

The potential of lactoferrin, ovotransferrin and lysozyme as antiviral and immune-modulating agents in COVID-19.

Running title: Lactoferrin and lysozyme in COVID-19

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Abstract

COVID-19, caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), is spreading rapidly with no established effective treatments. While most cases are mild, others experience uncontrolled inflammatory responses with oxidative stress, dysregulation of iron, and coagulation as features. Lactoferrin, ovotransferrin and lysozyme are abundant, safe antimicrobials that have wide antiviral as well as immunomodulatory properties. In particular, lactoferrin restores iron homeostasis and inhibits replication of SARS-CoV, which is closely related to SARS-CoV-2. Ovotransferrin has antiviral peptides and activities that are shared with lactoferrin. Both lactoferrin and lysozyme are “immune sensing” as they may stimulate immune responses or resolve inflammation. Mechanisms by which these antimicrobials may treat or prevent COVID-19, as well as sources and forms of these, are reviewed.

Lay summary

Severe COVID-19 is characterised by systemic inflammation, where unbound iron plays a significant role in tissue injury. There is no established effective treatment for COVID-19, and some treatment options being explored are unlikely to be widely available soon, especially in resource-limited settings. An abundant and safe antimicrobial that could act via oral ingestion to lower the risk of infection or prevent mild cases from progressing to severe disease would be ideal. Tear lactoferrin and lysozyme levels predict the risk of acquiring upper respiratory tract infections, and these antimicrobials are abundant in nature. Lactoferrin binds free iron and it has been shown to inhibit replication of the novel coronavirus. Ovotransferrin is closely related to lactoferrin and is more abundant (in hen egg white) – it shares iron-binding and antiviral properties of lactoferrin. Lactoferrin, ovotransferrin and lysozyme have several overlapping effects on microbes as well as the immune system, whereby they enhance or limit immune activation in a manner appropriate to the immune environment. Importantly, oral ingestion of these proteins in human and animal studies have shown safety as well as systemic effects, with the ability to limit immune pathology. Research into these products to prevent or treat severe COVID-19 disease is warranted.

Key words:

COVID-19, novel coronavirus, lactoferrin, ovotransferrin, lysozyme, glycerol

Introduction

A cluster of pneumonia cases of unknown etiology was first reported in December 2019 in Wuhan, China [1], and the disease, termed COVID-19, has subsequently spread rapidly throughout the world, posing a great threat to human health and the economy. The virus identified as the causative agent is the novel coronavirus - SARS-CoV-2 [2] – which is highly related to the coronaviruses in bats and, of the human coronaviruses identified to date, is most similar (79.5 % sequence identity) to SARS-CoV [3, 4]. Currently, there are no proven effective treatments or vaccines for novel human coronaviruses [1], though several clinical trials of repurposed existing drugs and new vaccines are underway to test for efficacy in treating or preventing SARS-CoV-2 infection [5, 6]. The aim of this review is to consider the potential of specific antimicrobial proteins that are abundant in nature to act as therapeutics in COVID-19 – as antivirals and/or counteracting the pathology - and to stimulate further research in this avenue.

Pathology of COVID-19: the role of free iron and oxidative stress in tissue injury

Both SARS-CoV and SARS-CoV-2 use human angiotensin-converting enzyme 2 (ACE2) as the receptor for entry and have a similar overall binding mode [7]. SARS-CoV-2 is transmitted predominantly through contact with respiratory droplets from an infected individual [4]. Primary viral replication is thought to occur in the upper respiratory tract followed by replication in the lower respiratory tract and gastrointestinal tract, and cells of the lung, heart, kidney and bladder may be infected. The infected individual may remain asymptomatic (estimated at 17.9 % in one study [8]) or develop symptoms. Those who experience symptoms may have mild disease (80.9 %), progress to severe disease (13.8 %), require critical care (4.7 %) or die (2.3 % in all reported cases) [4]. Those with severe disease experience an uncontrolled inflammatory response (a cytokine storm) that may lead to acute respiratory distress syndrome (ARDS; characterised by widespread inflammation in the lung) as the typical manifestation, or multi-organ failure through immune-induced damage [9], with ARDS occurring 7-14 days after symptom onset [10]. Manifestations are wide-ranging, however, including gastrointestinal [11], renal [12], skeletal muscle [13] and neurological symptoms [14] for example.

Severe COVID-19 is reminiscent of hyperferritinemic syndrome (ferritin levels reaching thousands of units, leukopenia, abnormal liver function tests, severe hypercytokinemia, coagulopathy), under which septic shock and macrophage activating syndrome (MAS) are also classified [15-17]. In

particular, the cytokine profile in COVID-19 most closely resembles MAS, where ferritin directly activates macrophages to release proinflammatory cytokines (including interleukin-6 [IL-6] and tumour necrosis factor [TNF]-alpha) and drives inflammation [16, 18]. MAS is a type of secondary haemophagocytic lymphohistiocytosis (sHLH), an underrecognised syndrome that may be triggered by viral infections and typically has pulmonary involvement (including ARDS) [19] – in this condition macrophages phagocytose red blood cells leading to anaemia [20]. The role of macrophages in the excessive inflammation in COVID-19 is further supported by a recent article [21]. It is well-documented that in conditions of systemic inflammation, oxidative stress mediates cell injury, and this is driven by increased free iron [15, 22, 23]. Further, there is disruption of iron homeostasis and increased free iron in the bronchoalveolar lavage fluid during ARDS and in other lung pathologies [24] [25, 26]. Free iron may directly react with oxygen to form superoxide radicals or with hydrogen peroxide (released by neutrophils and macrophages) to produce a highly toxic hydroxyl free radical [26]. Free radicals can also liberate iron from ferritin, the levels of which increase in response to sequester the reactive free iron as well as part of the acute phase response and due to leakage induced by immune damage [25, 27], leading to further radical formation [27]. Serum ferritin levels are increased in the majority of COVID-19 patients [28], with markedly higher levels of serum ferritin in non-survivors compared to survivors [29]. Another consequence of increased free iron and reactive oxygen species, is the promotion of the development of advanced glycation end products (AGEs) [30], which play a central role in the pathogenesis of ARDS and other pulmonary inflammatory diseases [31, 32]. Increased free iron also has direct effects on fibrinogen, fibrin and erythrocyte morphology and promotes a pro-coagulant state [22]. Intravascular coagulation is a frequent finding in the more severe COVID-19 cases, with D-dimer levels strongly linked to disease severity [33]. D-dimer levels rise prior to interleukin-6 (IL-6), and are not therefore simply secondary to systemic inflammation. Contributing to oxidative stress, and further immune activation, is increased angiotensin II (a product of the inflammation-driven activation of the renin-angiotensin system [RAS] [34]), which is potentiated by the binding of SARS-CoV-2 to the ACE2 receptor and is linearly associated with viral load and lung injury in COVID-19 [35]. In summary, systemic inflammation with associated oxidative stress, dysregulation of iron metabolism, and coagulation are key features of COVID-19.

Lactoferrin, ovotransferrin and lysozyme as potential therapeutics in COVID-19

A vaccine for SARS-CoV-2 is expected to be ready at minimum in 12-18 months from now. An abundantly available antimicrobial which could lower risk of infection or prevent mild disease from

becoming severe disease (where progression to this stage occurs in about 20% of symptomatic individuals and takes approximately 1-2 weeks after mild symptoms are experienced) would have great value. Drugs already approved for other uses are currently being tested in clinical trials, however there is currently little evidence to show that they are having much effect [5]. While remdesivir marginally shortens recovery time in hospitalised patients [6], it is unlikely to be widely available soon, especially in resource-limited settings.

Recently it was reported that tear lactoferrin and lysozyme are relevant biomarkers of mucosal immune competence and that the levels of these predict the risk of acquiring upper respiratory tract infections [36]. Lactoferrin and lysozyme concentration decrease with age [37, 38], potentially increasing risk for respiratory infections. Lactoferrin and lysozyme are among the most abundant antimicrobials found in nature that are widely distributed in animal tissues and secretions [39-41], and are considered among the most promising antimicrobials to become medicines for clinical use [42, 43]. They both act widely against bacteria, viruses, and fungi, as well as having positive stimulatory effects on the immune system yet dampening the pathological effects of an overreacting immune system. Lactoferrin and lysozyme are found in markedly high concentrations in tears compared to any other body fluid, and Lactoferrin is found in similarly high concentration in breastmilk and colostrum – this indicates the important role of these proteins in defence [39, 44, 45]. However, the usual concentrations are only just adequate and lower than normal levels in these secretions increase susceptibility to infection [44]. While SARS-CoV-2 is readily detected in throat swabs, nasal swabs, saliva and sputum, and in a third of patients in faeces [46-48], the virus is only infrequently detected in tears in a similar timeframe [49-51]. When detected in tears, this has been in patients that had conjunctivitis symptoms [49, 51]. A similar scenario is reported for SARS-CoV [52, 53]. The potential activity of lactoferrin and lysozyme against SARS-CoV-2 and against the immune-mediated pathology in COVID-19 (summarised in Figure 1 and Table 1) is considered. Since ovotransferrin is more abundantly available than lactoferrin and can substitute lactoferrin in many applications [54], its potential as a COVID-19 therapeutic is also reviewed.

Lactoferrin as an antiviral and immune modulator

Lactoferrin sequesters free iron, removing a substrate required for bacterial growth, however it also has antimicrobial effects independent of iron sequestration [39]. Lactoferrin is cationic (highly positively charged) and this enables interaction with various negatively charged microbial and viral

surfaces, DNA, as well as with cell surfaces that are required for bacterial and viral adhesion or for early interactions required for viral entry [39]. Lactoferrin may also exert antiviral effects intracellularly [55]. Potent antiviral effects of both human and bovine lactoferrin have been shown against both enveloped and naked viruses, such as cytomegalovirus, herpes simplex virus, and hepatitis B and C virus among others, whether in the metal saturated or apo form [39, 56]. Bovine lactoferrin may have higher antiviral activity than human lactoferrin – they are highly similar and possess identical multifunctions [43]. Therefore bovine lactoferrin is a good equivalent for human lactoferrin, especially since it is recognised by the European Food Safety Authority as a safe dietary supplement with medicinal properties and no contraindications [57]. Importantly, bovine lactoferrin inhibits SARS-CoV cell entry by binding to heparan sulphate proteoglycans (HSPGs) [58]. HSPGs on the cell surface provide an anchoring site on the cell surface and many viruses, including SARS-CoV, employ HSPGs for adhesion to susceptible cells. SARS-CoV-2 entry is highly similar to that of SARS-CoV [7] and was recently shown to be susceptible to lactoferrin-mediated inhibition of entry [59]. Lactoferrin inhibited both entry and post-entry steps of SARS-CoV-2 replication, and elevated interferon-stimulated genes [59].

