornwall, United Kingdom
- 78 <sup>40</sup>Departamento de Zoologia, Universidade Federal de Pernambuco, Recife, PE, Brazil
- 79 <sup>41</sup> Biology Department, California Lutheran University, Thousand Oaks CA USA
- 80 <sup>42</sup> School of Resource and Environmental Engineering, Anhui University, Hefei, China
- 81 <sup>43</sup>School of Anthropology, University of Arizona, Tucson, AZ, USA
- 82 <sup>44</sup> Department of Anthropology, University of Florida, Gainesville, FL, USA
- 83 <sup>45</sup> Department of Anthropology, Rutgers University, New Brunswick, NJ, USA
- 84 <sup>46</sup> Animal Microecology Institute, College of Veterinary, Sichuan Agricultural University,
- 85 Ya'an, China

\*corresponding author: katherine.amato@northwestern.edu

89 90

86 87

88

91 92

#### **ABSTRACT**

**Objectives:** Although fermented food use is ubiquitous in humans, the ecological and evolutionary factors contributing to its emergence are unclear. Here we investigated the ecological contexts surrounding consumption of fruits in the late stages of fermentation by wild primates to provide insight into its adaptive function. We hypothesized that climate, socio-ecological traits, and habitat patch size would influence the occurrence of this behavior due to effects on the environmental prevalence of late-stage fermented foods, the ability of primates to detect them, and potential nutritional benefits.

**Materials and Methods:** We compiled data from field studies lasting at least nine months to describe the contexts in which primates were observed consuming fruits in the late stages of fermentation. Using generalized linear mixed-effects models, we assessed the effects of 18 predictor variables on the occurrence of fermented food use in primates.

**Results:** Late-stage fermented foods were consumed by a wide taxonomic breadth of primates. However, they generally made up less than 3% of the annual diet and were limited to a subset of fruit species, many of which are reported to have mechanical and chemical defenses against herbivores when not fermented. Additionally, late-stage fermented food consumption was best predicted by climate and habitat patch size. It was more likely to occur in larger habitat patches with lower annual mean rainfall, higher annual mean maximum temperatures, and lower annual mean minimum temperatures.

**Discussion:** We posit that primates capitalize on the natural fermentation of some fruits as part of a nutritional strategy to maximize periods of fruit exploitation and/or access a wider range of plant species. We speculate that these factors contributed to the evolutionary emergence of the human propensity for fermented foods.

**Keywords:** fermentation, feeding ecology, climate, herbivore defense, human evolution

## **Research Highlights**

- Fruits in the late stages of fermentation are consumed by a wide range of primates but only a subset of available fruit species is consumed in this form.

- Primate late-stage fermented fruit consumption is more common in large habitat patches with lower annual mean rainfall and higher annual mean maximum temperatures.

- Consumption of late-stage fermented foods appears to be part of a primate nutritional strategy to increase dietary breadth both qualitatively and temporally.

#### 1 INTRODUCTION

140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

Food fermentation – the anaerobic microbial degradation of carbon compounds into ethanol and/or lactic acid – is a central part of human diet and culture (Tamang & Kailasapathy, 2010). Humans from many cultures regularly incite or direct microbial fermentation of a wide range of foods that include meat and dairy products, grains, fruits, and vegetables (Battcock & Azam-Ali, 1998; Campbell-Platt, 1994; Deshpande, 2000; Tamang, Holzapfel, Shin & Felis, 2017). Such foods make up 20 to 40% of the global food supply (Campbell-Platt, 1994). Although not all fermented foods contain ethanol, the majority of anthropological fermented food research to date targets ethanol as an indicator of fermentation (e.g. Dominy, 2015; Dudley, 2002; Garnier & Valamoti, 2016; Hayden, Canuel, & Shanse, 2013; Kuijt, 2009; Liu et al., 2018; Milton, 2004; Ross, Morgan, & Hill, 2002; Smalley et al., 2003). Directed fermentation by humans has early origins. There is archaeological evidence that humans have engaged in directed fermentation of fruits and grains and stored the resulting ethanol in large quantities since ~4300 BC, although some suggest a date as early as 12,500 cal BP (Garnier & Valamoti, 2016; Hayden et al., 2013). Evolutionary changes in human genes for processing ethanol and for interacting with a major lineage of fermenting bacteria (Lactobacillales) are compatible with an even earlier association with fermented foods, dating back to the divergence of hominids from other primates at ~10 Mya (Carrigan et al., 2015; Janiak, Pinto, Duytschaever, Carrigan, & Melin, 2020; Peters et al., 2019). Limited technology for processing and storing food at this time makes it likely that our hominid ancestors relied more heavily on naturally occurring fermented foods. However, some simple forms of directed fermentation, such as burying food items or submerging them in water (Speth 2017), may have been possible.

Why humans have incorporated fermented products so prominently into their diet across their evolutionary history is unclear. Fermentation is an effective food preservative since it produces locally high concentrations of ethanol and lactic acid that ultimately prevent microbial growth and associated food spoilage (Boulton, Singleton, Bisson, & Kunkee, 1999; Pretorius, 2000; Skinner, Passmore, & Davenport, 1980; Thomson et al., 2005). Additionally, the physiological effects of consuming ethanol (i.e. intoxication) are believed to have facilitated social gatherings and rituals (Liu et al., 2018). Accordingly, the modern and ancient contexts in which fermented food use has been documented often suggest central roles of food preservation and socially motivated ethanol acquisition in driving the ubiquity of human fermented food use (Dominy, 2015; Dudley, 2002; Kuijt, 2009; Liu et al., 2018; Milton, 2004; Ross, et al., 2002; Smalley et al., 2003). However, given genetic evidence that human adaptations for fermented food consumption emerged before the technology associated with its directed production and storage (Carrigan et al., 2015; Janiak et al., 2020; Peters et al., 2019), fermented food consumption may have provided another selective advantage earlier in our evolutionary history. Given their high sugar content, fruits often ferment naturally (Dominy, 2004; Duar et al., 2017; Dudley, 2002; Gorgus, Hittinger, & Schrenk, 2016; Martinson, Herre, Machado, & Arnold, 2012; Nyanga et al., 2007; Ruiz Rodriguez et al., 2019; Weaver, 2016) making it likely that all frugivorous animals consume some minimum amount of fermented foods. However, overripe fruits in late stages of fermentation commonly remain in food patches after other fruits have been depleted. As described above, fermentation is distinct from rot or decay in that it involves distinct microbes and precludes the production

of most toxic microbial byproducts (except ethanol). Therefore, it has been suggested that

fruits in the late stages of fermentation could have been a fallback food for increasingly

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

terrestrial hominids during periods of low food availability in patchy woodland environments (Carrigan et al., 2015).

188

189

191

195

198

200

201

203

204

205

207

208

209

210

190 Foods in the late stages of fermentation could also convey nutritional benefits that provide a selective advantage to consumers year-round. Compared to unfermented foods, 192 fermented foods have higher caloric, free amino acid, and vitamin content (NRC, 1998; 193 Gobbetti et al., 1994; LeBlanc et al., 2013; Mitchell & Herlong, 1986; Tamang, Shin, Jung 194 & Chae, 2016). In the wild, many fermented foods contain embedded insects, which provide an additional protein source (Barnett et al., 2017; Barnett et al., in press; Braham, 196 2015; Hodge & Arthur, 1996; Xiaoming, Ying, Hong, & Zhiyong, 2010). Also, 197 fermentation improves digestibility of food by breaking down resistant starch, soluble fiber, toxins, and secondary plant metabolites (Binita & Khetarpaul, 1997; Chaves-López et al., 199 2014; Gupta, Gangoliya, & Singh, 2015; Rollan, Gerez & Leblanc, 2019). For example, some toxic foods, such as blowfish and cassava, can only be consumed after fermentation (Akinrele, 1964; Anraku et al., 2013). Together these properties not only directly affect 202 consumer nutrient intake and balance but may also result in a more favorable balance among the nutrients of a food which in turn can play a critical role in food selection (Felton et al., 2009). Therefore, foods in the late stages of fermentation could have represented a critical nutritional resource to hominids, particularly as energetically expensive life history 206 traits such as long juvenile periods, short interbirth intervals, and large brains emerged across evolutionary time (Aiello & Key, 2002; Antón, Potts, & Aiello, 2014; Leonard & Robertson, 1992, 1997). The consumption of other high quality diet items such as meat and cooked foods has also been hypothesized to have provided essential nutritional resources for the development and maintenance of these traits in hominids (Aiello & Wells, 2002;

DeCasien, Williams, & Higham, 2017; Wrangham, 2009; Wrangham & Conklin-Brittain, 2003).

