Opinion paper of the ESC Working Group on Cellular Biology of the Heart:

COVID-19 related cardiac complications – from clinical evidences to basic mechanisms

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Abstract - The pandemic of Coronavirus disease (COVID)-19 is a global threat, causing high mortality, especially in the elderly. The main symptoms and the primary cause of death are related to interstitial pneumonia. Viral entry also into myocardial cells mainly via the angiotensin converting enzyme type 2 (ACE2) receptor and excessive production of pro-inflammatory cytokines, however, also make the heart susceptible to injury. In addition to the immediate damage caused by the acute inflammatory response, the heart may also suffer from long-term consequences of COVID-19, potentially causing a post-pandemic increase in cardiac complications.

Although the main cause of cardiac damage in COVID-19 remains coagulopathy with micro- (and to a lesser extent macro-) vascular occlusion, open questions remain about other possible modalities of cardiac dysfunction, such as direct infection of myocardial cells, effects of cytokines storm, and mechanisms related to enhanced coagulopathy. In this opinion paper, we focus on these lesser appreciated possibilities and propose experimental approaches that could provide a more comprehensive understanding of the cellular and molecular bases of cardiac injury in COVID-19 patients. We first discuss approaches to characterize cardiac damage caused by possible direct viral infection of cardiac cells, followed by formulating hypotheses on how to reproduce and investigate the hyperinflammatory and pro-thrombotic conditions observed in the heart of COVID-19 patients using experimental *in vitro* systems. Finally, we elaborate on strategies to discover novel pathology biomarkers using omics platforms.

Keywords: SARS-CoV-2; COVID-19; myocardial injury; disease modelling; infection; inflammation.

Introduction

Since the onset of the COVID-19 pandemic outbreak in Wuhan, China, patients with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection exhibited signs of severe acute myocardial injury, proven by significantly elevation in circulating cardiac troponin (cTn) T and -I levels, occasional heart failure with acute decrease in ejection fraction, arrhythmias, and high inhospital mortality¹⁻⁵. It is well known that elderly patients presenting with comorbidities or cardiovascular risk factors are more prone to cardiac complications of SARS-CoV-2 infection⁶. There are several possible links between COVID-19 and cardiac dysfunction. These include diffuse coagulopathy causing micro/macro-vascular occlusions and hypoxia, which may unmask underlying coronary artery disease; reduced lung compliance (the 'stiff lung'7), which impairs right and left ventricular function8; direct cytotoxicity due to infection of myocardial and/or endothelial cells or exposure to the so-called cytokine storm. Despite the prevalent causes of cardiac injury appear to be well characterized at present⁹, the cause-effect relationships existing between the severity of COVID-19 cardiac injury and cardiovascular risk factors remains elusive¹⁰. Indeed, the expression of various viral entry receptors (the so-called coronavirus-associated receptors and factors -SCARFs) in myocardial cells¹¹ suggests that exposure of the heart to the virus might, directly or indirectly, determine cytopathic effects even in healthy individuals. In line with this hypothesis, retrospective analyses in cohorts of COVID-19 patients have shown that the impact of infection on myocardial damage is not limited only to patients with pre-existing risk conditions (e.g., ischemic heart disease, heart failure), but it is also relevant in individuals with apparently healthy hearts suffering potentially persistent consequences¹²⁻¹⁴. Furthermore, in various autopsy reports¹⁵⁻²⁰ SARS-CoV-2 infection has been associated with signs of cardiomyocyte toxicity either directly, i.e., associated with the presence of viral particles, or indirectly, without detection of viral particles and apparently mediated by systemic inflammation^{21, 22}. Finally, the first prospective study aimed at identifying potential long-term cardiopulmonary damage after acute COVID-19 has described a high rate of diastolic dysfunction in moderate-to-severely ill COVID-19 patients, persisting months after the infection, similar to what has been observed after infection with the phylogenetically related SARS-CoV-1¹⁴. Together, this evidence suggests that SARS-CoV-2 damages myocardial cells by direct infection and that, for a better understanding of the relationships between cardiovascular risk factors, comorbidities and COVID-19-related cardiac complications, it is critical also to consider direct cytotoxicity effects²³.

This Opinion Paper from the Working Group on Cellular Biology of the Heart of the European Society of Cardiology will highlight possible experimental approaches that may be implemented to understand the direct and indirect modalities of cardiac damage due to SARS-CoV-2, to unravel the short- and long-term effects of the virus on myocardial cells, and discuss potential biomarkers to stage the degree of cardiac damage. This contribution parallels and integrates recent reports

focusing on other aspects of the pathophysiology of COVID-19, such as endothelial dysfunction^{24, 25}, to which interested readers are referred for more extensive information.

1. Modelling cardiac damage due to direct susceptibility of cardiomyocytes/noncardiomyocyte cells to SARS-CoV-2 infection

Despite the multiple clinical manifestations and the complexity of the underlying mechanisms, which are still not completely understood²⁶⁻²⁸, myocardial injury in COVID-19 could be also mediated by direct infection of myocardial cells. In fact, according to recent data produced with single-cell RNA sequencing, various cardiac cell types (cardiomyocytes, endothelial cells, smooth muscle cells, and fibroblasts) have been found to express the angiotensin converting enzyme-2 (ACE2) transmembrane protein^{29, 30}, one of the two major host cell receptors that mediate SARS-CoV-2 infection via interaction with the viral spike protein³¹. Although the Transmembrane Serine Protease 2 (TMPRSS2), crucial for spike protein priming and viral entry, is not expressed at significant levels in cardiac cells³², other receptors and peptidases, such as cathepsin-L (CTSL), furin and SCARFs are expressed in cardiomyocytes^{11, 33}, and neuropilin-1 [a vascular endothelial growth factor (VEGF) receptor] is expressed in endothelial cells^{34, 35}. These can compensate for the lack of TMPRSS2 and facilitate SARS-CoV-2 infection and replication in the myocardium. This picture is further complicated in case of concomitant cardiac diseases, such as aortic stenosis and heart failure, or in patients treated with antihypertensive therapies affecting the renin-angiotensin system, such as ACEinhibitors, which have been shown to increase myocardial susceptibility to the virus due to ACE2 upregulation^{29, 30, 36-39}. Collectively, these findings prompt the modelling of SARS-CoV-2 infection using primary or stem cell-derived myocardial cells with genetically-controlled levels of ACE2 and other SARS-CoV-2 co-receptors in order to reveal the likely complex interaction of the different molecular pathways involved in acute cardiac injury and, potentially, in post-infection myocardial fibrosis.

Suggested pathways for SARS-CoV-2 direct damage to the heart

The myocardium may be directly affected by SARS-CoV-2 by various modalities that might occur separately or in concert⁴⁰. These different modalities are represented in **Figure 1**.

- a) The cytotoxic effect of SARS-CoV-2 on the endothelium⁴¹ (**Figure 1**, **panel a**) may result in a pro-thrombotic status leading to diffuse micro-thrombosis in the heart, and conditions resembling type 2 myocardial infarction (MI)²⁸, or the Takotsubo syndrome^{42, 43}. The hypothesis on the role of endothelial dysfunction and SARS-CoV-2 infection has been extensively discussed elsewhere²⁴ and is supported, at least in part, by the direct observations of thrombi in the microcirculation of myocardial tissues in patients with COVID-19⁴⁴.
- **b)** Although there are relatively few reports of confirmed cardiomyocyte infection and myocarditis in COVID-19 patients⁴⁰, ultrastructural and molecular studies have shown the

presence of SARS-CoV-2 in the myocardial tissue of patients with COVID-19¹⁷ (**Figure 1**, **panel b**). On the other hand, exposure of induced pluripotent stem cell (iPSCs)-derived cardiomyocytes to the virus showed the ability of SARS-CoV-2 to cause cytotoxicity, cell death and cessation of cell contraction due to the ability of the virus to bind cardiomyocytes ^{20, 33, 39, 45}. Of course, *in vitro* uptake does not imply *in vivo* entrance of the virus into cardiomyocytes. Even more important, although the use of the anti-viral remdesivir was able to block damage to cardiomyocytes in culture²⁰, the efficacy of the drug for treating COVID-19 patients has been questioned^{46, 47}. This is a clear caveat in extending the implications of *in vitro* results to the clinical scenario.

- c) A third, relatively unexplored hypothesis is that, besides being exposed to viral cytotoxicity, the myocardium is a site for SARS-CoV-2 infection and replication within multiple noncontractile myocardial cell types such as endothelial cells, fibroblasts and pericytes (Figure 1, panels c, d). This aspect is particularly relevant for a systematic modelling of cardiac damage determined by infection, given the relevance of these cell types for myocardial inflammation⁴⁸⁻⁵¹. In support of this hypothesis, a recently published report has suggested that cardiac fibroblasts can be infected by the virus and increase its replication with an efficiency related to the level of expression of the ACE2 receptor. On the other hand, exposure to the virus also resulted in an ACE2-independent, sustained pro-inflammatory response, leading to upregulation of genes encoding inflammatory cytokines and extracellular matrix components involved in cardiac fibrosis⁵². More controversial is, to date, the possibility that endothelial cells become infected by SARS-CoV-2. In fact, while initial reports showed the presence of the virus in the endothelial cells of COVID-19 patients⁴¹, very recently the direct exposure of primary vascular endothelial cells to the virus did not give rise to productive infection, likely correlated to the absence of ACE2 expression in the tested cell lines⁵³.
- d) An important point that must be addressed concerns the possible modality of transmission of the virus in the heart. This aspect appears particularly relevant considering that viral infection could progress through the mode of direct diffusion from cell to cell (the so-called 'viral synapse'⁵⁴) other than the release of the virus in the extracellular compartment. It should also be considered that cells exposed to the virus could raise or induce a strong innate immune response through the activation of the nuclear factor-κB (NF-κB) pathway, due to the interaction of the spike protein with cellular receptors such as the Toll-like receptors (TLRs)^{55, 56}. Whether a synaptic-like intercellular viral propagation exists in the heart (Figure 1, panels c, d) and infection independent activation of pro-inflammatory pathways occurs in the myocardium⁵⁷ (see also section below) is thus far only a matter of speculation and should be further investigated *in vitro* and *in vivo*.

e) A final possibility, so far only speculative, but supported by evidence emerged from the autopsy reports on the lung as the primary target organ of SARS-CoV-2⁵⁸, and from in vitro reports on cell lines⁵⁹, is that expression of the viral Spike protein on the surface of infected cells might promote cell fusion of contractile and non-contractile cells (**Figure 1, panel e**) and the formation of syncytia with extensive cardiac damage. If a similar pathologic mechanism is experimentally confirmed in cardiac cells, the use of anti-syncytial drugs⁶⁰ could be an option to prevent cardiac damage in COVID-19.

Towards a further systematic understanding of direct myocardial injury by SARS-CoV-2

In order to explore how SARS-CoV-2 directly damages the myocardial tissue, we suggest the use of advanced in vitro systems, in which virus-exposed myocardial cells are mixed with non-exposed cells, both in physical connection and separated by barriers permeable to the virus and/or secreted factor (Figure 2). Using this setting, the ability of infected cells to 'pass' the virus or viral particles to neighbouring uninfected cells via membrane contacts, such as adherens junctions-associated pores and/or tunneling nanotubes (TNT), might be assessed and quantified using methods for re-isolation of cells from mixed cultures, followed by analysis of the intracellular viral bioprocess with conventional or single cell transcriptomics and/or proteomics. This approach is ideally tackled within an 'organoid' system, in which primary cells (e.g., fibroblasts, pericytes) from human cardiac explants are mixed with human iPSC-derived cardiomyocytes after in vitro infection, or vice-versa (Figure 2). A similar approach has been successfully applied to demonstrate the cellular effects of SARS-CoV-2 infection in human enterocytes⁶¹, or iPSC-derived hepatic/pancreatic organoids⁶², and at least partially in cardiac organoids²⁰. With the latter approach, infected and uninfected cells are co-cultured in bioreactors enabling medium exchange by fluidic connections. These systems, set to assess and quantify the delivery of drugs and/or metabolites to cells⁶³, enable monitoring of trans-infection between different types of myocardial cell, or even in co-cultured lung-cardiac organoids. The proposed systems can also be exploited to screen drugs that inhibit viral entry and intracellular replication directly into cardiac cells, thus contributing to assess the efficacy of cardioprotective strategies (Figure 2).

