

1 **Title: Evidence for ecological processes driving speciation among endemic lizards of**
2 **Madagascar**

3
4 **Running title:** Speciation among endemic lizards of Madagascar

5
6 **Authors:**

7 Laura A. Nunes^{1,2}, Christopher J. Raxworthy³ and Richard G. Pearson¹

8 1. Centre for Biodiversity & Environment Research, Department of Genetics, Evolution and
9 Environment, University College London, Gower Street, London WC1E 6BT, UK

10 2. Department of Forest and Wildlife Ecology, University of Wisconsin-Madison, 1630 Linden
11 Drive, Madison, WI 53706, USA

12 3. Department of Herpetology, The American Museum of Natural History, 200 Central Park
13 West, New York, NY 10024, USA

14 Correspondence should be addressed to L.A.N (lnunes@wisc.edu) or R.G.P
15 (richard.pearson@ucl.ac.uk).

16

17 **Author contributions**

18 L.A.N and R.G.P conceived of and designed the project. L.A.N and C.J.R compiled the species
19 locality and systematic data. L.A.N performed the analyses. L.A.N, R.G.P and C.J.R wrote the
20 paper.

21

22 **Acknowledgements**

23 We are grateful to Mark Wilkinson for helpful discussions and guidance during the early stages
24 of this study. We thank Victoria Palone for assistance in the collection of occurrence data from
25 literature sources. We also thank the Center for Computation and Visualization at Brown
26 University for granting us access to its Higher-Performance Computing facilities. We

27 acknowledge financial support from the following grants awarded to C.J.R and R.G.P: National
28 Science Foundation DEB 1257610, 0641023, 0423286, 9984496, 9625873, 9322600, and BSR
29 9024505. L.A.N was supported by a studentship co-funded by UCL and the Natural History
30 Museum.

31

32 **Data Accessibility Statement**

33 Species occurrence data are available through the Dryad Digital Repository database

34 (<https://doi.org/10.5061/dryad.rbnzs7hcb>). Custom R scripts for reproducing the paper's RTR

35 analysis are available at: <https://github.com/LauraANunes/NunesPearson2017>

36

37 **Abstract**

38 Although genetic patterns produced by population isolation during speciation are well
39 documented, the biogeographic and ecological processes that trigger speciation remain poorly
40 understood. Alternative hypotheses for the biogeography and ecology of speciation include
41 geographic isolation combined with niche conservatism (soft allopatry), or parapatric distribution
42 on an environmental gradient with niche divergence (ecological speciation). Here we utilize
43 species' distributions, environmental data and two null models (Random Translation and
44 Rotation, RTR, and the Background Similarity Test, BST) to test these alternative hypotheses
45 among 28 sister pairs of micro-endemic lizards in Madagascar. Our results demonstrate strong
46 bimodal peaks along a niche divergence-conservation spectrum, with at least 25 out of 28 sister
47 pairs exhibiting either niche conservatism or divergence, and the remaining pairs showing weak
48 ecological signals. Yet despite these significant results, we do not find strong associations of
49 niche conservatism with allopatric distributions, or niche divergence with parapatric distributions.
50 Our findings thus provide strong evidence of a role for ecological processes driving speciation,
51 rather than the classic expectation of speciation through geographic isolation, but demonstrate
52 that the link between ecological speciation and parapatry is complex and requires further analysis
53 of a broader taxonomic sample to fully resolve.

54 **Keywords:** speciation, biogeography, niche conservatism, allopatry

55

56 **Introduction**

57 Speciation is the process by which populations evolve to become distinct species. The
58 segregation of ancestral species' distributions during speciation is most commonly described as a
59 geographic separation (Doebeli and Dieckmann 2003; Fig. 1a). Examples of this 'hard allopatry'

