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Cardiac-induced liver deformation as a measure of liver stiffness using dynamic imaging without magnetisation tagging – preclinical proof-of-concept, clinical translation, reproducibility and feasibility in patients with cirrhosis --Manuscript Draft--

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Abstract:	<p>Purpose</p> <p>MR elastography and magnetisation-tagging use liver stiffness (LS) measurements to diagnose fibrosis but require physical drivers, specialist sequences and post-processing. Here we evaluate non-rigid registration of dynamic two-dimensional cine MRI images to measure cardiac-induced liver deformation (LD) as a measure of LS by (i) assessing preclinical proof-of-concept, (ii) clinical reproducibility and inter-reader variability, (iii) the effects of hepatic haemodynamic changes and (iv) feasibility in patients with cirrhosis.</p> <p>Methods</p> <p>Sprague-Dawley rats (n=21 bile duct ligated (BDL), n=17 sham-operated controls) and</p>	

	<p>fasted patients with liver cirrhosis (n=11) and healthy volunteers (HVs, n=10) underwent spoiled gradient-echo short-axis cardiac cine MRI studies at 9.4T (rodents) and 3.0T (humans). LD measurements were obtained from intrahepatic sub-cardiac regions-of-interest close to the diaphragmatic margin. One-week reproducibility and prandial-stress induced haemodynamic changes were assessed in healthy volunteers.</p> <p>Results</p> <p>Normalised LD was higher in BDL (1.304±0.062) compared with sham-operated rats (1.058±0.045, P=0.0031). HV seven-day reproducibility Bland-Altman (BA) limits-of-agreement (LoAs) were ±0.028 a.u. and inter-reader variability BA LoAs were ±0.030 a.u.. Post-prandial LD increases were non-significant (+0.0083±0.0076 a.u., P=0.3028) and uncorrelated with PV flow changes (r=0.42, p=0.2219). LD measurements successfully obtained from all patients were not significantly higher in cirrhotics (0.102±0.0099 a.u.) compared with HVs (0.080±0.0063 a.u., P=0.0847).</p> <p>Conclusion</p> <p>Cardiac-induced LD is a conceptually reasonable approach from preclinical studies, measurements demonstrate good reproducibility and inter-reader variability, are less likely to be affected by hepatic haemodynamic changes and are feasible in patients with cirrhosis.</p>
<p>Response to Reviewers:</p>	<p>Detailed in-line response to the reviewer comments can be found in the supplementary material, uploaded in a separate word document.</p>

Title:

Cardiac-induced liver deformation as a measure of liver stiffness using dynamic imaging without magnetisation tagging – preclinical proof-of-concept, clinical translation, reproducibility and feasibility in patients with cirrhosis

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Abstract:

Purpose:

MR elastography and magnetisation-tagging use liver stiffness (LS) measurements to diagnose fibrosis but require physical drivers, specialist sequences and post-processing. Here we evaluate non-rigid registration of dynamic two-dimensional cine MRI images to measure cardiac-induced liver deformation (LD) as a measure of LS by (i) assessing preclinical proof-of-concept, (ii) clinical reproducibility and inter-reader variability, (iii) the effects of hepatic haemodynamic changes and (iv) feasibility in patients with cirrhosis.

Methods:

Sprague-Dawley rats (n=21 bile duct ligated (BDL), n=17 sham-operated controls) and fasted patients with liver cirrhosis (n=11) and healthy volunteers (HVs, n=10) underwent spoiled gradient-echo short-axis cardiac cine MRI studies at 9.4T (rodents) and 3.0T (humans). LD measurements were obtained from intrahepatic sub-cardiac regions-of-interest close to the diaphragmatic margin. One-week reproducibility and prandial-stress induced haemodynamic changes were assessed in healthy volunteers.

Results:

Normalised LD was higher in BDL (1.304 ± 0.062) compared with sham-operated rats (1.058 ± 0.045 , $P=0.0031$). HV seven-day reproducibility Bland-Altman (BA) limits-of-agreement (LoAs) were ± 0.028 a.u. and inter-reader variability BA LoAs were ± 0.030 a.u.. Post-prandial LD increases were non-significant ($+0.0083 \pm 0.0076$ a.u., $P=0.3028$) and uncorrelated with PV flow changes ($r=0.42$, $p=0.2219$). LD measurements successfully obtained from all patients were not significantly higher in cirrhotics (0.102 ± 0.0099 a.u.) compared with HVs (0.080 ± 0.0063 a.u., $P=0.0847$).

Conclusion:

Cardiac-induced LD is a conceptually reasonable approach from preclinical studies, measurements demonstrate good reproducibility and inter-reader variability, are ~~robust~~ **less likely to be affected by** hepatic haemodynamic changes and are feasible in patients with cirrhosis.

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Keywords:

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Manil D Chouhan – no relevant conflicts of interest or competing interests to declare
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Steve Halligan – no relevant conflicts of interest or competing interests to declare
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Mark F Lythgoe – no relevant conflicts of interest or competing interests to declare
Rajeshwar P Mookerjee – no relevant conflicts of interest or competing interests to declare
Alex Menys – is the CEO of Motilent. No other relevant conflicts of interest or competing interests to declare.

Stuart A Taylor - has share options in Motilent and is a consultant at Alimentiv. No other relevant conflicts of interest or competing interests to declare.

Availability of data and material:

The authors confirm that the data supporting the findings of this study are available within the article. Raw data were generated at the UCL Centre for Medical Imaging/UCL Centre for Advanced Biomedical Imaging. Derived data supporting the findings of this study are available from the corresponding author on request.

Code availability:

Cardiac cine MRI data was analysed using GIQuant[®] (Motilent, London, UK).

Authors contributions:

Manil D Chouhan – guarantor of integrity of the entire study, study concepts and design, literature research, clinical studies, experimental pre-clinical studies, data-analysis, statistical analysis, manuscript preparation, manuscript editing

Heather E Fitzke – study concepts and design, experimental pre-clinical studies, data-analysis, manuscript editing

Alan Bainbridge – clinical studies, experimental pre-clinical studies, data-analysis

David Atkinson - clinical studies, experimental pre-clinical studies, data analysis, manuscript editing

Steve Halligan – statistical analysis, manuscript editing

Nathan Davies – experimental preclinical studies

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Alex Menys – study concepts and design, experimental pre-clinical studies, clinical studies, data-analysis, manuscript editing

Stuart A Taylor – study concepts and design, manuscript editing

Ethics approval:

Preclinical - approval obtained from the Animal Care Ethical Committee of University College London.