Finally, since post-acute evolution of the infection may also result in chronic activation of pro-fibrotic pathways in the heart, it is of importance to assess matrix remodelling activity (e.g., expression of matrix metalloproteinases) and the electromechanical coupling of cells exposed to the virus (e.g., cardiac fibroblasts and myocytes) alone or within organoids. This approach enables evaluation of the impact of the viral infection on matrix compaction, composition and remodelling, and their readout on the propagation of the action potentials that are required to maintain a synchronized heartbeat. In this regard, the combination of matrix components with cardiac cells exposed to virus or to recombinant SARS-CoV-2 proteins likely represent highly standardisable approaches^{64, 65}, also amenable to high-throughput screening of drugs with antiviral action or antifibrotic/antiarrhythmic effects. This approach can optimally benefit from the use of biological material (cardiomyocytes,

fibroblasts) obtained by iPSC reprogramming of cells from patients carrying genetic mutations for arrhythmic syndromes such as the long-QT syndrome⁶⁶, one of the conditions associated with severe arrhythmias caused by various COVID-19 medications, i.e. hydroxychloroquine with azithromycin^{67,68}, and the use of conventional or pathological experimental settings mimicking some common effects of the disease, such as electrolyte imbalance, altered pH, or hypoxemia⁶⁹.

2. Indirect effects of SARS-CoV-2 infection on pro-inflammatory and pro-fibrotic pathways in the cardiovascular system

Several mechanisms have been proposed for how SARS-CoV-2 could indirectly affect the cardiovascular system, i.e., without involving direct infection of cardiovascular cells (**Figure 1**, **panels a**, **b**). These include ACE2 downregulation/shedding, a SARS-CoV-2-elicited cytokine storm, activation of thrombotic mechanisms, i.e., the activation of platelets⁷⁰ and of the the so-called "neutrophil extracellular traps" (NETs)⁷¹, and profound changes in the immune profile^{40, 72}. These phenomena triggered by the virus can occur in parallel in case of viral damage, and interact with each other, exacerbating their effect.

ACE2 downregulation and shedding

After the initial interaction between SARS-CoV-2 spike protein and ACE2, the expression of ACE2 in the epithelial cells of the lung alveoli is strikingly reduced⁷³. Loss of ACE2 leads to the accumulation of angiotensin (Ang) II in the circulatory system, which plays a central role in the activation of the interleukin (IL)-6 amplifier, with the coactivation of NF-kB and the Janus kinases (JAK)-signal transducer and activator of transcription Signal Transducer And Activator Of Transcription 3 (STAT3) pathways. Therefore, SARS-CoV-2-infected patients fail to exert a robust, interferon (IFN)-mediated antiviral response, and exhibit exuberant inflammatory cytokine production⁷⁴. Ang II may further induce tumour necrosis factor (TNF) convertase (ADAM17), which leads to shedding of membrane-bound ACE275 and the release of soluble ACE2. Of interest, the expression of ADAM17 is negatively regulated by microRNA-145 and the administration of specific antagomirs targeting microRNA-145 has been found to increase the level of circulating ACE2, thereby reducing viral entry into cells⁷⁶. Beyond the local renin angiotensin aldosterone system (RAAS) activation in the lung, there is evidence that patients suffering from a severe course of SARS-CoV-2 infection have elevated levels of plasma Ang II, which correlate with total viral load and the degree of lung injury77. Loss of ACE2 and activation of the RAAS also result in a widespread endothelial dysfunction and multiple organ injury, including the heart, the kidney and the lung⁷⁸. In summary, the susceptibility of the heart and the vasculature to the hyperactive RAAS and proinflammatory cytokines appears to be the prevalent modality by which SARS-CoV-2 can indirectly affect the cardiovascular system.

Cytokine storm

SARS-CoV-2 infection induces a strong activation of the innate immune system, leading to elevated levels of several pro-inflammatory cytokines, including IL-6,IL-1, IL-2, TNF-α and IFN-γ. Besides a direct impact of SARS-CoV-2 on ACE2 and Ang II, the activation of the innate immune system is in part due to the activation of the IL-6 amplifier via TLR479. The resulting "cytokine storm-related hyperinflammation syndrome" underlies many of the severe manifestations of COVID-19 and is suggested to contribute to COVID-19-associated cardiovascular disease and death. IL-6 appears to play a central role here, with increased serum levels correlating with the onset of acute respiratory distress syndrome (ARDS) and with adverse clinical outcome. Besides elevated circulating IL-6, COVID-19 patients exhibit increased plasma levels of the soluble IL-6 receptor (sIL-6R) in plasma, reflecting its enhanced cleavage from the cell surface during infection⁸⁰. By binding to ubiquitously expressed cellular gp130, circulating IL-6/sIL-6R complexes can directly activate JAK-STAT signalling throughout the body. Such activation in endothelial cells may cause secretion of VEGF, a reduction of E-cadherin expression, and defective pericyte coverage⁸¹, contributing to vascular permeability and leakage⁸². Beyond these effects, which participate in the pathophysiology of hypotension and pulmonary dysfunction in ARDS, IL-6 induces oxidative stress and endothelial dysfunction through overexpression of the Ang II type-1 receptor⁸³. In a highly interdependent relationship with Ang II signalling, IL-6 further promotes vascular hypertrophy, vascular inflammation and stiffness, involving induction of matrix expansion^{84,85}. In the heart, both protective and harmful effects of IL-6 have been reported86. Myocarditis is a striking example of the dysregulation of the IL-6 response leading to a detrimental outcome. IL-6 is protective in the heart as far as it limits viral replication and thus cardiac damage⁸⁷, but long exposure to IL-6 can contribute to heart failure⁸⁸. IL-6 receptor antagonism with tocilizumab in experimental myocarditis has been shown to reduce cardiac inflammation and cardiac fibrosis, and to improve cardiomyocyte titin phosphorylation and thus myocardial stiffness⁸⁹. In summary, IL-6 appears to be a central player in the hyperinflammatory response to SARS-CoV-2. The efficacy of anti-IL-6R agent tocilizumab for the treatment of severe cases of COVID-19 is debated, due to the lack of confirmation of the efficacy of this drug in three randomized trials90, initially demonstrated in small observational studies^{91, 92}, and two large positive trials^{93, 94}. The effect

Immunothrombosis

Negative COVID-19 outcomes are associated with increased levels of fibrin degradation products (D-dimers) and lower platelet counts, which are markers for an activation of hemostatic pathways⁹⁰. Hypercoagulation is considered the main cause of organ failure in severe cases of COVID-19, supported by recent observations of micro-thrombi in the lungs, brain, heart and other organs⁹⁶⁻⁹⁹. Not only is the endothelium damaged in response to viral infection, but hyperactivated monocytes,

of anti-IL-6R agents is likely to depend on the severity of the inflammatory/cytokine storm response. The key factors that could play an important role in the effect of anti-IL-6R agents may be the timing [given before admission to the Intensive Care Unit (ICU)], and also the combination with other drugs, such as high doses corticosteroid, which is being evaluated in several ongoing trials⁹⁵ (**Figure 2**).

platelets and neutrophils may also play a pathophysiological role in this process. The coagulation cascade is induced by tissue factor, which is mainly expressed by circulating monocytes, but also exposed on activated endothelial cells, leading to fibrin deposition and blood clotting^{70, 100}. Neutrophils are recruited from activated endothelial cells and release NETs, consisting of DNA, histones, and granule protein¹⁰¹. NETs may serve as a scaffold for thrombus formation by capturing and activating platelets, red blood cells and procoagulant molecules 102, 103. COVID-19 patients have elevated circulating levels of NETs, measured as myeloperoxidase-DNA complexes, and their levels correlate with both the severity of disease and the occurrence of myocardial infarction⁷¹. The presence of platelets, neutrophils and NET-like structures in the lung and in cardiac microthrombi has been confirmed in COVID-19 autopsies, and potentially contributes to organ fibrosis¹⁰⁴. In support of enhanced immunothrombotic status in severe COVID-19 patients, serum or plasma from COVID-19 patients have been shown to trigger excessive NET formation in neutrophils in vitro, with enhanced NETosis found in neutrophils from COVID-19 patients^{98, 105, 106}. Finally, NETs released by SARS-CoV-2-activated neutrophils have been shown to promote lung epithelial cell death in vitro¹⁰⁷. Also, autoantibodies that recognize phospholipids could trigger NETs activation¹⁰⁸, thus exposing patients to risks of hyper-coagulation, similar to that occurring in the antiphospholipid antibody syndrome¹⁰⁹. Taken together, these data support NETs as potential effectors of thrombosis in organs affected by SARS-CoV-2, including the heart, where NETs activation could affect the microcirculation¹¹⁰, determining diffuse ischemic conditions. So far, NETs formation can only be assessed indirectly, based on the detection of circulating DNA, histone H3, and myeloperoxidase¹⁰⁵, ¹⁰⁷. In this regard, while neutrophil activation can be monitored using conventional techniques, such as antibody-based multiparametric flow cytometry¹¹¹, there is still a lack of validated assays to reveal the activity of enzymes linked to NET release (e.g., elastase, myeloperoxidase) directly on the cell surface. The use of enzymatic activity-sensitive probes¹¹² would be an advantage to unravel the relationships between neutrophil activation and NETs release upon stimulation of the cells with, for example, COVID-19 patient sera, or to readily detect circulating activated neutrophils at different times after infection or during recovery. This approach could finally benefit from the adoption of microfluidic devices to unravel whether the excessive NETosis in the microcirculation observed in several organs, including the heart, depends on an exacerbation of the shear forces action on hyperactivated neutrophils¹¹³ (Figure 2). This can be useful also for testing the efficacy of conventional and experimental strategies to reduce neutrophil activation and NETs release¹¹⁴ during the acute phase of infection, as well as the emerging post-COVID-19 syndrome¹¹⁵.

Altered immune cell profile

Emerging findings from the peripheral blood mononuclear cell immune profile comparing mild and moderate versus severe COVID-19 reveal profound changes in innate and adaptive immune cell compartments. Regardless of the severity of the disease, COVID-19 is associated with increased numbers of neutrophils and reduced number of T-lymphocytes, as well as a selective depletion of

non-classical monocytes¹¹⁶. Since patrolling, non-classical monocytes play a crucial role in endothelial cell homeostasis and repair, the loss of this monocyte population may contribute to microthrombosis and associated complications. Moreover, a COVID-19-specific alternative activation pattern of classical monocytes correlating with disease severity has been identified¹¹⁶. It remains merely speculative, however, whether an altered monocytes phenotype promotes enhanced cardiac infiltration of this leukocyte subset and subsequent cardiac damage.

Severe COVID-19 is associated with emergency granulopoiesis and increased frequency of immature and dysfunctional neutrophils. These immature myeloid cells may have immunosuppressive functions, as previously observed in cancer and sepsis $^{117-119}$. Moreover, a cluster of mature neutrophils expressing CD274 [Programmed Death-Ligand 1 ((PDL1)] was only detected in severe COVID-19 cases, suggesting that this receptor acts as an immune 'checkpoint', blocking T-cell activation 116 . Additional studies are necessary to clarify the role of these subsets of immature, potentially immunosuppressive neutrophils in cardiac inflammation following COVID-19. It has been recognized that life-threatening COVID-19 conditions can result from the presence of anti-IFN autoantibodies, which trigger reduction of innate immune response to SARS-CoV- 2120 . Since neutralization by autoantibodies in response to SARS-CoV-2 infection affects IFN- α but not the IFN- β subtypes, it is tempting to speculate that treating patients with IFN- β may ameliorate the disease. This latter conclusion also bears an important cardioprotective readout, given the beneficial effect of IFN- β on reduction of fibrosis-associated factors in cardiac fibroblasts 121 .

3. Novel biomarkers associated to SARS-Cov-2 infection

Since the beginning of the pandemic, several reports have investigated the important prognostic value of markers of acute cardiac injury [mainly cTn, pro-thrombotic state (D-dimer, fibrinogen), increased inflammatory response (C-reactive protein, lactate dehydrogenase, IL-6, procalcitonin, ferritin) and heart failure [brain natriuretic peptide (BNP) and its N-terminal pro-peptide, NT-proBNP] in patients with COVID-19^{1, 4, 6, 27, 122, 123}. Although multiple studies consistently demonstrate that several cardiac biomarkers correlate with the severity and prognosis of COVID-19 infection in critically ill patients, whether established cardiovascular biomarkers might provide additional prognostic information over clinical or physiological information in unselected patients hospitalized for COVID-19 with various degrees of disease severity has been recently questioned¹²⁴. Indeed, the levels of biomarker and the interpretation of the data depend on the severity of COVID-19, sex, age and the condition of new versus pre-existing cardiac disease¹²⁴. Importantly, the prognostic value of some of these blood-based biomarkers in the context of COVID-19 might differ from commonly used reference standards¹²⁵.

