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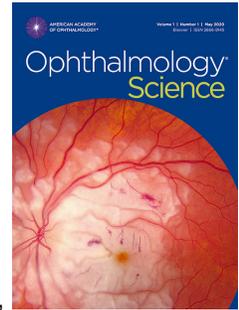
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Title

Comparing retinal structure in patients with achromatopsia and blue cone monochromacy using optical coherence tomography

Short title

Achromatopsia vs blue cone monochromacy: SD-OCT comparison

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52 **Abbreviations**

53 ACHM = achromatopsia

54 BCM = blue cone monochromacy

55 ELM = external limiting membrane

56 ERG = electroretinogram

57 EZ = ellipsoid zone

58 LCR = locus control region

59 LRP = longitudinal reflectivity profile

60 S-cone = short-wavelength-sensitive cone

61 SD-OCT = spectral domain optical coherence tomography

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62 Abstract

63 **Purpose:** To compare foveal hypoplasia and the appearance of the ellipsoid zone (EZ) at the
64 fovea in patients with genetically confirmed achromatopsia (ACHM) and blue cone
65 monochromacy (BCM).

66 **Design:** Retrospective, multi-center observational study.

67 **Subjects:** Molecularly confirmed patients with ACHM (n = 89) and BCM (n = 33).

68 **Methods:** We analyzed high-resolution spectral domain optical coherence tomography (SD-
69 OCT) images of the macula from aforementioned patients with BCM. Three observers
70 independently graded SD-OCT images for foveal hypoplasia (i.e. retention of one or more inner
71 retinal layers at the fovea) and four observers judged the integrity of the EZ at the fovea, based
72 on an established grading scheme. These measures were compared with previously published
73 data from the ACHM patients.

74 **Main Outcome Measures:** Presence of foveal hypoplasia and EZ grade.

75 **Results:** Foveal hypoplasia was significantly more prevalent in ACHM than in BCM
76 ($p < 0.001$). In addition, we observed a significant difference in the distribution of EZ grades
77 between ACHM and BCM, with grade II EZ being by far the most common phenotype in BCM
78 (61% of patients). In contrast, ACHM patients had a relatively equal prevalence of EZ grades
79 I, II, and IV. Interestingly, grade IV EZ was 2.6 times more prevalent in ACHM compared to
80 BCM, while grade V EZ (macular atrophy) was present in 3% of both the ACHM and BCM
81 cohorts.

82 **Conclusions:** The higher incidence of foveal hypoplasia in ACHM than BCM supports a role
83 for cone activity in foveal development. Although there are differences in EZ grades between
84 these conditions, the degree of overlap suggests EZ grade is not sufficient for definitive
85 diagnosis, in contrast to previous reports. Analysis of additional OCT features in similar
86 cohorts may reveal differences with greater diagnostic value. Finally, the extent to which foveal

87 hypoplasia or EZ grade is prognostic for therapeutic potential in either group remains to be
88 seen, but motivates further study.

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89 **Introduction**

90 Achromatopsia (ACHM) and blue cone monochromacy (BCM) are two congenital cone
91 dysfunction syndromes that are of great interest due to the emergence of novel therapeutic
92 approaches leading to clinical trials. While patients with ACHM typically lack function of all
93 three cone types, patients with BCM retain function of their short-wavelength-sensitive cones
94 (which comprise only 7-10% of the normal total cone population). Although ACHM is
95 autosomal recessive and BCM is X-linked, the inheritance pattern is not always clearly
96 discernible, especially in smaller families with few affected individuals. Moreover, clinical
97 symptoms are similar between the two pathologies, and inconsistent nomenclature throughout
98 the literature poses a further challenge to their differentiation.¹⁻⁴ As a result, diagnosis is not
99 straightforward, particularly in clinics that do not have access to, or funds for, genetic testing
100 or other specialized assessments. Accounting for the estimated prevalence of the known
101 underlying genetic causes of ACHM (40-50% *CNGB3*; 20-30% *CNGA3*; < 2% *GNAT2*;⁵
102 *PDE6C* and *PDE6H*)^{6,7} it is estimated that the genetic cause of at least 15% of ACHM cases
103 remains unknown (although some of these cases may represent missed intronic variants or
104 even misdiagnosed BCM);⁸ thus there is a need to develop methods to better differentiate these
105 conditions clinically.

106 Literature examining clinical differences in these populations is sparse,⁹⁻¹¹ especially
107 in molecularly-confirmed patients. Some differences in visual function have been found
108 between ACHM and BCM, but with limited discriminative abilities. Differences between these
109 groups have been found in eye movements using electro-oculography,¹² as well as in cone
110 responses using electroretinography (ERG),^{4,11} although ERG presentation in BCM and both
111 *GNAT2* and *PDE6C* -related ACHM can be similar, due to preservation of short-wavelength
112 sensitivity.^{13,14} Moreover, the procedures are not feasible for all patients, especially children,

113 and photopic ERG stimuli can be particularly uncomfortable for some patients, due to the
114 photoaversion that is characteristic of both conditions.

115 Color vision testing can offer a less vexatious alternative, with differences between
116 ACHM and BCM being evident on the Sloan achromatopsia test,¹⁵ albeit with limited
117 reliability, as well as the Berson test.^{10,16,17} However, the accuracy of any functional test is
118 dependent upon patient concentration and cooperation. Even for patients who perform reliably,
119 detection of any subtle differences in visual performance requires specialized expertise and
120 equipment, specific lighting conditions, and calibration of stimuli, making such methods
121 impracticable in most clinics. However, methods to assess cone structure that are widely
122 available, less dependent on patient performance, and readily interpreted, may offer an
123 alternative approach for discriminating BCM from ACHM.