Clinical - Research Ethics Committee (Joint RNOH/IOMS, reference number 08/H0724/35) approval was obtained.

Consent to participate:

Written consent obtained from all clinical study participants.

Consent for publication:

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Specific Remarks:

Not applicable

Introduction

1
2 Cirrhosis and chronic liver disease remain major causes of global morbidity and mortality[1]. Liver
3 fibrosis, a precursor to cirrhosis and common pathological phenotype of liver disease is
4 characterised by tissue accumulation of extracellular matrix proteins and remodelling[2]. Diagnosing
5 the presence and severity of liver fibrosis – particularly in early-stage liver disease – can direct
6 treatment and lifestyle changes that may enable recovery. Gold standard fibrosis measurements
7 however require invasive biopsy with procedural morbidity and occasional mortality, rendering
8 screening for early disease and repeated measurements unfeasible. Biopsy samples are also small
9 and prone to sampling error and significant inter-/intra-observer variability[3, 4].

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11 In response to chronic injury, fibrotic processes progressively alter the biomechanical
12 properties of the liver including tissue viscosity, elasticity and stiffness. These form the basis of
13 modern non-invasive biomechanical imaging methods to measure fibrosis [5]. By measuring tissue
14 response to an applied physical stress, biomechanical imaging methods such as ultrasound and MR
15 elastography (MRE) measure liver stiffness as a biomarker of fibrosis. US-based elastography
16 methods are most widely used but, like biopsy, only sample a small liver tissue volumes, often
17 blindly (in the case of transient elastography)[6, 7]. MRE potentially maps whole liver tissue
18 stiffness, but requires an external driver strapped to the abdomen to induce mechanical waves.
19 Modified phase-contrast MRI sequences are then used to map tissue stiffness, but the driver may
20 cause discomfort and measurements can be compromised in obese patients by field
21 inhomogeneities particularly at higher field strength and in patients with hepatic iron-overload[8, 9].

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23 MR strain imaging is a more recent biomechanical imaging method based on liver
24 deformation caused by physiological strain caused by cardiac or respiratory motion, using either cine
25 magnetisation tagging or strain-encoding imaging[10–12]. Most studies have used cardiac-induced
26 liver deformation with encouraging early results demonstrating differences between patients with
27 varying stages of histologically proven fibrosis[13, 14] and disease severity[15]. While these
28 approaches avoid additional hardware and mechanical drivers, both require specialist sequences and
29 post-processing that are not widely available.

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31 Non-rigid registration of dynamic two-dimensional cine MR images has been used to
32 successfully quantify small bowel motility[16] and vocal cord motion[17], and can assess Crohn's
33 disease and treatment response[18]. In tissue such as the liver, it is possible that deformation
34 reflects tissue stiffness, potentially yielding a more readily available biomarker of cirrhosis that could
35 be readily obtained from widely available MR sequences. We propose using non-rigid registration of
36 two-dimensional dynamic cine MR images of the liver to measure cardiac-induced liver deformation
37 (LD). Here we (a) evaluate LD measurement in a preclinical rodent model as a proof-of-concept,

1 before clinical translation to normal volunteers, with (b) assessment of reproducibility, (c) inter-
2 reader variability, (d) the effects of hepatic haemodynamic changes using prandial stress (as
3 haemodynamic changes could induce changes in tissue elasticity) and finally (e) feasibility in cirrhotic
4 patients.
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7 8 **Materials and Methods**

9 Preclinical proof-of-concept

10 *Subjects and preparation*

11 Experiments were conducted using <blinded> guidelines under <blinded>, with approval from the
12 Animal Care Ethical Committee of <blinded>. Healthy male Sprague-Dawley rats (<blinded>) with
13 normal liver function (weight 250-300g) underwent either sham laparotomy or bile duct ligation[19].
14 Animals were maintained for 4-5 weeks after the procedure to enable chronic liver disease to
15 develop in BDL rats as verified by previously reported histopathological and phenotypic changes at
16 our institution[20].
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19 Anaesthesia was induced and maintained at a constant level using isoflurane (2% in 1L
20 O₂/min). A rectal probe (SA Instruments, New York, NY) was used to monitor core temperature, that
21 was regulated while scanning using warm air and pipes with circulating warm water. A triple-
22 electrode single-lead system and respiratory bellows (SA Instruments) were used for respiratory and
23 cardiac monitoring respectively. All animal preparation was undertaken by the study coordinator
24 (<blinded>, a radiology research fellow qualified in animal handling). Other imaging data derived
25 from this same group of animals has been previously published[21].
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28 *Pre-clinical liver deformation measurement*

29 Animals were scanned using a 9.4-T MRI system (Agilent Technologies, Oxford, England) with
30 sequence parameters listed in table 1. Short-axis cardiac cine MRI studies were performed as
31 described previously[21]. Cardiac and respiratory gated gradient-echo coronal images of the thorax
32 and left-ventricular (LV) long-axis cardiac imaging were used to plan LV short-axis imaging.
33 Spoiled gradient-echo imaging with 1 mm contiguous LV short-axis slices planned from the LV apex
34 to the mitral valve orifice was undertaken using a 15° flip angle, 40x40 mm² field-of-view (FOV) and
35 128x64 (frequency encoding x phase encoding) matrix. Full LV coverage with data for at least 20
36 frames through the cardiac cycle (temporal resolution of approximately 7-9 milliseconds) was
37 typically obtained in under 12 minutes. Data was analysed using deformation analysis software
38 GIQuant® (Motilent, London, UK)[16, 22]. A single short-axis slice was selected mid-way between the
39 cardiac apex and mitral valve and a single hepatic parenchymal region-of-interest (ROI) was drawn
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1 close to the diaphragmatic margin and propagated automatically to other frames within the slice
2 cine-loop. As cardiac cine MRI images were acquired with the primary objective of measuring
3 dynamic changes in cardiac function in response to haemodynamic stress[21], imaging optimised to
4 shorten acquisition times for LV segmentation was noted to have consistent noise artefact in the
5 phase-encode direction (figure 1a and 1b). This was likely secondary to motion artefact introduced
6 by prospective gating errors. Given the small deformation signal being measured, the noise had the
7 potential to compromise deformation quantification, moreover the severity of the artefact varied
8 between subjects. To account for this in a subject-specific way, a second extra-corporeal ROI of
9 similar size to the parenchyma was placed within the imaged area outside of the body but parallel to
10 the parenchymal ROI (figure 1a and 1b). ROIs were placed blinded to subject cohort via joint
11 consensus between a radiology research fellow (<blinded>) and imaging scientist (<blinded>), both
12 with >5 years' experience of abdominal imaging and use of GIQuant™ software. LD measured in
13 arbitrary units (a.u.) was based on the standard deviation over time of each pixel's Jacobian
14 determinant, summarised as the ROI mean[22]. To account for potential noise artefact, preclinical
15 LD measurements were divided by the measurement obtained from the extra-corporeal ROI to
16 calculate normalised LD (nLD, equation 1):
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$$nLD \text{ (unitless)} = \frac{\text{preclinical } LD \text{ (a.u.)}}{\text{Extra-corporeal ROI deformation (a.u.)}} \quad (1)$$