Additional circulating biomarkers potentially associated with SARS-CoV-2 infection have been identified and proposed for future clinical use in COVID-19. For example, it has been recently shown that growth differentiation factor 15 (GDF-15) may represent a potential biomarker. GDF-15 is a member of the transforming growth factor β superfamily, which is induced by aging and several

diseases, including cardiovascular diseases, sepsis and cancer. GDF-15 has improved prognostic value compared to various cardiovascular and inflammatory biomarkers in unselected patients hospitalized with COVID-19¹²⁶. GDF-15 levels were elevated in the majority of COVID-19 hospitalized patients, and higher concentrations were associated with ICU admission and death during hospitalization, as well as SARS-CoV-2 viremia and hypoxemia¹²⁶. Although the precise mechanisms underlying the latter association are not yet completely understood, these results suggest that GDF-15 may provide important pathophysiological information in hospitalized patients with COVID-19 while contributing to risk stratification. SARS-CoV-2 binding to ACE2 leads to its internalization and cleavage to soluble ACE2 (sACE2), decreasing ACE2 tissue levels¹²⁷. It has been proposed that sACE2 levels might reflect a higher cellular content of ACE2 and thus greater susceptibility to COVID-19^{128, 129}. In addition, sACE2 might be a marker of the RAAS dysregulation¹³⁰. Consistent with this possibility, in two large, independent cohorts of elderly patients with atrial fibrillation and increased risk of stroke, higher levels of sACE2 were associated with male sex, cardiovascular disease, diabetes, and older age, which are also the main risk factors for complications and mortality of COVID-19 patients¹³¹. Interestingly, levels of GDF-15, NT-proBNP and high-sensitivity cTn had the strongest associations with sACE2 levels and the risk of death and cardiovascular complications¹³¹. More recently, in a large, prospective, global, community-based cohort of patients, increased levels of circulating sACE2 have been associated with a higher increased risk of total death, myocardial infarction, incident heart failure, stroke and diabetes 130. However, to what extent sACE2 levels in blood samples of COVID-19 patients reveal a prognostic role remains to be established. Also, further research is warranted to dissect the relationships between circulating sACE2 levels and ACE2 expression in various organs, and whether ACE2 and/or sACE2 levels might affect the risk of SARS-CoV-2 infection or severity¹³² (**Figure 2**).

Large-scale targeted/untargeted molecular screening technologies can also help to find novel measurable markers of cardiovascular risk in COVID-19 patients. For example, genome-wide association studies (GWAS) and Mendelian randomization can help identify host genetic variants associated with critical illness that enable identification of novel mechanistic targets for therapeutic development. Very recently, the Genetics Of Mortality In Critical Care (GenOMICC) study has discovered new variants in patients admitted to ICUs in the United Kingdom, plausibly associated with the immune-mediated phase of COVID-19, such as activated IFN signalling, monocyte activation and infiltration¹³³. Moreover, transcriptomic studies have identified a specific transcriptional signature induced by viral infection in cardiomyocytes, characterized by the induction of genes involved in IFN signalling, apoptotic cell death, reactive oxygen species production and disruption of structural proteins associated with myofibrillar fragmentation^{20, 134}. In this regard, non-coding RNA expression (especially miRNAs) may have prognostic value for their involvement in the regulation of the replication cycle/viral genome translation of RNA viruses, including COVID-19^{135, 136}, and in the control of risk conditions associated with a worse COVID-19 prognosis. Finally, COVID-19 may alter

the expression pattern of circulating miRNAs¹³⁷. If confirmed, this will suggest the existence of specific miRNA signatures characterizing the disease and possibly correlated to its severity in the heart and other organs¹³⁸ (**Figure 2**). On the other hand, it should be noted that there are currently substantial diagnostic challenges with miRNA expression pattern analysis, e.g., non-standardized test formats, lack of knowledge about normal miRNA levels, and how they may be affected by confounding factors¹³⁹.

Proteomic profiling of sera from COVID-19 patients is of interest as it provides valuable and novel information on disease progression and prognosis, leading to the identification of novel biomarkers or targets discriminating cardiovascular involvement¹⁴⁰⁻¹⁴². In addition, the definition of SARS-CoV-2-encoded proteome in relation to human genetics enables the unmasking of risk factors for adverse outcome in patients, as well as possible therapeutic interventions that may prevent infection 143. Untargeted proteomics may be utilized to identify key molecular effectors associated with viral infection in cells exposed to the virus in vitro. By using proteomics, an exhaustive map of the cellular pathways affected by the virus can be derived and crucial components of mRNA maturation, protein translation and metabolic control machineries can be identified, producing useful information for potential targeting key intracellular cascades implicated in viral replication¹⁴⁴. In line with these results, metabolomic and lipidomic serum profiling has shown that COVID-19 exerts remarkable effects on the metabolism by increasing, for example, the levels of ketone bodies and 2hydroxybutyric acid, indicating altered hepatic glutathione synthesis and oxidative stress, and promoting a redistribution of serum lipoproteins potentially enhancing atherosclerotic risk145, 146. Finally, it has been suggested that COVID-19 may also exacerbate age-related mitochondrial dysfunction with detrimental effects on inflammation, oxidative phosphorylation metabolism and antiviral response¹⁴⁷. Exposure of myocardial-derived cells to sera of patients, ideally sampled at various stages of the disease, could be an answer to the problem of potential alterations in cardiac metabolism leading to damage and cytotoxicity. Considering that molecular targets identified by omics analyses are an integral part of the pathophysiology in several diseases, the use of omics in COVID-19 can help in the identification of new possible therapeutic strategies for targeting, for example, i) viral proteins essential for virus entry into the host cell, ii) proteins involved in virus-host interaction, and iii) pathways involved in intracellular signalling elicited by virus entry or virus/cell interactions (Figure 2).

Conclusions

The quest for immediate answers to SARS-CoV-2 pandemic outbreak has prompted scientists around the world to make an unprecedented effort to understand the pathophysiological basis of the infection and its consequences for a systemic disease affecting various organs and organ systems. According to the fast growing availability of scientific literature on this topic, with unprecedented speed, and daily clinical experience, the heart appears as one of the elective targets of the virus, even if, to date, the nature of the damage and the persistence of long-term complications are unclear.

A possible consequence of COVID-19 in the post-pandemic period could be an increase in heart failure¹⁴⁸, with social costs additive to those already sustained for combatting the impact of the disease.

The introduction of SARS-CoV-2 experimental model systems may help understand the interplay between systemically acting factors (e.g., the cytokine storm) and tissue-specific responses that determine the multi-organ failure often observed in most severe cases 149, 150. Finally, an aspect that should be taken into account in this scenario is the relevance, in addition to age, of risk factors, including sex and frequently associated co-morbidities, in ACE2 regulation, inflammatory responses and thrombosis¹⁵¹⁻¹⁵⁸. In line with our recent recommendations for ischemic heart disease¹⁵⁹, it will be thus important to address the interaction of confounders such as sex and comorbidities in COVID-19 experimental settings, by including, for example, individuals and cells from both sexes. Moreover, in the event that sexual dimorphic phenotypes are observed, it should be determined experimentally whether they are dependent on the hormonal status, and if they are specific, or modified by genetics and sex. In this regard, in vivo preclinical models will be extremely helpful for addressing the role of sex by combining COVID-19 experimental conditions with specific comorbidity models in male and female animals, also including the most prevalent and relevant risk conditions for COVID-19, and the effects of their co-medications (Figure 2). Together with the results from in vitro modelling with human cells, integrated with unbiased multi-omics approaches and molecular network analyses, these efforts will contribute to a decisive advancement in precision medicine and genetic surveillance to predict new genetic variants of SARS-CoV-2, to redesign second generation vaccines for new variants and to improve prevention, diagnosis and treatment of COVID-19 cardiac complications more effectively. In this regard, the creation of a global genetic surveillance system for the virus appears necessary for the effective control of the pandemic. This will be key in the years to come to ensure that we will not be in a position where, at the time of herd immunity, the virus escapes from immunological control, and it becomes necessary to redesign vaccines for a new variant.

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Conflict of interest

PF is the founder and CEO of Pharmahungary Group, a group of R&D companies.

References

- 1. Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, Xiang J, Wang Y, Song B, Gu X, Guan L, Wei Y, Li H, Wu X, Xu J, Tu S, Zhang Y, Chen H, Cao B. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *The Lancet* 2020;**395**:1054-1062.
- 2. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q, Wang J, Cao B. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *The Lancet* 2020;**395**:497-506.
- 3. Wang DW, Hu B, Hu C, Zhu FF, Liu X, Zhang J, Wang BB, Xiang H, Cheng ZS, Xiong Y, Zhao Y, Li YR, Wang XH, Peng ZY. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. *Jama-J Am Med Assoc* 2020;**323**:1061-1069.
- 4. Guo T, Fan Y, Chen M, Wu X, Zhang L, He T, Wang H, Wan J, Wang X, Lu Z. Cardiovascular Implications of Fatal Outcomes of Patients With Coronavirus Disease 2019 (COVID-19). *JAMA Cardiology* 2020;**5**:811.
- 5. Giustino G, Croft LB, Stefanini GG, Bragato R, Silbiger JJ, Vicenzi M, Danilov T, Kukar N, Shaban N, Kini A, Camaj A, Bienstock SW, Rashed ER, Rahman K, Oates CP, Buckley S, Elbaum LS, Arkonac D, Fiter R, Singh R, Li E, Razuk V, Robinson SE, Miller M, Bier B, Donghi V, Pisaniello M, Mantovani R, Pinto G, Rota I, Baggio S, Chiarito M, Fazzari F, Cusmano I, Curzi M, Ro R, Malick W, Kamran M, Kohli-Seth R, Bassily-Marcus AM, Neibart E, Md GS, Perk G, Mancini D, Reddy VY, Pinney SP, Dangas G, Blasi F, Sharma SK, Mehran R, Condorelli G, Stone GW, Fuster V, Lerakis S, Goldman ME. Characterization of Myocardial Injury in Patients With COVID-19. *Journal of the American College of Cardiology* 2020;**76**:2043-2055.
- 6. Shi SB, Qin M, Shen B, Cai YL, Liu T, Yang F, Gong W, Liu X, Liang JJ, Zhao QY, Huang H, Yang B, Huang CX. Association of Cardiac Injury With Mortality in Hospitalized Patients With COVID-19 in Wuhan, China. *JAMA Cardiology* 2020;**5**:802-810.
- 7. Crimi E, Slutsky AS. Inflammation and the acute respiratory distress syndrome. *Best Practice* & *Research Clinical Anaesthesiology* 2004;**18**:477-492.
- 8. Caravita S, Baratto C, Di Marco F, Calabrese A, Balestrieri G, Russo F, Faini A, Soranna D, Perego GB, Badano LP, Grazioli L, Lorini FL, Parati G, Senni M. Haemodynamic characteristics of COVID-19 patients with acute respiratory distress syndrome requiring mechanical ventilation. An invasive assessment using right heart catheterization. *EUROPEAN JOURNAL OF HEART FAILURE* 2020;**22**:2228-2237.
- 9. Jaffe AS, Cleland JGF, Katus HA. Myocardial injury in severe COVID-19 infection. *European Heart Journal* 2020;**41**:2080-2082.
- 10. Cheng P, Zhu H, Witteles RM, Wu JC, Quertermous T, Wu SM, Rhee JW. Cardiovascular Risks in Patients with COVID-19: Potential Mechanisms and Areas of Uncertainty. *Current Cardiology Reports* 2020;**22**:34.
- 11. Singh M, Bansal V, Feschotte C. A Single-Cell RNA Expression Map of Human Coronavirus Entry Factors. *Cell Reports* 2020;**32**:108175.
- 12. Wang N, Cao J, Lal S. COVID-19: getting to the heart of the matter. *EUROPEAN JOURNAL OF HEART FAILURE* 2020;**22**:2216-2218.
- 13. Puntmann VO, Carerj ML, Wieters I, Fahim M, Arendt C, Hoffmann J, Shchendrygina A, Escher F, Vasa-Nicotera M, Zeiher AM, Vehreschild M, Nagel E. Outcomes of Cardiovascular Magnetic Resonance Imaging in Patients Recently Recovered From Coronavirus Disease 2019 (COVID-19). *JAMA Cardiology* 2020;**5**:1265.