Besides its direct antimicrobial effect, through sequestering free iron and restoring iron homeostasis, lactoferrin reduces oxidative stress and inflammation, which is pertinent to the COVID-19 pathology. Lactoferrin counteracts iron dysregulation through sequestering free iron and restoring levels of various proteins (ferroportin, ceruloplasmin, transferrin receptor 1 and ferritin) that are altered during inflammation [60, 61]. Lactoferrin reduces intracellular levels of reactive oxygen species as well as reducing oxidative-stress induced apoptosis [62], and short-term oral administration of bovine lactoferrin improves antioxidant capacity [63]. Importantly, lactoferrin can “sense” the immune activation status and respond accordingly [64]. For example, in individuals with high baseline immune activation bovine lactoferrin down-regulates IL-6 and TNF-alpha production by PBMCs (after 7 days of 40 mg per day oral administration) while in those with low immune activation, bovine lactoferrin upregulated these cytokines [64]. Lactoferrin suppresses extracellular traps released by neutrophils during inflammation [65], and has also been shown to stimulate proinflammatory macrophages (M1) to change to the anti-inflammatory type (M2) [43, 56]. Similarly, pasteurised whole cow’s milk has been shown to polarise macrophages from a proinflammatory M1 towards a pro-resolving M2 phenotype [66]. In addition, lactoferrin-derived peptides inhibit angiotensin II proinflammatory activity through binding to the AT1 receptor [67], and lactoferrin as well as other peptides in cow’s milk have an antithrombotic effect [68]. These effects of counteracting iron dysregulation, oxidative stress, neutrophil and macrophage-induced

inflammation, RAS-induced inflammation and thrombosis are highly relevant to COVID-19. Further, lactoferrin shows potential benefit in Alzheimer's disease through decreasing amyloid-beta aggregation (which leads to inflammation and neuron degeneration) [43, 69]. This aggregation may be induced by microbes and this has been suggested for SARS-CoV-2 [70] - a potential neuroprotective role of lactoferrin in COVID-19 is a hypothesis requiring further investigation.

Oral administration of lactoferrin, usually bovine lactoferrin, in human and animal studies of various inflammatory disease states shows safety [43]. In animal studies, oral bovine lactoferrin was shown to decrease inflammation and myeloperoxidase (a marker of neutrophil infiltration) in inflammatory bowel disease [43, 71]. In animal models of sepsis, a single oral dose of lactoferrin prior to insult protected against progression of insult-induced systemic inflammatory responses [62] and when orally administered after sepsis-induced lung injury, bovine lactoferrin was an effective therapeutic [72]. Further showing positive effects of lactoferrin in lung pathology, oral doses of human or mouse lactoferrin reduced *Mycobacterium tuberculosis*-induced lung pathology in a mouse model [73] and aerosolised bovine lactoferrin administered in a mouse model of cystic fibrosis with a *P. aeruginosa* lung infection resulted in decreased bacterial load, decreased infiltrated leukocytes and reduced iron overload [60].

In human studies, lactoferrin decreased late onset sepsis and necrotising enterocolitis in preterm infants [74], and oral bovine lactoferrin (250 mg/day for 3 months) decreased serum IL-6 and increased IL-10 as well as improved antioxidant activity in Alzheimer's disease [43, 75]. In pregnant women suffering from anaemia and/or thrombophilia, 100 mg of bovine lactoferrin taken orally twice a day improved haematological parameters, including serum iron, serum ferritin, haemoglobin and IL-6 levels, more effectively than the standard of care [76]. Clinical effect has also been observed following lactoferrin administration in viral diseases. In hepatitis C infected patients who responded to bovine lactoferrin monotherapy, when bovine lactoferrin was then combined with ribavirin and interferon, there was a sustained virologic response in 55 % of individuals compared to a sustained virologic response in 18 % of individuals who were treated with a combination of ribavirin and interferon alone [77]. Long-term oral consumption of bovine lactoferrin-containing products including yoghurt and milk (in the range of 100-500 mg lactoferrin per day) either reduces incidence or ameliorates symptoms of common viral infections, such as norovirus, likely through direct antiviral activity as well as the enhancement of systemic immunity (increased natural killer cell activity and Th1 cytokine responses) achieved by bovine lactoferrin consumption [78]. Importantly, Serrano *et*

al. (2020), reported that a liposomal bovine lactoferrin nutritional syrup administered at 256-384 mg lactoferrin/day resolved symptoms of COVID-19 patients within 4-5 days and considering their 256 contacts who received half this daily dose, none developed symptoms of the infection [57].

Ovotransferrin as an antiviral and immune modulator

Ovotransferrin shares many of the same activities as human/bovine lactoferrin and is more abundant than the latter [54]. Ovotransferrin combines the iron transport and defence functions of mammalian serum transferrin and lactoferrin, respectively, and shares about 50% sequence homology with each protein [79]. However, the structural analogy between ovotransferrin and lactoferrin is much closer than the sequence homology [80] and similar clusters of positively charged residues responsible for antiviral activity are found in the N-lobes of these proteins [81].

Ovotransferrin not only has antifungal activity [82] and a wide range of antibacterial activity through sequestration of iron and through binding to bacterial surfaces via cationic peptides [54, 83], but the antiviral activity of intact ovotransferrin was greater than that of intact bovine lactoferrin when studying Marek's disease virus [81]. Peptides in ovotransferrin that have high sequence homology with these bovine lactoferrin and human lactoferrin peptides acting against herpes simplex virus, human cytomegalovirus and adenovirus, were shown to have double the antiviral activity compared to the bovine lactoferrin peptides [81]. Recently, it was also reported that ovotransferrin upregulates antiviral interferon I in virus-infected macrophages [84].

Ovotransferrin has immunomodulatory, antioxidant and anti-inflammatory properties, and due to these properties it is being investigated as a therapeutic for cancer and cardiovascular disease [85, 86]. Ovotransferrin, as well as hydrolysates, are able to scavenge free radicals, with higher activity than other known antioxidants such as ascorbate (vitamin C) - ovotransferrin showed protective effects against oxidative stress-induced DNA damage, that was occurring via reaction of iron with hydrogen peroxide, in human leukocytes [87, 88]. Specifically, 16 antioxidant peptides are derived from egg white hydrolysate, where ovotransferrin peptides are in one of the most active fractions [89]. An ovotransferrin peptide attenuates TNF-alpha-induced inflammation and superoxide generation in endothelial cells [90]. Hydrolysates of ovotransferrin, as well as other egg white peptides, have also shown potent ACE inhibitory activity [87, 91] and as is the case for bovine lactoferrin, an ovotransferrin peptide blocked angiotensin II effects via the AT1 receptor [92], thereby

reducing inflammation potentiated by RAS activation. In an animal model study of peritonitis, ingestion of 40 mg/kg feed of an ovotransferrin peptide significantly attenuated the inflammatory responses: serum levels of TNF-alpha, IL-6 and myeloperoxidase activity were significantly reduced [93]. As is the case for bovine lactoferrin, these described activities of ovotransferrin, are highly pertinent to COVID-19 pathology.

Lysozyme as an antiviral and immune modulator

Lysozyme kills gram-positive bacteria through hydrolysing the β -1,4 glycosidic bond between *N*-acetylglucosamine (NAG)-*N*-acetylmuramic acid (NAM) in the bacterial cell wall [40]. However, besides its enzymatic activity, it exerts antimicrobial effects through its cationic nature which enables it to bind to negatively charged surfaces (as in the case of lactoferrin), thereby expanding its activity well beyond that of gram-positive bacteria [40, 41, 94]. The immunomodulatory function of lysozyme has only recently been appreciated [40]. Although lysozyme acting on microbes within neutrophils and macrophages increases their proinflammatory response, when it is released extracellularly by these cells as well as epithelial cells, it limits inflammation: it decreases the oxidative burst and chemotaxis in neutrophils [95], it significantly suppresses TNF-alpha and IL-6 production by macrophages [96], it binds and decreases circulating levels of AGEs (which are pro-oxidative) as well as enhancing their renal excretion [30], and exogenous lysozyme disrupts the ability of peptidoglycan to bind complement factors that act as anaphylotoxins [40]. Furthermore, when subjected to simulated gastrointestinal digestion, the hydrolysate of hen egg white lysozyme (HEWL) showed marked antioxidant and ACE-inhibitory activity [97]. As described earlier, the oxidative stress (including involvement of AGEs), inflammation induced by neutrophils and macrophages, the TNF-alpha and IL-6 cytokines, and an activated RAS system are features in ARDS and/or severe COVID-19. It is noteworthy that, as is the case for lactoferrin, lysozyme has a neuroprotective function in Alzheimer's disease through preventing amyloid-beta aggregation [98], which may have implications for the neurological manifestations in severe COVID-19.

From mouse and porcine models it is clear that lysozyme plays an important role in limiting inflammation systemically, resulting in decreased immune-driven pathology [40, 99, 100]. Human clinical trials with lysozyme are limited [41, 101], but have shown, mostly through oral administration of HEWL, antiviral effects against herpes (through oral administration of HEWL at 1 g/day [41, 102]), measles [101, 103] and hepatitis (60-170 mg/day of lysozyme chloride for 4-24 weeks significantly

reduced post-transfusion hepatitis incidence to 8% compared to 20% [104]), successful treatment of gum infections (750 mg/day [41, 105]) and skin ulcers [101, 106], improvement of immune responses in cancer patients with immune suppression [41], and rapid resolution of inflammatory foci and stabilisation of lysozyme levels in serum and stool of premature infants with diseases following 50 mg/L supplementation in milk for 2-3 weeks [101, 107]. Human lysozyme in combination with bovine lactoferrin (0.2 g lysozyme and 1.5 g bovine lactoferrin per day) reduced enteric dysfunction in Malawian children [108]. No local or systemic unfavourable effects have been reported in these human trials.