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

Fermented foods contain live microbes, substrates for microbial metabolism, and microbial metabolites, which may affect consumer health and fitness either directly or indirectly through impacts on the microbiome (Jacobsen et al., 1999; Kim et al., 2016; Maldonado-Gómez et al., 2016; Marco et al., 2017). Given the broad effects of the microbiome on host metabolism (Oliphant & Allen-Vercoe, 2019; Visconti et al., 2019), immune function (Al Nabhani & Eberl, 2020), and neuroendocrine dynamics (Cryan et al., 2019; Sylvia & Demas, 2018), fermented foods have the potential to affect consumer physiology in many ways. Beyond intoxication caused by excessive consumption of fermented foods with high ethanol content, none of these documented physiological effects are negative. Therefore, fermented food consumption could have provided a selective advantage to hominids in addition to the nutritional advantages discussed above. Indeed, studies of human fermented food use consistently demonstrate a range of improved health outcomes (e.g. Bourrie, Willing, & Cotter, 2016; Burton et al., 2017; Yartey, Nkrumah, Hori, Harrison, & Armar, 1995). However, the wide variety of positive health effects that fermented foods can produce via the microbiome make it difficult to predict specific scenarios in which these properties would be most evolutionarily advantageous based on current knowledge.

Even in the context of nutrition, modern human technology and cultural practices complicate our ability to evaluate the potential fitness benefits of human fermented food consumption. As a result, comparative data from non-human primates (hereafter primates) are essential for exploring the adaptive function of this behavior. By determining how pervasive consumption of late-stage fermented foods by wild primates is and the ecological

contexts in which it occurs, we can begin to more accurately assess the ecological and evolutionary forces that drive it and contextualize it within human evolutionary history.

Nevertheless, few studies on this subject have incorporated primate data.

235

236

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

A handful of comparative genetic analyses of physiological adaptations for fermented food consumption integrate data from multiple primate species (Carrigan et al., 2015; Janiak et al., 2020; Peters et al., 2019). Additionally, some behavioral research has investigated primate ethanol affinity in response to the Drunken Monkey Hypothesis (Dudley, 2002, 2004). This hypothesis posits that humans direct the production of fermented foods and consume them as a result of our affinity for ethanol, which stems from our evolutionary past as frugivorous primates that used ethanol as an olfactory and/or gustatory signal for energy-rich fruit (Dudley, 2002, 2004). Therefore, data from other primates have been used to test the relationship between frugivory and ethanol affinity. The results indicate that primates across the Order prefer solutions of 2-5% ethanol over water (Dausch Ibañez, Hernandez Salazar, & Laska, 2019; Gochman, Brown, & Dominy, 2016; Hockings et al., 2015; Kornet, Goosen, Ribbens, & Van Ree, 1990; Mandillo, Titchen, & Miczek, 1998). However, data from spider monkeys (Ateles geoffroyi) indicate that sweet solutions are preferred over ethanol regardless of calorie content (Dausch Ibañez et al., 2019). Outside of this context, fermented food consumption is rarely mentioned in studies of primate feeding ecology, despite the fact that not all fermented foods contain ethanol but all of them likely confer a range of nutritional and health benefits to consumers.

As a first step to address this knowledge gap, we compiled qualitative data describing overripe fruit consumption from primate field studies around the world to estimate the minimum prevalence of late-stage fermented foods in wild primate diets, regardless of ethanol content, and the ecological contexts in which the consumption of

these foods occurs. We hypothesized that local climate, primate socio-ecological traits, and habitat patch size (Table S1) would predict the prevalence of primate consumption of latestage fermented foods. First, climate affects both the rate of fermentation and the rate of ethanol evaporation (Isu & Njoku, 1998), thereby influencing the local prevalence of latestage fermented foods and the probability that primates will detect them via olfaction (Dominy, 2004; Nevo & Valenta, 2018; Melin et al., 2019). Therefore, we predicted that mean minimum annual temperature, mean maximum annual temperature, mean daily temperature, mean annual rainfall, elevation, and latitude and longitude would be associated with the occurrence of late-stage fermented food consumption in wild primates. Given that fruit ferments easily in nature (Dominy, 2004; Duar et al., 2017; Dudley, 2002; Gorgus, Hittinger, & Schrenk, 2016; Martinson, Herre, Machado, & Arnold, 2012; Nyanga et al., 2007; Ruiz Rodriguez et al., 2019; Weaver, 2016), we predicted that primate species and populations with high percentages of fruit in their diets and low percentages of leaves and invertebrates would be more likely to encounter and consume late-stage fermented food. Since home range, social group size, body size, and encephalization quotients are often correlated with diet (Clutton-Brock & Harvey, 1980; Dunbar & Shultz, 2007, 2017; Kudo & Dunbar, 2001), we also expected these variables to be associated with late-stage fermented food consumption. Finally, due to the relationship between habitat patch size and food availability more generally (Abbas et al., 2011; Fahrig, 2003; Laurance, Delamonica, Laurance, Vasconcelos, & Lovejoy, 2000), we predicted that habitat patch size would predict the prevalence of late-stage fermented foods and their consumption.

280

281

282

259

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

## 2 | MATERIALS AND METHODS

## 2.1 | Behavioral data collection

KRA, YZ, and TME identified a group of researchers who had completed a wild primate field study of at least nine consecutive months using multiple approaches. We searched two general online databases (<a href="https://scholar.google.com">https://scholar.google.com</a>,

<a href="http://xueshu.baidu.com">http://xueshu.baidu.com</a>) using specific key words such as "primate" and "diet" combined with primate family names one year at a time beginning with 2005. We also reviewed the literature cited in multiple primate ecology books (Brady & Carville, 2012; Campbell, Fuentes, MacKinnon, Bearder, & Stumpf, 2011; Davies & Oates, 1994; Dudley, 2014; NRC, 2003; Rowe & Myers, 2016; Strier, 2016). Finally, we flagged abstracts from the programs of primate conferences in 2018 and 2019, including the American Society of Primatologists and the American Association of Physical Anthropologists.

KRA asked 151 researchers with relevant field studies and current email contact information to report whether they had observed their study subjects consuming fermented foods (i.e., plant foods clearly overripe or fermenting based on their color, physical traits, smell, or other useful indicator traits). These food items could be found on the ground, but this was not necessary for a food to be deemed 'fermented.' Most fruits consumed by primates are likely to have undergone some degree of fermentation (Dominy, 2004), but only late stages of fermentation with higher concentrations of ethanol and other microbial products (Biale 1954) are likely to be identified using the sensorial cues we employed here. Therefore, it is likely that we are excluding a substantial amount of fermented foods (e.g. floral nectar and fruits with other levels of maturity (Aleksey Maro, personal communication; Wiens et al., 2008; Weaver, 2016) from our analysis. However, without systematic chemical and microbial analyses of primate food items, this conservative approach represents the current best available option.

Additionally, foods, particularly fruits, in late stages of fermentation are more likely to have physiological effects on consumers as a result of higher concentrations of microbes and/or microbial by-products (Tamang et al., 2016). As a result, we may be strengthening our statistical ability to detect ecological patterns by focusing on these late-stage fermented foods. Given that the relative concentrations of ethanol and other microbial products--and the likelihood of perception by human observers--are likely to vary by plant species, quantitative data describing these variables for a range of food items are necessary to test the extent to which primate ecology varies with food fermentation stage. However, that analysis is beyond the scope of the current paper, and independent efforts to achieve this goal are currently underway. We hope to address this question more effectively in the future once additional data are available.

We collated data for 40 species of primates inhabiting 50 research sites (Table S1). While these data encompass a small percentage of all extant primate species (7.9%, 40 out of 504 recognized species; Estrada et al., 2017), 11 out of the 16 extant primate families were represented across all continents inhabited by primates, and we included both tropical and temperate environments. Therefore, we believe that our database fairly represents the phylogenetic and geographic diversity of the order Primates. Study duration ranged from 9 to 312 months (median = 15 months), and we used data from multiple social groups or communities of 18 species distributed across 13 sites.

We included data describing the location and length of their study, the elevation, mean annual maximum and minimum temperatures, mean daily temperature, and mean annual rainfall of the study site, the mean contribution of fruits, leaves, and invertebrates to the diet of the study species, the frequency with which any fermented foods were consumed relative to total observation time, and any other relevant details about the types of foods

consumed or associated behavior, such as seasonality or specific handling behaviors (Table S1).