60 mode of speciation include single island endemics that became isolated via jump dispersal to
61 new islands (Ricklefs and Bermingham 2007). However, alternative hypotheses for the
62 biogeography of speciation bring into question the ubiquity of hard allopatry. One alternative
63 hypothesis, termed ‘soft allopatry’ (Pyron and Burbrink 2010), is that populations may become
64 geographically isolated when environmental disturbances, such as changes in climatic
65 conditions, cause an ancestral population to split its distribution (vicariance) because it is unable
66 to persist in intervening ecological conditions (Wiens 2004) (Fig. 1b). An example of soft
67 allopatry is observed in eastern North American salamanders, where sister pairs occupy
68 ecologically similar habitats in Appalachian montane regions but do not overlap geographically
69 because of unsuitable climates in intervening lowlands (Kozak and Wiens 2006). A third
70 hypothesis, termed ‘ecological speciation,’ describes a process that takes place without
71 immediate disruption to gene flow (Schluter 2001; Rundle and Nosil 2005; Nosil 2008) (Fig. 1c).
72 Ecological speciation has been proven to be possible both theoretically (Fisher 1930) and
73 experimentally (Doebeli and Dieckmann 2003; Berner et al. 2009), and speciation with gene
74 flow has been observed in cave salamanders (Niemiller et al. 2008) and in the flora of Lord
75 Howe Island (Papadopulos et al. 2014). Ecological speciation typically occurs along
76 environmental gradients (ecotones), which results in adjacent and overlapping geographic ranges
77 between descendent species (Caro et al. 2013), in contrast to the non-overlapping geographic
78 ranges of allopatric descendent species (Graham et al. 2004; Anacker and Strauss 2014).

79 Inferring speciation mechanisms from current geographic distributions is challenging
80 because of the likelihood of post-speciation range shifts (Losos and Glor 2003). Another
81 approach is to infer speciation mechanisms through the ecological processes associated with each
82 sister pair, which may be retained despite post-speciation range shifts (Losos and Glor 2003).

83 Given that soft allopatry results from the inability of ancestral species to survive in novel
84 ecological conditions, this speciation mechanism is associated with low ecological differentiation
85 between descendent species, termed phylogenetic niche conservatism (PNC, Fig. 1e; Peterson et
86 al. 1999; Wiens 2004). PNC is defined as the substantial retention of ecological traits through
87 time; thus, species within a sister pair will have similar ecological niches (Wiens and Graham
88 2005; Losos 2008). In contrast, substantial ecological differentiation between descent species,
89 termed phylogenetic niche divergence (PND, Fig 1f), is expected under ecological speciation
90 because the process is driven by ecological differentiation (Warren et al. 2008; Jezkova and
91 Wiens 2018). PND is defined as the differentiation of ecological traits through time, which
92 results in species within sister pairs having distinct ecological niches (Pyron et al. 2015; Nunes
93 and Pearson 2017). While soft allopatry and ecological speciation are mediated by intrinsic
94 environmental processes, hard allopatry takes place primarily through a process of isolation due
95 to discrete geographic features which limit population dispersal between potentially favorable
96 habitats (Pyron and Burbrink 2010). In this case, ecological differentiation between sister pairs
97 will not be due to environmental conditions selected by the species but rather be related to the
98 ecological conditions available in the landscape, or else represent the random independent
99 evolution of niches subsequent to isolation. As a result, we would not expect to find evidence of
100 either PNC or PND in cases of hard allopatry (Fig. 1d).

101 Several previous studies have tested for associations between speciation processes and
102 PNC/PND by combining species distribution data, fine-scale global climate data and statistical
103 null models (Warren et al. 2008; McCormack et al. 2009; Nunes and Pearson 2017; Jezkova and
104 Wiens 2018). Methods focus on testing sister pairs, which are useful for assessing speciation
105 processes because their recent divergence results in less time for range and niche shifts to have