Regarding the investigation of lysozyme in lung diseases, in Eastern Europe, HEWL has been used successfully in combination with antibiotics to treat bronchitis and pneumonia in humans with no respiratory or systemic toxicity [109, 110]. Administration of lysozyme through aerosols to treat pneumonia has been investigated in animal models [111, 112]. A 1% solution of aerosolised human lysozyme in hamsters with *P. aeruginosa*-induced pneumonia resulted in decreased lung histopathological changes, alveolar septal apoptosis, neutrophils and other leukocytes in the bronchiolar lavage fluid as well as increased activity of lysozyme in that fluid [111]. However, it should be noted that lysozyme impairs the ability of hyaluronan to prevent elastase injury to elastic fibres through binding of the lysozyme to the elastic fibres, and thus on inhalation of lysozyme in an animal model of emphysema, airspaces further increased [113], which cautions against the inhalation route of administration of lysozyme in similar disease states.

Sources, forms and practical use of lactoferrin, ovotransferrin and lysozyme

Cow's milk is the most readily available source of lactoferrin, with an average concentration of 0.174 g/L in low heat pasteurised cow's milk (and 1.2 g/kg in semi-hard cheese produced from that milk) [114], which is in good agreement with other studies [115-117], though the range experienced (0.03-0.486 g/L) is dependent on several factors [118]. The concentration in colostrum is higher, but varies greatly between breeds and may be anything between 0.3 g/L and 5 g/L, and is typically at the lower end of the range [119-121]. The degradation of bovine lactoferrin in milk with low heat pasteurisation (72 °C for 5 seconds) is minimal [114, 122], while ultra-high temperature (UHT) processes significantly denature the protein [123]. Large scale isolation of bovine lactoferrin is performed from cheese whey [124] where only 19% of the total bovine lactoferrin in milk is found [114], and the cost of purified bovine lactoferrin remains high – hence methods to achieve large scale production of lactoferrin are

being developed [125, 126]. Bovine lactoferrin is sold in bulk powder form, capsules (typically 250-300 mg), liposomal syrups (32 mg/10 ml), or as a liposomal lactoferrin nebulizer [57, 108, 127]. The majority of bovine lactoferrin taken orally can be considered to survive gastric transit (62 % for the apo form and 79 % for the more stable iron-bound form [128]) and thereafter enter the intestine from where it is absorbed into the circulation, but liposomalisation or encapsulation has been shown to enhance availability and effect [129, 130]. It is also important to note that digestion with enzymes in the gastrointestinal tract (pepsin, trypsin or chymotrypsin) yields lactoferrin fragments that are still able to bind iron [131], and that fragments of lactoferrin have antimicrobial activity [39] which may be stronger than that of the intact protein [132, 133]. Peptides of lactoferrin are considered promising antivirals, but isolation costs and stability pose challenges to reach the clinical phase [42], thus, at present the whole intact protein or food products/supplements with high content of lactoferrin are more accessible. A hindrance for use in medicine is the classification of lactoferrin products (as well as egg white powder and lysozyme discussed below) as food supplements, where these are not intended to treat disease, there is no controlled system for reporting effectiveness, and the active ingredient is not always of the same quality or integrity [127].

Ovotransferrin is abundant in hen egg white (12 g/L egg white) [134]. Methods to pasteurise egg whites use temperatures that minimise damage to heat sensitive proteins in the egg white, such as lysozyme and ovotransferrin [135-137]. Dried egg white powder, where 250 egg whites are equivalent to 1 kg powder, sold as a supplement is a compressed source of these proteins. Although 0.5-2.5 % of children have an allergy to egg white, about 70 % outgrow the allergy - nevertheless, many medicines and vaccines have ingredients derived from egg [138]. Iron-bound ovotransferrin is more resistant to gastrointestinal digestion [139, 140], with iron-bound ovotransferrin well-absorbed after ingestion [141]. Ovotransferrin is more readily digested by pepsin in the stomach compared to lysozyme [142], however the bioactive peptides (antimicrobial, antioxidant, anti-inflammatory, ACE-inhibitory) of ovotransferrin, as well as other egg white proteins, described earlier have mostly been produced by digestion that simulates that occurring in the gastrointestinal tract [143], and these peptides resist further digestion [144] and are readily transported into human intestinal cells [145]. Ovotransferrin does however lose iron-binding activity after hydrolysis [134]. Simpler protocols with better yield and purity as well as low cost, will enable the use of isolated egg white proteins such as ovotransferrin or their peptides as pharmaceuticals [87], while presently the most readily available source is egg white powder.

Egg white is also the most readily available source of lysozyme (3.8 g/L egg white) [134]. Isolated HEWL is commercially available and is labelled as a food supplement by the European Commission. It is however available as lysozyme hydrochloride tablets (10 mg, 30 mg or 90 mg), granules (10% or 20%) and syrup (0.5%) in Japan, and is prescribed by doctors to improve expectoration in bronchitis, bronchial asthma and bronchiectasis [146]. It is also sold as a food or pharmaceutical grade powder, and is widely used in the food industry as well as in pharmaceutical products (*e.g.* eye drops, wound healing creams, oral health products and over-the-counter drugs). Wider spread use is hindered somewhat by the isolation cost (\$2.05/g) and there is significant effort being made around the world for commercial production of lysozyme, especially human lysozyme [147, 148]. Methods to produce human lysozyme, which has higher enzymatic activity than HEWL and is therefore preferred, are under development (*e.g.* using transgenic rice), although these have a higher production cost than for HEWL [147]. However, the antiviral and immunomodulatory effects do not derive from the enzymatic activity, and the most available form of lysozyme currently is HEWL, whether isolated or in egg white. Oral administration of HEWL results in systemic effects - after oral administration of 90-900 mg HEWL in human subjects peak plasma concentrations are reached within an hour (with overnight fasting increasing absorption by 7-fold) and return to undetectable levels after 2 days [146, 149]. While HEWL is fairly resistant to digestion in the stomach and partially resistant to digestion in the duodenum [142, 150], enzymatic hydrolysis does produce antimicrobial fragments and broadens the antimicrobial spectrum [151].

Susceptibility of proteins to proteolytic digestion is very strongly related to protein stability [152], and polyols or their derivatives are commonly used to enhance protein stability in formulations [153]. An easily accessible and safe polyol may therefore be considered to improve stability of lactoferrin, ovotransferrin and HEWL following ingestion, and here it is suggested that glycerol may be a particularly suitable supportive solvent for the powdered sources of lactoferrin, ovotransferrin and HEWL. Glycerol is a low cost, readily available, sweet-tasting polyol, with excellent solvent and emulsifying properties, which is safe for ingestion and widely used in pharmaceutical applications (such as cough syrups) [154-156]. It is known to effectively stabilise proteins as well as refold denatured proteins [157, 158], thereby restoring activities of enzymes that were inactivated by diverse processes [157]. In particular, glycerol was already shown to protect ovotransferrin and lysozyme when these proteins were subjected to stresses [137, 153], and to partially restore the structure/activities of these proteins after denaturation [137, 159]. Other properties may add further benefit, including anti-inflammatory [160] and antiviral effects [161, 162], as well as the ability to inhibit ACE activity and decrease angiotensin II [163].

Future perspective

In view of (i) the direct antiviral effects of lactoferrin, ovotransferrin and lysozyme against a wide range of viruses (including SARS-CoV for lactoferrin) and their antimicrobial effects against a wide range of bacteria and fungi that may cause secondary infections in COVID-19 patients [164], (ii) their immunomodulatory properties which stimulate antimicrobial responses yet promote resolution of inflammation, and in particular their previously shown beneficial effects in counteracting pathological neutrophil infiltration, macrophage activation, free iron overload, oxidative stress, AGE effects, excessive proinflammatory cytokine production (IL-6 and TNF-alpha in particular), and thrombus formation, which all feature in severe COVID-19, and (iii) their abundance and good safety profile; further testing of their potential role in prevention of SARS-CoV-2 infection or prevention of severe COVID-19 is suggested. The main suggestion is to use these antimicrobials upon presentation of symptoms to prevent non-critical cases from progressing to critical cases, although they may also be considered as a preventative for those at high risk of infection where lower quantities could be taken as a means of lowering risk of infection. Since the number of SARS-CoV-2 infection cases is growing so rapidly, the most expedient way to achieve this is through oral administration, which is suitable in the case of lactoferrin, ovotransferrin and lysozyme as these substances have systemic effects following ingestion. It is further suggested that, in the current circumstances of the COVID-19 pandemic, good quality non-isolated forms of these (such as egg white powder, bovine colostrum powder and other non-UHT milk products with appreciable lactoferrin content) should also be tested while ensuring the desired concentrations of each antimicrobial are met, especially in settings where the isolated forms may not be as readily accessible. In favour of this suggestion, studies using lactoferrin-containing milk or lactoferrin-supplemented yoghurt have shown clinical efficacy in viral diseases [78, 165], pasteurised whole milk has shown the effect of switching macrophages from M1 to M2 [66], several peptides in milk are antithrombotic [68], and several peptides in egg white besides those in ovotransferrin show supportive antioxidant as well as ACE-inhibitory effects [87, 89]. However, in those individuals who are already critically ill and on ventilators, more care may need to be taken with the approach. Here, perhaps lactoferrin and lysozyme could be considered for intravenous administration or nebulisation – a liposomal bovine lactoferrin nebulizer product is available. The accessibility and reasonable cost (in comparison to some of the other drugs - such as remdesivir and tocilizumab – under investigation to treat COVID-19) make these antimicrobials attractive as a therapeutic option and we therefore call for their rapid testing in clinical trials.

Executive summary

Pathology of COVID-19: the role of free iron and oxidative stress in tissue injury

- Severe COVID-19 is reminiscent of hyperferritinemic syndrome.
- Ferritin contributes to the inflammation by directly activating macrophages, and free iron may be liberated from ferritin by free radicals.
- Free iron reacts with oxygen or hydrogen peroxide to form free radicals, thereby driving oxidative stress and leading to tissue injury.

Lactoferrin, ovotransferrin and lysozyme as potential therapeutics in COVID-19

- There is no established effective treatment for COVID-19, and some treatment options being explored are unlikely to be widely available soon, especially in resource-limited settings.
- An abundant and safe antimicrobial that could act via oral ingestion to lower the risk of infection or prevent mild cases from progressing to severe disease would be ideal.
- Tear lactoferrin and lysozyme levels predict the risk of acquiring upper respiratory tract infections, and these antimicrobials are abundant in natural secretions.
- Ovotransferrin is more abundant than lactoferrin and can substitute lactoferrin in many applications.