#### 2.2 | Physical and chemical traits of the late-stage fermented foods consumed

For all late-stage fermented foods, we compiled data from each study site or the literature describing presence/absence of a tough husk or skin, relative fiber content, and presence/absence of secondary metabolites and their concentrations. We evaluated tough husks qualitatively. A relative assessment of fiber content compared to other fruits at the same site was possible for 35 fruits, and for 25 of these fruits the nutritional data were available for that study site specifically. Secondary metabolite data were more difficult to compile. Quantitative data were available for 11 fruit species at three sites. For the rest of the fruits, we searched the literature using the fruit species name combined with terms such as 'toxin' and 'secondary metabolite.' Because data describing secondary metabolite content in fruits is sparse, in many cases we had to rely on literature describing medicinal use that implied increased concentrations of identified or unidentified secondary metabolites. Using this approach, we were able to find evidence of the occurrence of secondary metabolites for 34 fruit species (Table 3).

#### 2.3 | Data analysis

We assessed the influence of 18 predictor variables (Table 1) on the occurrence of latestage fermented food consumption via generalized linear mixed-effects models (GLMM; Zuur, Ieno, Walker, Saveliev, & Smith, 2009) with a binomial distribution and logit link function using the function 'lmer' of the R package lme4 (Bates et al., 2015). We specified the occurrence of late-stage fermented food consumption as a binary response variable, all the predictor variables as fixed factors, and study site as a random factor to account for data from repeated measures of the same species in different social groups at the same site. In addition to socio-ecological, climate, and habitat patch size variables, we included study length in all of our models to determine if shorter studies were biased against what we assumed would be a relatively rare behavior (Souza-Alves et al., 2019). To avoid overparameterization and problems of convergence with the global model, we did not consider variable interactions (see Grueber, Nakagawa, Laws, & Jamieson, 2011). We also did not include variables describing primate taxonomy because limited replication of species reduced the power of the analysis to assess the impact of these variables.

Given differences in scale among the predictor variables, we standardized them using the 'standardize' function of the package MuMIn (Barton, 2020) as recommended by Grueber et al. (2011). We avoided multicollinearity problems by only including those variables with Variance Inflation Factors (VIF) <3 into the models (Zuur et al., 2009) using the 'vifstep' function of the package usdm (Naimi, Hamm, Groen, Skidmore, & Toxopeus, 2014). The seven variables with VIF >3 that we excluded from the global model were female body mass, male body mass, male relative encephalization quotient, percentage of leaves in the diet, percentage of invertebrates in the diet, mean daily temperature, and elevation.

We selected models with an  $\Delta AICc < 2$  as the most parsimonious (Grueber et al., 2011). Given the occurrence of multiple equally parsimonious models, we also performed full-model averaging on all models with an  $\Delta AICc < 2$  to account for model uncertainty and to identify the best predictors of patterns of late-stage fermented food consumption in our data set (Grueber et al., 2011). We used the 'model average' function of the R package MuMIn

to identify the averaged model and the predictor weight ( $\sum w_i$ ) of each variable. We determined the coefficient of determination for each model with  $\Delta AICc < 2$  using the MuMIn 'r.squaredGLMM' function. We performed all statistical analyses in R 3.6.3 (R CoreTeam, 2020).

## 3 | RESULTS

## 3.1 | Occurrence of late-stage fermented food consumption in wild primates

Out of 40 species of wild primates studied at 50 sites, 15 species (37.5%) were reported to consume late-stage fermented foods at 23 sites in 12 countries across four continents. Overall, late-stage fermented food consumption occurred infrequently (Table 2, S1). We estimated that it constituted from 0.01% to 3% of the annual diet in most groups, although there were seasonal differences. For example, we found that late-stage fermented fruits could account for as much as 15% of the feeding records of *Cebus imitator* and *Alouatta guariba clamitans* during some seasons. For some primates, such as *A. guariba clamitans*, these seasons represented periods of low food availability (VBF personal observation), whereas for many others, such as *C. imitator*, they did not (EKM personal observation). While we recorded late-stage fermented food consumption in all our *Pan paniscus* and *C. imitator* social groups (three and seven, respectively), not all populations or social groups of the other species studied exhibited this behavior.

#### 3.2 | Main sources of fermented foods and behavioral strategies used

Late-stage fermented food consumption was limited to fruits (Tables 2, 3, S1). The richness of late-stage fermented fruits exploited ranged from one to nine fruit species for a given primate species (Tables 2, 3, S1). *Pan paniscus* exhibited the highest richness of late-stage fermented fruit species in the diet (N=9 fruit species), followed by *Ateles geoffroyi* (N=8), *Alouatta guariba clamitans* (N=7), and *Cebus imitator* (N=5; Table 2). The remaining primate species exploited between one and three fruit species (Table 2).

At least 31 of the 44 fruit species that were consumed in late stages of fermentation have defenses in the form of difficult-to-break tough husks/skins (N=16) or secondary metabolites, such as alkaloids, acetogens, saponins, and tannins (N=25; Table 3). Almost all fruits (95%) were consumed both ripe/unfermented and overripe/fermented (Table 3). In some cases, late-stage fermented fruits were only consumed when the patch was depleted of ripe fruits (Table S1). We also reported cases in which very ripe or late-stage fermented fruits appeared to be preferred over semi-ripe and unripe fruits. Specifically, Cebus imitator at La Suerte, Costa Rica, was observed frequently knocking ripe *Dipteryx oleifera* (Fabaceae) fruits to the ground and returning up to two weeks later to consume them (up to 15% of feeding time seasonally, EKM personal observation). These fruits were never consumed unfermented by the capuchins. Eulemur fulvus at Ampijiroa, Madagascar (up to 5% of feeding time seasonally, PTR personal observation) and Ateles geoffroyi at Punta Laguna, Mexico (up to 1% of feeding time seasonally, BPG personal observation) were also reported to drop fruits to the ground and return to feed on them later. However, unlike the capuchins, both lemurs and spider monkeys consumed the target fruits in different stages of ripening, although the lemurs appeared to prefer fallen fruits over those on the trees since they would consume fallen fruits first when both were available.

424

401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

422

## 3.3 | Main primate predictors of late-stage fermented fruit consumption

Only climate and habitat patch size were significant factors predicting late-stage fermented food consumption in wild primates. Other socio-ecological traits did not have a significant effect in any of our top ranked models. We found six GLMMs equally parsimonious ( $\Delta$ AIC <2) for explaining the observed patterns in late-stage fermented food consumption (Table 4). These models included mean maximum and minimum annual temperature, mean annual rainfall, habitat patch size, mean minimum annual temperature, longitude, home range size, and female relative encephalization quotient and explained approximately 99% of the observed variance (Table 4). However, only mean annual maximum temperature, rainfall, and habitat patch size were present in all six models. The model with the strongest empirical support ( $\Delta$ AICc = 0.00) included these three variables and mean minimum annual temperature (Table 4). The averaged model explained 99% of the observed variance, and late-stage fermented food consumption was only significantly negatively influenced by annual mean rainfall and mean annual minimum temperature, and positively influenced by mean annual maximum temperature and habitat patch size (Table 4).

#### 4 | DISCUSSION

We found that wild primates from all major evolutionary lineages consume foods in the later stages of fermentation, although the behavior is relatively infrequent and limited to only a few species of fruits at the sites where we recorded it. Additionally, climatic and environmental variables generally predict the occurrence of late-stage fermented food consumption better than socioecological variables. Specifically, late-stage fermented food

consumption is more common in hotter, drier environments and in larger, presumably less fragmented, habitats. These findings provide an important foundation for understanding the ecological and evolutionary forces that drive fermented food consumption in primates and offer new insights into the emergence of this behavior in humans.

## 4.1 | Occurrence of fermented food consumption in wild primates

First, although reports of fermented food consumption are rare in most studies of wild primate feeding ecology, this behavior is probably pervasive across the Order. We observed late-stage fermented food consumption in more than one third of the primate species for which we received data. However, given that our data were biased toward late-stage fermentation and many fermented foods consumed by primates cannot be identified by researchers without chemical analyses, it is likely that the prevalence of fermented food consumption among wild primates is even higher. Fruits consumed by primates commonly ferment naturally despite no clear signs to observers that fermentation has occurred (Dominy, 2004; Dudley, 2002; Aleksey Maro, personal communication; Weaver, 2016). Given that most primates, even those considered leaf-eaters, rely heavily on fruit during at least part of the year (Campbell et al., 2011; Rowe, 2018; Sussman, 1991), it is likely that most primates regularly consume fermented foods. This scenario becomes more probable when we consider the fact that other foods such as nectar or gums may also often ferment despite being difficult to observe (e.g. Wiens et al., 2008)

However, we do not expect that all primates consume fermented fruits. For instance, primates of the subfamily Colobinae, which are physically unable to consume large amounts of ripe fruits as a result of their sacculated foregut (Davies & Oates, 1994), as well

as immature fruit specialists, such as the Neotropical Pitheciinae, were not observed consuming fermented fruit (at least not clearly overripe fruits) in any context in this study. Additionally, the physical nature of some habitats can reduce access to fermented fruits. For example, while not represented in our data set, swamps and riverbank forests reduce opportunities for fruit fermentation on the ground, and fruits in these habitats are often water-dispersed and rarely fleshy and easily fermentable (López, 2001).