Human translation

Subjects and preparation

40 Research Ethics Committee approval was obtained and all participants provided informed written
41 consent. Healthy volunteers were recruited by University campus advertisement. Exclusion criteria
42 were (a) known contraindication to MRI, (b) any long-term medication (excluding oral
43 contraceptives) and, (c) any previous history of liver or gastrointestinal disease. Patients with
44 histologically confirmed cirrhosis undergoing invasive transjugular procedures for routine clinical
45 care were recruited from hepatology outpatient clinics or elective transjugular intrahepatic
46 portosystemic shunt (TIPSS) lists for refractory ascites, excepting participants who (a) had known
47 MRI contraindication, (b) were unable to provide informed consent or, (c) were on specific
48 treatment for portal hypertension (e.g. beta-blockers). All participants were asked to avoid
49 caffeinated fluids and fast for six hours prior to MRI.
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Clinical liver deformation measurement

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Imaging was performed using a 3.0T scanner (Achieva, Philips Healthcare, Best, Netherlands) with a 16-channel body coil (SENSE XL-Torso, Philips Healthcare, Best, Netherlands) with sequence parameters listed in table 1. Short-axis cardiac cine MRI studies were planned using thoracic coronal and LV long-axis anatomical imaging. Images were obtained in expiratory breath-hold with pulse-oximetry signal used for cardiac gating. Spoiled gradient-echo imaging with 8 mm contiguous LV short-axis slices planned from the LV apex to the mitral orifice was undertaken using a 45° flip angle, 320x320 mm FOV and 256x256 matrix. Full LV coverage with data for approximately 30 cardiac cycle phases was typically obtained within 7 breath-holds. Data from a single short-axis slice selected mid-way between the cardiac apex and mitral valve was analysed using GIQuant™ by drawing a single ROI on the subdiaphragmatic hepatic parenchyma (avoiding vessels and ducts) and automatic propagation to other frames within the slice cine-loop (figure 1c and d). ROIs were placed independently by two imaging scientists blinded to clinical details (<blinded> and <blinded>), both with >5 years' experience of abdominal imaging and use of GIQuant™ software. LD was measured in arbitrary units (a.u.) as described above for preclinical subjects[22].

Hepatic haemodynamic assessment

Post-prandial stress was used to stimulate haemodynamic change. Phase-contrast MRI (PCMRI) measurements of portal venous (PV) flow were used to assess post-prandial haemodynamic response. Anatomical upper abdominal coronal steady-state free precession (SSFP) imaging was used to plan further SSFP imaging parallel to the PV, which together were used to plan two-dimensional PCMRI studies orthogonal to the vessel (sequence parameters listed in table 1). PV PCMRI studies were performed in expiratory breath-hold with pulse-oximetry based cardiac gating, using a velocity encoding setting (V_{enc}) of 40 cm/s, 10° flip angle, 5 mm slice thickness, 271x210 mm FOV and 336x336 matrix. Seven phases were obtained through the cardiac cycle and PCMRI measurements were performed in triplicate. Flow quantification was performed using Segment (Medviso, Lund, Sweden), with averaging of triplicate PV bulk flow measurements. Upper abdominal coronal SSFP imaging was used to estimate liver mass, with volumetric segmentation performed manually using Amira (Amira Resolve RT, Visage Imaging, Berlin, Germany) and assumption of 1.0 g/ml tissue density[23]. Triplicate PV bulk flow measurements normalised to estimated liver mass were averaged, with all PCMRI vessel and hepatic volumetric segmentation undertaken by the study coordinator (<blinded>).

Reproducibility, inter-reader variability and prandial stress studies

1 For reproducibility studies (n=8), subjects were fasted for 6 hours and scanned on the same scanner
2 using the same MRI protocol 7 days later at a similar time of day. Assessment was based on
3 measurements made by the more senior imaging scientist (<blinded>), blinded to
4 baseline/reproducibility group or post-prandial state.
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7 Independent clinical LD measurements from healthy volunteers at baseline (including
8 reproducibility measurements) from both readers (<blinded> and <blinded>) were used for inter-
9 reader variability studies. For prandial stress studies (n=10), healthy volunteers ingested 440 mls of
10 oral Ensure Plus® (Abbot Laboratories, Illinois, USA) after the initial scan. MRI was then repeated 45-
11 60 minutes after ingestion.
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16 17 *Statistical analysis*

18 Statistical analyses used Prism (Graphpad Software Inc., version 6.01), with calculation of intra-class
19 correlation coefficients using SPSS Statistics (IBM, version 26.0). Distribution normality was
20 confirmed with Shapiro-Wilk testing. Comparisons between animal groups were undertaken with
21 unpaired Student t-tests, with Welch's correction to account for group differences in standard
22 deviation. Where appropriate (and indicated in the presented results), non-parametric Mann-
23 Whitney U-tests were used for parameters that were not normally distributed. Clinical LD
24 measurement reproducibility and inter-reader variability in healthy volunteers was assessed using
25 Bland-Altman analysis with calculation of 95% limits-of-agreement (LoAs), intra-class correlation
26 coefficient (ICC) estimation (using the mean of both measurements, absolute agreement and a two-
27 way mixed-effects model) and within-subject Coefficient of Variation (CoV). The effects of prandial
28 stress in healthy volunteers was assessed by comparing baseline and post-prandial PV flow and LD
29 measurements using paired Student t-tests, with Welch's correction to account for group differences
30 in standard deviation. Correlations between post-prandial change in PV flow and LD were assessed
31 using Pearson's correlation coefficient. Baseline LD measurements in healthy volunteers were
32 compared with liver disease patients using unpaired Student t-tests with Welch's correction. Data
33 are expressed as means \pm standard errors, with significance at the 5% threshold.
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51 **Results**