Lactoferrin as an antiviral and immune modulator

- Bovine lactoferrin has been shown to inhibit SARS-CoV cell entry, which is similar to that of SARS-CoV-2.
- Lactoferrin restores iron homeostasis through sequestering free iron and modulating levels of proteins involved in controlling iron balance between blood and tissues.
- Lactoferrin reduces oxidative stress and inflammation, and it is immune “sensing” with its effect dependent on the environment.
- Oral administration of lactoferrin in animal models and human studies of viral diseases, as well as various inflammatory disease states, shows beneficial effects and safety.

Ovotransferrin as an antiviral and immune modulator

- Ovotransferrin has antiviral peptides that are conserved with those found in human and bovine lactoferrin, and ovotransferrin may have a more potent antiviral effect.
- Ovotransferrin has immunomodulatory, antioxidant, anti-inflammatory, and ACE-inhibitory activities.

Lysozyme as an antiviral and immune modulator

- Lysozyme exhibits antiviral activity via its cationic peptides and has immunomodulatory, antioxidant and ACE-inhibitory properties.
- Oral administration of lysozyme in animal models and human studies shows its ability to limit inflammation systemically, resulting in decreased immune-driven pathology.

Sources, forms and practical use of lactoferrin, ovotransferrin and lysozyme

- Lactoferrin is abundant in cow's milk, while ovotransferrin and lysozyme are abundant in hen egg white.
- High costs of isolation have limited wider-spread use of purified forms of these antimicrobials.
- Isolation costs and stability pose challenges for bioactive peptides of these antimicrobials to reach the clinical phase.

Future perspective

- These antimicrobials could be used upon presentation of symptoms to prevent non-critical cases from progressing to critical cases, and lower quantities could be taken to lower risk of infection in those at high risk.
- Good quality non-isolated forms of these should also be tested while ensuring the desired concentrations of each antimicrobial are met, especially in settings where the isolated forms may not be as readily accessible.
- The accessibility and reasonable cost make these antimicrobials attractive as a therapeutic option and we therefore call for their rapid testing in clinical trials.

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References

1. Li X, Zai J, Wang X, Li Y. Potential of large "first generation" human-to-human transmission of 2019-nCoV. *J. Med. Virol.* 92(4), 448-454 (2020).
2. Zhu N, Zhang D, Wang W *et al.* A novel coronavirus from patients with pneumonia in China, 2019. *N. Engl. J. Med.* 382(8), 727-733 (2020).
3. Lu R, Zhao X, Li J *et al.* Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. *Lancet* 395(10224), 565-574 (2020).
4. Jin Y, Yang H, Ji W *et al.* Virology, Epidemiology, Pathogenesis, and Control of COVID-19. *Viruses* 12(4), 372 (2020).

5. Tu YF, Chien CS, Yarmishyn AA *et al.* A Review of SARS-CoV-2 and the Ongoing Clinical Trials. *Int. J. Mol. Sci.* 21(7), 2657 (2020).
6. Beigel JH, Tomashek KM, Dodd LE *et al.* Remdesivir for the Treatment of Covid-19 - Preliminary Report. *N. Engl. J. Med.* doi: 10.1056/NEJMoa2007764 (2020). [Epub ahead of print]
7. Wang Q, Zhang Y, Wu L *et al.* Structural and Functional Basis of SARS-CoV-2 Entry by Using Human ACE2. *Cell* 181(4), 894-904 (2020).
8. Mizumoto K, Kagaya K, Zarebski A, Chowell G. Estimating the asymptomatic proportion of coronavirus disease 2019 (COVID-19) cases on board the Diamond Princess cruise ship, Yokohama, Japan, 2020. *Euro. Surveill.* 25(10), 2000180 (2020).
9. Cao X. COVID-19: immunopathology and its implications for therapy. *Nat. Rev. Immunol.* 20(5), 269-270 (2020).
10. Lin L, Lu L, Cao W, Li T. Hypothesis for potential pathogenesis of SARSCoV-2 infection—a review of immune changes in patients with viral pneumonia. *Emerg. Microbes Infect.* 9(1), 727-732 (2020).
11. Yang L, Tu L. Implications of gastrointestinal manifestations of COVID-19. *The Lancet* 5(7), 629-630 (2020).
12. Kunutsor SK, Laukkanen JA. Renal complications in COVID-19: a systematic review and meta-analysis. *Ann. Med.* 52(7), 345-353 (2020).
13. Samies NL, Pinninti S, James SH. Rhabdomyolysis and Acute Renal Failure in an Adolescent With Coronavirus Disease 2019. *J. Pediatric Infect. Dis. Soc.* doi.org/10.1093/jpids/piaa1083. (2020). [Epub ahead of print]
14. Mao L, Jin H, Wang M *et al.* Neurologic Manifestations of Hospitalized Patients With Coronavirus Disease 2019 in Wuhan, China. *JAMA Neurol.* 77, 683-690 (2020).
15. Colafrancesco S, Alessandri C, Conti F, Priori R. COVID-19 Gone Bad: A New Character in the Spectrum of the Hyperferritinemic Syndrome? *Autoimmun. Rev.* 19(7), 102573 (2020).
- * The similarities of COVID-19 with hyperferritinemic syndrome, and the underlying pathology, is discussed.
16. Shoenfeld Y. Corona (COVID-19) time musings: Our involvement in COVID-19 pathogenesis, diagnosis, treatment and vaccine planning. *Autoimmun. rev.* 19(6), 102538 (2020).
17. Rosário C, Zandman-Goddard G, Meyron-Holtz EG, D'cruz DP, Shoenfeld Y. The Hyperferritinemic Syndrome: macrophage activation syndrome, Still's disease, septic shock and catastrophic antiphospholipid syndrome. *BMC Med.* 11, 185 (2013).
18. Mcgonagle D, Sharif K, O'regan A, Bridgewood C. The Role of Cytokines including Interleukin-6 in COVID-19 induced Pneumonia and Macrophage Activation Syndrome-Like Disease. *Autoimmun. Rev.* 19(6), 102537 (2020).
19. Mehta P, McAuley DF, Brown M *et al.* COVID-19: consider cytokine storm syndromes and immunosuppression. *The Lancet* 395 (10229), 1033-1034 (2020).
20. Zoller EE, Lykens JE, Terrell CE *et al.* Hemophagocytosis causes a consumptive anemia of inflammation. *J. Exp. Med.* 208(6), 1203–1214 (2011).
21. Park MD. Macrophages: a Trojan horse in COVID-19? *Nat. Rev. Immunol.* 20(6), 351 (2020).
22. Pretorius E, Kell DB. Diagnostic Morphology: Biophysical Indicators for Iron-Driven Inflammatory Diseases. *Integr. Biol. (Camb)* 6(5), 486-510 (2014).
23. Jomova K, Valko M. Importance of Iron Chelation in Free Radical-Induced Oxidative Stress and Human Disease. *Curr. Pharm. Des.* 17(31), 3460-3473 (2011).
24. Ghio AJ, Carter JD, Richards JH, Richer LD, Grissom CK, Elstad MR. Iron and iron-related proteins in the lower respiratory tract of patients with acute respiratory distress syndrome. *Crit. Care Med.* 31(2), 395-400 (2003).
25. Kim J, Wessling-Resnick M. The Role of Iron Metabolism in Lung Inflammation and Injury. *J. Allergy Ther.* 3(Suppl 4), 004 (2012).

26. Khiroya H, Turner AM. The role of iron in pulmonary pathology. *Multidiscip. Respir. Med.* 10, 34 (2015).
27. Connelly KG, Moss M, Parsons PE *et al.* Serum ferritin as a predictor of the acute respiratory distress syndrome. *Am. J. Respir. Crit. Care Med.* 155(1), 21-25 (1997).
28. Chen N, Zhou M, Dong X *et al.* Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet* 395(10223), 507-513 (2020).
29. Zhou F, Yu T, Du R *et al.* Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet* 395(10229), 1054-1062 (2020).
30. Aldini G, Vistoli G, Stefek M *et al.* Molecular strategies to prevent, inhibit, and degrade advanced glycoxidation and advanced lipoxidation end products. *Free Radic. Res.* 47(Suppl 1), 93-137 (2013).
31. Oczypok EA, Perkins TN, Oury TD. All the "RAGE" in lung disease: The receptor for advanced glycation endproducts (RAGE) is a major mediator of pulmonary inflammatory responses. *Paediatr. Respir. Rev.* 23, 40-49 (2017).
32. Wang H, Wang T, Yuan Z *et al.* Role of Receptor for Advanced Glycation End Products in Regulating Lung Fluid Balance in Lipopolysaccharide-induced Acute Lung Injury and Infection-Related Acute Respiratory Distress Syndrome. *Shock* 50(4), 472-482 (2018).
33. Oudkerk M, Büller HR, Kuijpers D *et al.* Diagnosis, Prevention, and Treatment of Thromboembolic Complications in COVID-19: Report of the National Institute for Public Health of the Netherlands. *Radiology* 201629. doi: 201610.201148/radiol.2020201629 (2020). [Epub ahead of print]
34. Satou R, Penrose H, Navar LG. Inflammation as a Regulator of the Renin-Angiotensin System and Blood Pressure. *Curr. Hypertens. Rep.* 20(12), 100 (2018).
35. Liu Y, Yang Y, Zhang C *et al.* Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury. *Sci. China Life Sci.* 63(3), 364-374 (2020).
36. Hanstock HG, Edwards JP, Walsh NP. Tear Lactoferrin and Lysozyme as Clinically Relevant Biomarkers of Mucosal Immune Competence. *Front. Immunol.* 10, 1178 (2019).
37. Kawashima M, Kawakita T, Inaba T *et al.* Dietary lactoferrin alleviates age-related lacrimal gland dysfunction in mice. *PLoS One* 7(3), e33148 (2012).
38. Mcgill JI, Liakos GM, Goulding N, Seal DV. Normal tear protein profiles and age-related changes. *Br. J. Ophthalmol.* 68, 316-320 (1984).
39. Berlutti F, Pantanella F, Natalizi T *et al.* Antiviral Properties of Lactoferrin—A Natural Immunity Molecule. *Molecules* 16, 6992-7018 (2011).
- ** The mechanisms of the antiviral activity of lactoferrin are explained, and its effect against a range of different viruses is reviewed.
40. Ragland SA, Criss AK. From bacterial killing to immune modulation: Recent insights into the functions of lysozyme. *PLoS Pathog.* 13(9), e1006512 (2017).
- ** The immunomodulatory actions of lysozyme are described in detail.
41. Sava G. Pharmacological aspects and therapeutic applications of lysozymes. In: *Lysozymes: Model Enzymes in Biochemistry and Biology.*, Jolles P (Ed.^(Eds). Birkhauser Verlag Germany (1996).
42. Vilas Boas LCP, Campos ML, Berlanda RLA, De Carvalho Neves N, Franco OL. Antiviral peptides as promising therapeutic drugs. *Cell Mol. Life Sci.* 76(18), 3525-3542 (2019).
43. Lepanto MS, Rosa L, Paesano R, Valenti P, Cutone A. Lactoferrin in Aseptic and Septic Inflammation. *Molecules* 24(7), 1323 (2019).
- ** The ability of lactoferrin to act as an immune "sensing" agent is described, and the evidence for reduction of immune pathology in a wide range of disease models following lactoferrin administration is reviewed.