# 4.2 | Ecological contexts associated with late-stage fermented food consumption by wild primates

Despite how relatively common late-stage fermented fruit consumption appears to be throughout the Order Primates, we found that it is selectively employed in specific ecological contexts. Although most primates include many fruit species in their diets, in most cases only one or two fruit species were consumed in the late stages of fermentation by a given primate population or social group. In some cases, this pattern appeared to be a result of primates extending the utility of a fruit patch. For example, in the rare instances when *Pongo pygmaeus* was observed consuming late-stage fermented fruits, it was after the patch had been depleted by other frugivores (ERV personal observation). Alternatively, some primates, such as groups of *Alouatta guariba clamitans* in Santa Maria municipality, Southern Brazil, appeared to rely on late-stage fermented fruits during periods of low or altered food availability (VBF personal observation). Similarly, *Ateles geoffroyi* on Barro Colorado Island, Panama utilized late-stage fermented *Quararibea asterolepis* during a period of unusual fruiting patterns associated with the previous year's El Niño event, as did other frugivorous mammals and birds (Campbell, 2000). These potential uses of late-stage

fermented foods as fallback foods are in line with previous hypotheses in other contexts (Carrigan et al., 2015).

Other primates appeared to use fermentation to increase fruit edibility. Many fruits contain secondary metabolites, and in some cases they may reach sufficient levels to have meaningful physiological effects if consumed in large quantities (Cipollini & Levey, 1997; Janzen, 1983). At least two-thirds of the fruit species consumed in the late stages of fermentation by wild primates in this study had mechanical or chemical herbivore defenses when unfermented. For seven of these species, primates were reported to reject fruits unless they were very ripe or fermented. *Pan troglodytes* has been previously shown to preferentially consume ripe fruits of plant species whose unripe fruits have high levels of tannins since ripening reduces tannin content (Wrangham & Waterman, 1983). Therefore, it is possible that fermentation was used by some of our study subjects in a similar way to break down plant herbivore defenses. For example, *Dipteryx oleifera*, has a hard husk that can only be breached by *Cebus imitator* when fermented (EKM personal observation).

Together, these patterns are compatible with the use of targeted consumption of late-stage fermented fruits in multiple ways by primates as part of a broader nutritional strategy to increase food availability and expand their dietary niches. We found preliminary support for this interpretation. As predicted, our models indicated that late-stage fermented food consumption was associated with climate and habitat patch size. In particular, late-stage fermented food consumption was more common in drier environments with more extreme mean annual maximum and minimum temperatures, as well as in larger habitat patches. Habitats with higher mean annual maximum temperatures, lower mean annual minimum temperatures, and lower annual rainfall are potentially more nutritionally stressful for primates due to both chronic and seasonal reductions in food availability, as

well as distinct plant growth strategies that result in increased mechanical and/or chemical defenses against herbivory (Coley & Barone, 1996; Onoda et al., 2011; Poorter & Kitajima, 2007; Zhao, Hartmann, Trumbore, Ziegler, & Zhang, 2013). In such environments, a primate foraging strategy that relied more heavily on late-stage fermented foods could well enhance survival during lean periods by both extending the utility of depleted food patches and increasing digestibility of heavily defended plant foods. We do not have quantitative data relating food availability or plant herbivore defenses to late-stage fermented food consumption across sites, precluding our ability to rigorously test this hypothesis here. However, future explorations of this relationship are warranted by our findings.

Our results also indicate other potentially important mechanisms driving patterns of primate late-stage fermented food consumption. To some extent, it appears that late-stage fermented food consumption occurs with more prevalence in habitats where primates are more likely to come into contact with fruit in the late stages of fermentation. Higher mean annual maximum temperatures are likely to result in more rapid rates of fermentation and ethanol evaporation (Isu & Njoku, 1998), increasing the local prevalence of late-stage fermented foods and the probability that primates will detect them via olfaction (Dominy, 2004; Nevo & Valenta, 2018; Melin et al., 2019). Furthermore, larger, potentially less fragmented, habitats are often associated with an increased abundance and diversity of fruiting trees (Abbas et al., 2011; Fahrig, 2003; Laurance, Delamonica, Laurance, Vasconcelos, & Lovejoy, 2000). Therefore, there may be a higher probability that primates in these habitats will encounter fermenting fruits. However, we found that late-stage fermented food consumption was a function of both low mean annual minimum temperatures and high annual maximum temperatures, and in our dataset, the effect of habitat patch size appears to be driven by two particularly large sites, Goualougo and

Mandika in the Republic of Congo. As a result, it remains unclear whether factors influencing the availability of late-stage fermented foods to primates truly shape patterns of consumption more globally.

548

549

547

545

546

## 4.3 | Potential evolutionary benefits of late-stage fermented food consumption

550

551

552

553

554

555

556

557

558

559

560

561

562

563

564

565

566

567

568

The aforementioned relationships open up new perspectives on the emergence of food fermentation as an important component of the human diet. If late-stage fermented food consumption is part of an extant primate strategy for extending the time over which a particular type of fruit can be fed on and/or increasing the nutritional accessibility of foods, particularly in nutritionally harsh environments or environments with high levels of interspecific feeding competition, it may have served a similar role for our hominin ancestors. As hominids diverged from other primates, they began to more consistently occupy a more terrestrial niche (Sponheimer et al., 2013). It has been suggested that fermented fruits may have emerged as a fallback food in this context (Carrigan et al., 2015), and the patterns we observed in extant non-human primates provide some support for this hypothesis. Additionally, hominins including *Paranthropus* and *Australopithecus* are believed to have incorporated substantial amounts of hard and abrasive food items, as well as underground plant storage organs, in their diets (Dominy, 2012; Kay, 1985; Plummer, 2004; Teaford & Ungar, 2000). Underground plant storage organs are mechanically challenging, contain more starch and fiber compared to most ripe fruits, and expose foragers to potentially high amounts of diverse secondary plant metabolites that are toxic or can interfere with digestion (Buonocore & Silano, 1986; Dominy, Vogel, Yeakel, Constantino, & Lucas, 2008; Stahl et al., 1984; Waterman, 1984).

Fermentation could have reduced both the fiber and toxin levels in these food items. In fact, fermentation is commonly used to process tubers in modern human contexts (Akinrele, 1964; Ray & Sivakumar, 2009). While the transition to more settled, agrarian communities is often associated with the advent of human fermented food production for food preservation and ritual (Kuijt, 2009; Liu et al., 2018; Ross et al., 2002), the potential nutritional benefits of fermentation should not be underestimated. We found evidence that these benefits may be important drivers of late-stage fermented food consumption across the Order Primates.

569

570

571

572

573

574

575

576

577

578

579

580

581

582

583

584

585

586

587

588

589

590

591

592

Other nutritional and non-nutritional factors that we could not quantify should also be considered as proximate drivers of late-stage fermented food consumption in primates. First, the nutritional benefits of late-stage fermented fruits could be further improved by the presence of insects. Whilst generally composed of small individuals, insect assemblages in fermenting fruit can be diverse and abundant (Braham, 2015; Feinstein, Mori, & Berkov, 2007; Hodge & Arthur, 1996). Insects can provide fat, protein, vitamins and amino acids (Barnett et al., in press; Xiaoming et al., 2010), and fruit infested with them are known to be selected by some primate species in other contexts (Barnett et al., 2017). Additionally, fermentation is likely to alter food taste. Anecdotal researcher taste tests in our study indicated positive changes in taste with fruit fermentation. Fermentation is generally associated with sour or acid tastes, and humans tend to prefer sweet-sour tastes (Breslin, 2013; Katz, 2012). Little is known about sour taste receptors in primates and other animals—or even sour taste preference (Montell, 2018; Roper, 2007). However, it is likely that primates share an affinity for sour taste with humans. Taste has not been systematically examined in wild primate foods, but it will likely provide additional insight into primate food choices, both fermented and unfermented.

Finally, fermented foods are likely to provide health benefits to consumers as a result of probiotic and prebiotic properties (Bourrie et al., 2016; Burton et al., 2017; Löwenadler & Linberg, 1994; Marco et al., 2017; Summer et al., 2017; Tamang et al., 2016; Veiga et al., 2014; Yartey et al., 1995). These properties are likely to be stronger in late-stage fermented foods as a result of increased microbial activity, which may explain why these foods are targeted by some primates. Currently, without chemical and microbial data from primate foods as well as physiological and microbial data from primates, it is impossible to assess these potential relationships. However, rapidly emerging evidence of the importance of microbes for primate ecology and evolution (Amato, 2016; Amato, Jeyakumar, Poinar, & Gros, 2019; Davenport et al., 2017; Dunn et al., 2020; Gaulke et al., 2018) suggests that these interactions should not be overlooked.