124 Spectral domain optical coherence tomography (SD-OCT) is used widely in clinical
125 settings and enables visualization of the retinal layers as distinct reflective bands. The second
126 hyperreflective outer retinal band has been shown to correspond to photoreceptor integrity, and
127 the reflective signal has been hypothesized to originate from either the mitochondria-rich
128 ellipsoid zone (EZ), or the junction between the inner and outer segment of photoreceptors. For
129 simplicity, we hereon refer to the second band as the EZ. Discontinuities in the EZ have been
130 observed at the fovea in patients with BCM, suggesting disruption of photoreceptor
131 structure.^{11,18-20} Similarly, there is variable disruption of the EZ at the fovea in patients with
132 ACHM (ranging from normal-appearing to complete absence). While this variability does not
133 correlate with visual function,²¹ it does broadly correlate with remnant foveal cone density, as
134 assessed using adaptive optics imaging.²² Comparison between the two pathologies using
135 longitudinal reflectivity profile (LRP) analysis of time-domain OCT images showed reduced
136 total foveal thickness in BCM compared to ACHM,¹¹ although subsequent SD-OCT studies
137 have reported retinal thinning in both BCM and ACHM.^{18,23} In addition, Barthelmes et al.

138 (2006)¹¹ reported an absence of the EZ in ACHM and an absence of the external limiting
139 membrane (ELM) in BCM, suggesting this is an absolute biomarker for distinguishing the two
140 conditions. Importantly, the patients used in that study were not genotyped, but instead were
141 classified using best-corrected visual acuity, ERG and color-plate testing.

142 Here we use SD-OCT to assess foveal hypoplasia and the appearance of the EZ at the
143 fovea in patients with genetically confirmed BCM, and compare with previously reported data
144 from patients with genetically confirmed ACHM.

145

146 **Methods**

147 **Patients**

148 Images from 33 male patients with genetically confirmed BCM were used for analysis. The
149 genotype and clinical phenotype for each patient is shown in **Table 1**. Thirteen patients had a
150 deletion of the locus control region (LCR) and 20 had the Cys203Arg substitution affecting the
151 only opsin gene or at least the first two genes in the *OPNILW/OPNIMW* array. LCR deletions
152 preclude expression of all *OPNILW/OPNIMW* genes, while genes with the Cys203Arg mutant
153 encode a nonfunctional opsin that is toxic to the cones that express it. ACHM data for 89
154 patients was drawn from two previously published studies: 38 patients with *CNGA3*-related
155 ACHM (21 M; 17 F) from Georgiou et al. (2019)²⁴ and 51 with *CNGB3*-related ACHM (30 M;
156 21 F) from Langlo et al. (2016).²² This study followed the tenets of the Declaration of Helsinki
157 and was approved by local institutional review boards (MCW: PRO17439 & PRO30741;
158 UCL/Moorfields reference: 67979). Informed consent was obtained from all patients, after the
159 nature and possible consequences of the study were explained.

160

161 **SD-OCT Imaging**

162 High resolution SD-OCT images of the macula were acquired using the Bioptigen Envisu
163 R2200 (MCW) or C2300 (UCL/Moorfields) SD-OCT systems (Leica Microsystems). High
164 density horizontal line scans (either 750 or 1000 A-scans/B-scan, 100–150 repeated B scans)
165 were acquired through the foveal center. Line scans were registered and averaged to reduce
166 speckle noise in the image, as previously described.²⁵ Images from both eyes for each patient
167 were reviewed by a single rater (EJP) and the eye with better image quality was then selected
168 for further analysis. For the patients with ACHM, SD-OCT images from the right eye of
169 patients included in two previously reported studies were used for analysis.^{22,24}

170 For the patients with BCM, foveal hypoplasia was assessed in a binary fashion (*i.e.*,
171 presence or absence) independently by three raters (EJP, CSL, MG), with the consensus grade
172 being used for all images. For the patients with ACHM, their previously reported foveal
173 hypoplasia status was used in our analysis. For the patients with BCM, the EZ integrity at the
174 fovea was assessed by four raters (EJP, CSL, MG, JC). We used Sundaram et al's (2014)²¹ five
175 categories for grading, whereby: I) continuous EZ, II) EZ disruption, III) EZ absence, IV)
176 presence of a hyporeflective zone, or V) outer retinal atrophy (including loss of retinal pigment
177 epithelium). Any assessment that did not reach a consensus across raters was reviewed and
178 discussed by EJP and JC for a final determination. For the patients with ACHM, their
179 previously reported EZ grade was used in our analysis. Statistical analysis was performed using
180 GraphPad Prism (version 9.0.0, GraphPad Software, La Jolla, CA), R (The R Foundation,
181 Vienna, Austria) and SAS (version 9.4, The SAS Institute, Cary, NC). A Shapiro-Wilk test was
182 used to test for normality. As the data was found to have a non-normal distribution, non-
183 parametric tests were used to test for statistical significance.

184

185 **Results**

186 Foveal hypoplasia judgements were identical between eyes for all BCM patients. EZ grading
187 was identical between eyes for all BCM patients except JC_11033, whose right eye was graded
188 as grade V and left eye as grade III by a single rater (EJP), demonstrating high interocular
189 symmetry in BCM. The eye with better image quality was used for further analysis. Foveal
190 hypoplasia judgements were also identical between eyes for all ACHM patients. Four of 51
191 ACHM patients had interocular differences in EZ grade, again demonstrating high interocular
192 symmetry.

193

194 **Foveal Hypoplasia**

195 Sixty-two out of the total 89 ACHM patients (70%) had foveal hypoplasia, compared to 11 out
196 of 33 BCM patients (33%). Examples of foveal hypoplasia in ACHM and BCM are shown in
197 **Figure 1**. A Fisher's Exact test revealed that foveal hypoplasia was significantly more
198 prevalent in ACHM than BCM ($p < 0.001$). Within each condition, we found no association
199 between the underlying genotype and the prevalence of hypoplasia (ACHM: *CNGA3* vs.
200 *CNGB3*, $p = 0.64$; BCM: LCR deletions vs. Cys203Arg, $p = 0.71$).