52 *Preclinical cohort and proof-of-concept*

53 Similar weights were observed in both cohorts (250-300g) at baseline, but four weeks following
54 surgery, wet liver mass was higher in BDL (31 \pm 1g, n=21) compared with sham-operated animals
55 (15 \pm 1g, n=17, Mann-Whitney U-test, P<0.0001) but whole body weight was lower in BDL (413 \pm 9g)
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1 compared with sham-operated animals ($474\pm 5\text{g}$, $P<0.0001$). Measurements from five subjects were
2 not possible due to wrap-around artefact resulting in insufficient area for appropriate placement of
3 an extra-corporeal ROI or suboptimal image quality. Acceptable measurements were ultimately
4 obtained from fifteen sham and eighteen BDL animals (figure 2a). Normalised LD was higher in BDL
5 (1.304 ± 0.062) compared with sham-operated rats (1.058 ± 0.045 , Mann-Whitney U-test, $P=0.0031$,
6 table 2, figure 3).
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10 *Healthy volunteer cohort and clinical LD reproducibility and inter-reader variability*

11 Ten healthy volunteers were recruited (5 males, 5 females, aged 28.6 ± 1.6 years, figure 2b). Mean
12 baseline LD was 0.080 ± 0.0063 a.u. (table 2) and normal volunteer reproducibility studies ($n=8$)
13 demonstrated a mean LD difference of 0.0043 a.u. with Bland-Altman 95% LoAs of ± 0.028 a.u.
14 (figure 4), ICC of 0.771 ($P=0.008$) and within-subject CoV of 12.7% after seven days.
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21 Inter-reader variability studies using measurements obtained at baseline and after seven
22 days ($n=8$ subjects, with 16 measurements made by each reader) demonstrated a mean LD
23 difference of 0.0014 a.u. with Bland-Altman 95% LoAs of ± 0.030 a.u. (figure 5), ICC of 0.787
24 ($P<0.0001$) and a CoV of 11.8% between both readers.
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30 *Prandial stress studies*

31 All ten healthy volunteers participated in prandial stress studies. Post-prandial increases in PCMRI
32 PV flow were observed (55.3 ± 4.5 ml/min/100g in the fasted state compared with 106.0 ± 3.9
33 ml/min/100g in the post-prandial state, mean difference $+50.7\pm 4.6$ ml/min/100g, $P<0.0001$, figure
34 6a). Post-prandial LD was higher than in the fasted state, but this was not statistically significant
35 (0.080 ± 0.0063 a.u. in the fasted state compared with 0.088 ± 0.0078 a.u. in the post-prandial state,
36 mean difference $+0.0083\pm 0.0076$ a.u., $P=0.3028$, table 2, figure 6b). No correlation was
37 demonstrated between post-prandial change in LD and post-prandial change in PCMRI PV flow
38 ($r=0.42$, $p=0.2219$, figure 6c).
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48 *Feasibility in patients with cirrhosis*

49 Eleven patients participated in feasibility studies (10 males, 1 female, aged 53 ± 3 years). Cirrhotic
50 aetiologies included alcohol ($n=8$), non-alcoholic steatohepatitis cirrhosis ($n=2$) and hereditary
51 haemochromatosis ($n=1$), with predominantly stable disease at the time of scanning (Child-Pugh
52 score $A=7$, $B=4$) (figure 2c). LD measurements were obtained successfully from all patients. Although
53 LD was higher in cirrhotic patients (0.102 ± 0.0099 a.u.) compared with healthy volunteers
54 (0.080 ± 0.0063 a.u.), this difference was not statistically significant ($P=0.0847$, table 2, figure 7).
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Discussion

We demonstrated the use of non-rigid registration of dynamic two-dimensional cardiac short-axis cine MR imaging to measure cardiac-induced liver deformation (LD). As a proof-of-concept we found that in a rat model of advanced liver disease, normalised LD measurements were significantly different to those obtained from control animals. Because imaging had been optimised for LV segmentation, varying levels of phase-encode direction noise artefact were addressed by normalising animal LD measurements to a second parallel extra-corporeal ROI. As this noise was not appreciable on clinical scans, human LD measurements were not normalised. Alternative magnetisation-tagging approaches to measure cardiac-induced liver motion have not reported preclinical data for validation/proof-of-concept but there have been several preclinical canine and porcine liver and splenic MRE studies demonstrating an association between stiffness, hepatic venous pressure gradient[24, 25] and fibrosis[26].

Following human translation, measured ICCs for LD reproducibility on the same scanner after 7 days in similar scanning conditions suggest measurement reliability, with reasonable within-subject CoVs. Repeatability studies (three measurements within the same scanning session) of respiratory-induced liver motion using magnetisation-tagging have reported comparable ICCs[10] and while formal reproducibility data for measurement of cardiac-induced liver motion using magnetisation-tagging has not been reported, a meta-analysis of MRE liver stiffness measurements across 274 patients has demonstrated a smaller within-subject CoV of 8%[27].

Inter-reader variability studies have also demonstrated good measurement reliability also with reasonable within-subject CoVs, despite the relatively small ROIs used and potential for noise contamination. Measured LD inter-reader variability ICCs, CoVs and Bland-Altman LoAs in this study were similar to LD reproducibility statistics, therefore suggesting that reader differences are unlikely to introduce variation outside of that expected from repeat measurement. Inter-reader variability was superior to that reported for cardiac-induced liver motion with magnetisation tagging (ICC of 0.584 and CoV of 29.7%)[13].

Post-prandial stress was used to stimulate haemodynamic change with the premise that this could induce changes in tissue elasticity and therefore LD. Although healthy volunteer post-prandial PV flow almost doubled, change in LD was not significant, and was much smaller than Bland-Altman LoAs for LD reproducibility and inter-reader variability. Furthermore, we found no statistically significant association between post-prandial change in PV flow and LD, although a general positive trend was noted between post-prandial change in LD and PV flow, suggesting that further studies would be needed before concluding that LD measurements are unlikely to be affected by