#### Conclusion

We find that late-stage fermented fruits are consumed by a variety of non-human primates globally. This behavior generally targets a specific subset of fruit species, some of which contain herbivore defenses that are likely degraded by bacterial fermentation. It also occurs more often in drier environments with more extreme mean annual temperatures, and in larger habitat patches. As a result, we suggest that primate late-stage fermented food consumption may be part of a nutritional strategy that increases food availability by increasing the duration across which a particular fruit patch can be used, and expands dietary niche space by degrading some toxins in ripe fruit and providing easily accessible nutrients. It is possible that the human propensity for fermented food consumption is rooted on this ancestral primate strategy, which was favored during the course of human evolution by periods of nutritional stress caused by climate change events and migration to unknown

617 or unfavorable landscapes. Future studies should pair systematic assessments of spatial and 618 temporal patterns of wild primate fermented food consumption with nutritional and 619 microbial analyses of fermented and unfermented food items to further investigate these 620 relationships. 621 622 **ACKNOWLEDGEMENTS** 623 We thank Jessica Rothman for contributing data from C. ascanius, C. guereza, L. albigena, 624 P. anubis, and P. rufomitratus at Kibale National Park and G. beringei at Bwindi 625 Impenetrable Forest (funding through NSF 1528521), as well as for her input on earlier 626 versions of the manuscript. The authors would also like to acknowledge a long list of 627 funders, permitting agencies, and people for supporting the fieldwork associated with each 628 field site listed. This list can be accessed in the Supplementary Material. KRA is supported 629 as a fellow in the CIFAR 'Humans and the Microbiome' program. She would also like to 630 thank the organizers of the Wenner-Gren Symposium #160 'Cultures of Fermentation,' 631 held on October 11-17, 2019 (C. Warinner, J. Hendy, M. Aldenderfer, M. Rest) for 632 sparking the idea for this paper 633 634 635 636 **REFERENCES** 637 Abbas, F., Morellet, N., Hewison, A. J. M., Merlet, J., Cargnelutti, B., Lourtet, B., . . . 638 Verheyden, H. (2011). Landscape fragmentation generates spatial variation of diet composition and quality in a generalist herbivore. *Oecologia*, 167(2), 401–411. 639

Aiello, L. C., & Key, C. (2002). Energetic consequences of being a *Homo erectus* female.

American Journal of Human Biology, 14, 551-565.

640

- Aiello, L. C., & Wells, J. C. K. (2002). Energetics and the evolution of the genus Homo.
- 643 Annual Review of Anthropology, 31(1), 323–338.
- 644 Akinrele, I. (1964). Fermentation of cassava. Journal of the Science of Food and
- 645 Agriculture, 15(9), 589–594.
- 646 Al Nabhani, Z., & Eberl, G. (2020). Imprinting of the immune system by the microbiota
- early in life. *Mucosal Immunology*, 13, 183–189.
- Amato, K. R. (2016). Incorporating the gut microbiota into models of human and non-
- human primate ecology and evolution. Yearbook of Physical Anthropology, 159, S196-
- 650 S215.
- Amato, K. R., Jeyakumar, T., Poinar, H., & Gros, P. (2019). Shifting Climates, Foods, and
- Diseases: The Human Microbiome through Evolution. *Bioessays*, 41(10), 1900034.
- Anraku, K., Nonaka, K., Yamaga, T., Yamamoto, T., Shin, M., Wakita, M., . . . Akaike, N.
- 654 (2013). Removal of toxin (tetrodotoxin) from puffer ovary by traditional fermentation.
- 655 *Toxins*, 5(1), 193–202.
- Antón, S. C., Potts, R., & Aiello, L. C. (2014). Evolution of early Homo: an integrated
- 657 biological perspective. *Science*, *345*(6192), 1236828.
- Barnett, A., Ronchi-Teles, B., Silva, W., Andrade, R., Almeida, T., Bezerra, B., . . . Ross,
- 659 C. (2017). Covert carnivory? A seed-predating primate, the golden-backed uacari, shows
- preferences for insect-infested fruits. *Journal of Zoological Research*, 1, 16–33.
- Barnett, A., Stone, A., Shaw, P., Ronchi-Teles, B., Pimenta, N., Kinup, N., . . . Wenzel, J.
- 662 (in press). When food fights back: at-nest predation of larval paper-wasps by
- Neotropical Cebid primates, the high-energy yield of high-risk foraging. *American*
- 664 Journal of Primatology.
- Barton, K. (2020). R package MuMIn: model selection inference. R package version 1.43.7.
- https://cran.r-project.org/web/packages/MuMIn/MuMIn.pdf
- Bates, D., Maechler, M., Bolker, B., & Walker, S. (2015). Fitting Linear Mixed-Effects
- Models using lme4. *Journal of Statistical Software*, 67(1), 1–48.
- Battcock, M., & Azam-Ali, S. (1998). Fermented fruits and vegetables: a global
- 670 perspective. Rome: Food & Agriculture Organization of the United Nations.
- Biale. J.B. (1954). The ripening of fruit. Scientific American. 190:40-45.
- Binita, R., & Khetarpaul, N. (1997). Probiotic fermentation: Effect on antinutrients and
- digestibility of starch and protein of indigenously developed food mixture. *Nutrition and*
- 674 *health*, 11(3), 139–147.
- Boulton, R. B., Singleton, V. L., Bisson, L. F., & Kunkee, R. E. (1999). Yeast and
- biochemistry of ethanol fermentation. In R. B. Boulton, V. L. Singleton, L. F. Bisson,
- & R. E. Kunkee (Eds.), *Principles and practices of winemaking* (pp. 102–192). New
- 678 York: Springer.
- Bourrie, B. C., Willing, B. P., & Cotter, P. D. (2016). The microbiota and health promoting
- characteristics of the fermented beverage kefir. Frontiers in Microbiology, 7, 647.

- Brady, A. G., & Carville, A. A. (2012). Digestive system diseases of nonhuman primates.
- In C. R. Abee, K. Mansfield, S. Tardiff, & T. Morris (Eds.), Nonhuman primates in
- *biomedical research* (pp. 589–627). New York: Elsevier Inc.
- Braham, M. (2015). Insect larvae associated with dropped pomegranate fruits in an organic orchard in Tunisia. *Journal on Entomology and Nematology*, 7, 5–10.
- 686 Breslin, P. A. (2013). An evolutionary perspective on food and human taste. *Current Biology*, 23(9), R409–R418.
- Buonocore, V., & Silano, V. (1986). Biochemical, nutritional and toxicological aspects of
- alpha-amylase inhibitors from plant foods. *Advanced Experimental Medicine and Biology*, *199*, 483–507.
- Burton, K. J., Rosikiewicz, M., Pimentel, G., Bütikofer, U., Von Ah, U., Voirol, M.-J., . . .
- McTernan, P. G. (2017). Probiotic yogurt and acidified milk similarly reduce
- postprandial inflammation and both alter the gut microbiota of healthy, young men.
- 694 British Journal of Nutrition, 117(9), 1312–1322.
- 695 Campbell, C.J. (2000). The reproductive biology of black-handed spider monkeys (Ateles
- 696 *geoffroyi*): integrating behavior and endocrinology. Ph.D. Thesis, University of
- 697 California, Berkeley.
- 698 Campbell, C.J., Fuentes, A., MacKinnon, K., Bearder, S., & Stumpf, R. (2011). *Primates in Perspective* (Second Edition ed.). New York: Oxford University Pres.
- Campbell-Platt, G. (1994). Fermented foods—a world perspective. *Food Research International*, 27(3), 253–257.
- Carrigan, M. A., Uryasev, O., Frye, C. B., Eckman, B. L., Myers, C. R., Hurley, T. D., &
- Benner, S. A. (2015). Hominids adapted to metabolize ethanol long before human-
- directed fermentation. *Proceedings of the national Academy of Sciences*, 112(2), 458–463.
- 706 Chaves-López, C., Serio, A., Grande-Tovar, C. D., Cuervo-Mulet, R., Delgado-Ospina, J.,
- We Paparella, A. (2014). Traditional fermented foods and beverages from a
- microbiological and nutritional perspective: the Colombian heritage. Comprehensive
- 709 Reviews in Food Science and Food Safety, 13(5), 1031–1048.
- Cipollini, M. L., & Levey, D. J. (1997). Why are some fruits toxic? Glycoalkaloids in solanumand fruit choice by vertebrates. *Ecology*, 78(3), 782–798.
- 712 Clutton-Brock, T. H., & Harvey, P. H. (1980). Primates, brains and ecology. *Journal of Zoology*, *190*(3), 309–323.
- Coley, P. D., & Barone, J. (1996). Herbivory and plant defenses in tropical forests. *Annual Review of Ecology and Systematics*, 27(1), 305–335.
- 716 Cryan, J. F., O'Riordan, K. J., Cowan, C. S., Sandhu, K. V., Bastiaanssen, T. F., Boehme,
- M., ... Golubeva, A. V. (2019). The microbiotagut-brain axis. *Physiological Reviews*,
- 718 99(4), 1877–2013.
- Dausch Ibañez, D., Hernandez Salazar, L. T., & Laska, M. (2019). Taste responsiveness of
- spider monkeys to dietary ethanol. *Chemical senses*, 44(8), 631–638.