201 Given that the majority of ACHM patients had foveal hypoplasia and the majority of
202 BCM patients did not, it was of interest to determine the predictive value of the presence of
203 hypoplasia. The sensitivity of foveal hypoplasia as a diagnostic sign for differentiating
204 between ACHM and BCM was 70% (95% confidence interval {CI} = 59%-78%) and the
205 specificity was 67% (95% CI = 50%-80%), with a positive predictive value of 85% (95% CI =
206 75%-91%) and negative predictive value of 45% (95% CI = 32%-59%).

207

208 **EZ Integrity**

209 A breakdown of the relative prevalence of the different EZ grades within BCM and ACHM is
210 shown in **Figure 2**. Of note is the large proportion of BCM patients with grade II EZ (61%)

211 compared to ACHM (36%), as well as the higher prevalence of grade I and IV in ACHM (25%
212 and 31% respectively) than BCM (12% and 12%), and of grade III in BCM (12%) than ACHM
213 (4%). Grade V accounted for 3% of retinas for both ACHM and BCM. A Fisher's Exact test
214 revealed a significant difference in the distribution of grades between pathologies ($p = 0.02$),
215 with a Cramér's V yielding a moderate effect size of 0.30.

216 Due to the low prevalence of EZ grades III and V, patients with these grades were
217 excluded from the following analysis. The distribution of EZ grades between pathologies
218 remained significantly different ($p = 0.01$, Pearson's Chi-Square test), with a Cramér's V
219 yielding an effect size of 0.28. Grades I and IV were significantly more prevalent in ACHM
220 than BCM ($p < 0.004$, Fisher's Exact test). The sensitivity of grades I and IV as a diagnostic
221 sign of ACHM was 61% (95% CI = 50%-72%) and the specificity was 71% (95% CI = 51%-
222 87%), with a PPV of 86% (95% CI = 75%-94%) and NPV of 39% (95% CI = 25%-54%).

223 Multivariable exact logistic regression showed that both hypoplasia ($p = 0.004$) and EZ
224 grade (with 3 levels, $p = 0.026$) had significant predictive value when controlling for the other
225 factor. The area under the curve in the multivariate model was 0.669 for hypoplasia (95% CI =
226 0.566-0.772), 0.667 for EZ grade (95% CI = 0.564-0.771), and 0.743 with both factors
227 combined (95% CI = 0.642-0.844), which represented a significantly better predictive value
228 than either factor alone ($p < 0.0001$). Examination of the classification table allows evaluation
229 of sensitivity and specificity when using a decision rule based on a given cut-point probability
230 of ACHM (Table 2).

231

232 **Examining Possible Sex Differences**

233 All BCM patients were male, so it was important to establish that sex differences in the ACHM
234 group were not contributing to any differences found between conditions. A Fisher's Exact test
235 showed no statistically significant difference in the prevalence of foveal hypoplasia between

236 males and females across the ACHM group ($p = 0.17$). In addition, there was no significant
237 difference in age between ACHM and BCM groups ($p = 0.46$, Mann-Whitney test). Thus the
238 differences in hypoplasia and grade distribution between ACHM and BCM appear to be due to
239 differences in the underlying disease mechanism.

240

241 **Discussion**

242 In this study we compared patients with genetically confirmed BCM and ACHM, to determine
243 whether their SD-OCT images revealed distinguishable features that could aid differential
244 diagnosis between the two patient populations. We found moderate differences in the
245 distribution of EZ grades between ACHM and BCM, with ACHM patients being more likely
246 than BCM to have grade I or IV EZ, and BCM patients being more likely than ACHM to have
247 grade II or III EZ. In contrast to Barthelmes et al. (2006),¹¹ who reported absence of the EZ
248 (which they labelled P2) and presence of the ELM (which they labelled P3) in all ACHM
249 patients, we observed several cases of EZ presence in ACHM, and three cases of ELM absence
250 (all grade V). The same study reported the opposite pattern for all BCM patients, a presence of
251 the EZ (their P2) and absence of the ELM (their P3); however, we observed several cases of
252 EZ absence, and noted ELM presence in all but one BCM patient, who had macular atrophy
253 (grade V). We believe that it is very unlikely for all six of Barthelmes' BCM patients to have
254 lacked ELM while retaining EZ. Of the four bands they measured, the ELM (their P3) typically
255 yields the smallest LRP peak; this, combined with the poorer lateral and axial resolution of
256 time-domain OCT (compared to SD-OCT), as well as the inherent difficulty of obtaining sharp
257 images in these populations, may have led to misidentification of retinal bands in some
258 patients. In addition, they used the LRP at a single, precisely placed retinal location for grading
259 the EZ, as opposed to the holistic EZ grading used in our study. Many BCM patients have a
260 focal disruption of the EZ (**Figure 3**, JC_10558), which is hypothesized to represent the S-cone

261 free zone,¹⁸ although this disruption does not always align axially with the foveal reflex (**Figure**
262 **3**, JC_0184) and therefore LRP analysis at the foveal center may miss a bona fide EZ
263 disruption. More generally, dependence of LRP measurements on the precise placement of the
264 LRP makes analysis susceptible to variation due to differences in signal, tilt in the OCT scan,
265 or a lack of scanning frames at the exact foveal center. Furthermore, the steps required to
266 overcome these issues often necessitate post-acquisition manipulation, which is not feasible in
267 the clinic. Thus, while a categorical grading scheme has its own disadvantages, we feel it
268 provides a more accurate depiction of the EZ status of a given fovea than the isolated LRP
269 approach.