1 haemodynamic changes. Post-prandial changes in magnetisation-tagging derived measures of liver
2 motion have not been reported, but small statistically significant increases in MRE liver stiffness and
3 poor positive but significant correlations between change in MRE liver stiffness and PV flow have
4 been demonstrated in the setting of a similar meal challenge[28]. This could suggest lower LD
5 sensitivity to haemodynamic changes relative to MRE liver stiffness.
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8 Finally, we demonstrated that LD measurements were feasible in patients with established
9 liver disease and while LD increased relative to healthy volunteers, this difference was not
10 statistically significant. This could reflect a type II error arising from the small sample size and larger
11 scale studies are definitely needed to understand potential for disease severity stratification. It may
12 also however reflect lower sensitivity in patients with milder more stable disease, as no data was
13 collected in patients with decompensated cirrhosis, for example. Although sample size was small,
14 demonstration of feasibility in patients with liver disease, including in the setting of iron-overload is
15 an important strength. Patients often struggle with breath-holds, imaging is prone to motion
16 artefact, and difficulties arising from the use of MRE external drivers or quantification errors in the
17 setting of iron-overload are recognised limitations of alternative MR biomechanical imaging
18 methods. We do acknowledge that only one patient in our sample had significant hepatic siderosis,
19 so further work would be required to confirm if LD measurements are definitively robust in the
20 setting of iron-overload. Interestingly, several studies measuring cardiac-induced liver motion with
21 magnetisation-tagging have demonstrated reducing liver 'strain' with advancing liver disease[11, 13,
22 29], opposing our finding of increased normalised/absolute LD in the setting of liver disease. This
23 may have arisen from methodological differences, and we postulate that higher LD values in liver
24 disease may reflect greater sensitivity to increased translational motion (as a result of stiffer liver
25 tissue), as opposed to greater elastic motion as seen in non-diseased liver (which strain methods are
26 potentially more sensitive to). It is also worth noting that we studied a largely stable disease cohort
27 (predominantly Child-Pugh A) – while this could account for a non-significantly higher patient LD,
28 biomechanical properties of liver tissue are arguably more dependent upon the degree of
29 fibrosis/cirrhosis severity rather than disease stability. Previous studies using magnetisation-tagging
30 based measures of liver motion for example, were still able to demonstrate significantly different
31 measurements in similarly sized patient cohorts with predominantly stable Child-Pugh A liver
32 disease[29], suggesting that LD may have lower sensitivity.
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35 Low sensitivity of LD measurements may reflect difficulties obtaining sufficient signal-to-
36 noise, highlighted by our animal data and the use of small ROIs. Despite availability of data from the
37 entirety of liver imaged within a single slice, the lack of signal further away from the left ventricle
38 restricted measurements to a sub-cardiac location (negating the advantage of whole-liver coverage
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1 and quantification) and conversely increasing the risk of noise contamination. It is worth noting that
2 despite this, reproducibility and inter-reader variability were reasonable. Measured tissue
3 deformation/motion is however directly related to the energy of applied stress, which likely varies
4 between patients. For cardiac-induced motion, energy released by ventricular contraction is likely to
5 vary in liver disease as cirrhotic cardiomyopathy is common[30]. . MRE for example, avoids this
6 limitation by using an external driver so that stress is consistent between patients. Formal clinical
7 translation is therefore likely to require use of a means of measuring the energy applied to the liver
8 for normalisation of deformation measurements. It is worth noting that alternative approaches,
9 including the tracking dynamic wave propagation through the liver during the cardiac cycle remain
10 novel approaches that could potentially be used to calculate liver stiffness from cine MRI data.

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18 Finally, despite these limitations, our findings remain relevant because they demonstrate
19 the potential of LD in the assessment of liver disease. An important strength is the use of non-
20 specialist MRI sequences, readily available on most scanners, simple to use, and easy for
21 radiographers/technologists to acquire. We do recognise this benefit is partly mitigated by the need
22 to use third party image registration software – in this work for example, we used a proprietary
23 (commercial) image analysis tool to obtain measurements. Because measurements rely on signal-
24 intensity based tissue deformation, measures are less likely to be affected by scanner vendors or
25 field-strength. Avoiding additional hardware (such as a driver) decreases cost, increases
26 acceptability and maximises patient comfort. Analyses are also an advantage, the GIQuant™
27 software used here is CE marked for clinical use (albeit in Crohn’s Disease) and integrates directly
28 with hospital PACS. GIQuant™ already includes respiratory motion correction features that may be
29 used to exploit dynamic imaging through the respiratory cycle to derive a respiratory LD score.

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39 This small-scale translational study has highlighted some important strengths and
40 weaknesses of LD measurements, including the need for alternative approaches to the application
41 and measurement of physical stress, and the need for appropriately powered patient studies. In
42 summary, cardiac-induced liver deformation using non-rigid registration of dynamic two-
43 dimensional cine MRI imaging is a conceptually reasonable approach from preclinical studies, LD
44 measurements demonstrate good reproducibility and inter-reader variability, are robust to hepatic
45 haemodynamic changes and are feasible in patients with liver disease.

46 47 48 49 50 51 52 53 **References:**

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Figure captions:

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Fig. 1: Short-axis cardiac cine images of representative slices and ROIs (yellow-dashed line) used for LD quantification in rats (a, b) and humans (c, d). Anatomical images are shown on the left (a, c) with overlaid deformation maps shown on the right (b, d). Note the presence of an additional extra-corporeal ROI parallel to the hepatic ROI used for normalisation to account for phase-encode direction noise artefact (a)

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Fig. 2: Study cohorts

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Fig. 3: Normalised Liver Deformation in sham control (■) and BDL rats (△)

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Fig. 4: Seven-day reproducibility of LD measurements in healthy volunteers, with (a) Bland-Altman analysis of agreement (bias 0.0043 a.u, solid line; 95% LoAs ± 0.028 a.u., dashed lines) and (b) raw data scatterplots

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Fig. 5: Inter-reader variability of LD measurements in healthy volunteers, with (a) Bland-Altman analysis of agreement (bias 0.0014 a.u, solid line; 95% LoAs ± 0.030 a.u., dashed lines) and (b) raw data scatterplots

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Fig. 6: Healthy volunteer prandial stress studies with post-prandial changes in (a) PV flow, (b) LD and (c) correlation between changes in PV flow and LD

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Fig. 7: Liver Deformation in healthy volunteer controls (■) and patients with cirrhosis (△)

Figures:

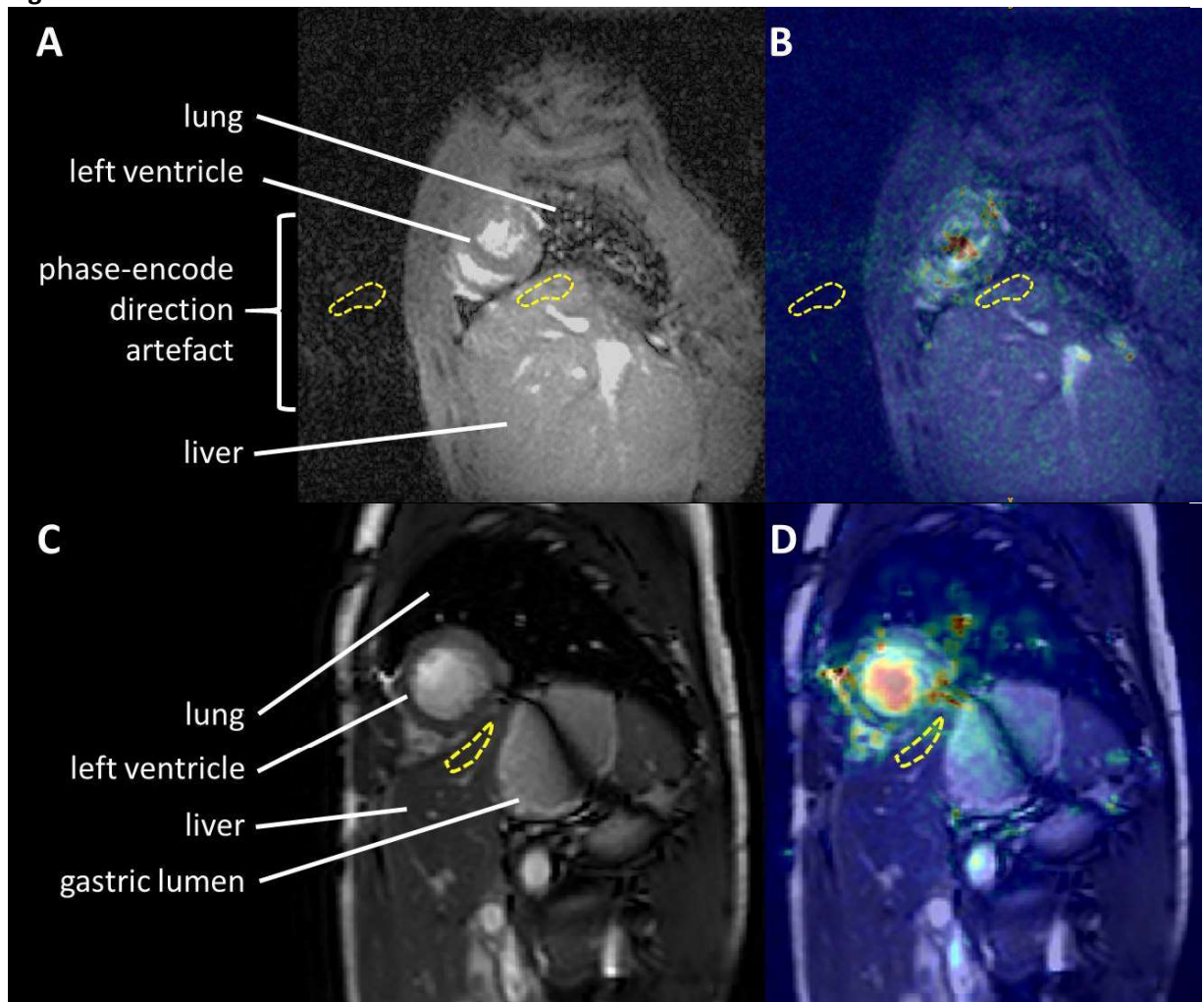


Figure 1

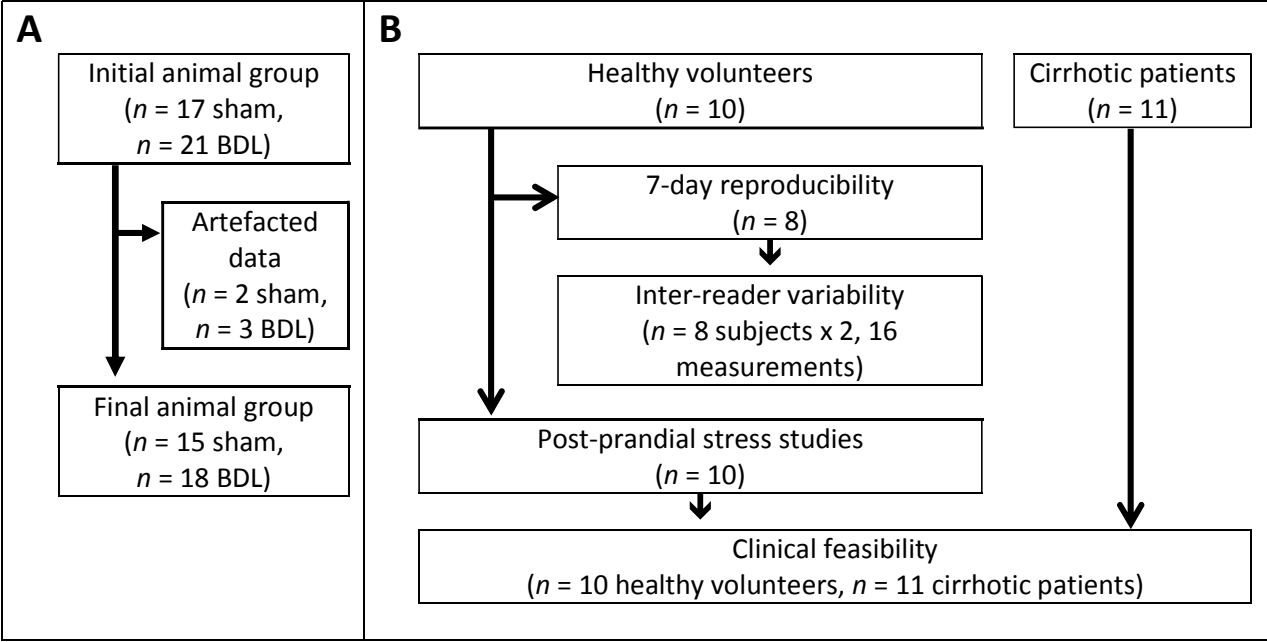


Figure 2

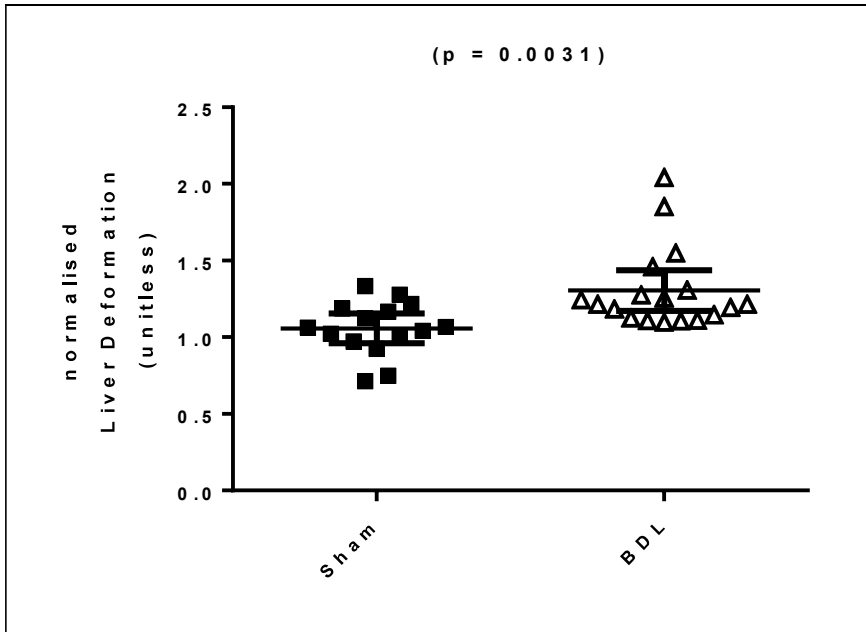


Figure 3

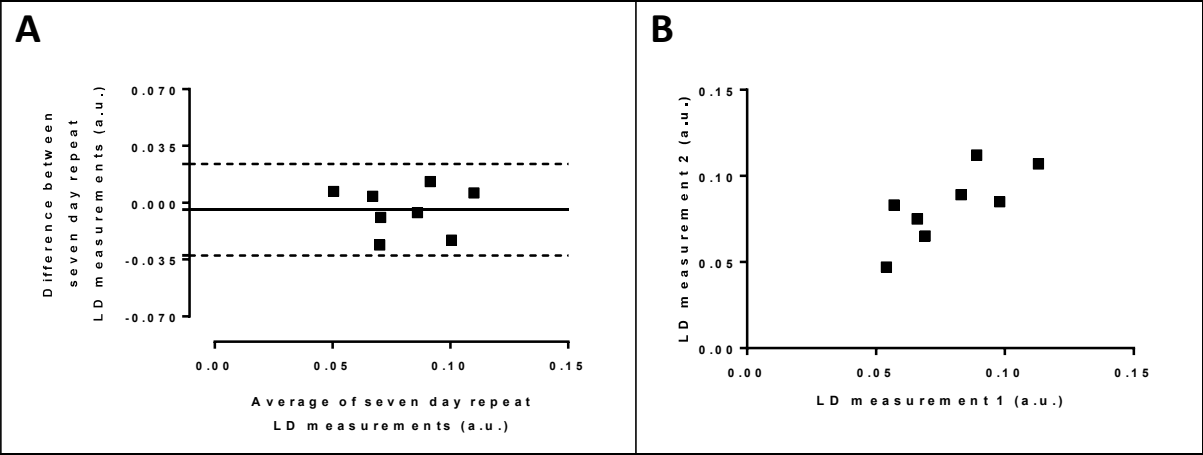