- Davenport, E. R., Sanders, J. G., Song, S. J., Amato, K. R., Clark, A. G., & Knight, R.
- 722 (2017). The human microbiome in evolution. *BMC Biology*, 15, 127.
- 723 Davies, A., & Oates, J. (1994). Colobine Monkeys: their ecology, behavior and evolution.
- New York: Cambridge University Press.
- DeCasien, A. R., Williams, S. A., & Higham, J. P. (2017). Primate brain size is predicted
- by diet but not sociality. *Nature Ecology & Evolution*, 1(5), 1–7.
- 727 Deshpande, S. (2000). Fermented grain legumes, seeds and nuts: a global perspective (Vol.
- 728 142). Rome: Food & Agriculture Organization of the United Nations.
- 729 Dominy, N. J. (2015). Ferment in the family tree. Proceedings of the national Academy of
- 730 *Sciences*, 112(2), 308–309.
- 731 Dominy, N. (2012). Hominins living on the sedge. Proceedings of the National Academy of
- 732 *Science*, 109(50), 20171–20172.
- 733 Dominy, N. J. (2004). Fruits, fingers, and fermentation: the sensory cues available to
- foraging primates. *Integrative and Comparative Biology*, 44(4), 295–303.
- 735 Dominy, N., Vogel, E. R., Yeakel, J. D., Constantino, P., & Lucas, P. W. (2008).
- Mechanical properties of plant underground storage organs and implications for dietary
- models of early hominins. *Evolutionary Biology*, 35(3), 159–175.
- Duar, R. M., Lin, X. B., Zheng, J., Martino, M. E., Grenier, T., Pérez–Muñoz, M. E., . . .
- Walter, J. (2017). Lifestyles in transition: evolution and natural history of the genus
- 740 Lactobacillus. FEMS Microbiology Reviews, 41, S27–S48.
- 741 Dudley, R. (2014). The drunken monkey: why we drink and abuse alcohol. Los Angeles:
- 742 University of California Press.
- Dudley, R. (2004). Ethanol, fruit ripening, and the historical origins of human alcoholism in
- primate frugivory. *Integrative and Comparative Biology*, 44(4), 315–323.
- Dudley, R. (2002). Fermenting fruit and the historical ecology of ethanol ingestion: is
- alcoholism in modern humans an evolutionary hangover? *Addiction*, 97(4), 381–388.
- Dunbar, R., & Shultz, S. (2017). Why are there so many explanations for primate brain
- evolution? *Philosophical Transactions of the Royal Society B: Biological Sciences*,
- 749 *372*(1727), 20160244.
- 750 Dunbar, R. I. M., & Shultz, S. (2007). Understanding primate brain evolution.
- 751 Philosophical Transactions of the Royal Society B, 362, 649–658.
- Dunn, R. R., Amato, K. R., Archie, E. A., Arandjelovic, M., Crittenden, A. N., & Nichols,
- L. M. (2020). The Internal, external and extended microbiomes of hominins. *Frontiers*
- 754 in Ecology and Evolution, 8, 25.
- Estrada, A., Garber, P. A., Rylands, A. B., Roos, C., Fernandez–Duque, E., Di Fiore, A., . .
- Lambert, J. E. (2017). Impending extinction crisis of the world's primates: why
- primates matter. Science Advances, 3(1), e1600946.
- 758 Fahrig, L. (2003). Effects of habitat fragmentation on biodiversity. *Annual Review of*
- 759 Ecological and Evolutionary Systems, 34, 487–515.
- Feinstein, J., Mori, S., & Berkov, A. (2007). Saproflorivory: a diverse insect community in
- fallen flowers of Lecythidaceae in French Guiana. *Biotropica*, 39(4), 549–554.

- Felton, A., Felton, A., Wood, J., Foley, W., Raubenheimer, D., Wallis, I., & Lindenmayer,
- D. (2009). Nutritional ecology of *Ateles chamek* in lowland Bolivia: how macronutrient
- balancing influences food choices. *International Journal of Primatology*, 30(5), 675–
- 765 696.
- Garnier, N., & Valamoti, S. M. (2016). Prehistoric wine-making at Dikili Tash (northern
- Greece): integrating residue analysis and archaeobotany. *Journal of Archaeological*
- 768 Science, 74, 195–206.
- Gaulke, C. A., Arnold, H. K., Humphreys, I. R., Kembel, S. W., O'Dwyer, J. P., &
- Sharpton, T. J. (2018). Ecophylogenetics clarifies the evolutionary association between
- mammals and their gut microbiota. *mBio*, 9(5), e01348–01318.
- Gobbetti, M., Simonetti, M., Rossi, J., Cossignani, L., Corsetti, A., & Damiani, P. (1994).
- Free D-and L-amino acid evolution during sourdough fermentation and baking. *Journal*
- 774 of Food Science, 59(4), 881–884.
- Gochman, S. R., Brown, M. B., & Dominy, N. J. (2016). Alcohol discrimination and
- preferences in two species of nectar-feeding primate. Royal Society Open Science, 3(7),
- 777 160217.
- Gorgus, E., Hittinger, M., & Schrenk, D. (2016). Estimates of ethanol exposure in children
- from food not labeled as alcohol-containing. *Journal of Analytical Toxicology*, 40(7),
- 780 537–542.
- 781 Grueber, C., Nakagawa, S., Laws, R., & Jamieson, I. (2011). Multimodel inference in
- ecology and evolution: challenges and solutions. Journal of Evolutionary Biology, 24(4),
- 783 699–711.
- Gupta, R. K., Gangoliya, S. S., & Singh, N. K. (2015). Reduction of phytic acid and
- enhancement of bioavailable micronutrients in food grains. *Journal of Food Science and*
- 786 Technology, 52(2), 676–684.
- Hayden, B., Canuel, N., & Shanse, J. (2013). What was brewing in the Natufian? An
- archaeological assessment of brewing technology in the Epipaleolithic. *Journal of*
- 789 Archaeological Method and Theory, 20(1), 102–150.
- Hockings, K. J., Bryson–Morrison, N., Carvalho, S., Fujisawa, M., Humle, T., McGrew, W.
- 791 C., ... Yamakoshi, G. (2015). Tools to tipple: ethanol ingestion by wild chimpanzees
- using leaf-sponges. Royal Society Open Science, 2(6), 150150.
- Hodge, S., & Arthur, W. (1996). Insect invasion sequences: systematic or stochastic?
- 794 *Ecological Entomology*, 21(2), 150–154.
- 795 Isu, N., & Njoku, H. (1998). Studies on the influence of temperature, relative humidity and
- microenvironment on the natural fermentation of African oil bean seeds to 'Ugba'. *Plant*
- 797 *Foods for Human Nutrition*, 52(4), 337–351.
- Jacobsen, C. N., Nielsen, V. R., Hayford, A., Møller, P. L., Michaelsen, K., Paerregaard,
- A., ... Jakobsen, M. (1999). Screening of probiotic activities of forty-seven strains of
- 800 Lactobacillus spp. by in vitro techniques and evaluation of the colonization ability of
- five selected strains in humans. *Appl. Environ. Microbiol.*, 65(11), 4949–4956.