106 occurred (Losos and Glor 2003; Anacker and Strauss 2014). To test for evidence of PNC/PND,
107 we apply two approaches: the Random Translation and Rotation null model (RTR; Nunes and
108 Pearson 2017) and the Background Similarity Test (BST; Warren et al. 2008). These approaches
109 provide complementary ways to assess whether the observed niche overlap between two closely
110 related species is higher or lower than by chance. The RTR model has several features that make
111 it suitable for this test, including: i) it preserves the range size and shape of each species; ii) it
112 maintains the underlying spatial autocorrelation of the climatic data while decoupling the link
113 between distribution and environment (Cardillo et al. 2019); iii) it avoids reciprocal testing
114 between two species, which may lead to conflicting estimates of speciation mechanisms (Warren
115 et al. 2008; Jezkova and Wiens 2018); and iv) it does not require the fitting of ecological niche
116 models which may be unreliable for species with small number of occurrences (Elith and
117 Graham 2009; Proosdij et al. 2016; Nunes and Pearson 2017; Soutan and Safi 2017). BST
118 complements the RTR method in that it incorporates local environmental heterogeneity between
119 species occurrences and their surrounding landscapes (Warren et al. 2008) and thus compares the
120 ecological suitability of each species within species-specific accessible areas (Barve 2011). Both
121 tests enable the testing of three distinct categorical outcomes: ecological niche overlap may be
122 significantly higher than random (PNC), lower than random (PND), or not significant; and they
123 can contextualize the observed niche overlap along a continuous niche divergence-conservation
124 spectrum.

125 We use the RTR and BST null tests to infer three possible outcomes: (1) when observed
126 niche overlap is not statistically distinct from the null expectation, the sister pair shows neither
127 PNC or PND, which is consistent with geographic drivers of hard allopatry (Fig. 1a,d) or may
128 reflect limitations of the data and/or methods (see Discussion); (2) when niche overlap is

129 statistically higher than the null expectation, the sister pair demonstrates PNC and is associated
130 with ecological drivers of soft allopatry (Fig. 1b,e); and (3) when niche overlap is statistically
131 lower than the null expectation, the sister pair demonstrates PND and is associated with
132 divergence due to ecological speciation (Fig. 1c,f). Using a metric for the degree of conservatism
133 (where low degrees of niche conservatism reflect niche divergence), we expect that pairs that speciated
134 through soft allopatry would cluster toward the top end of this distribution (i.e., statistical signal close to
135 or significant for PNC) and pairs that speciated through ecological speciation would cluster toward the
136 bottom end of the distribution (i.e., statistical signal close to or significant for PND). To account for the
137 potential accumulation of ecological differences and range shifts through time (Culumber and Tobler
138 2016), we also tested for associations between speciation age and a suite of biogeographic traits within
139 our sampled taxa.

140 Malagasy lizards stand out as suitable candidates for testing speciation processes and
141 local endemism because: i) ectothermic organisms are more sensitive to climatic conditions (Hu
142 et al. 2016) and are therefore more susceptible to ecological-mediated processes; ii) they are
143 relatively young, with radiations emerging during the Cenozoic period (Yoder and Nowak 2006)
144 within a relatively stable and isolated tropical environment and therefore have less potential for
145 post-speciation range and ecological shifts compared to sister pairs of older radiations (Blair et
146 al. 2013); iii) almost resolved phylogenetic trees are available (Townsend et al. 2009; Pyron et al.
147 2013; Zheng and Wiens 2016) with high sister pair richness (Raxworthy et al. 2008); and iv)
148 there is a high number of range restricted species, which indicates low dispersal abilities (Brown
149 et al. 1996; Lester et al. 2007) and therefore high potential for ranges to be stable through time.

150 Madagascar is an ideal setting for studying evolutionary processes because it has been
151 isolated from the mainland since the Mesozoic and therefore harbors multiple monophyletic
152 lineages and unique patterns of local endemism (Wilme 2006; Raxworthy et al. 2008; Vences et

153 al. 2009; Brown et al. 2014). Allopatric speciation has been proposed to be the dominant
154 speciation mechanism driving micro-endemism in Madagascar (Wilme 2006), yet cross-taxon
155 analyses question this and point to multiple potential mechanisms (Pearson and Raxworthy
156 2009).