- 14. Fayol A, Livrozet M, Boutouyrie P, Khettab H, Betton M, Tea V, Blanchard A, Bruno RM, Hulot JS, Grp FCCS. Cardiac performance in patients hospitalized with COVID-19: a 6 month follow-up study. *ESC Heart Failure* 2021;**n/a**.
- 15. Lindner D, Fitzek A, Brauninger H, Aleshcheva G, Edler C, Meissner K, Scherschel K, Kirchhof P, Escher F, Schultheiss HP, Blankenberg S, Puschel K, Westermann D. Association of Cardiac Infection With SARS-CoV-2 in Confirmed COVID-19 Autopsy Cases. *JAMA Cardiology* 2020;**5**:1281-1285.
- 16. Buja LM, Wolf DA, Zhao BH, Akkanti B, McDonald M, Lelenwa L, Reilly N, Ottaviani G, Elghetany MT, Trujillo DO, Aisenberg GM, Madjid M, Kar B. The emerging spectrum of cardiopulmonary pathology of the coronavirus disease 2019 (COVID-19): Report of 3 autopsies from Houston, Texas, and review of autopsy findings from other United States cities. *Cardiovascular Pathology* 2020;**48**:107233.
- 17. Tavazzi G, Pellegrini C, Maurelli M, Belliato M, Sciutti F, Bottazzi A, Sepe PA, Resasco T, Camporotondo R, Bruno R, Baldanti F, Paolucci S, Pelenghi S, Iotti GA, Mojoli F, Arbustini E. Myocardial localization of coronavirus in COVID-19 cardiogenic shock. *EUROPEAN JOURNAL OF HEART FAILURE* 2020;**22**:911-915.
- 18. Wenzel P, Kopp S, Gobel S, Jansen T, Geyer M, Hahn F, Kreitner KF, Escher F, Schultheiss HP, Munzel T. Evidence of SARS-CoV-2 mRNA in endomyocardial biopsies of patients with clinically suspected myocarditis tested negative for COVID-19 in nasopharyngeal swab. *Cardiovascular Research* 2020;**116**:1661-1663.
- 19. Escher F, Pietsch H, Aleshcheva G, Bock T, Baumeier C, Elsaesser A, Wenzel P, Hamm C, Westenfeld R, Schultheiss M, Gross U, Morawietz L, Schultheiss HP. Detection of viral SARS-CoV-2 genomes and histopathological changes in endomyocardial biopsies. *ESC Heart Failure* 2020;**7**:2440-2447.
- 20. Bojkova D, Wagner JUG, Shumliakivska M, Aslan GS, Saleem U, Hansen A, Luxán G, Günther S, Pham MD, Krishnan J, Harter PN, Ermel UH, Frangakis AS, Milting H, Zeiher AM, Klingel K, Cinatl J, Dendorfer A, Eschenhagen T, Tschöpe C, Ciesek S, Dimmeler S. SARS-CoV-2 infects and induces cytotoxic effects in human cardiomyocytes. *Cardiovascular Research* 2020;**116**:2207-2215.
- 21. Zheng YY, Ma YT, Zhang JY, Xie X. COVID-19 and the cardiovascular system. *Nature Reviews Cardiology* 2020;**17**:259-260.
- 22. Akhmerov A, Marban E. COVID-19 and the Heart. Circulation Research 2020;126:1443-1455.
- 23. Fox SE, Li G, Akmatbekov A, Harbert JL, Lameira FS, Brown JQ, Vander Heide RS. Unexpected Features of Cardiac Pathology in COVID-19 Infection. *Circulation* 2020;**142**:1123-1125.
- 24. Evans PC, Rainger GE, Mason JC, Guzik TJ, Osto E, Stamataki Z, Neil D, Hoefer IE, Fragiadaki M, Waltenberger J, Weber C, Bochaton-Piallat M-L, Bäck M. Endothelial dysfunction in COVID-19: a position paper of the ESC Working Group for Atherosclerosis and Vascular Biology, and the ESC Council of Basic Cardiovascular Science. *Cardiovascular Research* 2020;**116**:2177-2184.
- 25. Libby P, Luscher T. COVID-19 is, in the end, an endothelial disease. *European Heart Journal* 2020;**41**:3038-3044.
- 26. Hendren NS, Drazner MH, Bozkurt B, Cooper LT. Description and Proposed Management of the Acute COVID-19 Cardiovascular Syndrome. *Circulation* 2020;**141**:1903-1914.
- 27. Stefanini GG, Chiarito M, Ferrante G, Cannata F, Azzolini E, Viggiani G, De Marco A, Briani M, Bocciolone M, Bragato R, Corrada E, Gasparini GL, Marconi M, Monti L, Pagnotta PA, Panico C, Pini D, Regazzoli D, My I, Kallikourdis M, Ciccarelli M, Badalamenti S, Aghemo A, Reimers B, Condorelli G, Force HC-T. Early detection of elevated cardiac biomarkers to optimise risk stratification in patients with COVID-19. *Heart (British Cardiac Society)* 2020;**106**:1512-1518.

- 28. Guzik TJ, Mohiddin SA, Dimarco A, Patel V, Savvatis K, Marelli-Berg FM, Madhur MS, Tomaszewski M, Maffia P, D'Acquisto F, Nicklin SA, Marian AJ, Nosalski R, Murray EC, Guzik B, Berry C, Touyz RM, Kreutz R, Wang DW, Bhella D, Sagliocco O, Crea F, Thomson EC, McInnes IB. COVID-19 and the cardiovascular system: implications for risk assessment, diagnosis, and treatment options. *Cardiovasc Res* 2020;**116**:1666-1687.
- 29. Nicin L, Abplanalp WT, Mellentin H, Kattih B, Tombor L, John D, Schmitto JD, Heineke J, Emrich F, Arsalan M, Holubec T, Walther T, Zeiher AM, Dimmeler S. Cell type-specific expression of the putative SARS-CoV-2 receptor ACE2 in human hearts. *European Heart Journal* 2020;**41**:1804-1806.
- 30. Xiong C, Feng Y, Chen M, Li X, Chen L. The ACE2 expression in human heart indicates new potential mechanism of heart injury among patients infected with SARS-CoV-2. *Cardiovascular Research* 2020;**116**:1097-1100.
- 31. Wang QH, Zhang YF, Wu LL, Niu S, Song CL, Zhang ZY, Lu GW, Qiao CP, Hu Y, Yuen KY, Wang QS, Zhou H, Yan JH, Qi JX. Structural and Functional Basis of SARS-CoV-2 Entry by Using Human ACE2. *Cell* 2020;**181**:894.
- 32. Liu PP, Blet A, Smyth D, Li HL. The Science Underlying COVID-19 Implications for the Cardiovascular System. *Circulation* 2020;**142**:68-78.
- 33. Liu HN, Gai SJ, Wang XY, Zeng JT, Sun C, Zhao Y, Zheng Z. Single-cell analysis of SARS-CoV-2 receptor ACE2 and spike protein priming expression of proteases in the human heart. *Cardiovascular Research* 2020;**116**:1733-1741.
- 34. Daly JL, Simonetti B, Klein K, Chen KE, Williamson MK, Anton-Plagaro C, Shoemark DK, Simon-Gracia L, Bauer M, Hollandi R, Greber UF, Horvath P, Sessions RB, Helenius A, Hiscox JA, Teesalu T, Matthews DA, Davidson AD, Collins BM, Cullen PJ, Yamauchi Y. Neuropilin-1 is a host factor for SARS-CoV-2 infection. *Science* 2020;**370**:861.
- 35. Cantuti-Castelvetri L, Ojha R, Pedro LD, Djannatian M, Franz J, Kuivanen S, van der Meer F, Kallio K, Kaya T, Anastasina M, Smura T, Levanov L, Szirovicza L, Tobi A, Kallio-Kokko H, Österlund P, Joensuu M, Meunier FA, Butcher SJ, Winkler MS, Mollenhauer B, Helenius A, Gokce O, Teesalu T, Hepojoki J, Vapalahti O, Stadelmann C, Balistreri G, Simons M. Neuropilin-1 facilitates SARS-CoV-2 cell entry and infectivity. *Science* 2020;**370**:eabd2985.
- 36. Goulter AB, Goddard MJ, Allen JC, Clark KL. ACE2 gene expression is up-regulated in the human failing heart. *BMC medicine* 2004;**2**:19.
- 37. Breidenbach JD, Dube P, Ghosh S, Abdullah BN, Modyanov NN, Malhotra D, Dworkin LD, Haller ST, Kennedy DJ. Impact of Comorbidities on SARS-CoV-2 Viral Entry-Related Genes. *J Pers Med* 2020;**10**:146.
- 38. Oz M, Lorke DE, Kabbani N. A comprehensive guide to the pharmacologic regulation of angiotensin converting enzyme 2 (ACE2), the SARS-CoV-2 entry receptor. *Pharmacology & Therapeutics* 2021;**221**:107750.
- 39. Bristow MR, Zisman LS, Altman NL, Gilbert EM, Lowes BD, Minobe WA, Slavov D, Schwisow JA, Rodriguez EM, Carroll IA, Keuer TA, Buttrick PM, Kao DP. Dynamic Regulation of SARS-Cov-2 Binding and Cell Entry Mechanisms in Remodeled Human Ventricular Myocardium. *Jacc-Basic Transl Sc* 2020;**5**:871-883.
- 40. Van Linthout S, Klingel K, Tschöpe C. SARS-CoV-2-related myocarditis-like syndromes Shakespeare's question: what's in a name? *EUROPEAN JOURNAL OF HEART FAILURE* 2020;**22**:922-925.
- 41. Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, Mehra MR, Schuepbach RA, Ruschitzka F, Moch H. Endothelial cell infection and endotheliitis in COVID-19. *Lancet* 2020;**395**:1417-1418.