270 We also found that patients with ACHM were significantly more likely to have foveal
271 hypoplasia than patients with BCM. Barthelmes et al. (2006)¹¹ did not explicitly comment on
272 hypoplasia, however the broader internal limiting membrane peak (which they called P4)
273 reported in ACHM than both normal and BCM suggests that their P4 may also have
274 incorporated other inner retinal bands, such as the plexiform layers; this thereby makes it highly
275 likely that hypoplasia was present in their ACHM population. The finding that foveal
276 hypoplasia is more prevalent in ACHM than BCM has important implications for the
277 mechanisms underlying human foveal development. In the immature eye, all the retinal layers
278 are still present at the fovea.²⁶ Histological and *in vivo* studies have shown a lateral shift of
279 inner retinal layers away from the fovea *in utero*, which continues throughout the first few
280 months after birth.^{27,28} Its failure to occur in most ACHM patients suggests that cone function
281 helps to guide this process. Additionally, the finding that peripheral migration of inner retinal
282 layers occurs in most BCM patients suggests that retained function of a single minority cone
283 class may be sufficient to prevent severe hypoplasia. The fact that S-opsin expression precedes
284 L/M opsin and rhodopsin expression, as well as foveal cone migration and Henle fiber
285 elongation, lends support for this hypothesis.^{29,30}

286 One issue raised in the process of conducting this study is the ambiguity in classifying
287 OCT images. For example, the extent to which the EZ must be “disrupted” to warrant a grade
288 II (as opposed to grade I) is arguable and, to some extent, arbitrary – must the disruption extend
289 the full height of the EZ band at the fovea (**Figure 3**, MP_10097 and JC_11237), or is it
290 sufficient for it to simply have altered reflectivity (**Figure 3**, MM_0186)? Differentiating
291 between grades II and IV can be particularly problematic. Literature using Sundaram’s (2014)²¹
292 grading scheme appears to classify a vitread bowing of the ELM (in combination with a
293 hyporeflective zone) as grade IV, although this is not explicitly stated. One feature often
294 observed in BCM is a small “pocket” of hyporeflectivity at or near the fovea (**Figure 3**,
295 JC_10558) – the threshold at which this pocket becomes a hyporeflective “zone” is not clearly
296 defined. Moreover, many patients with BCM lack a foveal bulge,²⁰ whereby the ELM inclines
297 inwards (i.e. upwards in our images) at the foveal center. This feature (**Figure 3**, JC_0184), or
298 lack thereof (**Figure 3**, MP_10100), may influence one’s interpretation of the term,
299 “hyporeflective zone”, which is used to describe the foveal cavitation in grade IV. This grading
300 scheme may therefore be less suitable for BCM than for ACHM in its current form, but could
301 perhaps benefit from further clarification within each grading category. Foveal cavitation has
302 been observed in a number of inherited retinal dystrophies,^{31,32} and is likely to be indicative of
303 outer segment loss,³¹ rather than cone loss, as adaptive optics imaging has revealed remnant
304 inner segments within these areas.²² Future work combining OCT with *en face* adaptive optics
305 imaging may help to elucidate the cellular origin of abnormal patterns of reflectivity observed
306 in OCT, particularly in the photoreceptor layers. Such clarity could facilitate the development
307 of anatomically and clinically relevant grading schemes.

308 One notable limitation of the current study is that differences between pathologies may
309 have been lost through binary classification of foveal hypoplasia. Although not assessed
310 quantitatively, it was noted that there was a trend towards a greater number or thickness of

311 preserved inner retinal layers at the fovea in ACHM than in BCM (**Figure 1**). Not only does
312 binary assessment ignore this potentially important difference, but it also increases uncertainty
313 when categorising images from BCM patients. Future work may benefit from quantifying the
314 number or thickness of retained inner retinal layers, which could be facilitated by utilizing
315 directional OCT. The reflectivity of the Henle fiber layer changes depending on the pupil entry
316 position, which could help to disambiguate hypoplasia judgements. Furthermore, given recent
317 advances in deep learning techniques and their successful application to ocular images, it is
318 also possible that by using training data consisting of SD-OCT images classified simply by
319 genotype, a convolutional neural network may be able to distinguish between the pathologies.

320 Accurate diagnosis is critical, not only for the welfare of the individual patient but also
321 for estimations of disease prevalence. There has been renewed interest in congenital cone
322 disorders, thanks to recent advances in gene therapy efforts to restore cone function. However,
323 motivation to target a given disease will be influenced by its prevalence. The prevalence of
324 each pathology has been somewhat “lost in translation” throughout the literature; no doubt
325 exacerbated by ambiguous descriptions and use of terms,^{1-3,33} as well as a misunderstanding of
326 the genetic origin in earlier work. BCM has variably been referred to as “incomplete” or
327 “atypical” achromatopsia, although both terms have also been used to describe different
328 conditions. Estimates for “total color blindness” (*i.e.*, ACHM and BCM combined) range from
329 1/20,000 to 1/100,000 of the total population,^{33,34} with the majority consisting of autosomal
330 recessive ACHM.¹ BCM is generally considered to affect around 1/100,000 individuals,³⁵
331 although early estimates quote as few as 1/100 million people,² and even 1/100 million
332 percent.¹ Misdiagnosis of BCM for ACHM could potentially contribute to an underestimation
333 of BCM, making it a less favorable target for gene therapy efforts. It is therefore crucial to
334 ensure accurate diagnosis and to continually update estimates of prevalence based on emerging
335 research.