- Janiak, M. C., Pinto, S. L., Duytschaever, G., Carrigan, M. A., & Melin, A. D. (2020).
- Genetic evidence of widespread variation in ethanol metabolism among mammals:
- revisiting the 'myth'of natural intoxication. *Biology Letters*, 16(4), 20200070.
- Janzen, D. H. (1983). Physiological ecology of fruits and their seeds. In O. Lange, P.
- Nobel, C. Osmond, & H. Ziegler (Eds.), *Physiological plant ecology III* (pp. 625–655).
- 807 Berlin: Springer.
- Katz, S. E. (2012). *The art of fermentation: an in-depth exploration of essential concepts* and processes from around the world. White River Junction: Chelsea Green Publishing.
- Kay, R. F. (1985). Dental evidence for the diet of Australopithecus. *Annual Review of Anthropology*, *14*(1), 315–341.
- 812 Kim, J., Choi, E., Hong, Y., Song, Y., Han, J., Lee, S., . . . Cho, K. (2016). Changes in
- Korean adult females' intestinal microbiota resulting from kimchi intake. *Journal of*
- Nutrition and Food Science, 6, 4172.
- Kornet, M., Goosen, C., Ribbens, L. G., & Van Ree, J. M. (1990). Analysis of spontaneous
- alcohol drinking in rhesus monkeys. *Physiology & behavior*, 47(4), 679–684.
- Kudo, H., & Dunbar, R. I. M. (2001). Neocortex size and social network size in primates.
- 818 *Animal Behavior*, 62(4), 711–722.
- Kuijt, I. (2009). What do we really know about food storage, surplus, and feasting in
- preagricultural communities? *Current Anthropology*, 50(5), 641–644.
- Laurance, W. F., Delamonica, P., Laurance, S. G., Vasconcelos, H. L., & Lovejoy, T. E.
- 822 (2000). Rainforest fragmentation kills big trees. *Nature*, 404, 806.
- LeBlanc, J. G., Milani, C., de Giori, G. S., Sesma, F., van Sinderen, D., & Ventura, M.
- 824 (2013). Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Current Opinion in Biotechnology*, 24(2), 160–168.
- 826 Leonard, W. R., & Robertson, J. (1997). Comparative primate energetics and hominid
- evolution. American Journal of Physical Anthropology, 102(2), 265–281.
- Leonard, W. R., & Robertson, M. (1992). Nutritional requirements and human evolution: a
- bioenergetics model. *American Journal of Human Biology*, 4, 179–195.
- Liu, L., Wang, J., Rosenberg, D., Zhao, H., Lengyel, G., & Nadel, D. (2018). Fermented
- beverage and food storage in 13,000 y-old stone mortars at Raqefet Cave, Israel:
- Investigating Natufian ritual feasting. Journal of Archaeological Science: Reports, 21,
- 833 783–793. ie
- 834 López, O. (2001). Seed flotation and postflooding germination in tropical terra firme and
- seasonally flooded forest species. *Functional Ecology*, 763–771.
- Löwenadler, J., & Linberg, C. L. (1994). Study of locally prepared lactic acid fermented
- 837 weaning food in Tanzania -Presence of diarrhoeal pathogens. A minor field study.
- [Working Paper]. International Rural Development Center, Swedish University of
- Agricultural Sciences, Sweden.
- Maldonado-Gómez, M. X., Martínez, I., Bottacini, F., O'Callaghan, A., Ventura, M., van
- Sinderen, D., . . . Hutkins, R. W. (2016). Stable engraftment of *Bifidobacterium longum*

- AH1206 in the human gut depends on individualized features of the resident
- 843 microbiome. *Cell Host & Microbe*, 20(4), 515–526.
- Mandillo, S., Titchen, K., & Miczek, K. A. (1998). Ethanol drinking in socially housed
- squirrel monkeys. *Behavioural pharmacology*, 9(4), 363–368.
- 846 Marco, M. L., Heeney, D., Binda, S., Cifelli, C. J., Cotter, P. D., Foligné, B., . . . Pihlanto,
- A. (2017). Health benefits of fermented foods: microbiota and beyond. *Current Opinion* in *Biotechnology*, 44, 94–102.
- Martinson, E. O., Herre, E. A., Machado, C. A., & Arnold, A. E. (2012). Culture-free
- survey reveals diverse and distinctive fungal communities associated with developing
- figs (Ficus spp.) in Panama. Microbial Ecology, 64(4), 1073–1084.
- Melin, A.D., Nevo, O., Shirasu, M., Williamson, R.E., Garrett, E.C., Endo, M., Sakurai, K.,
- Matsushita, Y., Touhara, K., Shoji, K. (2019). Fruit scent and observer colour vision
- shape food-selection strategies in wild capuchin monkeys. *Nature Communications*, 1,
- 855 2407.
- Milton, K. (2004). Ferment in the family tree: does a frugivorous dietary heritage influence
- contemporary patterns of human ethanol use? *Integrative and Comparative Biology*,
- 858 44(4), 304–314.
- Mitchell, M. C., & Herlong, H. F. (1986). Alcohol and nutrition: caloric value,
- bioenergetics, and relationship to liver damage. Annual Review of Nutrition, 6(1), 457–
- 861 474.
- 862 Montell, C. (2018). pHirst sour taste channels pHound? *Science*, *359*(6379), 991–992.
- Naimi, B., Hamm, N. A., Groen, T. A., Skidmore, A. K., & Toxopeus, A. G. (2014). Where
- is positional uncertainty a problem for species distribution modelling? *Ecography*, 37(2),
- 865 191–203.
- Nayak, B. S., Dinda, S. C., & Ellaiah, P. (2013). Evaluation of diuretic activity of Gmelina
- arborea Roxb. fruit extracts. Asian Journal of Pharmaceutical and Clinical Research,
- 868 *6*(1), 111–113.
- Nevo, O., & Valenta, K. (2018). The ecology and evolution of fruit odor: implications for
- primate seed dispersal. *International Journal of Primatology*, 39(3), 338–355.
- NRC (1998). Lost Crops of Africa: Volume I: Grains. National Resource Council,
- Washington, D.C.: National Academies Press.
- NRC (2003). Nutrient requirements of nonhuman primates. National Resource Council,
- Washington, D.C.: National Academies Press.
- Nyanga, L. K., Nout, M. J., Gadaga, T. H., Theelen, B., Boekhout, T., & Zwietering, M. H.
- 876 (2007). Yeasts and lactic acid bacteria microbiota from masau (*Ziziphus mauritiana*)
- fruits and their fermented fruit pulp in Zimbabwe. *International Journal of Food*
- 878 *Microbiology*, 120(1–2), 159–166.
- 879 Oliphant, K., & Allen-Vercoe, E. (2019). Macronutrient metabolism by the human gut
- microbiome: major fermentation by-products and their impact on host health.
- 881 *Microbiome*, 7(1), 91.

- Onoda, Y., Westoby, M., Adler, P. B., Choong, A. M., Clissold, F. J., Cornelissen, J. H., . .
- 883 . Enrico, L. (2011). Global patterns of leaf mechanical properties. *Ecology Letters*,
- 884 *14*(3), 301–312.
- Peters, A., Krumbholz, P., Jäger, E., Heintz-Buschart, A., Çakir, M. V., Rothemund, S., . . .
- Stäubert, C. (2019). Metabolites of lactic acid bacteria present in fermented foods are
- highly potent agonists of human hydroxycarboxylic acid receptor 3. PLoS Genetics,
- 888 *15*(5), e1008145.
- Plummer, T. (2004). Flaked stones and old bones: Biological and cultural evolution at the
- dawn of technology. Yearbook of Physical Anthropology, 47, 118–164.
- Poorter, L., & Kitajima, K. (2007). Carbohydrate storage and light requirements of tropical
- moist and dry forest tree species. *Ecology*, 88(4), 1000–1011.
- Pretorius, I. S. (2000). Tailoring wine yeast for the new millennium: novel approaches to
- the ancient art of winemaking. Yeast, 16(8), 675–729.
- R Core Team. (2020). R: a language and environment for statistical computing (Version
- 3.6.3). Vienna, Austria: R Foundation for Statistical Computing.
- 897 Ray, R. C., & Sivakumar, P. S. (2009). Traditional and novel fermented foods and
- beverages from tropical root and tuber crops. International Journal of Food Science &
- 899 Technology, 44(6), 1073–1087.
- Rollan GC, Gerez CL, and LeBlanc JG. 2019. Lactic fermentation as a strategy to improve
- the nutritional and functional values of pseudocereals. Frontiers in Nutrition, 6, 1–16.
- 902 Roper, S. D. (2007). Signal transduction and information processing in mammalian taste
- buds. *Pflügers Archiv-European Journal of Physiology*, 454(5), 759–776.
- Ross, R. P., Morgan, S., & Hill, C. (2002). Preservation and fermentation: past, present and
- 905 future. *International Journal of Food Microbiology*, 79, 3–16.
- Rowe, M. H. (2018). Trichromatic color vision in primates. *Physiology*.
- Rowe, N., & Myers, M. (2016). All the world's primates. Charlestown: Pogonias Press.
- Ruiz Rodriguez, L. G., Mohamed, F., Bleckwedel, J., Medina, R. B., De Vuyst, L., Hebert,
- 909 E. M., & Mozzi, F. (2019). Diversity and functional properties of lactic acid bacteria
- 910 isolated from wild fruits and flowers present in Northern Argentina. Frontiers in
- 911 *Microbiology, 10,* 1091.
- 912 Skinner, F. A., Passmore, S. M., & Davenport, R. (1980). *Biology and activities of yeasts*.
- 913 London: Academic Press.
- 914 Smalley, J., Blake, M., Chavez, S. J., DeBoer, W. R., Eubanks, M. W., Gremillion, K. J., . .
- 915 . Piperno, D. R. (2003). Sweet beginnings: stalk sugar and the domestication of maize.
- 916 *Current Anthropology*, *44*(5), 675–703.
- 917 Souza-Alves, J. P., Mourthé, Í., Hilário, R. R., Bicca-Marques, J. C., Rehg, J., Gestich, C.
- 918 C.... Berthet, M. (2019). Terrestrial behavior in titi monkeys (*Callicebus*, *Cheracebus*,
- and *Plecturocebus*): potential correlates, patterns, and differences between genera.
- 920 International Journal of Primatology, 40, 553–572.