- 42. Sala S, Peretto G, Gramegna M, Palmisano A, Villatore A, Vignale D, De Cobelli F, Tresoldi M, Cappelletti AM, Basso C, Godino C, Esposito A. Acute myocarditis presenting as a reverse Tako-Tsubo syndrome in a patient with SARS-CoV-2 respiratory infection. *European Heart Journal* 2020;**41**:1861-1862.
- 43. Giustino G, Croft LB, Oates CP, Rahman K, Lerakis S, Reddy VY, Goldman M. Takotsubo Cardiomyopathy in COVID-19. *Journal of the American College of Cardiology* 2020;**76**:628-629.
- 44. Bois MC, Boire NA, Layman AJ, Aubry M-C, Alexander MP, Roden AC, Hagen CE, Quinton RA, Larsen C, Erben Y, Majumdar R, Jenkins SM, Kipp BR, Lin PT, Maleszewski JJ. COVID-19—Associated Nonocclusive Fibrin Microthrombi in the Heart. *Circulation* 2021;**143**:230-243.
- 45. Sharma A, Garcia G, Wang Y, Plummer JT, Morizono K, Arumugaswami V, Svendsen CN. Human iPSC-Derived Cardiomyocytes Are Susceptible to SARS-CoV-2 Infection. *Cell Reports Medicine* 2020;**1**:100052.
- 46. Santoro MG, Carafoli E. Remdesivir: From Ebola to COVID-19. *Biochemical and Biophysical Research Communications* 2021;**538**:145-150.
- 47. Pan H, Peto R, Henao-Restrepo AM, Preziosi MP, Sathiyamoorthy V, Abdool Karim Q, Alejandria MM, Hernández García C, Kieny MP, Malekzadeh R, Murthy S, Reddy KS, Roses Periago M, Abi Hanna P, Ader F, Al-Bader AM, Alhasawi A, Allum E, Alotaibi A, Alvarez-Moreno CA, Appadoo S, Asiri A, Aukrust P, Barratt-Due A, Bellani S, Branca M, Cappel-Porter HBC, Cerrato N, Chow TS, Como N, Eustace J, García PJ, Godbole S, Gotuzzo E, Griskevicius L, Hamra R, Hassan M, Hassany M, Hutton D, Irmansyah I, Jancoriene L, Kirwan J, Kumar S, Lennon P, Lopardo G, Lydon P, Magrini N, Maguire T, Manevska S, Manuel O, McGinty S, Medina MT, Mesa Rubio ML, Miranda-Montoya MC, Nel J, Nunes EP, Perola M, Portolés A, Rasmin MR, Raza A, Rees H, Reges PPS, Rogers CA, Salami K, Salvadori MI, Sinani N, Sterne JAC, Stevanovikj M, Tacconelli E, Tikkinen KAO, Trelle S, Zaid H, Røttingen JA, Swaminathan S. Repurposed Antiviral Drugs for Covid-19 Interim WHO Solidarity Trial Results. *New England Journal of Medicine* 2021;**384**:497-511.
- 48. Humeres C, Frangogiannis NG. Fibroblasts in the Infarcted, Remodeling, and Failing Heart. *Jacc-Basic Transl Sc* 2019;**4**:449-467.
- 49. Van Linthout S, Miteva K, Tschope C. Crosstalk between fibroblasts and inflammatory cells. *Cardiovascular Research* 2014;**102**:258-269.
- 50. Steffens S, Van Linthout S, Sluijter JPG, Tocchetti CG, Thum T, Madonna R. Stimulating proreparative immune responses to prevent adverse cardiac remodelling: consensus document from the joint 2019 meeting of the ESC Working Groups of cellular biology of the heart and myocardial function. *Cardiovascular Research* 2020;**116**:1850-1862.
- 51. Beltrami AP, Madeddu P. Pericytes and cardiac stem cells: Common features and peculiarities. *Pharmacol Res* 2018;**127**:101-109.
- 52. Amendola A, Garoffolo G, Songia P, Nardacci R, Ferrari S, Bernava G, Canzano P, Myasoedova V, Colavita F, Castilletti C, Sberna G, Capobianchi MR, Piacentini M, Agrifoglio M, Colombo GI, Poggio P, Pesce M. Human cardiosphere-derived stromal cells exposed to SARS-CoV-2 evolve into hyper-inflammatory/pro-fibrotic phenotype and produce infective viral particles depending on the levels of ACE2 receptor expression. *Cardiovascular Research* 2021;10.1093/cvr/cvab082.
- 53. McCracken IR, Saginc G, He L, Huseynov A, Daniels A, Fletcher S, Peghaire C, Kalna V, Andaloussi-Mäe M, Muhl L, Craig NM, Griffiths SJ, Haas JG, Tait-Burkard C, Lendahl U, Birdsey GM, Betsholtz C, Noseda M, Baker AH, Randi AM. Lack of Evidence of Angiotensin-Converting Enzyme 2 Expression and Replicative Infection by SARS-CoV-2 in Human Endothelial Cells. *Circulation* 2021;**143**:865-868.

- 54. Zhong P, Agosto LM, Munro JB, Mothes W. Cell-to-cell transmission of viruses. *Current Opinion in Virology* 2013;**3**:44-50.
- 55. Dosch SF, Mahajan SD, Collins AR. SARS coronavirus spike protein-induced innate immune response occurs via activation of the NF-κB pathway in human monocyte macrophages in vitro. *Virus Research* 2009;**142**:19-27.
- 56. Brandão SCS, Ramos JdOX, Dompieri LT, Godoi ETAM, Figueiredo JL, Sarinho ESC, Chelvanambi S, Aikawa M. Is Toll-like receptor 4 involved in the severity of COVID-19 pathology in patients with cardiometabolic comorbidities? *Cytokine & Growth Factor Reviews* 2021;**58**:102-110.
- 57. Ammirati E, Wang DW. SARS-CoV-2 inflames the heart. The importance of awareness of myocardial injury in COVID-19 patients. *INTERNATIONAL JOURNAL OF CARDIOLOGY* 2020;**311**:122-123.
- 58. Bussani R, Schneider E, Zentilin L, Collesi C, Ali H, Braga L, Volpe MC, Colliva A, Zanconati F, Berlot G, Silvestri F, Zacchigna S, Giacca M. Persistence of viral RNA, pneumocyte syncytia and thrombosis are hallmarks of advanced COVID-19 pathology. *EBioMedicine* 2020;**61**:103104.
- 59. Buchrieser J, Dufloo J, Hubert M, Monel B, Planas D, Rajah MM, Planchais C, Porrot F, Guivel-Benhassine F, Van der Werf S, Casartelli N, Mouquet H, Bruel T, Schwartz O. Syncytia formation by SARS-CoV-2-infected cells. *Embo Journal* 2020;**39**:e106267.
- 60. Braga L, Ali H, Secco I, Chiavacci E, Neves G, Goldhill D, Penn R, Jimenez-Guardeño JM, Ortega-Prieto AM, Bussani R, Cannatà A, Rizzari G, Collesi C, Schneider E, Arosio D, Shah AM, Barclay WS, Malim MH, Burrone J, Giacca M. Drugs that inhibit TMEM16 proteins block SARS-CoV-2 Spike-induced syncytia. *Nature* 2021;10.1038/s41586-021-03491-6.
- 61. Lamers MM, Beumer J, van der Vaart J, Knoops K, Puschhof J, Breugem TI, Ravelli RBG, van Schayck JP, Mykytyn AZ, Duimel HQ, van Donselaar E, Riesebosch S, Kuijpers HJH, Schipper D, van de Wetering WJ, de Graafl M, Koopmans M, Cuppen E, Peters PJ, Haagmans BL, Clevers H. SARS-CoV-2 productively infects human gut enterocytes. *Science* 2020;**369**:50-54.
- 62. Yang LL, Han YL, Nilsson-Payant BE, Gupta V, Wang PF, Duan XH, Tang XM, Zhu JJ, Zhao ZP, Jaffre F, Zhang T, Kim TW, Harschnitz O, Redmond D, Houghton S, Liu CY, Naji A, Ciceri G, Guttikonda S, Bram Y, Nguyen DHT, Cioffi M, Chandar V, Hoagland DA, Huang YX, Xiang J, Wang H, Lyden D, Borczuk A, Chen HJ, Studer L, Pan FC, Ho DD, tenOever BR, Evans T, Schwartz RE, Chen SB. A Human Pluripotent Stem Cell-based Platform to Study SARS-CoV-2 Tropism and Model Virus Infection in Human Cells and Organoids. *Cell Stem Cell* 2020;27:125-+.
- 63. Marrella A, Buratti P, Markus J, Firpo G, Pesenti M, Landry T, Ayehunie S, Scaglione S, Kandarova H, Aiello M. In Vitro Demonstration of Intestinal Absorption Mechanisms of Different Sugars Using 3D Organotypic Tissues in a Fluidic Device. *Altex-Altern Anim Ex* 2020;**37**:255-264.
- 64. Garoffolo G, Ferrari S, Rizzi S, Barbuto M, Bernava G, Pesce M. Harnessing Mechanosensation in Next Generation Cardiovascular Tissue Engineering. *Biomolecules* 2020;**10**:1419.
- 65. Garoffolo G, Madonna R, de Caterina R, Pesce M. Cell based mechanosensing in vascular patho-biology: More than a simple go-with the flow. *Vascular Pharmacology* 2018;**111**:7-14.
- 66. Itzhaki I, Maizels L, Huber I, Zwi-Dantsis L, Caspi O, Winterstern A, Feldman O, Gepstein A, Arbel G, Hammerman H, Boulos M, Gepstein L. Modelling the long QT syndrome with induced pluripotent stem cells. *Nature* 2011;**471**:225-U113.
- 67. Manolis AS, Manolis AA, Manolis TA, Apostolopoulos EJ, Papatheou D, Melita H. COVID-19 infection and cardiac arrhythmias. *Trends in Cardiovascular Medicine* 2020;**30**:451-460.

- 68. Stillitano F, Hansen J, Kong CW, Karakikes I, Faunck-Brentano C, Geng L, Scott S, Reynier S, Wu M, Valogne Y, Desseaux C, Salem JE, Jeziorowska D, Zahr N, Li R, Iyengar R, Hajjar RJ, Hulot JS. Modeling susceptibility to drug-induced long QT with a panel of subject-specific induced pluripotent stem cells. *eLife* 2017;6.
- 69. Dherange P, Lang J, Qian P, Oberfeld B, Sauer WH, Koplan B, Tedrow U. Arrhythmias and COVID-19 A Review. *Jacc-Clin Electrophy* 2020;**6**:1193-1204.
- 70. Canzano P, Brambilla M, Porro B, Cosentino N, Tortorici E, Vicini S, Poggio P, Cascella A, Pengo MF, Veglia F, Fiorelli S, Bonomi A, Cavalca V, Trabattoni D, Andreini D, Sale EO, Parati G, Tremoli E, Camera M. Platelet and Endothelial Activation as Potential Mechanisms Behind the Thrombotic Complications of COVID-19 Patients. *Jacc-Basic Transl Sc* 2021;**6**:202-218.
- 71. Blasco A, Coronado MJ, Hernandez-Terciado F, Martin P, Royuela A, Ramil E, Garcia D, Goicolea J, Del Trigo M, Ortega J, Escudier JM, Silva L, Bellas C. Assessment of Neutrophil Extracellular Traps in Coronary Thrombus of a Case Series of Patients With COVID-19 and Myocardial Infarction. *JAMA Cardiology* 2020;10.1001/jamacardio.2020.7308.
- 72. Pearce L, Davidson SM, Yellon DM. The cytokine storm of COVID-19: a spotlight on prevention and protection. *Expert Opinion on Therapeutic Targets* 2020;**24**:723-730.
- 73. Kuba K, Imai Y, Rao SA, Gao H, Guo F, Guan B, Huan Y, Yang P, Zhang YL, Deng W, Bao LL, Zhang BL, Liu G, Wang Z, Chappell M, Liu YX, Zheng DX, Leibbrandt A, Wada T, Slutsky AS, Liu DP, Qin CA, Jiang CY, Penninger JM. A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus-induced lung injury. *Nature Medicine* 2005;**11**:875-879.
- 74. Blanco-Melo D, Nilsson-Payant BE, Liu WC, Uhl S, Hoagland D, Moller R, Jordan TX, Oishi K, Panis M, Sachs D, Wang TT, Schwartz RE, Lim JK, Albrecht RA, tenOever BR. Imbalanced Host Response to SARS-CoV-2 Drives Development of COVID-19. *Cell* 2020;**181**:1036-+.
- 75. Lambert DW, Yarski M, Warner FJ, Thornhill P, Parkin ET, Smith AI, Hooper NM, Turner AJ. Tumor Necrosis Factor-α Convertase (ADAM17) Mediates Regulated Ectodomain Shedding of the Severe-acute Respiratory Syndrome-Coronavirus (SARS-CoV) Receptor, Angiotensin-converting Enzyme-2 (ACE2). *Journal of Biological Chemistry* 2005;**280**:30113-30119.
- 76. Rizzo P, Vieceli Dalla Sega F, Fortini F, Marracino L, Rapezzi C, Ferrari R. COVID-19 in the heart and the lungs: could we "Notch" the inflammatory storm? *Basic Res Cardiol* 2020;**115**:31.
- 77. Liu YX, Yang Y, Zhang C, Huang FM, Wang FX, Yuan J, Wang ZQ, Li JX, Li JM, Feng C, Zhang Z, Wang LF, Peng L, Chen L, Qin YH, Zhao DD, Tan SG, Yin L, Xu J, Zhou CZ, Jiang CY, Liu L. Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury. *Sci China Life Sci* 2020;**63**:364-374.
- 78. Kassiri Z, Zhong JC, Guo D, Basu R, Wang XH, Liu PP, Scholey JW, Penninger JM, Oudit GY. Loss of Angiotensin-Converting Enzyme 2 Accelerates Maladaptive Left Ventricular Remodeling in Response to Myocardial Infarction. *Circ-Heart Fail* 2009;**2**:446-455.
- 79. Imai Y, Kuba K, Neely GG, Yaghubian-Malhami R, Perkmann T, van Loo G, Ermolaeva M, Veldhuizen R, Leung YHC, Wang HL, Liu HL, Sun Y, Pasparakis M, Kopf M, Mech C, Bavari S, Peiris JSM, Slutsky AS, Akira S, Hultqvist M, Holmdahl R, Nicholls J, Jiang CY, Binder CJ, Penninger JM. Identification of oxidative stress and toll-like receptor 4 signaling as a key pathway of acute lung injury. *Cell* 2008;**133**:235-249.
- 80. de Queiroz TM, Lakkappa N, Lazartigues E. ADAM17-Mediated Shedding of Inflammatory Cytokines in Hypertension. *Frontiers in Pharmacology* 2020;**11**.
- 81. Gopinathan G, Milagre C, Pearce OMT, Reynolds LE, Hodivala-Dilke K, Leinster DA, Zhong H, Hollingsworth RE, Thompson R, Whiteford JR, Balkwill F. Interleukin-6 Stimulates Defective Angiogenesis. *Cancer research* 2015;**75**:3098-3107.