157

158 **Methods**

159

160 We identified sister taxa within monophyletic lineages of Malagasy lizards from phylogenetic
161 studies (Townsend et al. 2009; Zheng and Wiens 2016). These monophyletic lineages included
162 the chameleons (*Brookesia*, *Calumma* and *Furcifer*), geckos (*Blaesodactylus*, *Geckolepsis*,
163 *Paroedura*, *Phelsuma* and *Uroplatus*), iguanas (*Oplurus*), plated lizards (*Tracheloptychus* and
164 *Zonosaurus*) and skinks (*Amphiglossus*, *Madascincus*, *Pygomeles* and *Voeltzkowia*). We
165 collected distribution data for 68 lizard sister pairs that are endemic to Madagascar from the
166 Global Biodiversity Information Facility (GBIF.org), VertNet (Constable et al. 2010) and
167 available literature (Pearson and Raxworthy 2009; Brown et al. 2014; Jezkova and Wiens 2018)
168 and Raxworthy (pers.comms.) and corrected for locality errors by review of the occurrence
169 dataset with reference data (IUCN Red List) and expert knowledge (Raxworthy). We removed
170 sister pairs that occupied more than 50% of the total area of the island to focus our analysis on
171 sister pairs that exhibit a localized distribution pattern and allow sufficient geographic space for
172 random sampling within the island (Nunes and Pearson 2017). Using range-restricted taxa also
173 provides higher confidence in species delimitations because broadly ranged reptiles may actually
174 contain cryptic complexes of species (e.g., Florio and Raxworthy 2016; Ruane et al. 2018) and
175 therefore will confound analysis of speciation processes (Raxworthy et al. 2007). A total of 28

176 lizard species pairs were kept (Supporting Information Table 1). Data on divergence dating of
177 speciation was obtained from Zheng and Wiens (2016). This study was selected due to the use of
178 a large genetic data set and primary fossil calibrations (Title and Rabosky 2016). The inclusion
179 of our sampled taxa within this phylogenetic tree was based on 12 (of 52 total) gene markers
180 used to construct the supermatrix phylogeny, with similar data coverage across all sister pairs
181 (see Supporting Information Table 6).

182 Species occurrence data and global climate data provide insight into the climatic
183 conditions that species are known to survive (i.e., species realized climatic niche *sensu* Soberón,
184 2007). Despite being an incomplete representation of a species' fundamental ecological niche
185 (Saupe et al. 2018; Owens et al. 2020), occurrence data are the most available data to generate
186 ecological models for a variety of species (Guillera-Arroita et al. 2015; Warren et al. 2020). By
187 comparing the similarity between the realized climatic niches of sister species and their
188 geographic mode of speciation, we analyze speciation in the context of ecological niche
189 differentiation across monophyletic phylogenies.

190 We conducted the analyses at a spatial resolution of 2.5 arc minutes (~4.5km grid cells)
191 to account for potential errors in locality measurements which could lead to errors in realized
192 climatic condition (Graham et al. 2007). To construct the realized climatic niche of each of the
193 sister taxa, we extracted climatic data on temperature and precipitation from the locality data
194 (WorldClim v2, Fick and Hijmans 2017). We selected seven climatic variables that are
195 associated with the physiology and life history requirements of reptiles (Pearson et al. 2014):
196 temperature seasonality, maximum temperature of warmest month, mean temperature of wettest
197 quarter, mean temperature of coldest quarter, annual precipitation, precipitation of driest month
198 and precipitation seasonality.

199 To characterize the biogeographic distributions and distinguish between allopatric and
200 parapatric pairs, we drew minimum convex polygons (MCPs) and calculated the minimum
201 distance (isolation distance) and the amount of overlap between polygons of sister taxa. The
202 isolation distance was measured as the minimum distance between occurrences of each sister
203 species using the `spDistN1` function from the 'sp' R package (Pebesma and Bivand 2005). The
204 range overlap of each species was calculated by dividing the total area of intersecting sister pairs
205 MCP by the area of the smaller ranged species (Anacker and Strauss 2014). Sister taxa with non-
206 overlapping MCPs were classified as having allopatric distributions, while those with
207 overlapping MCPs were classified as having parapatric distributions. Using minimum convex
208 polygons to describe biogeographic patterns in complex landscapes presents potential caveats,
209 such as inflating the amount of range overlap between two geographic distributions. As a result,
210 we found 4 pairs with apparent substantial range overlap, which could indicate sympatric
211 speciation (Table 1 and Table S1). However, we did not consider these pairs to be strong
212 candidates for sympatric speciation because they do not have any observed sympatry in the field
213 (Raxworthy pers. comms.) and some (i.e., *B. superciliaris* – *B. therezieni*) were distributed along
214 an elevational gradient. Therefore, these pairs were considered to be parapatric. We calculated
215 the relative occurrence area of each sister pair (ROA; Jiménez-Valverde et al. 2008; Lobo et al.
216 2008), which is a measure of the degree of endemism of each sister pair, with decreasing ROA
217 values indicating increasing endemism (e.g., a ROA of 0.3 suggest that the sister pair occupied
218 30% of the island of Madagascar). The ROA of each sister pair was measured as the ratio
219 between the area occupied by each sister pair relative to the extent of the island of Madagascar
220 by dividing the combined area occupied by each polygon of a sister pair by the total area of the
221 island of Madagascar.