44. Ridley F. Lysozyme: An Antibacterial Body present in Great Concentration in Tears, and its Relation to Infection of the Human Eye. *Proc. R. Soc. Med.* 21(9), 1495-1506 (1928).
45. Hankiewicz J, Swierczek E. Lysozyme in human body fluids. *Clin. Chim. Acta* 57(3), 205-209 (1974).
46. To KK, Tsang OT, Chik-Yan Yip C *et al.* Consistent detection of 2019 novel coronavirus in saliva. *Clin. Infect. Dis.* 71(15), 841-843 (2020).
47. Lin C, Xiang J, Yan M, Li H, Huang S, Shen C. Comparison of throat swabs and sputum specimens for viral nucleic acid detection in 52 cases of novel coronavirus (SARS-Cov-2) infected pneumonia (COVID-19). *Clin. Chem. Lab. Med.* 58(7), 1089-1094 (2020).
48. Wang W, Xu Y, Gao R *et al.* Detection of SARS-CoV-2 in Different Types of Clinical Specimens. *JAMA.* 323(18), 1843-1844 (2020).
49. Wu P, Duan F, Luo C *et al.* Characteristics of Ocular Findings of Patients With Coronavirus Disease 2019 (COVID-19) in Hubei Province, China. *JAMA Ophthalmol.* 138(5), 575-578 (2020).
50. Seah IYJ, Anderson DE, Kang AEZ *et al.* Assessing Viral Shedding and Infectivity of Tears in Coronavirus Disease 2019 (COVID-19) Patients. *Ophthalmology* 127(7), 977-979 (2020).
51. Xia J, Tong J, Liu M, Shen Y, Guo D. Evaluation of coronavirus in tears and conjunctival secretions of patients with SARS-CoV-2 infection. *J. Med. Virol.* 92(6), 589-594 (2020).
52. Chan WM, Yuen KS, Fan DS, Lam DS, Chan PK, Sung JJ. Tears and conjunctival scrapings for coronavirus in patients with SARS. *Br. J. Ophthalmol.* 88(7), 968-969 (2004).
53. Loon SC, Teoh SC, Oon LL *et al.* The severe acute respiratory syndrome coronavirus in tears. *Br. J. Ophthalmol.* 88(7), 861-863 (2004).
54. Chiurciu C, Chiurciu V, Oporanu M *et al.* PC2 Ovotransferrin: Characterization and Alternative Immunotherapeutic Activity. *Evid. Based Complement. Alternat. Med.* 2017, 8671271 (2017).
55. Picard-Jean F, Bouchard S, Larivée G, Bisailon M. The intracellular inhibition of HCV replication represents a novel mechanism of action by the innate immune Lactoferrin protein. *Antiviral Res.* 111, 13-22 (2014).
56. Wisgrill L, Wessely I, Spittler A, Förster-Waldl E, Berger A, Sadeghi K. Human lactoferrin attenuates the proinflammatory response of neonatal monocyte-derived macrophages. *Clin. Exp. Immunol.* 192(3), 315–324 (2018).
57. Serrano G, Kochergina I, Albors A *et al.* Liposomal Lactoferrin as Potential Preventative and Cure for COVID-19. *Int. J. Res. Health Sci.* 8(1), 8-15 (2020).
58. Lang J, Yang N, Deng J *et al.* Inhibition of SARS pseudovirus cell entry by lactoferrin binding to heparan sulfate proteoglycans. *PLoS One* 6(8), e23710 (2011).
- ** Bovine lactoferrin is shown to inhibit SARS-CoV cell entry by binding to heparan sulphate proteoglycans (HSPGs), which the virus uses as an anchoring site on the cell surface, at an IC50 of 0.7 uM.
59. Mirabelli C, Wotring JW, Zhang CJ *et al.* Morphological Cell Profiling of SARS-CoV-2 Infection Identifies Drug Repurposing Candidates for COVID-19. *bioRxiv* <https://doi.org/10.1101/2020.1105.1127.117184> (2020). [preprint]
- ** Bovine lactoferrin is shown to inhibit SARS-CoV-2 replication *in vitro*, inhibiting both entry and post-entry steps in replication.
60. Cutone A, Lepanto MS, Rosa L *et al.* Aerosolized Bovine Lactoferrin Counteracts Infection, Inflammation and Iron Dysbalance in A Cystic Fibrosis Mouse Model of Pseudomonas aeruginosa Chronic Lung Infection. *Int. J. Mol. Sci.* 20(9), E2128 (2019).
61. Bonaccorsi Di Patti MC, Cutone A, Polticelli F *et al.* The ferroportin-ceruloplasmin system and the mammalian iron homeostasis machine: regulatory pathways and the role of lactoferrin. *Biomaterials* 31(3), 399-414 (2018).
62. Actor JK, Hwang S, Kruzel ML. Lactoferrin as a Natural Immune Modulator. *Curr. Pharm. Des.* 15(17), 1956–1973 (2009).

63. Mulder AM, Connellan PA, Oliver C, Morris CA, Stevenson LM. Bovine lactoferrin supplementation supports immune and antioxidant status in healthy human males. *Nutr. Res.* 28(9), 583-589 (2008).
64. Kruzel ML, Zimecki M, Actor JK. Lactoferrin in a Context of Inflammation-Induced Pathology. *Front. Immunol.* 8, 1438 (2017).
65. Okubo K, Kamiya M, Urano Y *et al.* Lactoferrin Suppresses Neutrophil Extracellular Traps Release in Inflammation. *EBioMedicine* 10, 204-215 (2016).
66. Panahipour L, Kochergina E, Kreissl A, Haiden N, Gruber R. Milk modulates macrophage polarization in vitro. *Cytokine* 1(2), 100009 (2019).
67. Fernández-Musoles R, Salom JB, Martínez-Maqueda D, López-Díez JJ, Recio I, Manzanares P. Antihypertensive effects of lactoferrin hydrolyzates: Inhibition of angiotensin- and endothelin-converting enzymes. *Food Chem.* 139(1-4), 994-1000 (2013).
68. Rutherford KJ, Gill HS. Peptides affecting coagulation. *Br. J. Nutr.* 84(Suppl 1), S99-102 (2000).
69. Guo C, Yang ZH, Zhang S *et al.* Intranasal Lactoferrin Enhances alpha-Secretase-Dependent Amyloid Precursor Protein Processing via the ERK1/2-CREB and HIF-1 Pathways in an Alzheimer's Disease Mouse Model. *Neuropsychopharmacology* 42(13), 2504-2515 (2017).
70. Naughton SX, Raval U, Pasinetti GM. Potential Novel Role of COVID-19 in Alzheimer's Disease and Preventative Mitigation Strategies. *J. Alzheimers Dis.* 76, 21-25 (2020).
71. Togawa J, Nagase H, Tanaka K *et al.* Oral administration of lactoferrin reduces colitis in rats via modulation of the immune system and correction of cytokine imbalance. *J. Gastroenterol. Hepatol.* 17, 1291-1298 (2002).
72. Han H, Li H, Li G, Shen Y, Fei M, Nan Y. Effect of bovine lactoferrin as a novel therapeutic agent in a rat model of sepsis-induced acute lung injury. *AMB Expr.* 9, 177 (2019).
73. Hwang SA, Kruzel ML, Actor JK. Oral recombinant human or mouse lactoferrin reduces Mycobacterium tuberculosis TDM induced granulomatous lung pathology. *Biochem. Cell Biol.* 95(1), 148-154 (2017).
74. Pammi M, Abrams SA. Oral lactoferrin for the prevention of sepsis and necrotizing enterocolitis in preterm infants. *Cochrane Database Syst Rev* (2), CD007137 (2015).
75. Mohamed WA, Salama RM, Schaalán MF. A pilot study on the effect of lactoferrin on Alzheimer's disease pathological sequelae: Impact of the p-Akt/PTEN pathway. *Biomed. Pharmacother.* 111, 714-723 (2019).
76. Rosa L, Cutone A, Lepanto MS, Paesano R, Valenti P. Lactoferrin: A Natural Glycoprotein Involved in Iron and Inflammatory Homeostasis. *Int. J. Mol. Sci.* 18(9), 1985 (2017).
77. Kaito M, Iwasa M, Fujita N *et al.* Effect of lactoferrin in patients with chronic hepatitis C: combination therapy with interferon and ribavirin. *J. Gastroenterol. Hepatol.* 22(11), 1894-1897 (2007).
78. Wakabayashi H, Oda H, Yamauchi K, Abe F. Lactoferrin for prevention of common viral infections. *J. Infect. Chemother.* 20(11), 666-671 (2014).
79. Giansanti F, Rossi P, Massucci MT *et al.* Antiviral activity of ovotransferrin discloses an evolutionary strategy for the defensive activities of lactoferrin. *Biochem. Cell Biol.* 80(1), 125-130 (2002).
80. Kurokawa H, Dewan JC, Mikami B, Sacchettini JC, Hirose M. Crystal structure of hen apo-ovotransferrin. Both lobes adopt an open conformation upon loss of iron. *J. Biol. Chem.* 274(40), 28445-28452 (1999).
81. Giansanti F, Massucci MT, Giardi MF *et al.* Antiviral activity of ovotransferrin derived peptides. *Biochem. Biophys. Res. Commun.* 331(1), 69-73 (2005).