Figure 4

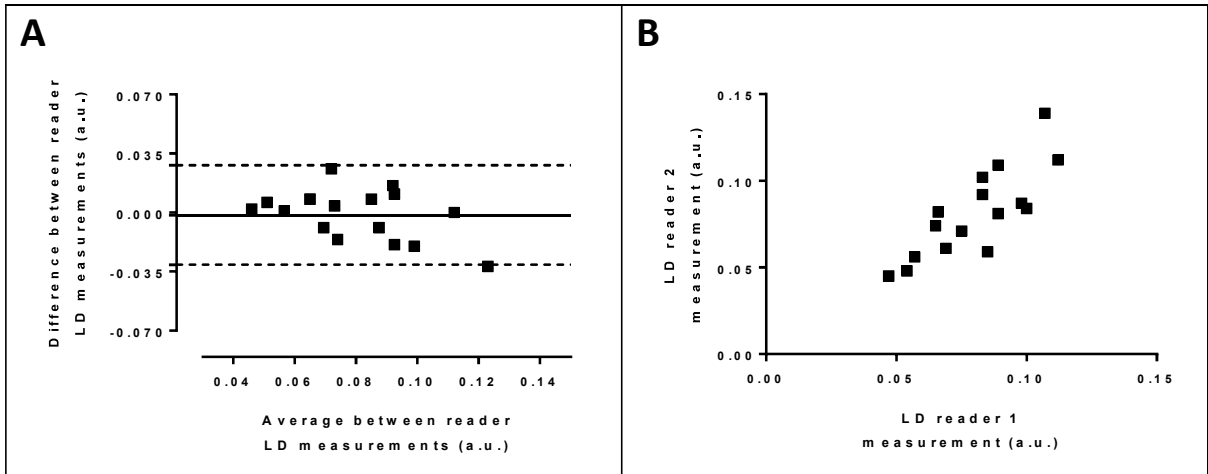
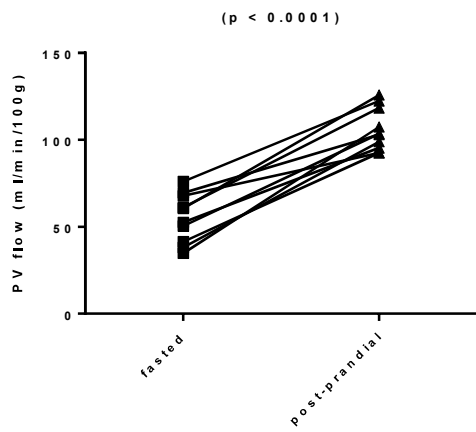
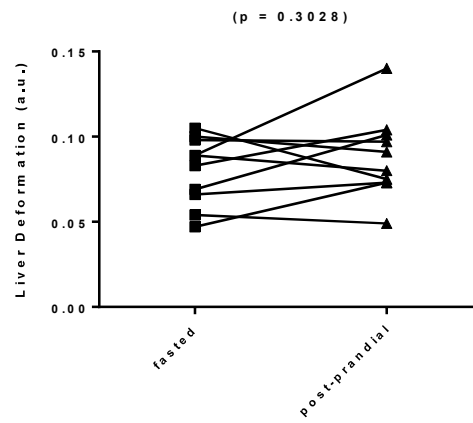
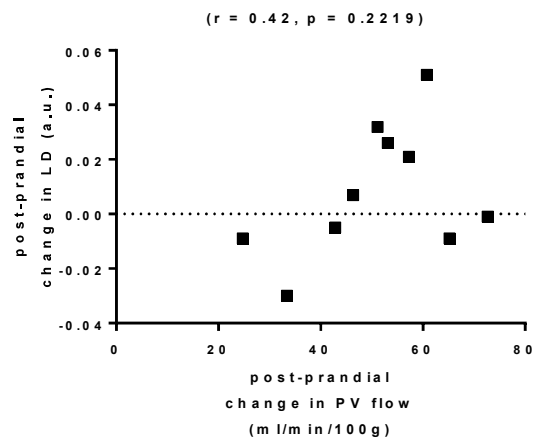