- 82. Alsaffar H, Martino N, Garrett JP, Adam AP. Interleukin-6 promotes a sustained loss of endothelial barrier function via Janus kinase-mediated STAT3 phosphorylation and de novo protein synthesis. *American Journal of Physiology-Cell Physiology* 2018;**314**:C589-C602.
- 83. Wassmann S, Stumpf M, Strehlow K, Schmid A, Schieffer B, Bohm M, Nickenig G. Interleukin-6 induces oxidative stress and endothelial dysfunction by overexpression of the angiotensin II type 1 receptor. *Circulation Research* 2004;**94**:534-541.
- 84. Coles B, Fielding CA, Rose-John S, Scheller J, Jones SA, O'Donnell VB. Classic interleukin-6 receptor signaling and interleukin-6 trans-signaling differentially control angiotensin II-dependent hypertension, cardiac signal transducer and activator of transcription-3 activation, and vascular hypertrophy in vivo. *AMERICAN JOURNAL OF PATHOLOGY* 2007;171:315-325.
- 85. Brasier AR. The nuclear factor-kappa B-interleukin-6 signalling pathway mediating vascular inflammation. *Cardiovascular Research* 2010;**86**:211-218.
- 86. Fontes JA, Rose NR, Cihakova D. The varying faces of IL-6: From cardiac protection to cardiac failure. *Cytokine* 2015;**74**:62-68.
- 87. Kanda T, McManus JEW, Nagai R, Imai S, Suzuki T, Yang DC, McManus BM, Kobayashi I. Modification of viral myocarditis in mice by interleukin-6. *Circulation Research* 1996;**78**:848-856.
- 88. Tanaka T, Kanda T, McManus BM, Kanai H, Akiyama H, Sekiguchi K, Yokoyama T, Kurabayashi M. Overexpression of Interleukin-6 Aggravates Viral Myocarditis: Impaired Increase in Tumor Necrosis Factor-α. *Journal of Molecular and Cellular Cardiology* 2001;**33**:1627-1635.
- 89. Savvatis K, Muller I, Frohlich M, Pappritz K, Zietsch C, Hamdani N, Grote K, Schieffer B, Klingel K, Van Linthout S, Linke WA, Schultheiss HP, Tschope C. Interleukin-6 receptor inhibition modulates the immune reaction and restores titin phosphorylation in experimental myocarditis. *Basic Res Cardiol* 2014;**109**:449.
- 90. Zhang Y, Xiao M, Zhang SL, Xia P, Cao W, Jiang W, Chen H, Ding X, Zhao H, Zhang HM, Wang CY, Zhao J, Sun XF, Tian R, Wu W, Wu D, Ma J, Chen Y, Zhang D, Xie J, Yan XW, Zhou X, Liu ZY, Wang JL, Du B, Qin Y, Gao P, Qin XZ, Xu YC, Zhang W, Li TS, Zhang FC, Zhao YQ, Li YZ, Zhang SY. Coagulopathy and Antiphospholipid Antibodies in Patients with Covid-19. *New England Journal of Medicine* 2020;**382**:e38.
- 91. Tay MZ, Poh CM, Renia L, MacAry PA, Ng LFP. The trinity of COVID-19: immunity, inflammation and intervention. *Nature Reviews Immunology* 2020;**20**:363-374.
- 92. Oberfeld B, Achanta A, Carpenter K, Chen P, Gilette NM, Langat P, Said JT, Schiff AE, Zhou AS, Barczak AK, Pillai S. SnapShot: COVID-19. *Cell* 2020;**181**:954-+.
- 93. Hermine O, Mariette X, Tharaux PL, Resche-Rigon M, Porcher R, Ravaud P, Grp C-C. Effect of Tocilizumab vs Usual Care in Adults Hospitalized With COVID-19 and Moderate or Severe Pneumonia: A Randomized Clinical Trial (vol 181, pg 32, 2021). *JAMA Internal Medicine* 2021;**181**:144-144.
- 94. Salama C, Han J, Yau LD, Reiss WG, Kramer B, Neidhart JD, Criner GJ, Kaplan-Lewis E, Baden R, Pandit L, Cameron ML, Garcia-Diaz J, Chavez V, Mekebeb-Reuter M, de Menezes FL, Shah R, Gonzalez-Lara MF, Assman B, Freedman J, Mohan SV. Tocilizumab in Patients Hospitalized with Covid-19 Pneumonia. *New England Journal of Medicine* 2021;**384**:20-30.
- 95. Parr JB. Time to Reassess Tocilizumab's Role in COVID-19 Pneumonia. *JAMA Internal Medicine* 2021;**181**:12.
- 96. Connors JM, Levy JH. COVID-19 and its implications for thrombosis and anticoagulation. *Blood* 2020;**135**:2033-2040.
- 97. Iba T, Levy JH, Levi M, Thachil J. Coagulopathy in COVID-19. *Journal of Thrombosis and Haemostasis* 2020;**18**:2103-2109.

- 98. Nicolai L, Leunig A, Brambs S, Kaiser R, Weinberger T, Weigand M, Muenchhoff M, Hellmuth JC, Ledderose S, Schulz H, Scherer C, Rudelius M, Zoller M, Hochter D, Keppler O, Teupser D, Zwissler B, von Bergwelt-Baildon M, Kaab S, Massberg S, Pekayvaz K, Stark K. Immunothrombotic Dysregulation in COVID-19 Pneumonia Is Associated With Respiratory Failure and Coagulopathy. *Circulation* 2020;**142**:1176-1189.
- 99. Guagliumi G, Sonzogni A, Pescetelli I, Pellegrini D, Finn AV. Microthrombi and ST-Segment-Elevation Myocardial Infarction in COVID-19. *Circulation* 2020;**142**:804-809.
- 100. Satoh K, Satoh T, Yaoita N, Shimokawa H. Recent Advances in the Understanding of Thrombosis. *Arterioscl Throm Vas* 2019;**39**:E159-E165.
- 101. Döring Y, Libby P, Soehnlein O. Neutrophil Extracellular Traps Participate in Cardiovascular Diseases. *Circulation Research* 2020;**126**:1228-1241.
- 102. Fuchs TA, Brill A, Duerschmied D, Schatzberg D, Monestier M, Myers DD, Wrobleski SK, Wakefield TW, Hartwig JH, Wagner DD. Extracellular DNA traps promote thrombosis. *Proceedings of the National Academy of Sciences of the United States of America* 2010;**107**:15880-15885.
- 103. von Brühl M-L, Stark K, Steinhart A, Chandraratne S, Konrad I, Lorenz M, Khandoga A, Tirniceriu A, Coletti R, Köllnberger M, Byrne RA, Laitinen I, Walch A, Brill A, Pfeiler S, Manukyan D, Braun S, Lange P, Riegger J, Ware J, Eckart A, Haidari S, Rudelius M, Schulz C, Echtler K, Brinkmann V, Schwaiger M, Preissner KT, Wagner DD, Mackman N, Engelmann B, Massberg S. Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice in vivo. *Journal of Experimental Medicine* 2012;209:819-835.
- 104. Martinod K, Witsch T, Erpenbeck L, Savchenko A, Hayashi H, Cherpokova D, Gallant M, Mauler M, Cifuni SM, Wagner DD. Peptidylarginine deiminase 4 promotes age-related organ fibrosis. *Journal of Experimental Medicine* 2017;**214**:439-458.
- 105. Zuo Y, Zuo M, Yalavarthi S, Gockman K, Madison JA, Shi H, Woodard W, Lezak SP, Lugogo NL, Knight JS, Kanthi Y. Neutrophil extracellular traps and thrombosis in COVID-19. *Journal of thrombosis and thrombolysis* 2021;**51**:446-453.
- 106. Middleton EA, He XY, Denorme F, Campbell RA, Ng D, Salvatore SP, Mostyka M, Baxter-Stoltzfus A, Borczuk AC, Loda M, Cody MJ, Manne BK, Portier I, Harris ES, Petrey AC, Beswick EJ, Caulin AF, Iovino A, Abegglen LM, Weyrich AS, Rondina MT, Egeblad M, Schiffman JD, Yost CC. Neutrophil extracellular traps contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome. *Blood* 2020;**136**:1169-1179.
- 107. Veras FP, Pontelli MC, Silva CM, Toller-Kawahisa JE, de Lima M, Nascimento DC, Schneider AH, Caetite D, Tavares LA, Paiva IM, Rosales R, Colon D, Martins R, Castro IA, Almeida GM, Lopes MIF, Benatti MN, Bonjorno LP, Giannini MC, Luppino-Assad R, Almeida SL, Vilar F, Santana R, Bollela VR, Auxiliadora-Martins M, Borges M, Miranda CH, Pazin A, da Silva LLP, Cunha LD, Zamboni DS, Dal-Pizzol F, Leiria LO, Li SY, Batah S, Fabro A, Mauad T, Dolhnikoff M, Duarte-Neto A, Saldiva P, Cunha TM, Alves JC, Arruda E, Louzada P, Oliveira RD, Cunha FQ. SARS-CoV-2-triggered neutrophil extracellular traps mediate COVID-19 pathology. *Journal of Experimental Medicine* 2020;**217**.
- 108. Zuo Y, Estes SK, Ali RA, Gandhi AA, Yalavarthi S, Shi H, Sule G, Gockman K, Madison JA, Zuo M, Yadav V, Wang JT, Woodard W, Lezak SP, Lugogo NL, Smith SA, Morrissey JH, Kanthi Y, Knight JS. Prothrombotic autoantibodies in serum from patients hospitalized with COVID-19. *Science translational medicine* 2020;**12**.
- 109. Urbanus RT. Recent Developments in Antiphospholipid Antibodies and the Antiphospholipid Syndrome Preface. *Seminars in Thrombosis and Hemostasis* 2018;**44**:417-418.