336 Despite our finding that the distribution of EZ grades is significantly different between
337 diseases and that foveal hypoplasia is more prevalent in ACHM than BCM, these population
338 differences likely cannot be used to definitively diagnose an individual patient, in contrast to
339 previous reports.¹¹ However, OCT findings could be used to guide diagnosis or decisions
340 concerning genetic testing, as *OPNILW/OPNIMW* sequencing is not widespread. Moreover,
341 as our understanding of how OCT disruptions relate to the underlying cone structure improves,
342 accurate classification/grading of images will be of great importance in interpreting progressive
343 changes or responses to therapeutic intervention.

344 **References**

- 345 1. Pitt FHG. Monochromatism. *Nature*. 1944;154:466-468.
- 346 2. Krill AE. Congenital color vision defects. In: Krill AE, Archer DB, eds. *Krill's*
347 *Hereditary Retinal and Choroidal Diseases, Vol II. Clinical Characteristics*. Harper &
348 Row; 1977:355-390.
- 349 3. Pokorny J, Smith VC, Verriest G. Congenital color defects. In: Pokorny J, Smith VC,
350 Verriest G, Pinckers AJLG, eds. *Congenital and Acquired Color Vision Defects*. Grune
351 & Stratton; 1979:183-241.
- 352 4. Andréasson S, Tornqvist K. Electroretinograms in patients with achromatopsia. *Acta*
353 *Ophthalmol*. 2009;69(6):711-716.
- 354 5. Kohl S, Baumann B, Rosenberg T, et al. Mutations in the cone photoreceptor G-
355 protein α -subunit gene GNAT2 in patients with achromatopsia. *Am J Hum Genet*.
356 2002;71(2):422-425.
- 357 6. Thiadens AAHJ, den Hollander AI, Roosing S, et al. Homozygosity mapping reveals
358 PDE6C mutations in patients with early-onset cone photoreceptor disorders. *Am J Hum*
359 *Genet*. 2009;85(2):244-247.
- 360 7. Kohl S, Coppieters F, Meire F, et al. A nonsense mutation in PDE6H causes
361 autosomal-recessive incomplete achromatopsia. *Am J Hum Genet*. 2012;91(3):527-
362 532.
- 363 8. Weisschuh N, Sturm M, Baumann B, et al. Deep- intronic variants in CNGB3 cause
364 achromatopsia by pseudoexon activation. *Hum Mutat*. 2020;41(1):255-264.
- 365 9. Haegerstrom-Portnoy G, Schneck ME, Verdon WA, Hewlett SE. Clinical vision
366 characteristics of the congenital achromatopsias. I. Visual acuity, refractive error, and
367 binocular status. *Optom Vis Sci*. 1996;73(7):446-456.
- 368 10. Haegerstrom-Portnoy G, Schneck ME, Verdon WA, Hewlett SE. Clinical vision

- 369 characteristics of the congenital achromatopsias. II. Color vision. *Optom Vis Sci.*
370 1996;73(7):457-465.
- 371 11. Barthelmes D, Sutter FK, Kurz-Levin MM, et al. Quantitative analysis of OCT
372 characteristics in patients with achromatopsia and blue-cone monochromatism. *Invest*
373 *Ophthalmol Vis Sci.* 2006;47(3):1161-1166.
- 374 12. Yee RD, Farley MK, Bateman JB, Martin DA. Eye movement abnormalities in rod
375 monochromatism and blue-cone monochromatism. *Graefe's Arch Clin Exp*
376 *Ophthalmol.* 1985;223(2):55-59.
- 377 13. Michaelides M, Aligianis IA, Holder GE, et al. Cone dystrophy phenotype associated
378 with a frameshift mutation (M280fsX291) in the α -subunit of cone specific transducin
379 (GNAT2). *Br J Ophthalmol.* 2003;87(11):1317-1320.
- 380 14. Georgiou M, Robson AG, Singh N, et al. Deep phenotyping of PDE6C-associated
381 achromatopsia. *Invest Ophthalmol Vis Sci.* 2019;60(15):5112-5123.
- 382 15. Sloan LL. Congenital achromatopsia: a report of 19 cases. *J Opt Soc Am.*
383 1954;44(2):117-127.
- 384 16. Berson EL, Sandberg MA, Rosner B, Sullivan PL. Color plates to help identify
385 patients with blue cone monochromatism. *Am J Ophthalmol.* 1983;95(6):741-747.
- 386 17. Pinckers A. Berson test for blue cone monochromatism. *Int Ophthalmol.*
387 1992;16(3):185-186.
- 388 18. Carroll J, Dubra A, Gardner JC, et al. The effect of cone opsin mutations on retinal
389 structure and the integrity of the photoreceptor mosaic. *Invest Ophthalmol Vis Sci.*
390 2012;53(13):8006-8015.
- 391 19. Cideciyan A V., Hufnagel RB, Carroll J, et al. Human cone visual pigment deletions
392 spare sufficient photoreceptors to warrant gene therapy. *Hum Gene Ther.*
393 2013;24(12):993-1006.