* Ovotransferrin peptides that are conserved with peptides from bovine lactoferrin (which are also active against herpes simplex virus) were shown to be more potent than bovine lactoferrin peptides in their effect against Marek's disease virus.

82. Valenti P, Visca P, Antonini G, Orsi N. Antifungal activity of ovotransferrin towards genus *Candida*. *Mycopathologia* 89(3), 169-175 (1985).
83. Ibrahim HR, Sugimoto Y, Aoki T. Ovotransferrin antimicrobial peptide (OTAP-92) kills bacteria through a membrane damage mechanism. *Biochim. Biophys. Acta* 1523(2-3), 196-205 (2000).
84. Zhou Y, Tang Q, Du H *et al.* Antiviral effect of ovotransferrin in mouse peritoneal macrophages by up-regulating type I interferon expression. *Food Agric. Immunol.* 29(1), 600-614 (2018).
85. Chen S, Jiang H, Peng H, Wu X, Fang J. The Utility of Ovotransferrin and Ovotransferrin-Derived Peptides as Possible Candidates in the Clinical Treatment of Cardiovascular Disease. *Oxid. Med. Cell Longev.* 2017, 6504518 (2017).
86. Réhault-Godbert S, Guyot N, Nys Y. The Golden Egg: Nutritional Value, Bioactivities, and Emerging Benefits for Human Health. *Nutrients* 11(3), E684 (2019).
- ** The potential of egg white proteins, including lysozyme and ovotransferrin, and their bioactive peptides to be used in preventing/curing diseases is reviewed.
87. Abeyrathne EDNS, Huang X, Ahn DU. Antioxidant, angiotensin-converting enzyme inhibitory activity and other functional properties of egg white proteins and their derived peptides – A review. *Poult. Sci.* 97, 1462–1468 (2018).
88. Kim J, Moon SH, Ahn DU, Paik HD, Park E. Antioxidant effects of ovotransferrin and its hydrolysates. *Poult. Sci.* 91(11), 2747-2754 (2012).
89. Nimalaratne C, Bandara N, Wu J. Purification and characterization of antioxidant peptides from enzymatically hydrolyzed chicken egg white. *Food Chem.* 88, 467-472 (2015).
90. Majumder K, Chakrabarti S, Davidge ST, Wu J. Structure and activity study of egg protein ovotransferrin derived peptides (IRW and IQW) on endothelial inflammatory response and oxidative stress. *J. Agric. Food Chem.* 61(9), 2120-2129 (2013).
91. Moon SH, Lee JH, Kim JH, Paik HD, Ahn DU. In vitro cytotoxic and ACE-inhibitory activities of promod 278P hydrolysate of ovotransferrin from chicken egg white. *Poult. Sci.* 96(6), 1982-1987 (2017).
92. Liao W, Fan H, Wu J. Egg White-Derived Antihypertensive Peptide IRW (Ile-Arg-Trp) Inhibits Angiotensin II-Stimulated Migration of Vascular Smooth Muscle Cells via Angiotensin Type I Receptor. *J. Agric. Food Chem.* 66(20), 5133-5138 (2018).
93. Jiao H, Zhang Q, Lin Y, Gao Y, Zhang P. The Ovotransferrin-Derived Peptide IRW Attenuates Lipopolysaccharide-Induced Inflammatory Responses. *Biomed. Res. Int.* 2019, 8676410 (2019).
94. Woods CM, Hooper DN, Ooi EH, Tan LW, Carney AS. Human lysozyme has fungicidal activity against nasal fungi. *Am. J. Rhinol. Allergy* 25(4), 236-240 (2011).
95. Gordon LI, Douglas SD, Kay NE, Yamada O, Osserman EF, Jacob HS. Modulation of neutrophil function by lysozyme. Potential negative feedback system of inflammation. *J. Clin. Invest.* 64(1), 226-232 (1979).
96. Tagashira A, Nishi K, Matsumoto S, Sugahara T. Anti-inflammatory effect of lysozyme from hen egg white on mouse peritoneal macrophages. *Cytotechnology* 70(3), 929-938 (2018).
97. Rao S, Sun J, Liu Y, Zeng H, Su Y, Yang Y. ACE inhibitory peptides and antioxidant peptides derived from in vitro digestion hydrolysate of hen egg white lysozyme. *Food Chem.* 135(3), 1245-1252 (2012).
98. Helmfors L, Boman A, Civitelli L *et al.* Protective properties of lysozyme on β -amyloid pathology: implications for Alzheimer disease. *Neurobiol. Dis.* 83, 122-133 (2015).
99. Ganz T, Gabayan V, Liao H *et al.* Increased inflammation in lysozyme M-deficient mice in response to *Micrococcus luteus* and its peptidoglycan. *Blood* 101(6), 2388-2392 (2003).
100. Lee M, Kovacs-Nolan J, Yang C, Archbold T, Fan MZ, Mine Y. Hen egg lysozyme attenuates inflammation and modulates local gene expression in a porcine model of dextran sodium sulfate (DSS)-induced colitis. *J. Agric. Food Chem.* 57(6), 2233-2240 (2009).

101. Artym J, Zimecki M. Milk-derived proteins and peptides in clinical trials. *Postepy Hig. Med. Dosw. (Online)* 67, 800-816 (2013).
102. De Douder C, Morias J. On lysozyme therapy. I. Lysozyme tablets in treatment of some localized and generalized viral skin diseases. *Medickon* 3, 19-20 (1974).
103. Glück U. The antimicrobial effect of lysozyme on nasal mucosa. *HNO* 37, 207-210 (1989).
104. Sato M, Oe H, Nakano M, Kawasaki H, Hirayama C. A random controlled study of the prophylactic effect of lysozyme chloride on post-transfusion hepatitis. *Hepatogastroenterology* 28, 135-138 (1981).
105. Rubbini A, Rizzi S, Ruozi C. Use of lysozyme in parodontopathy. Controlled clinical trials. *Riv. It. Biol. Med.* 9, 45-49 (1989).
106. Gąsior-Chrzan B. Clinical trial of lysozyme treatment of crural ulcers in humans. *Przegl. Dermatol.* 75, 435-438 (1988).
107. Bol'shakova AM, Medvedeva MM, T.P. Z. Lysozyme in the feeding of premature infants with mixed pathology. *Antibiotiki* 29, 784-790 (1984).
108. Cheng WD, Wold KJ, Bollinger LB *et al.* Supplementation With Lactoferrin and Lysozyme Ameliorates Environmental Enteric Dysfunction: A Double-Blind, Randomized, Placebo-Controlled Trial. *Am. J. Gastroenterol.* 114(4), 671-678 (2019).
109. Luniakin AA, Bogomaz TA. Lysozyme in the overall treatment of children with an influenza infection and pneumonia. *Pediatr. Akus. Ginekol.* 1, 11-13 (1977).
110. Gavrilenko TI, Siurin SA, Lolaeva LT, Savchenko VM. The characteristics of lysozyme and carbenicillin action on the clinico-immunological status of patients with chronic bronchitis. *Lik. Sprava* 8, 42-45 (1992).
111. Bhavsar T, Liu M, Hardej D, Liu X, Cantor J. Aerosolized recombinant human lysozyme ameliorates *Pseudomonas aeruginosa*-induced pneumonia in hamsters. *Exp. Lung Res.* 36(2), 94-100 (2010).
112. Griswold KE, Bement JL, Teneback CC, Scanlon TC, Wargo MJ, Leclair LW. Bioengineered lysozyme in combination therapies for *Pseudomonas aeruginosa* lung infection. *Bioengineered* 5(2), 143-147 (2014).
113. Cantor JO, Shteyngart B, Cerreta JM, Turino GM. The effect of lysozyme on elastase-mediated injury. *Exp. Biol. Med. (Maywood)* 227(2), 108-113 (2002).
114. Dupont D, Arnould C, Rolet-Repecaud O *et al.* Determination of bovine lactoferrin concentrations in cheese with specific monoclonal antibodies. *International Dairy Journal* 16(9), 1081-1087 (2006).
115. Hagiwara S, Kawai K, Anri A, Nagahata H. Lactoferrin concentrations in milk from normal and subclinical mastitic cows. *J. Vet. Med. Sci.* 65, 319-323 (2003).
116. Chen PW, Mao FC. Detection of lactoferrin in bovine and goat milk by enzyme-linked immunosorbent assay. *J. Food Drug. Anal.* 12, 133-139 (2004).
117. Campanella L, Martini E, Pintore M, Tomassetti M. Determination of Lactoferrin and Immunoglobulin G in Animal Milks by New Immunosensors. *Sensors* 9, 2202-2221 (2009).
118. Cheng JB, Wang JQ, Bu DP *et al.* Factors Affecting the Lactoferrin Concentration in Bovine Milk. *J. Dairy Sci.* 3, 970-976 (2008).
119. Mcgrath BA, Fox PF, Mcsweeney PLH, Kelly AL. Composition and properties of bovine colostrum: a review. *Dairy Sci. Technol.* 96, 133-158 (2015).
120. Tsuji S, Hirata Y, Mukai F, Ohtagaki S. Comparison of Lactoferrin Content in Colostrum Between Different Cattle Breeds. *J. Dairy Sci.* 73(1), 125-128 (1990).
121. Yoshida S, Wei Z, Shinmura Y, Fukunaga N. Separation of Lactoferrin-A and -B From Bovine Colostrum. *J. Dairy Sci.* 83(10), 2211-2215 (2000).
122. Abe H, Saito H, Miyakawa H *et al.* Heat stability of bovine lactoferrin at acidic pH. *J. Dairy Sci.* 74, 65-71 (1991).
123. Brick T, Ege M, Boeren S *et al.* Effect of Processing Intensity on Immunologically Active Bovine Milk Serum Proteins. *Nutrients* 9(9), E963 (2017).