- 110. Kasal DA, De Lorenzo A, Tibiriçá E. COVID-19 and Microvascular Disease: Pathophysiology of SARS-CoV-2 Infection With Focus on the Renin-Angiotensin System. *Heart, Lung and Circulation* 2020;**29**:1596-1602.
- 111. Orfao A, Matarraz S, Perez-Andres M, Almeida J, Teodosio C, Berkowska MA, van Dongen JJM, EuroFlow. Immunophenotypic dissection of normal hematopoiesis. *Journal of Immunological Methods* 2019;**475**:112684.
- 112. Rios MR, Garoffolo G, Rinaldi G, Megia-Fernandez A, Ferrari S, Robb CT, Rossi AG, Pesce M, Bradley M. A fluorogenic peptide-based smartprobe for the detection of neutrophil extracellular traps and inflammation. *Chemical Communications* 2021;**57**:97-100.
- 113. Yu X, Tan J, Diamond SL. Hemodynamic force triggers rapid NETosis within sterile thrombotic occlusions. *Journal of Thrombosis and Haemostasis* 2018;**16**:316-329.
- 114. Lee YY, Park HH, Park W, Kim H, Jang JG, Hong KS, Lee JY, Seo HS, Na DH, Kim TH, Choy YB, Ahn JH, Lee W, Park CG. Long-acting nanoparticulate DNase-1 for effective suppression of SARS-CoV-2-mediated neutrophil activities and cytokine storm. *Biomaterials* 2021;**267**:120389.
- 115. Sawadogo SA, Dighero-Kemp B, Ouedraogo DD, Hensley L, Sakande J. How NETosis could drive "Post-COVID-19 syndrome" among survivors. *Immunology Letters* 2020;**228**:35-37.
- 116. Schulte-Schrepping J, Reusch N, Paclik D, Bassler K, Schlickeiser S, Zhang BW, Kramer B, Krammer T, Brumhard S, Bonaguro L, De Domenico E, Wendisch D, Grasshoff M, Kapellos TS, Beckstette M, Pecht T, Saglam A, Dietrich O, Mei HE, Schulz AR, Conrad C, Kunkel D, Vafadarnejad E, Xu CJ, Horne A, Herbert M, Drews A, Thibeault C, Pfeiffer M, Hippenstiel S, Hocke A, Muller-Redetzky H, Heim KM, Machleidt F, Uhrig A, de Jarcy LB, Jurgens L, Stegemann M, Glosenkamp CR, Volk HD, Goffinet C, Landthaler M, Wyler E, Georg P, Schneider M, Dang-Heine C, Neuwinger N, Kappert K, Tauber R, Corman V, Raabe J, Kaiser KM, Vinh MT, Rieke G, Meisel C, Ulas T, Becker M, Geffers R, Witzenrath M, Drosten C, Suttorp N, von Kalle C, Kurth F, Handler K, Schultze JL, Aschenbrenner AC, Li Y, Nattermann J, Sawitzki B, Saliba AE, Sander LE, Initiative DC-O. Severe COVID-19 Is Marked by a Dysregulated Myeloid Cell Compartment. *Cell* 2020;**182**:1419.
- 117. Ng LG, Ostuni R, Hidalgo A. Heterogeneity of neutrophils. *Nature Reviews Immunology* 2019;**19**:255-265.
- 118. Dinh HQ, Eggert T, Meyer MA, Zhu YFPP, Olingy CE, Llewellyn R, Wu RP, Hedrick CC. Coexpression of CD71 and CD117 Identifies an Early Unipotent Neutrophil Progenitor Population in Human Bone Marrow. *Immunity* 2020;**53**:319.
- 119. Kwok I, Becht E, Xia Y, Ng M, Teh YC, Tan L, Evrard M, Li JLY, Tran HTN, Tan YR, Liu DH, Mishra A, Liong KH, Leong K, Zhang YN, Olsson A, Mantri CK, Shyamsunder P, Liu ZY, Piot C, Dutertre CA, Cheng H, Bari S, Ang N, Biswas SK, Koeffler HP, Tey HL, Larbi A, Su IH, Lee B, John AS, Chan JKY, Hwang WYK, Chen JM, Salomonis N, Chong SZ, Grimes HL, Liu B, Hidalgo A, Newell EW, Cheng T, Ginhoux F, Ng LG. Combinatorial Single-Cell Analyses of Granulocyte-Monocyte Progenitor Heterogeneity Reveals an Early Uni-potent Neutrophil Progenitor. *Immunity* 2020;53:303.
- 120. Bastard P, Rosen LB, Zhang Q, Michailidis E, Hoffmann HH, Zhang Y, Dorgham K, Philippot Q, Rosain J, Beziat V, Manry J, Shaw E, Haljasmagi L, Peterson P, Lorenzo L, Bizien L, Trouillet-Assant S, Dobbs K, de Jesus AA, Belot A, Kallaste A, Catherinot E, Tandjaoui-Lambiotte Y, Le Pen J, Kerner G, Bigio B, Seeleuthner Y, Yang R, Bolze A, Spaan AN, Delmonte OM, Abers MS, Aiuti A, Casari G, Lampasona V, Piemonti L, Ciceri F, Bilguvar K, Lifton RP, Vasse M, Smadja DM, Migaud M, Hadjadj J, Terrier B, Duffy D, Quintana-Murci L, van de Beek DV, Roussel L, Vinh DC, Tangye SG, Haerynck F, Dalmau D, Martinez-Picado J, Brodin P, Nussenzweig MC, Boisson-Dupuis S, Rodriguez-Gallego C, Vogt G, Mogensen TH, Oler AJ, Gu JW, Burbelo PD,

- Cohen JI, Biondi A, Bettini LR, D'Angio M, Bonfanti P, Rossignol P, Mayaux J, Rieux-Laucat F, Husebye ES, Fusco F, Ursini MV, Imberti L, Sottini A, Paghera S, Quiros-Roldan E, Rossi C, Castagnoli R, Montagna D, Licari A, Marseglia GL, Duval X, Ghosn J, Tsang JS, Goldbach-Mansky R, Kisand K, Lionakis MS, Puel A, Zhang SY, Holland SM, Gorochov G, Jouangu E, Rice CM, Cobat A, Notarangelo LD, Abel L, Su HC, Casanova JL, Lab H, COVID N-UIR, Clinicians C, Clinicians C-S, Grp IC, Grp FCCS, Consortium MI, Cohort C-C, Biobank AUC-, Effort CHG. Autoantibodies against type I IFNs in patients with life-threatening COVID-19. *Science* 2020;**370**:423-+.
- 121. Bolivar S, Espitia-Corredor JA, Olivares-Silva F, Valenzuela P, Humeres C, Anfossi R, Castro E, Vivar R, Salas-Hernández A, Pardo-Jiménez V, Díaz-Araya G. In cardiac fibroblasts, interferonbeta attenuates differentiation, collagen synthesis, and TGF-β1-induced collagen gel contraction. *Cytokine* 2020;**138**:155359.
- 122. Ni WT, Yang XW, Liu J, Bao J, Li R, Xu Y, Guo W, Hu Y, Gao ZC. Acute Myocardial Injury at Hospital Admission Is Associated With All-Cause Mortality in COVID-19. *Journal of the American College of Cardiology* 2020;**76**:124-125.
- 123. Sandoval Y, Januzzi JL, Jaffe AS. Cardiac Troponin for Assessment of Myocardial Injury in COVID-19 JACC Review Topic of the Week. *Journal of the American College of Cardiology* 2020;**76**:1244-1258.
- 124. Omland T, Prebensen C, Roysland R, Sovik S, Sorensen V, Rosjo H, Svensson M, Berdal JE, Berdal JE, Myhre PL. Established Cardiovascular Biomarkers Provide Limited Prognostic Information in Unselected Patients Hospitalized With COVID-19. *Circulation* 2020;**142**:1878-1880.
- 125. Qin JJ, Cheng X, Zhou F, Lei F, Akolkar G, Cai JJ, Zhang XJ, Blet A, Xie J, Zhang P, Liu YM, Huang ZZ, Zhao LP, Lin LJ, Xia M, Chen MM, Song XH, Bai LJ, Chen Z, Zhang XY, Xiang D, Chen J, Xu QB, Ma XL, Touyz RM, Gao C, Wang HT, Liu LM, Mao WM, Luo PC, Yan YQ, Ye P, Chen MH, Chen GH, Zhu LH, She ZG, Huang XD, Yuan YF, Zhang BH, Wang YB, Liu PP, Li HL. Redefining Cardiac Biomarkers in Predicting Mortality of Inpatients With COVID-19. *Hypertension* 2020;**76**:1104-1112.
- 126. Myhre PL, Prebensen C, Strand H, Roysland R, Jonassen CM, Rangberg A, Sorensen V, Sovik S, Rosjo H, Svensson M, Berdal JE, Omland T. Growth Differentiation Factor 15 Provides Prognostic Information Superior to Established Cardiovascular and Inflammatory Biomarkers in Unselected Patients Hospitalized With COVID-19. *Circulation* 2020;**142**:2128-2137.
- 127. Michaud V, Deodhar M, Arwood M, Al Rihani SB, Dow P, Turgeon J. ACE2 as a Therapeutic Target for COVID-19; Its Role in Infectious Processes and Regulation by Modulators of the RAAS System. *Journal of Clinical Medicine* 2020;9:2096.
- 128. Gheblawi M, Wang K, Viveiros A, Nguyen Q, Zhong J-C, Turner AJ, Raizada MK, Grant MB, Oudit GY. Angiotensin-Converting Enzyme 2: SARS-CoV-2 Receptor and Regulator of the Renin-Angiotensin System. *Circulation Research* 2020;**126**:1456-1474.
- 129. Danser AHJ, Epstein M, Batlle D. Renin-Angiotensin System Blockers and the COVID-19 Pandemic At Present There Is No Evidence to Abandon Renin-Angiotensin System Blockers. *Hypertension* 2020;**75**:1382-1385.
- 130. Narula S, Yusuf S, Chong M, Ramasundarahettige C, Rangarajan S, Bangdiwala SI, van Eikels M, Leineweber K, Wu A, Pigeyre M, Pare G. Plasma ACE2 and risk of death or cardiometabolic diseases: a case-cohort analysis. *Lancet* 2020;**396**:968-976.
- 131. Wallentin L, Lindback J, Eriksson N, Hijazi Z, Eikelboom JW, Ezekowitz MD, Granger CB, Lopes RD, Yusuf S, Oldgren J, Siegbahn A. Angiotensin-converting enzyme 2 (ACE2) levels in relation to risk factors for COVID-19 in two large cohorts of patients with atrial fibrillation. *European Heart Journal* 2020;**41**:4037-4046.

- 132. Kornilov SA, Lucas I, Jade K, Dai CZL, Lovejoy JC, Magis AT. Plasma levels of soluble ACE2are associated with sex, Metabolic Syndrome, and its biomarkers in a large cohort, pointing to a possible mechanism for increased severity in COVID-19. *Critical Care* 2020;**24**:452-452.
- 133. Pairo-Castineira E, Clohisey S, Klaric L, Bretherick AD, Rawlik K, Pasko D, Walker S, Parkinson N, Fourman MH, Russell CD, Furniss J, Richmond A, Gountouna E, Wrobel N, Harrison D, Wang B, Wu Y, Meynert A, Griffiths F, Oosthuyzen W, Kousathanas A, Moutsianas L, Yang ZJ, Zhai RR, Zheng CQ, Grimes G, Beale R, Millar J, Shih B, Keating S, Zechner M, Haley C, Porteous DJ, Hayward C, Yang J, Knight J, Summers C, Shankar-Hari M, Klenerman P, Turtle L, Ho A, Moore SC, Hinds C, Horby P, Nichol A, Maslove D, Ling L, McAuley D, Montgomery H, Walsh T, Pereira AC, Renieri A, Shen X, Ponting CP, Fawkes A, Tenesa A, Caulfield M, Scott R, Rowan K, Murphy L, Openshaw PJM, Semple MG, Law A, Vitart V, Wilson JF, Baillie JK, Investigators G, Investigators IC, Initiative C-HG, Investigators a, Investigators B, Investigators G-C. Genetic mechanisms of critical illness in COVID-19. *Nature* 2021;**591**:92-+.
- 134. Pérez-Bermejo JA, Kang S, Rockwood SJ, Simoneau CR, Joy DA, Ramadoss GN, Silva AC, Flanigan WR, Li H, Nakamura K, Whitman JD, Ott M, Conklin BR, McDevitt TC. SARS-CoV-2 infection of human iPSC-derived cardiac cells predicts novel cytopathic features in hearts of COVID-19 patients. *bioRxiv* 2020;10.1101/2020.08.25.265561.
- 135. Devi A, Chaitanya NSN. Targeting SARS CoV2 (Indian isolate) genome with miRNA: an in silico study. *IUBMB Life* 2020;**72**:2454-2468.
- 136. Bhattacharyya P, Biswas SC. Small Non-coding RNAs: Do They Encode Answers for Controlling SARS-CoV-2 in the Future? *Frontiers in Microbiology* 2020;**11**.
- 137. Nishiga M, Wang DW, Han YL, Lewis DB, Wu JC. COVID-19 and cardiovascular disease: from basic mechanisms to clinical perspectives. *Nature Reviews Cardiology* 2020;**17**:543-558.
- 138. Li CX, Hu X, Li LL, Li JH. Differential microRNA expression in the peripheral blood from human patients with COVID-19. *Journal of Clinical Laboratory Analysis* 2020;**34**:e23590.
- 139. Backes C, Meese E, Keller A. Specific miRNA Disease Biomarkers in Blood, Serum and Plasma: Challenges and Prospects. *Molecular Diagnosis & Therapy* 2016;**20**:509-518.
- 140. Di BA, Jia HL, Luo OJ, Lin FQ, Li KBA, Zhang YL, Wang HD, Liang HY, Fan J, Yang ZC. Identification and validation of predictive factors for progression to severe COVID-19 pneumonia by proteomics. *Signal Transduction and Targeted Therapy* 2020;**5**:217.
- 141. Yu HX, Li CY, Wang X, Duan JY, Yang N, Xie LJ, Yuan Y, Li SZ, Bi CH, Yang B, Li YB. Techniques and Strategies for Potential Protein Target Discovery and Active Pharmaceutical Molecule Screening in a Pandemic. *Journal of Proteome Research* 2020;**19**:4242-4258.
- 142. Demichev V, Tober-Lau P, Nazarenko T, Thibeault C, Whitwell H, Lemke O, Röhl A, Freiwald A, Szyrwiel L, Ludwig D, Correia-Melo C, Helbig ET, Stubbemann P, Grüning N-M, Blyuss O, Vernardis S, White M, Messner CB, Joannidis M, Sonnweber T, Klein SJ, Pizzini A, Wohlfarter Y, Sahanic S, Hilbe R, Schaefer B, Wagner S, Mittermaier M, Machleidt F, Garcia C, Ruwwe-Glösenkamp C, Lingscheid T, de Jarcy LB, Stegemann MS, Pfeiffer M, Jürgens L, Denker S, Zickler D, Enghard P, Zelezniak A, Campbell A, Hayward C, Porteous DJ, Marioni RE, Uhrig A, Müller-Redetzky H, Zoller H, Löffler-Ragg J, Keller MA, Tancevski I, Timms JF, Zaikin A, Hippenstiel S, Ramharter M, Witzenrath M, Suttorp N, Lilley K, Mülleder M, Sander LE, Ralser M, Kurth F. A time-resolved proteomic and diagnostic map characterizes COVID-19 disease progression and predicts outcome. medRxiv 2020;10.1101/2020.11.09.20228015:2020.2011.2009.20228015.
- 143. Sirpilla O, Bauss J, Gupta R, Underwood A, Qutob D, Freeland T, Bupp C, Carcillo J, Hartog N, Rajasekaran S, Prokop JW. SARS-CoV-2-Encoded Proteome and Human Genetics: From Interaction-Based to Ribosomal Biology Impact on Disease and Risk Processes. *Journal of Proteome Research* 2020;**19**:4275-4290.