- 394 20. Sumaroka A, Garafalo A V., Cideciyan A V., et al. Blue cone monochromacy caused
395 by the C203R missense mutation or large deletion mutations. *Invest Ophthalmol Vis*
396 *Sci.* 2018;59(15):5762-5772.
- 397 21. Sundaram V, Wilde C, Aboshiha J, et al. Retinal structure and function in
398 achromatopsia. *Ophthalmology.* 2014;121(1):234-245.
- 399 22. Langlo CS, Patterson EJ, Higgins BP, et al. Residual foveal cone structure in CNGB3-
400 associated achromatopsia. *Invest Ophthalmol Vis Sci.* 2016;57(10):3984-3995.
- 401 23. Thiadens AAHJ, Somervuo V, van den Born LI, et al. Progressive Loss of cones in
402 achromatopsia: an imaging study using spectral-domain optical coherence
403 tomography. *Invest Ophthalmol Vis Sci.* 2010;51(11):5952-5957.
- 404 24. Georgiou M, Litts KM, Kalitzeos A, et al. Adaptive optics retinal imaging in CNGA3-
405 associated achromatopsia: retinal characterization, interocular symmetry, and
406 intrafamilial variability. *Invest Ophthalmol Vis Sci.* 2019;60(1):383-396.
- 407 25. Tanna H, Dubis AM, Ayub N, et al. Retinal imaging using commercial broadband
408 optical coherence tomography. *Br J Ophthalmol.* 2010;94(3):372-376.
- 409 26. Vinekar A, Mangalesh S, Jayadev C, Maldonado RS, Bauer N, Toth CA. Retinal
410 imaging of infants on spectral domain optical coherence tomography. *Biomed Res Int.*
411 2015;2015:782420.
- 412 27. Hendrickson AE, Yuodelis C. The morphological development of the human fovea.
413 *Ophthalmology.* 1984;91(6):603-612.
- 414 28. Maldonado RS, O'Connell R V., Sarin N, et al. Dynamics of human foveal
415 development after premature birth. *Ophthalmology.* 2011;118(12):2315-2325.
- 416 29. Cornish EE, Xiao M, Yang Z, Provis JM, Hendrickson AE. The role of opsin
417 expression and apoptosis in determination of cone types in human retina. *Exp Eye Res.*
418 2004;78(6):1143-1154.

- 419 30. Hendrickson A, Zhang C. Development of cone photoreceptors and their synapses in
420 the human and monkey fovea. *J Comp Neurol*. 2019;527(1):38-51.
- 421 31. Leng T, Marmor MF, Kellner U, et al. Foveal cavitation as an optical coherence
422 tomography finding in central cone dysfunction. *Retina*. 2012;32(7):1411-1419.
- 423 32. Parodi MB, Cicinelli MV, Iacono P, Bolognesi G, Bandello F. Multimodal imaging of
424 foveal cavitation in retinal dystrophies. *Graefe's Arch Clin Exp Ophthalmol*.
425 2017;255(2):271-279.
- 426 33. Judd DB. Facts of color-blindness. *J Opt Soc Am*. 1943;33(6):294-307.
- 427 34. Nathans J, Davenport CM, Maumenee IH, et al. Molecular genetics of human blue
428 cone monochromacy. *Science*. 1989;245(4920):831-838.
- 429 35. Gardner JC, Michaelides M, Holder GE, et al. Blue cone monochromacy: causative
430 mutations and associated phenotypes. *Mol Vis*. 2009;15:876-884.

431 **Figure Captions**

432 **Figure 1:** Examples of foveal hypoplasia in ACHM and BCM. Shown are processed
433 Bioptigen SD-OCT images of two patients with *CNGA3*-related ACHM and two patients
434 with Cys203Arg-related BCM. Subjective assessment reveals that foveal hypoplasia is more
435 severe in ACHM than BCM, as there is greater retention of inner retinal layers. Images in this
436 figure were rotated to negate tilt for aesthetic purposes.

437

438 **Figure 2:** Percentage of each EZ grade in ACHM and BCM. The frequency of each grade is
439 shown within or above each bar. We observed a significant difference in the distribution of
440 grades between ACHM and BCM, with a grade II EZ being the commonest phenotype in
441 BCM. ACHM patients were more than twice as likely to have a grade IV EZ than BCM,
442 suggesting that functional S-cones in BCM may help to prevent development of a
443 hyporeflective zone at the fovea.

444

445 **Figure 3:** Examples of OCT images demonstrating the significant heterogeneity of grade II EZ
446 in BCM. **MP_10097** and **JC_11237** are fairly typical examples of grade II, with both patients
447 having disruption that extends the full height of the EZ, although **MP_10097** has a focal
448 disruption and **JC_11237** shows broader mottling of the EZ. There was some debate as to
449 whether **MM_0186** was grade I or II as, although there was a small focal disruption of the EZ
450 just nasal of the foveal center, it did not extend the full height of the band. It was decided that
451 any altered reflectivity constituted “EZ disruption”. **JC_10558** has a small pocket of
452 hyporeflectivity, which may represent the S-cone free zone. There was contention between
453 graders as to whether **JC_0184** was grade II or IV, as the region of hyporeflectivity is small,
454 and it was debatable as to whether the ELM was bowing upwards (which would indicate grade
455 IV) or whether it had a normal contour (indicating grade II). Although BCM patients often lack

456 the foveal bulge, it was decided that JC_0184 had a normal ELM contour. **MP_10100** had
457 abnormal hyperreflectivity between the EZ and ELM, which gives the impression of a dipping
458 ELM (perhaps indicating grade III), but it was decided that the ELM was intact, leaving the
459 source of the abnormal hyperreflectivity unclear.

460

461 **Supplemental Figure 1:** Pedigrees for Families 5, 9, 16 and 17, as indicated in Table 1.