124. Karav S, German JB, Rouquié C, Le Parc A, Barile D. Studying Lactoferrin N-Glycosylation. *Int. J. Mol. Sci.* 18(4), 870 (2017).
125. Wang M, Sun Z, Yu T *et al.* Large-scale production of recombinant human lactoferrin from high-expression, marker-free transgenic cloned cows. *Sci. Rep.* 7, 10733 (2017).
126. Ho CL, Hwang IY, Loh K, Chang MW. Matrix-immobilized yeast for large-scale production of recombinant human lactoferrin. *Med. Chem. Commun.* 6, 486-491 (2015).
127. Rosa L, Cutone A, Lepanto MS *et al.* Physico-chemical properties influence the functions and efficacy of commercial bovine lactoferrins. *Biometals* 3, 301-312 (2018).
128. Troost FJ, Steijns J, Saris WH, Brummer RJ. Gastric Digestion of Bovine Lactoferrin in Vivo in Adults. *J. Nutr.* 131(8), 2101-2104 (2001).
129. Kilic E, Novoselova MV, Lim SH *et al.* Formulation for Oral Delivery of Lactoferrin Based on Bovine Serum Albumin and Tannic Acid Multilayer Microcapsules. *Sci. Rep.* 7, 44159 (2017).
130. Ishikado A, Imanaka H, Takeuchi T, Harada E, Makino T. Liposomalization of Lactoferrin Enhanced It's Anti-Inflammatory Effects via Oral Administration. *Biol. Pharm. Bull.* 28(9), 1717-1721 (2005).
131. Bluard-Deconinck JM, Williams J, Evans RW, Van Snick J, Osinski PA, Masson PL. Iron-binding Fragments From the N-terminal and C-terminal Regions of Human Lactoferrin. *Biochem. J.* 171(2), 321-327 (1978).
132. Saito H, Takase M, Tamura Y, Shimamura S, Tomita M. Physicochemical and Antibacterial Properties of Lactoferrin and its Hydrolysate Produced by Heat Treatment at Acidic pH. *Adv. Exp. Med. Biol.* 219–226 (1994).
133. Bellamy W, Takase M, Yamauchi K, Wakabayashi H, Kawase K, Tomita M. Identification of the bactericidal domain of lactoferrin. *Biochim. Biophys. Acta* 1121, 130-136 (1992).
134. Abeyrathne EDNS, Lee HY, Ahn DU. Egg white proteins and their potential use in food processing or as nutraceutical and pharmaceutical agents—A review. *Poult. Sci.* 92(12), 3292-3299 (2013).
135. Froning GW, Peters D, Muriana P, Eskridge K, Travnicsek D, Sumner SS. International egg pasteurization manual. (2002).
136. Venkataramani S, Truntzer J, Coleman DR. Thermal stability of high concentration lysozyme across varying pH: A Fourier Transform Infrared study. *J. Pharm. Bioallied Sci.* 5(2), 148–153 (2013).
137. Khan MV, Ishtikhar M, Rabbani G, Zaman M, Abdelhameed AS, Khan RH. Polyols (Glycerol and Ethylene glycol) mediated amorphous aggregate inhibition and secondary structure restoration of metalloproteinase-conalbumin (ovotransferrin). *Int. J. Biol. Macromol.* 94(Pt A), 290-300 (2017).
138. Caubet J, Wang J. Current understanding of egg allergy. *Pediatr. Clin. North Am.* 58(2), 427–443 (2011).
139. Brock JH, Arzabe F, Lampreave F, Piñeiro A. The Effect of Trypsin on Bovine Transferrin and Lactoferrin. *Biochim. Biophys. Acta* 446(1), 214-225 (1976).
140. Azari PR, Feeney RE. Resistance of Metal Complexes of Conalbumin and Transferrin to Proteolysis and to Thermal Denaturation. *J. Biol. Chem.* 232(1), 293-302 (1958).
141. Pazzucconi F, Barbi S, Baldassarre D, Colombo N, Dorigotti F, Sirtori CR. Iron-ovotransferrin Preparation Does Not Interfere With Ciprofloxacin Absorption. *Clin. Pharmacol. Ther.* 59(4), 418-422 (1996).
142. Martos G, López-Fandiño R, Molina E. In vitro digestions and IgE binding of proteins from white and whole hen's egg. *Clin. Transl. Allergy* 1(Suppl 1), O8 (2011).
143. Chakrabarti S, Guha S, Majumder K. Food-Derived Bioactive Peptides in Human Health: Challenges and Opportunities. *Nutrients.* 10(11), E1738 (2018).
144. Majumder K, Wu J. Purification and characterisation of angiotensin I converting enzyme (ACE) inhibitory peptides derived from enzymatic hydrolysate of ovotransferrin. *Food Chem.* 126(4), 1614-1619 (2011).

145. Ding L, Wang L, Zhang Y, Liu J. Transport of Antihypertensive Peptide RVPSSL, Ovotransferrin 328-332, in Human Intestinal Caco-2 Cell Monolayers. *J. Agric. Food Chem.* 63(37), 8143-8150 (2015).
146. Hashida S, Ishikawa E, Nakamichi N, Sekino H. Concentration of egg white lysozyme in the serum of healthy subjects after oral administration. *Clin. Exp. Pharmacol. Physiol.* 29(1-2), 79-83 (2002).
147. Ercan D, Demirci A. Recent advances for the production and recovery methods of lysozyme. *Crit. Rev. Biotechnol.* 36(6), 1078-1088 (2016).
148. Wu T, Jiang Q, Wu D *et al.* What Is New in Lysozyme Research and Its Application in Food Industry? A Review. *Food Chem.* 274, 698-709 (2019).
149. Yuzuriha T, Katayama K, Tsutsumi J. Studies on biotransformation of lysozyme. IV. Radioimmunoassay of lysozyme and its evaluation. *Chem. Pharm. Bull.* 26, 908-914 (1978).
150. Martos G, López-Fandiño R, Molina E. Immunoreactivity of Hen Egg Allergens: Influence on in Vitro Gastrointestinal Digestion of the Presence of Other Egg White Proteins and of Egg Yolk. *Food Chem.* 136(2), 775-781 (2013).
151. Mine Y, Ma F, Lauriau S. Antimicrobial Peptides Released by Enzymatic Hydrolysis of Hen Egg White Lysozyme. *J. Agric. Food Chem.* 52(5), 1088-1094 (2004).
152. Ahmad S, Kumar V, Ramanand KB, Rao NM. Probing protein stability and proteolytic resistance by loop scanning: A comprehensive mutational analysis. *Protein Sci.* 21(3), 433-446 (2012).
153. Singh S, Singh J. Effect of Polyols on the Conformational Stability and Biological Activity of a Model Protein Lysozyme. *AAPS PharmSciTech.* 4(3), E42 (2003).
154. Suleyman P, Yalçın H, Boyali E. The Effect of Glycerol Supplements on Aerobic and Anaerobic Performance of Athletes and Sedentary Subjects. *J. Hum. Kinet.* 34, 69-79 (2012).
155. Deutsche Forschungsgemeinschaft. Glycerin [MAK Value Documentation, 2007]. In: *The MAK-Collection Part I, MAK Value Documentations*, (Ed.^(Eds).Wiley <https://doi.org/10.1002/3527600418.mb3527605681kske3527604215> (2015).
156. Eccles R, Mallefet P. Soothing Properties of Glycerol in Cough Syrups for Acute Cough Due to Common Cold. *Pharmacy (Basel)* 5(1), 4 (2017).
157. Zancan P, Sola-Penna M. Trehalose and glycerol stabilize and renature yeast inorganic pyrophosphatase inactivated by very high temperatures. *Arch. Biochem. Biophys.* 444(1), 52-60 (2005).
158. Sato S, Ward CL, Krouse ME, Wine JJ, Kopito RR. Glycerol reverses the misfolding phenotype of the most common cystic fibrosis mutation. *J. Biol. Chem.* 271(2), 635-638 (1996).
159. Rariy RV, Klivanov AM. Correct protein folding in glycerol. *PNAS* 94(25), 13520-13523 (1997).
160. Szél E, Polyánka H, Szabo K, Hartmann P. Anti-irritant and anti-inflammatory effects of glycerol and xylitol in sodium lauryl sulphate-induced acute irritation. *J. Eur. Acad. Dermatol.* 29(12), 2333-2341 (2015).
161. Van Baare J, Buitenwerf J, Hoekstra MJ, Du Pont JS. Virucidal effect of glycerol as used in donor skin preservation. *Burns* 20(Suppl 1), S77-80 (1994).
162. Marshall L, Ghosh MM, Boyce SG, Macneil S, Freedlander E, Kudesia G. Effect of glycerol on intracellular virus survival: implications for the clinical use of glycerol-preserved cadaver skin. *Burns* 21(5), 356-361 (1995).
163. Liu YY, Zeng SY, Leu YL, Tsai TY. Antihypertensive Effect of a Combination of Uracil and Glycerol Derived from *Lactobacillus plantarum* Strain TWK10-Fermented Soy Milk. *J. Agric. Food Chem.* 63(33), 7333-7342 (2015).
164. Cox MJ, Loman N, Bogaert D, O'grady J. Co-infections: potentially lethal and unexplored in COVID-19. *Lancet* 1(1), e1 (2020).
165. El-Fakharany EM, El-Baky NA, Linjawi MH *et al.* Influence of camel milk on the hepatitis C virus burden of infected patients. *Exp. Ther. Med.* 13(4), 1313-1320 (2017).