- 144. Bojkova D, Klann K, Koch B, Widera M, Krause D, Ciesek S, Cinatl J, Munch C. Proteomics of SARS-CoV-2-infected host cells reveals therapy targets. *Nature* 2020;**583**:469-+.
- 145. Blasco H, Bessy C, Plantier L, Lefevre A, Piver E, Bernard L, Marlet J, Stefic K, Benz-de Bretagne I, Cannet P, Lumbu H, Morel T, Boulard P, Andres CR, Vourc'h P, Herault O, Guillon A, Emond P. The specific metabolome profiling of patients infected by SARS-COV-2 supports the key role of tryptophan-nicotinamide pathway and cytosine metabolism. *Scientific Reports* 2020;**10**:16824.
- 146. Bruzzone C, Bizkarguenaga M, Gil-Redondo R, Diercks T, Arana E, de Vicuna AG, Seco M, Bosch A, Palazon A, San Juan I, Lain A, Gil-Martinez J, Bernardo-Seisdedos G, Fernandez-Ramos D, Lopitz-Otsoa F, Embade N, Lu S, Mato JM, Millet O. SARS-CoV-2 Infection Dysregulates the Metabolomic and Lipidomic Profiles of Serum. *iScience* 2020;**23**:101645.
- 147. Moreno Fernández-Ayala DJ, Navas P, López-Lluch G. Age-related mitochondrial dysfunction as a key factor in COVID-19 disease. *Experimental Gerontology* 2020;**142**:111147.
- 148. Thum T. SARS-CoV-2 receptor ACE2 expression in the human heart: cause of a post-pandemic wave of heart failure? *European Heart Journal* 2020;**41**:1807-1809.
- 149. Leist SR, Dinnon KH, Schafer A, Tse LV, Okuda K, Hou YXJ, West A, Edwards CE, Sanders W, Fritch EJ, Gully KL, Scobey T, Brown AJ, Sheahan TP, Moorman NJ, Boucher RC, Gralinski LE, Montgomery SA, Baric RS. A Mouse-Adapted SARS-CoV-2 Induces Acute Lung Injury and Mortality in Standard Laboratory Mice. Cell 2020;183:1070-+.
- 150. Cleary SJ, Pitchford SC, Amison RT, Carrington R, Robaina Cabrera CL, Magnen M, Looney MR, Gray E, Page CP. Animal models of mechanisms of SARS-CoV-2 infection and COVID-19 pathology. *British Journal of Pharmacology* 2020;**177**:4851-4865.
- 151. Viveiros A, Rasmuson J, Vu J, Mulvagh SL, Yip CYY, Norris CM, Oudit GY. Sex differences in COVID-19: candidate pathways, genetics of ACE2, and sex hormones. *American Journal of Physiology-Heart and Circulatory Physiology* 2021;**320**:H296-H304.
- 152. Ritter O, Kararigas G. Sex-Biased Vulnerability of the Heart to COVID-19. *Mayo Clinic Proceedings* 2020;**95**:2332-2335.
- 153. Bienvenu LA, Noonan J, Wang XW, Peter K. Higher mortality of COVID-19 in males: sex differences in immune response and cardiovascular comorbidities. *Cardiovascular Research* 2020;**116**:2197-2206.
- 154. Bae S, Kim SR, Kim MN, Shim WJ, Park SM. Impact of cardiovascular disease and risk factors on fatal outcomes in patients with COVID-19 according to age: a systematic review and meta-analysis. *Heart (British Cardiac Society)* 2021;**107**:373.
- 155. Gadi N, Wu SC, Spihlman AP, Moulton VR. What's Sex Got to Do With COVID-19? Gender-Based Differences in the Host Immune Response to Coronaviruses. *Frontiers in immunology* 2020;**11**.
- 156. Robinson EL, Alkass K, Bergmann O, Maguire JJ, Roderick HL, Davenport AP. Genes encoding ACE2, TMPRSS2 and related proteins mediating SARS-CoV-2 viral entry are upregulated with age in human cardiomyocytes. *Journal of Molecular and Cellular Cardiology* 2020;**147**:88-91.
- 157. Takahashi T, Ellingson MK, Wong P, Israelow B, Lucas C, Klein J, Silva J, Mao TY, Oh JE, Tokuyama M, Lu PW, Venkataraman A, Park A, Liu FM, Meir A, Sun J, Wang EY, Casanovas-Massana A, Wyllie AL, Vogels CBF, Earnest R, Lapidus S, Ott IM, Moore AJ, Shaw A, Fournier JB, Odio CD, Shelli F, Dela Cruz C, Grubaugh ND, Schulz WL, Ring AM, Ko AI, Omer SB, Iwasaki A, Team YIR. Sex differences in immune responses that underlie COVID-19 disease outcomes. *Nature* 2020;**588**:315-320.
- 158. Takahashi T, Iwasaki A. Sex differences in immune responses. *Science* 2021;**371**:347-348.
- 159. Perrino C, Ferdinandy P, Botker HE, Brundel BJJM, Collins P, Davidson SM, den Ruijter HM, Engel FB, Gerdts E, Girao H, Gyongyosi M, Hausenloy DJ, Lecour S, Madonna R, Marber M,

Murphy E, Pesce M, Regitz-Zagrosek V, Sluijter JPG, Steffens S, Gollmann-Tepekoylu C, Van Laake LW, Van Linthout S, Schulz R, Ytrehus K. Improving translational research in sexspecific effects of comorbidities and risk factors in ischaemic heart disease and cardioprotection: position paper and recommendations of the ESC Working Group on Cellular Biology of the Hear. *Cardiovascular Research* 2021;**117**:367-385.

Figures

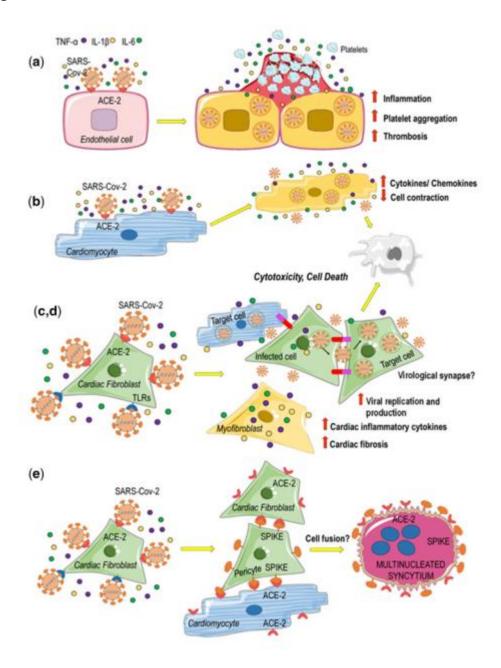


Figure 1. Proposed mechanisms of myocardial damage by SARS-CoV-2. (a) Infection of endothelial cells and exposure to circulating cytokines may cause increased platelet activation and micro-thrombosls. In the heart, this can lead to diffuse clotting, causing conditions similar to type-2 myocardial infarction²⁴. (b) Direct damage of cardiomyocytes may occur as a result of ACE2 receptor expression. Direct infection of these cells may cause decreased cardiac contractility and cell death due to cardiotoxicity¹⁶. (c, d) Infection of stromal cells, recently demonstrated *in vitro*⁵², could damage the heart through infection-independent differentiation into myofibroblasts and ACE2-dependent intracellular replication. In addition to the release of the virus in the intercellular space by infected and death cells, the possibility of spreading the virus to adjacent cells by the so-called 'viral synapse'⁴³ is discussed as a further modality of virus propagation in cardiac tissue and myocardial

injury. **(e)** Formation of multinucleated syncytia⁵⁹ might cause extensive cardiac damage due to the fusion of contractile and non-contractile myocardial cells, both of which express ACE2 receptor. This is still an experimentally unsupported mechanism of cardiac damage that could be addressed with in vitro studies using patient-derived and iPSCs-derived cardiac cells.

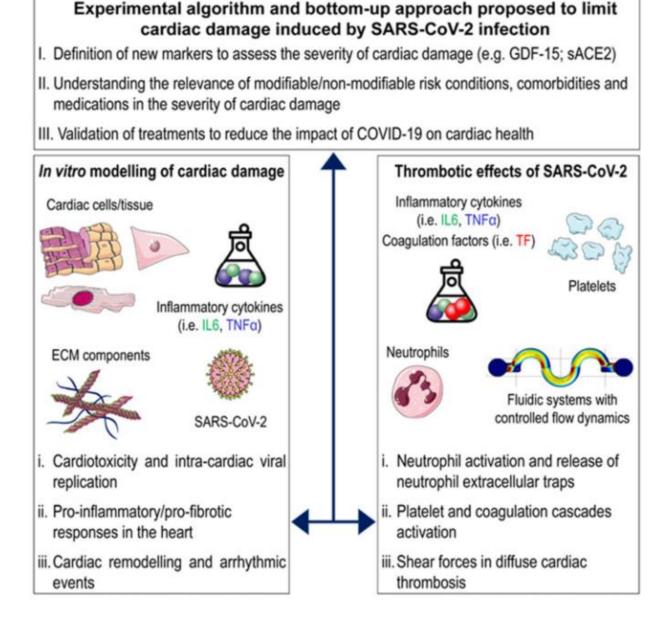


Figure 2. Experimental algorithm and bottom-up approach proposed to limit cardiac damage induced by SARS-CoV-2 infection. The disease caused by SARS-CoV-2 infection leads to a sharp elevation in the level of circulating inflammatory cytokines and to an increase in thrombotic events, especially in the microcirculation. The left box describes materials that could be combined in vitro to systematically approach the problem of cardiac damage from direct infection of myocardial cells, or by mimicking the pro-inflammatory effects of the cytokine storm. This approach aims to *i*) clarify

the cardiotoxicity and intra-cardiac viral replication of the virus, *ii*) assess the pro-inflammatory/pro-fibrotic responses in the heart due to direct/indirect effects and, *iii*) dissect cardiac remodelling and arrhythmic events. The box on the right indicate materials and tools that could be used to investigate in vitro the problem of the hypercoagulation found in COVID-19 patients with reference to *i*) neutrophil activation and release of the neutrophil extracellular traps, *ii*) mechanisms of platelet and coagulation cascades activation and, *iii*) the effects of shear forces in COVID-19-dependent diffuse cardiac thrombosis. The results emerging from these two experimental research areas should be finally integrated (arrows) with results from 'omics research performed directly with patient samples, and with results of epidemiologic/genetic studies and clinical trials. The aims of this bottom-up approach are, *i*) to define new markers to assess the severity of cardiac damage, *ii*) to understand the relevance of modifiable/non-modifiable risk conditions, comorbidities and drugs in the severity of cardiac damages and *iii*) to validate treatments reducing the impact of COVID-19 on cardiac health.