462 Asterisks denote patients included in this study.

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Table 1 - A summary of the genotype and clinical phenotype of subjects with Blue Cone Monochromacy

Family	Subject	Age (yrs)	Disease-causing variant	Eye	OCT Grade	Foveal hypoplasia
F1	JC_0078	27	LCR deletion	OS	3	No
F2	MM_0223	13	LCR deletion	OS	2	Yes
F3	JC_0611	34	LCR deletion	OD	3	Yes
F4	JC_0613	14	LCR deletion	OD	2	No
F5: IV-1	JC_0909†	7	LCR deletion	OS	2	Yes
F5: III-4	JC_0911†	41	LCR deletion	OD	2	No
F5: II-8	JC_0912†	58	LCR deletion	OS	4	No
F6	KS_10992	25	LCR deletion	OD	2	No
F7	JC_11033	53	LCR deletion	OS	3	No
F8	JC_11230	8	LCR deletion	OS	2	Yes
F9: IV-3	JC_11237†	6	LCR deletion	OD	2	Yes
F9: II-1	JC_11239†	75	LCR deletion	OS	3	No
F9: III-8	JC_11266†	35	LCR deletion	OS	2	No
F10	MM_0151	54	M _{C203R}	OD	5	No
F11	MM_0177	10	M _{C203R}	OD	1	No
F12	JC_0183*	24	M _{C203R}	OD	2	No
F12	JC_0184*	21	M _{C203R}	OS	2	No
F13	MM_0187	21	M _{C203R}	OD	1	Yes
F14	MM_0235	16	M _{C203R}	OD	2	No
F15	JC_11532*	49	M _{C203R}	OS	2	No
F15	JC_11585*	54	M _{C203R}	OS	4	No
F16: IV-1	JC_10066†	24	L _{C203R} -L _{C203R}	OS	2	No
F16: IV-3	JC_10067†	13	L _{C203R} -L _{C203R}	OD	2	No
F16: III-7	MP_10100†	35	L _{C203R} -L _{C203R}	OS	2	No
F17: IV-7	MP_10097†	43	L _{C203R} -M _{C203R}	OS	2	Yes
F17: V-2	MP_10116†	10	L _{C203R} -M _{C203R} ‡	OS	1	Yes
F18	MM_0186	11	M _{C203R} -M _{C203R}	OD	2	No
F19	JC_0440*	18	M _{C203R} -M _{C203R}	OD	2	Yes
F19	JC_0441*	18	M _{C203R} -M _{C203R}	OS	2	No
F20	JC_10557*	16	M _{C203R} -M _{C203R}	OS	4	No
F20	JC_10558*	16	M _{C203R} -M _{C203R}	OD	2	Yes
F21	JC_10561	50	M _{C203R} -M _{C203R}	OS	4	Yes
F22	JC_11919	20	M _{C203R} -M _{C203R}	OD	1	No

C203R = Cys203Arg. Yrs = years.

For simplicity, only the first two genes within the *OPN1LW/OPN1MW* array are reported.

* The following are brothers: JC_0183 and JC_0184; JC_11532 and JC_11585; JC_0440 and JC_0441; JC_10557 and JC_10558.

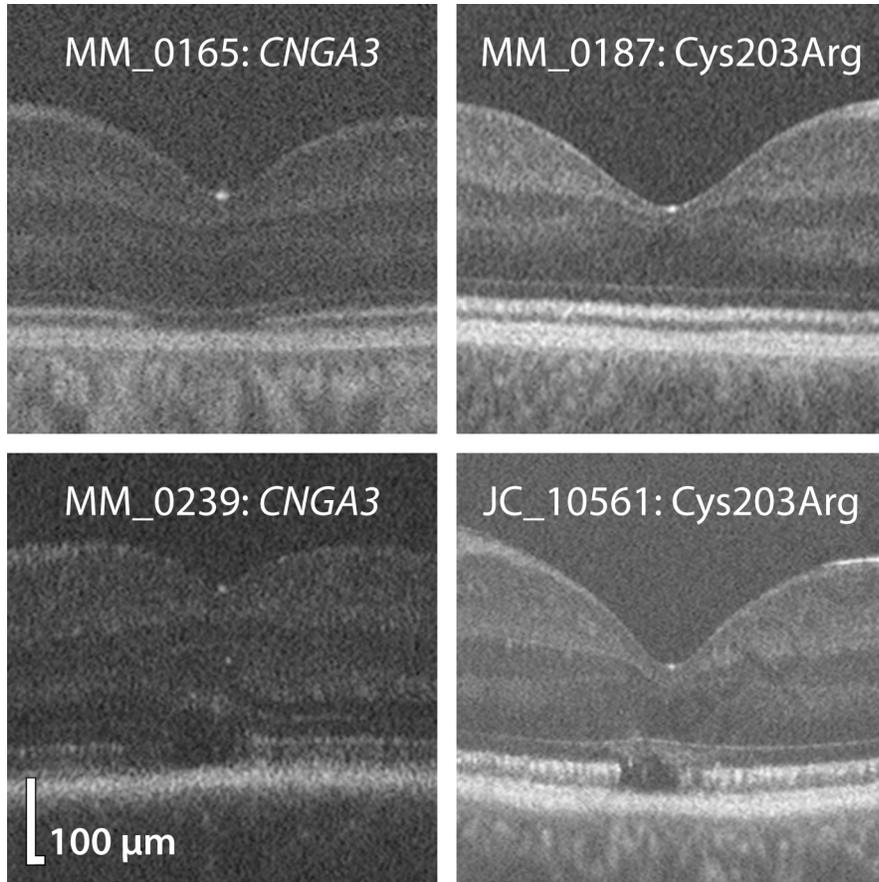
† Pedigrees shown in Supplemental Figure 1.

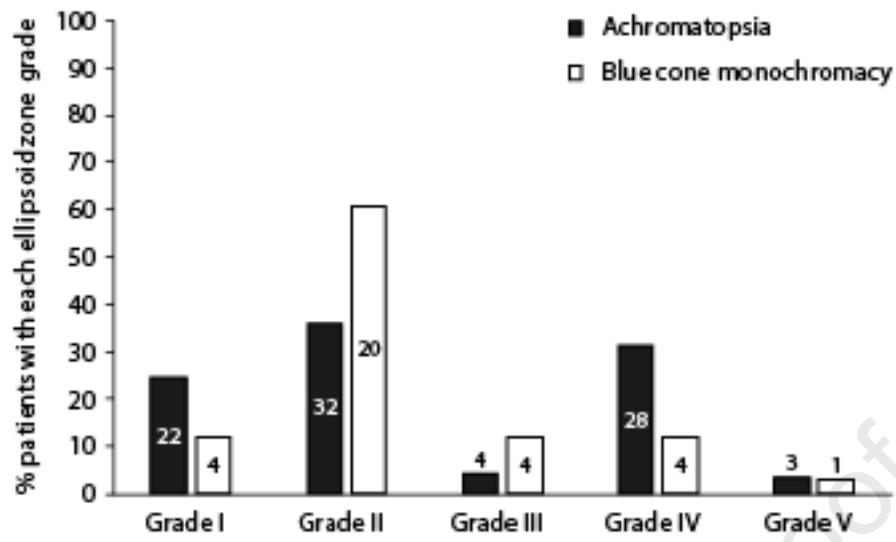
‡ Genotype inferred from MP_10097.