166. Ishikawa H, Awano N, Fukui T, Sasaki H, Kyuwa S. The protective effects of lactoferrin against murine norovirus infection through inhibition of both viral attachment and replication. . *Biochem. Biophys. Res. Commun.* 434, 791-796 (2013).
167. Ellison RTI, Giehl TJ, La Force FM. Damage of the outer membrane of enteric gram negative bacteria by lactoferrin and transferrin. *Infect. Immun.* 56, 2774-2781 (1988).
168. Fernandes KE, Carter DA. The Antifungal Activity of Lactoferrin and Its Derived Peptides: Mechanisms of Action and Synergy with Drugs against Fungal Pathogens. *Front. Microbiol.* 8, 2 (2017).
169. Artym J, Zimecki M, Kuryszko J, Kruzel ML. Lactoferrin accelerates reconstitution of the humoral and cellular immune response during chemotherapy-induced immunosuppression and bone marrow transplant in mice. *Stem Cells Dev.* 14, 548-555 (2005).
170. Valenti P, Visca P, Antonini G, Orsi N. Interaction between lactoferrin and ovotransferrin and *Candida* cells. *FEMS Microbiol. Lett.* 33(2-3), 271-275 (1986).
171. Lee JH, Ahn DU, Paik HD. In Vitro Immune-Enhancing Activity of Ovotransferrin from Egg White via MAPK Signaling Pathways in RAW 264.7 Macrophages. *Korean J. Food Sci. Anim. Resources* 38(6), 1226-1236 (2018).
172. Zhu G, Luo J, Du H *et al.* Ovotransferrin enhances intestinal immune response in cyclophosphamide-induced immunosuppressed mice. *Int. J. Biol. Macromol.* 120(Pt A), 1-9 (2018).
173. Steinrauf LK, Shiuan D, Yang WJ, Chiang MY. Lysozyme association with nucleic acids. *Biochem. Biophys. Res. Commun.* 266(2), 366-370 (1999).
174. Behbahani M, Nosrati M, Mohabatkar H. Inhibition of Human Immunodeficiency Type 1 Virus (HIV-1) Life Cycle by Different Egg White Lysozymes. *Appl. Biochem. Biotechnol.* 185(3), 786-798 (2018).
175. Lee-Huang S, Maiorov V, Huang PL *et al.* Structural and functional modeling of human lysozyme reveals a unique nonapeptide, HL9, with anti-HIV activity. *Biochemistry* 44(12), 4648-4655 (2005).
176. Lampis G, Deidda D, Pinza M, Pompei R. Enhancement of anti-herpetic activity of glycyrrhizic acid by physiological proteins. *Antivir. Chem. Chemother.* 12, 125-131 (2001).
177. Sebaa S, Hizette N, Boucherit-Otmani Z, Courtois P. Dose-dependent effect of lysozyme upon *Candida albicans* biofilm. *Mol. Med. Rep.* 15(3), 1135-1142 (2017).
178. Ibrahim HR, Imazato K, Ono H. Human lysozyme possesses novel antimicrobial peptides within its N-terminal domain that target bacterial respiration. *J. Agric. Food Chem.* 59(18), 10336-10345 (2011).

Figure legends

Figure 1. Potential benefits of lactoferrin, ovotransferrin and lysozyme in SARS-CoV-2 infection.

Potential antiviral (A) and immunomodulatory (B) effects of these proteins in SARS-CoV-2 infection are illustrated. The mechanisms are further detailed in Table 1. The antiviral effects of lactoferrin against SARS-CoV-2 replication have been demonstrated. The antiviral mechanisms of ovotransferrin and lysozyme are inferred from their known effects on other viruses, however their effects against SARS-CoV-2 are currently unknown.

RAS – renin-angiotensin system; AGEs – advanced glycation end products; ACE – angiotensin converting enzyme ; HPSGs – heparan sulphate proteoglycans; IFN – type I interferon

Table 1. The overlapping antimicrobial and immunomodulatory effects of lactoferrin, ovotransferrin and lysozyme

Protein	Form	Effect	Mechanisms	Ref.
Lactoferrin	Intact and/or peptides	Antiviral*	Interaction with virus surface, DNA or cell surfaces required for virus entry, and induction of type I interferons	[39, 55, 58, 59, 166]
	Intact and peptides	Antibacterial	Iron sequestration Direct interaction with bacterial surface	[39, 132, 133, 167]
	Intact and peptides	Antifungal	Iron sequestration Direct interaction with fungal surface	[168]
	Intact	Immune enhancement, immune suppression reversal ^a	Enhances natural killer cell activity Enhances T cell responses (helper and cytotoxic T cell responses) Elevation of antibody response	[62, 63, 169]
	Intact and/or peptides	Anti-inflammatory ^a	Suppresses extracellular traps released by neutrophils Polarises macrophages to anti-inflammatory type (M2) Down-regulates IL-6 and TNF-alpha in those with high immune activation Binds to AT1 receptor to inhibit angiotensin II pro-inflammatory activity	[43, 56, 64, 65, 67]
	Intact and large fragments	Iron homeostasis ^b	Sequestering free iron Restoring levels of iron-binding proteins	[60]
	Intact and/or large fragments	Anti-oxidant ^b	Sequestering free iron Reduces intracellular levels of reactive oxygen species Increases anti-oxidant capacity of serum	[62, 63]
	Intact and peptides	Anti-thrombotic	Through sequestering free iron (free iron induces pro-coagulant state) Inhibits platelet aggregation	[68]
Ovotransferrin	Intact and peptides	Antiviral	Upregulation of type 1 interferon in virus-infected macrophages Restriction of virus entry Antiviral peptides that have sequence homology with antiviral lactoferrin peptides	[81, 84]
	Intact and peptides	Antibacterial	Iron sequestration Direct interaction with bacterial surface	[54, 83]
	Intact and peptides	Antifungal	Iron sequestration Direct interaction with fungal surface	[82, 170]
	Intact	Immune enhancement, immune restoration in immunosuppression model	Enhanced phagocytic activity as well as cytokine production of macrophages Enhanced intestinal immune responses: dendritic cell maturation, Th1/Th2 balance restored, humoral immunity promoted	[171, 172]
	Peptides	Anti-inflammatory	Down-regulates IL-6 and TNF-alpha and myeloperoxidase activity in peritonitis	[87, 91-93]

			Binds to AT1 receptor to inhibit angiotensin II pro-inflammatory activity ACE inhibitory activity (anti-hypertensive)	
	Intact	Iron-binding activity ^b	Sequestering free iron	[79]
	Intact and peptides	Anti-oxidant ^b	Sequestering free iron Free radical scavenging	[87, 88]
Lysozyme	Intact and peptides	Antiviral	Inhibits viral entry by binding to cell receptors or virus – cationic and hydrophobic nature is required rather than enzymatic activity Binds nucleic acids Inhibits virus-induced cell fusion Affects cell signalling, including NFκB pathway, to influence susceptibility to infection	[173-176]
	Intact and/or peptides	Antibacterial	Hydrolyses cell wall of gram positive bacteria (enzyme activity) Insert into and form pores in negatively charged bacterial membranes	[40]
	Intact and/or peptides	Antifungal	Enzymatic activity Cationic nature leading to membrane destabilisation Agglutination effect	[94, 177, 178]
	Intact and/or peptides	Enhance or limit immune responses ^a	Lysozyme in bacteria-containing phagosomes activates the pro-inflammatory responses of neutrophils and macrophages Decreases chemotaxis in neutrophils Suppresses TNF-alpha and IL-6 production by macrophages Facilitates excretion of AGEs Disrupts binding of peptidoglycans to complement ACE inhibitory activity Anti-oxidant activity	[30, 40, 95-97]

* Specific anti-coronavirus activity has been demonstrated: inhibits SARS-CoV cell entry by binding to HSPGs; inhibits entry and post-entry steps of SARS-CoV-2 replication and elevates interferon-stimulated genes in SARS-CoV-2-infected cells.

SARS-CoV – severe acute respiratory syndrome coronavirus; HSPGs – heparan sulphate proteoglycans; IL-6 – interleukin 6; TNF-alpha – tumour necrosis factor-alpha; ADP – adenosine diphosphate; ACE – angiotensin-converting enzyme; NFκB - nuclear factor kappa-light-chain-enhancer of activated B cells; AGE – advanced glycation end product

^a The action is of immune homeostasis – its action is appropriate in the context of the immune environment [64].

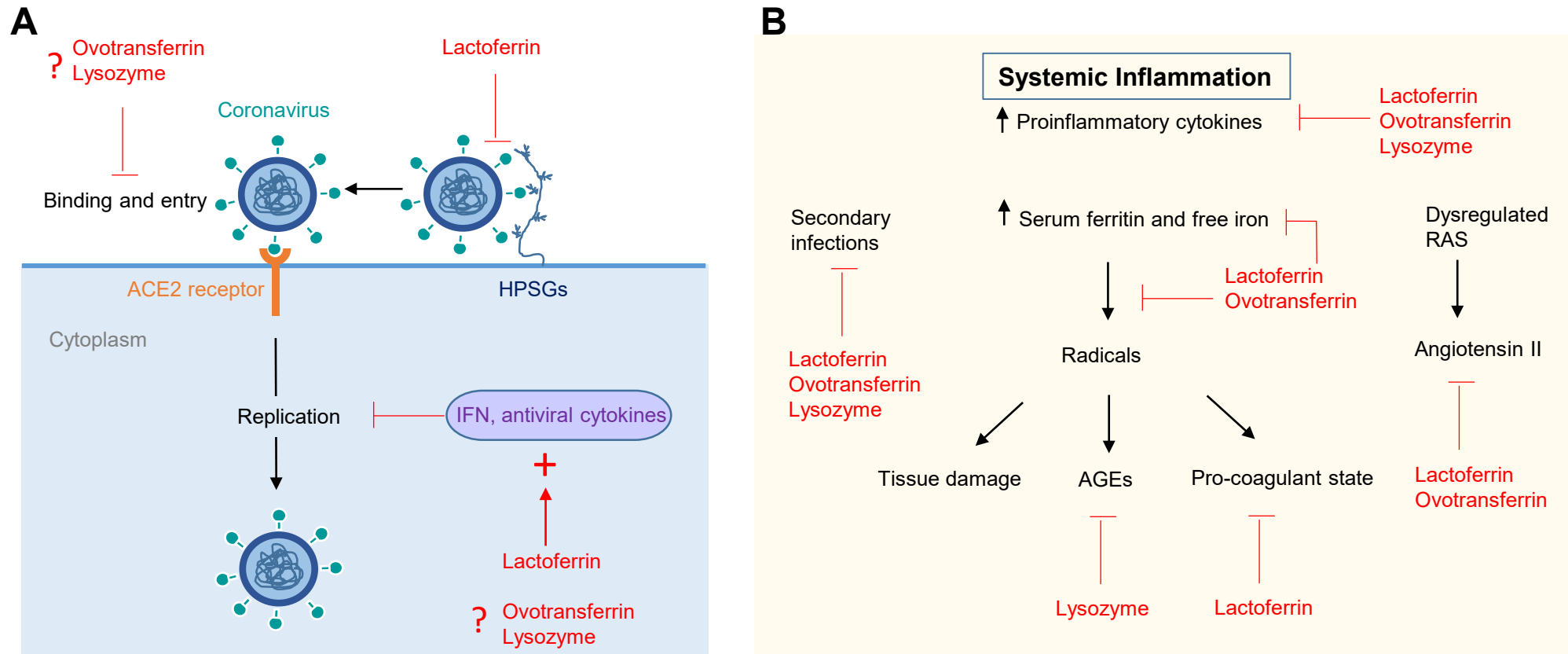


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