- 921 Speth, J.D. (2017). Putrid meat and fish in the eurasian middle and upper paleolithic: Are
- we missing a key part of neanderthal and modern human diet? *PaleoAnthropology*.
- 923 *2017*, 44-72.
- 924 Sponheimer, M., Alemseged, Z., Cerling, T. E., Grine, F. E., Kimbal, W. H., Leakey, M.
- 925 G., . . . Wood, B. A. (2013). Isotopic evidence of early hominin diets. *Proceedings of the*
- 926 *National Academy of Science, 110,* 10513–10518.
- 927 Stahl, A. B., Dunbar, R., Homewood, K., Ikawa-Smith, F., Kortlandt, A., McGrew, W., . . .
- Sugardjito, J. (1984). Hominid dietary selection before fire [and Comments and Reply].
- 929 *Current Anthropology*, 25(2), 151–168.
- 930 Strier, K. B. (2016). *Primate behavioral ecology*. New York: Routledge.
- 931 Summer, A., Formaggioni, P., Franceschi, P., Di Frangia, F., Righi, F., & Malacarne, M.
- 932 (2017). Cheese as functional food: the example of parmigiano reggiano and grana
- padano. Food Technology and Biotechnology, 55(3), 277–289.
- 934 Sussman, R. W. (1991). Primate origins and the evolution of Angiosperms. American
- 935 *Journal of Primatology*, 23(4), 209–223.
- 936 Sylvia, K. E., & Demas, G. E. (2018). A gut feeling: microbiome-brain-immune
- 937 interactions modulate social and affective behaviors. Hormones and Behavior, 99, 41-
- 938 49.
- 939 Tamang, J. P., Holzapfel, W. H., Shin, D. H., & Felis, G. E. (2017). Microbiology of ethnic
- 940 fermented foods and alcoholic beverages of the world. Frontiers in Microbiology, 8,
- 941 1377.
- Tamang, J., & Kailasapathy, K. (2010). Fermented foods and beverages of the world. Boca
- 943 Raton: CRC Press.
- Tamang, J. P., Shin, D.-H., Jung, S.-J., & Chae, S.-W. (2016). Functional properties of
- microorganisms in fermented foods. Frontiers in Microbiology, 7, 578.
- Teaford, M., & Ungar, P. (2000). Diet and the evolution of the earliest human ancestors.
- *Proceedings of the National Academy of Science*, 97(25), 13506–13511.
- Thomson, J. M., Gaucher, E. A., Burgan, M. F., De Kee, D. W., Li, T., Aris, J. P., &
- Benner, S. A. (2005). Resurrecting ancestral alcohol dehydrogenases from yeast. *Nature*
- 950 *Genetics*, 37(6), 630.
- Veiga, P., Pons, N., Agrawal, A., Oozeer, R., Guyonnet, D., Brazeilles, R., . . . Whorwell,
- P. J. (2014). Changes of the human gut microbiome induced by a fermented milk
- product. Scientific Reports, 4, 6328.
- Visconti, A., Le Roy, C. I., Rosa, F., Rossi, N., Martin, T. C., Mohney, R. P., . . . Venter, J.
- 955 C. (2019). Interplay between the human gut microbiome and host metabolism. *Nature*
- 956 *Communications*, 10(1), 1–10.
- Waterman, P. G. (1984). Food acquisition and processing as a function of plant chemistry.
- In D. J. Chivers, B. A. Wood, & A. Bilsborough (Eds.), Food Acquisition and
- 959 *Processing in Primates* (pp. 177–211). Boston: Springer.

- 960 Weaver, V. R. (2016). Dietary ethanol ingestion by free ranging spider monkeys (Ateles
- 961 geoffroyi): An evalution of the drunken monkey hypothesis. (unpublished master's
- thesis), California State University, Northridge.
- Wiens, F., Zitzmann, A., Lachance, M.-A., Yegles, M., Pragst, F., Wurst, F. M., . . .
- Spanagel, R. (2008). Chronic intake of fermented floral nectar by wild treeshrews.
- *Proceedings of the National Academy of Sciences, 105*(30), 10426–10431.
- 966 Wrangham, R. W. (2009). *Catching fire: how cooking made us human*. New York: Basic Books.
- Wrangham, R., & Waterman, P. (1983). Condensed tannins in fruits eaten by chimpanzees.
   *Biotropica*, 15(3), 217–222.
- 970 Wrangham, R. W., & Conklin–Brittain, N. L. (2003). Cooking as a biological trait.
- 971 *Comparative Biochemistry and Physiology, 136*(1), 35–46.
- Yiaoming, C., Ying, F., Hong, Z., & Zhiyong, C. (2010). Review of the nutritive value of
- edible insects. In P. B. Drust, D. V. Johnson, R. N. Leslie, & K. Shono (Eds.), Forest
- 974 insects as food: humans bite back (pp. 85–92). Bangkok, Thailand: Food and
- 975 Agriculture Organization of the United Nations.
- 976 Yartey, J., Nkrumah, F., Hori, H., Harrison, K., & Armar, D. (1995). Clinical trial of
- 977 fermented maize-based oral rehydration solution in the management of acute diarrhoea
- 978 in children. Annals of Tropical Paediatrics, 15(1), 61–68.
- 279 Zhao, J., Hartmann, H., Trumbore, S., Ziegler, W., & Zhang, Y. (2013). High temperature
- causes negative whole-plant carbon balance under mild drought. *New phytologist*,
- 981 200(2), 330–339.
- 982 Zuur, A. F., Ieno, E. N., Walker, N. J., Saveliev, A. A., & Smith, G. M. (2009). GLM and
- GAM for count data. In A. Zuur, E. N. Ieno, N. Walker, A. A. Saveliev, & G. M. Smith
- 984 (Eds.), *Mixed effects models and extensions in ecology with R* (pp. 209–243). New York:
- 985 Springer.

986

#### FIGURE LEGENDS

**Figure 1.** Wild primates consuming fermented fruits. (a) *Chlorocebus djadjamensis* consuming *L. abyssinica* at Kokosa, Ethiopia; credit Addisu Mekonnen (b) *Cebus capucinus imitator* consuming *D. oleifera* at La Suerte Biological Field Station, Costa Rica; credit: Liz Rasheed (c) *Pan paniscus* consuming *A. mannii* at LuiKatole, Democratic Republic of Congo; credit Gottfried Hohmann (d) *Ateles geoffroyi* consuming *M. zapota* at Punta Laguna, Mexico; credit Fabrizio Dell'Anna (e) *Alouatta guariba clamitans* consuming *P. guajava* at Parque São Paulo, Brazil; credit Claudio Godoy (f) *Macaca assamensis* consuming *N. cadamba* at Phu Khieo Wildlife Sanctuary, Thailand; credit Oliver Schülke (g) *Hapalemur meridionalis* consuming *Uapaca* sp. at Mandena, Madagascar; credit Tim Eppley (h) *Callithrix jacchus* consuming *P. pachycladus* at Baracuhy Biological Field Station, Brazil; credit: Filipa Abreu.

**Figure 2.** Fruits consumed fermented by wild primates. (a) *Lagenaria abyssinica*, credit: Addisu Mekonnen (b) *Stemmadenia obovata*, credit: Amanda Melin (c) *Vangueria madagascariensis*, credit: Tojotanjona Razanaparany (d) *Spondias mombin*, credit: Amanda Melin (e) *Landolphia myrtifolia*, credit Tojotanjona Razanaparany (f) *Diospoyros kaki*, credit Bingua Sun.