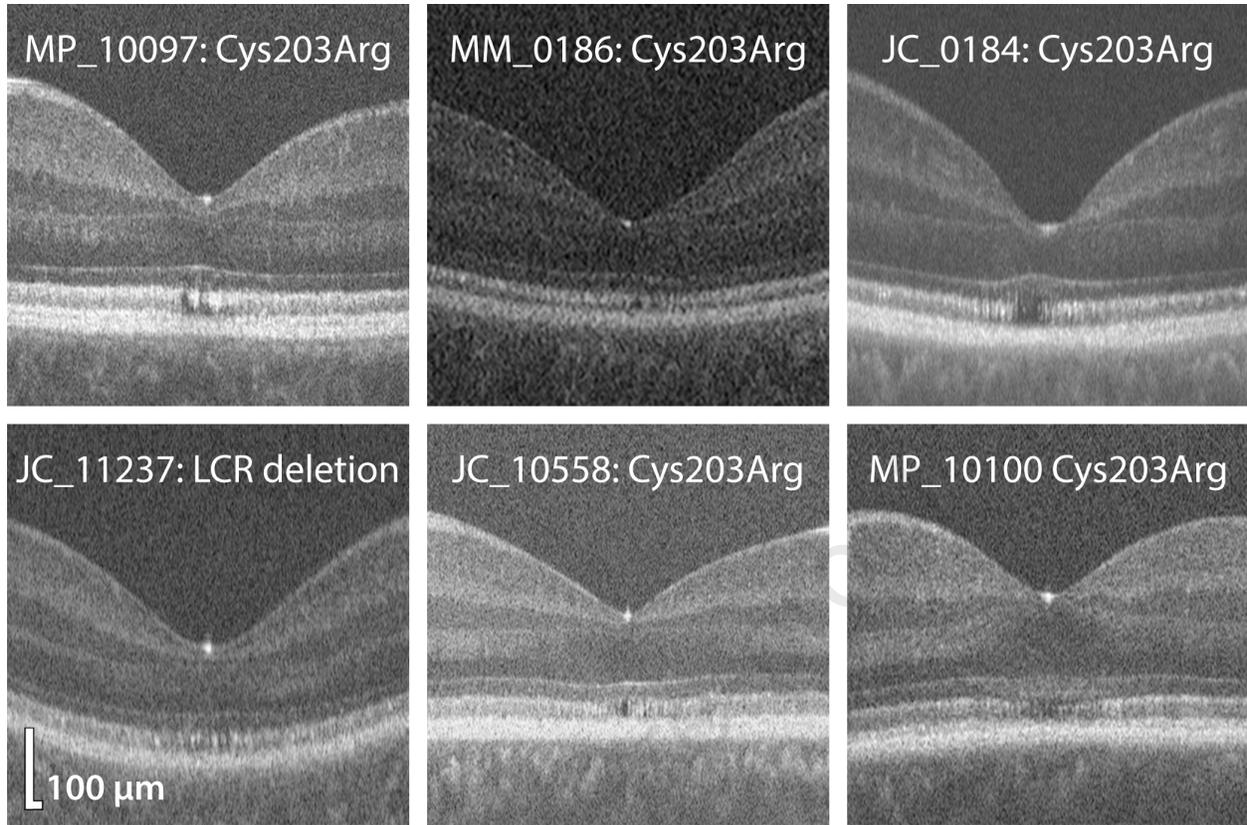
Table 2 - Classification table from multivariate logistic regression

Hypoplasia	EZ grade	n	Predicted probability of ACHM	Sensitivity	Specificity	PPV	NPV	Sensitivity + specificity
No	1, 2, or 4	23	0.4372	1.0000	0.0000	0.7455	--	1.0000
No	1 or 4	11	0.7442	0.8780	0.4643	0.8276	0.5652	1.3423
No	4	9	0.7510	0.7683	0.5357	0.8289	0.4412	1.3040
Yes	1, 2, or 4	29	0.7567	0.6951	0.6429	0.8507	0.4186	1.3380
Yes	1 or 4	15	0.9209	0.4268	0.8929	0.9211	0.3472	1.3197
Yes	4	23	0.9235	0.2683	0.9643	0.9565	0.3103	1.2326

Rows are ordered by predicted probability of achromatopsia (ACHM). Sensitivity, specificity, positive predictive value (PPV), and negative predictive value (NPV) apply to a decision rule based on a cut-point probability. For example, a cut-point at $p = 0.7567$ predicts that all patients with hypoplasia and any ellipsoid zone (EZ) grade have ACHM with sensitivity = 69.5%, specificity = 64.3%, PPV = 85.1%, and NPV = 41.9%. A cut-point at $p = 0.7442$ minimized classification error (which is statistically optimal, although may not be clinically optimal).







Précis

Optical coherence tomography reveals greater prevalence of foveal hypoplasia in achromatopsia than blue cone monochromacy, as well as significant differences in ellipsoid zone integrity between conditions.

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