222 For the RTR approach, we quantified the amount of niche overlap using the MO metric
223 (Nunes and Pearson 2017), which averages the overlap between the climatic conditions occupied
224 by each species of a sister pair. This approach retains the biological meaning of the climatic
225 variables (instead of transforming the climatic variables into PCA axis, Broennimann et al. 2012)
226 and avoids assumptions of climatic suitability obtained from species distribution models (Warren
227 et al. 2008; Jezkova and Wiens 2018). We then used a null model to test the degree of niche
228 conservatism (the RTR null biogeographic test, Nunes and Pearson 2017). The RTR null
229 biogeographic model tests whether the observed niche overlap is likely to be higher or lower
230 than expected by random, given the available climate of a region of interest. Given that several
231 wide-ranging endemic reptiles occupy almost the entire island of Madagascar, we constrained
232 our region of interest to the whole island thus including areas within the dispersal range of the
233 broader taxa. We considered cases where observed niche overlap was within the top or bottom
234 2.5% of the null distributions as cases of significant PNC or PND respectively.

235 To further assess our hypotheses, we also applied the BST in environmental space using
236 ordination statistics (Warren et al. 2008; Broennimann et al. 2012). Unlike the RTR model, BST
237 requires a reciprocal approach to assess whether the observed niche overlap is lower or higher
238 than the simulated niche overlaps (derived from comparison of the observed niche of one species
239 against the simulated niche from the background of the other species). Therefore, we conducted
240 two BST analyses for each sister pair (one for each reciprocal test). The environmental space was
241 calibrated using the same climatic variables as in the RTR model and the background climate of
242 each species was calculated based on a minimum convex polygon around the occurrence data for
243 each species including a 5km buffer zone (Hu et al. 2016) to reflect the low dispersal ability of
244 these range-restricted species. Species occurrence densities were projected onto a gridded PCA

245 environment with a resolution of 100x100 cells and niche overlap was calculated using
246 Schoener's *D* statistic (Warren et al. 2008). Statistical significance of niche overlap was tested
247 using a two-sided background similarity test (i.e., either lower or higher than by chance) against
248 100 niche simulations per species background (Broennimann et al. 2012; Hu et al. 2016).

249 We used these approaches to test three possible outcomes: ecological niche overlap may be
250 significantly higher than random (PNC), lower than random (PND), or not significant. We also
251 define the degree of niche conservatism as the cumulative frequency distribution of the observed
252 niche overlap within the null distribution, thus generating a continuous measure of ecological
253 niche conservatism from 0 to 1, where values closer or equal to 1 represent high degree of niche
254 conservatism and values closer or equal to 0 suggest low degree of niche conservatism. We
255 analyzed the tendency for sister pairs to cluster at either ends of the PND-PNC spectrum (i.e.,
256 suggesting either ecological or soft allopatric speciation but not hard allopatry) using Hartigan's
257 dip test for unimodal distributions from the 'dipTest' R package (Maechler 2016). A significant
258 deviation from a unimodal distribution indicates a multimodal distribution. We also calculated
259 the location of the two modes and the antimode of the bimodal distribution using the 'loemode'
260 function from the 'multimode' R package (Ameijeiras-Alonso et al. 2021). We hypothesized that
261 sister pairs centered around the upper and lower modes would suggest potential ecological
262 processes during speciation (soft allopatry and ecological speciation respectively) while sister
263 pairs clustered around the antimode would suggest either no ecological processes during
264 speciation (and therefore a hard allopatry mode of speciation) or that any ecological signal has
265 been lost over time or is undetectable using our methods (see Discussion). To test for differences
266 in observed niche overlap and degree of niche conservatism between sister pairs exhibiting
267 allopatric and parapatric distributions, we conducted a permutation test. In this permutation test,

268 observed differences between the medians of the two speciation groups are calculated (with the
269 highest median being subtracted by the lowest median). This observed difference was then tested
270 against mean differences generated from randomly allocated sister pairs to either group but
271 keeping the sample size of each group equal to the original dataset. We conducted generalized
272 linear models to test for significant relationships correlations between speciation age, range
273 overlap and degree of niche conservatism with other biogeographic traits. The analyses were
274 carried out in R (R Core Team 2020).