Figure 5

A**B****C****Figure 6**

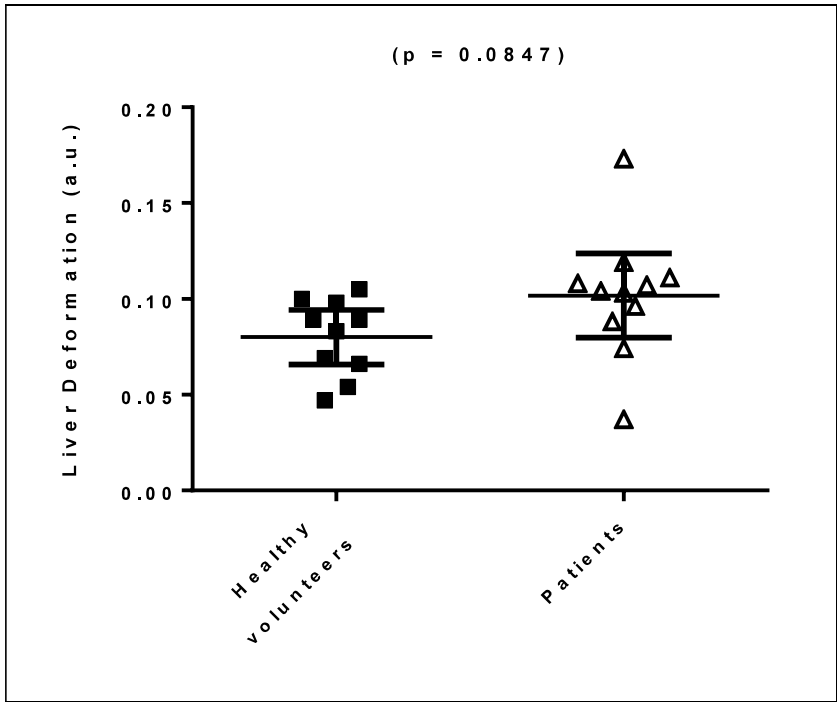


Figure 7

Tables:

	PRECLINICAL (9.4T)		CLINICAL (3.0T)	
	ANATOMICAL PLANNING IMAGES (GE)	CARDIAC CINE MRI (GE)	ANATOMICAL PLANNING IMAGES (SSFP)	CARDIAC CINE MRI (GE)
<i>TR/TE (milliseconds)</i>	8.2/5.6	7.5/1.2	2.47/1.23	3.40/1.70
<i>Flip angle (°)</i>	20	15	45	45
<i>Matrix size (pixels)</i>	128 x 128	128 x 64	352 x 352	256 x 256
<i>Field-of-view (mm)</i>	80 x 80	40 x 40	350 x 350	320 x 320
<i>Spatial resolution (mm²)</i>	0.625 x 0.625	0.313 x 0.625	0.994 x 0.994	1.25 x 1.25
<i>Slice thickness (mm)</i>	2	1	5	8
<i>Slice gap (mm)</i>	4.5	0	5.5	8
<i>Cardiac cycle phases</i>	-	≥20	-	30

(GE – gradient echo, SSFP – steady-state free precession)

Table 1: Sequence parameters

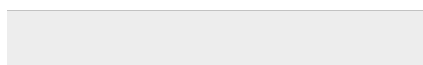
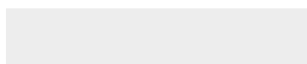
	RAT (nLD)		HUMAN (LD)	
	MEAN±SE	P-VALUE [†]	MEAN±SE	P-VALUE [†]
<i>Control group</i> [‡]	1.058±0.045	-	0.080±0.0063	-
<i>Post-prandial stress</i>	-	-	0.088±0.0078	0.3028
<i>Disease group</i> ^{††}	1.304±0.062	0.0031*	0.102±0.0099	0.0847

([†]from comparison with control group data; [‡] sham-operated rats/healthy volunteers; ^{††}BDL rats/patients with cirrhosis; *P<0.05)

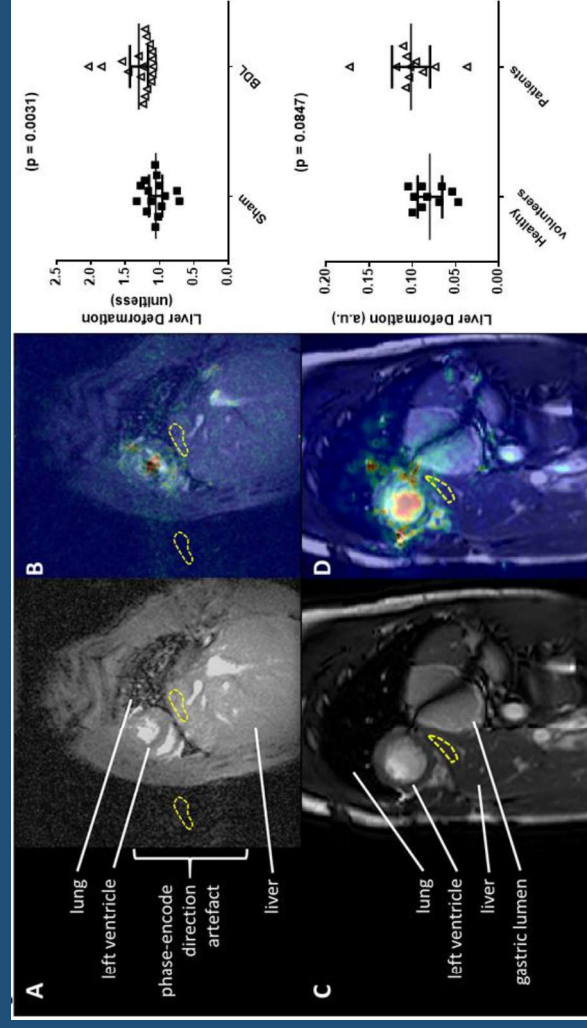
Table 2: nLD/LD measurements by study group



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Supplementary Material
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Cardiac-induced liver deformation as a measure of liver stiffness using dynamic imaging without magnetisation tagging – preclinical proof-of-concept, clinical translation, reproducibility and feasibility in patients with cirrhosis



Liver deformation was higher in cirrhotic rats ($p=0.0031$), reproducible in healthy volunteers and feasible but not significantly different in a cirrhotic patients ($p=0.0847$).