275

276 **Results**

277 Both the RTR and BST tests point to a bimodal distribution in the distribution of degree of
278 conservatism across our sample of sister pairs (RTR Hartingan's $D = 0.11062$ and $P < 0.05$, Fig.
279 2; BST Hartingan's $D = 0.98391, 0.12963$ and $P < 0.05$, Fig. S2, Fig. S4;). These results suggest
280 a tendency for sister pairs to tend towards either PNC or PND, thus supporting a role for
281 ecological processes in driving speciation for the majority of our sample taxa. With RTR, we
282 found that among sister pairs clustered around the upper mode of the distribution (i.e., suggesting
283 high degree of niche conservatism), 8 pairs have allopatric distributions and 5 have parapatric
284 distributions and around the lower mode (low degree of niche conservatism), 6 have allopatric
285 distributions and 8 have parapatric distributions. With BST, the upper modes consisted of 6
286 allopatric pairs and 9 parapatric pairs, and the lower modes consistent of 7 allopatric pairs and 4
287 parapatric pairs (Figs. S2, S4). Among pairs with distributions that are currently parapatric, the
288 RTR model showed one pair with significant PND (as expected) but one pair showed significant
289 PNC, which is the opposite of our expectation for parapatric pairs (Fig. 1c,f). Among allopatric
290 pairs, the RTR model showed two pairs with significant PNC (as expected) and three with

291 significant PND, which is the opposite of our expectation for allopatric pairs (Fig. 1b,e). BST
292 showed 0-1 allopatric pair with significant PNC and 2 parapatric pairs with significant PND (as
293 expected) but 5 allopatric pairs showed significant PND and 2-3 parapatric pairs showed
294 significant PNC. BST results were reciprocal between all pairs (i.e., no pairs had signals for both
295 PNC and PND, Supporting Information Table 2). We also found pairs with consistent ecological
296 signals between the RTR and BST tests (Table S2). Specifically, PND signals were consistent in
297 *U. pietschmanni*- *U. alluaudi* and *P. dubia* – *P. ravenala* and PNC signals were consistent in *L.*
298 *guibei* – *L. miops*. Only one pair, *C. boettgeri* - *C. nasutum*, had PND signal with the RTR test
299 but PNC signals with the BST test (Table S2).

300 For both the RTR model and the BST test we also found that observed niche overlap is
301 higher in parapatric pairs than allopatric pairs ($P < 0.05$; Fig. 3a, Fig. S3a; Fig S5a), which is
302 expected given that parapatric pairs have partially overlapping geographic ranges and therefore
303 are more likely to experience similar climatic conditions (climate overlap increases with
304 geographic range overlap; Supporting Information Table 3); however, when observed niche
305 overlap was tested against the null RTR model, we found that allopatric pairs tend to have niches
306 that are more similar than expected by chance (median = 0.663), whereas parapatric pairs have
307 niches that are more divergent (median = 0.281; Fig. 3b). The opposite was found when observed
308 niche overlap was assessed with BST, with allopatric pairs tending to have niches more
309 divergent than expected by chance (median = 0.26 and 0.32) than parapatric pairs (median = 0.69
310 and 0.73; Fig. S3b, Fig. S5b). For all sets of analysis, these findings do not confirm our
311 expectation that pairs with allopatric distributions tend toward niche conservatism, and pairs with
312 parapatric distributions tend toward niche divergence ($P = 0.223$; Fig. 3b and $P = 0.964$ and
313 0.865 ; Fig. S3b; Fig. S5b), but they suggest a complex role for ecological processes and point to

314 a need for further analysis of a larger number of sister pairs. We also found no significant
315 relationship between estimated divergence date for speciation (based on Zheng and Wiens, 2016)
316 and any of the other variables given in Table 1 (Supporting Information Table 3-5). These
317 results suggest that post-speciation range shifts (which otherwise should be more pronounced in
318 older sister pairs) were not contributing a major source of signal in our analyses.

319

320 **Discussion**

321 Several studies of reptiles support either speciation with PNC or PND (Wollenberg Valero
322 et al. 2019), which aligns with mixed evidence for PNC in other taxa (Peterson 2011). Examples
323 of mixed evidence among reptiles include a study of 11 Caribbean Anolis lizards that found no
324 evidence that niche conservatism was more common in closely related species than in distantly
325 related species (Knouft et al. 2006) and a survey of 49 allopatric squamate reptiles that found
326 niche divergence to be 3 times more likely than niche conservatism (Jezkova and Wiens 2018).

327 Our findings provide a first systematic assessment of speciation processes for a sample of
328 lizard sister taxa in Madagascar using two complementary null model approaches. Overall, both
329 approaches suggest tendencies for sister pairs to cluster at either end of the niche divergence-
330 conservation spectrum, thus providing robust support for ecologically-mediated speciation,
331 including both soft allopatry and ecological speciation (Fig. 2). These results highlight a strong
332 influence of ecological processes on speciation events of Malagasy reptiles. Various mechanisms
333 have been proposed to explain diversification patterns in Madagascar including forest
334 contractions (Raxworthy and Nussbaum 1995), riverine barriers (Pastorini et al. 2003),
335 watersheds (Wilme 2006), topography (Townsend et al. 2009) and selection along ecotones
336 (Pearson and Raxworthy 2009). However, individual mechanisms have weak support for

337 explaining speciation in multiple taxa, such as the influence of riverine barriers on the
338 diversification of reed frogs (Gehring et al. 2012) and lemurs (Blair et al. 2013) while evidence
339 of both watershed barriers and ecotone selection were found among reptiles and lemurs (Pearson
340 and Raxworthy 2009). In this study, we also found evidence of multiple speciation processes
341 among range-restricted reptiles which further supports the notion that not a single mechanism
342 may fully explain diversification patterns in amphibians and reptiles of Madagascar (Brown et al.
343 2014). By testing for multiple speciation hypothesis, including both ecological (i.e., soft
344 allopatry and ecological speciation) and non-ecological processes (i.e., hard allopatry), our study
345 also provides novel insights into the high prevalence of ecological niche divergence and
346 conservatism among closely-related lineages.

347 Neither test showed a significant difference in the tendency for niche conservatism
348 between allopatric and parapatric sister pairs (Fig 3b; Fig. S3b; Fig. S5b). Higher degree of niche
349 conservatism among pairs with allopatric distributions would support the hypothesis of limited
350 ecological divergence during soft allopatric speciation (Peterson et al. 1999; Graham et al. 2004)
351 and thus a role for PNC in promoting soft allopatric speciation (Wiens 2004). However, the lack
352 of statistically significant differences in the degree of niche conservatism between the two
353 groups concurs with findings of non-statistical niche divergence among allopatric pairs of
354 squamate reptiles worldwide (Jezkova and Wiens 2018). Building on from our study, potential
355 future work could apply these analyses to older lineages to further test for the consistency of
356 these patterns across deeper nodes of this phylogeny, with the caveat that older lineages allow for
357 longer times for post-speciation shifts to take place, and also recognizing the challenges of
358 estimating niches for ancestral lineages. Future work could also use additional Bayesian

359 phylogenetic methods to further test the robustness of the patterns observed from the supermatrix
360 phylogeny of Zheng and Wiens (2016).

361 Sister pairs that cluster around the antimode of the niche divergence-conservation spectrum
362 suggest a potential hard allopatric model of speciation with no role of ecological processes. One
363 of the pairs whose ecological signal was closely centered around the antimode in the RTR test, *P.*
364 *quadriocellata* – *P. antanosy*, is a strong candidate for hard allopatry, given its lack of sympatry
365 in the field and specialization to distinct palm tree species (Raxworthy pers. obs.), which would
366 be an example of jump dispersal and consequent geographic isolation due to specialization of
367 habitat but not of climatic niche preferences. However, there are several alternative explanations
368 for why an ecological signal may not be detected. For example: i) our tests may not have
369 included the relevant climatic variables or non-climatic traits such as soil microhabitats (Blair et
370 al. 2013; Wollenberg Valero et al. 2019); ii) the climatic baseline we used may not be
371 representative of the climatic conditions during speciation (Roubicek et al. 2010); iii) ecological
372 processes may have operated at a finer spatial resolution than we are able to capture (Hurlbert
373 and Jetz 2007); iv) speciation events were not related to either environmental conditions or
374 geographic barriers (for example, speciation may have been driven by sexual selection (Lande
375 1982) or socially-mediated factors (Wollenberg Valero et al. 2019)); or v) low statistical power
376 due to model inadequacy, low sample sizes or, in the case of the BST, a narrow buffer distance
377 which could lead to conservative estimates of species dispersal ability. Furthermore, non-
378 biogeographical factors could also lead to non-overlapping species distributions and thus
379 confound analysis of ecological niche evolution. For example, one possible explanation for the
380 maintenance of non-overlapping ranges through time could be biotic interactions, such as
381 interspecific territoriality between closely-related species rather than geographic or climatic

382 barriers (Freeman et al. 2019; Cowen et al. 2020). Secondly, human activities have resulted in
383 differences between current reptile distributions and pre-human distributions in Madagascar, as a
384 result of anthropogenic habitat modification (Raxworthy 2003). Given that there are these many
385 possible reasons for not finding an ecological signal in our analyses, it is thus especially
386 noteworthy that we *did* find ecological signal, suggesting that ecological processes played a role
387 during speciation and that those signals remain detectable.

388 Both methods that we used to measure niche overlap are sensitive to the choice of
389 background area (for a comparison see Brown and Carnaval 2019). Increasing the background
390 area generates random niches that may be more dissimilar to observed niches, resulting in higher
391 likelihood of type I errors (false rejection of the null hypothesis, Nunes and Pearson 2017).
392 However, the methods use distinct approaches for sampling background climates: the RTR test
393 samples climates within the largest contiguous geographical area available to the studied taxa
394 without dispersal constraints (i.e., the entire island of Madagascar) whereas the BST samples the
395 climatic space that is accessible to the species based on a dispersal buffer. Given these
396 differences, the two approaches may be expected to result in different outcomes for the same
397 sister pair (Nunes and Pearson 2017) yet, remarkably, while we found some sister pairs with
398 different ecological signals between methods, overall patterns were consistent across methods,
399 particularly the significant bimodal curves that resulted from the clustering of sister pairs in
400 opposite ends of the niche divergence-conservation spectrum.

401 Although we aimed to minimize the influence of post-speciation range shifts by our
402 selection of recently-diverged and range-restricted sister taxa, evolving in a relatively more
403 stable tropical environment; and we found no evidence for the effect of speciation age on any
404 biogeographic traits (Table S5); the potential for range shifts to confound these types of analyses

405 remains (Losos and Glor 2003). For instance, extirpations may have resulted in distribution
406 contractions and thus post-speciation loss of a contact zone in the *U. pietschmanni*- *U. alluaudi*
407 pair, both species of which are considered to be biogeographic relicts (Raxworthy et al. 2008). In
408 contrast, range expansion and secondary contact after speciation might explain the current
409 parapatric distributions and unexpected PNC signal in the pair *P. androyensis* – *P. picta*. To
410 more definitively pinpoint post-speciation range shift events and historical gene flow, these non-
411 concordant pairs should be targets for future genetic work to look for evidence of gene flow
412 during speciation (Martin et al. 2013; Sousa and Hey 2013; Seehausen et al. 2014) and for post-
413 speciation population expansion (Gehring et al. 2013; Rakotoarisoa et al. 2013a,b; Gehara et al.
414 2017).

415 In conclusion, despite there being several factors that will confound our ability to identify
416 ecological signals, and there being a broad set of circumstances under which ecological
417 speciation could occur, our analyses find evidence of a tendency for sister pairs to cluster at
418 either end of the niche divergence-conservation spectrum. This supports a strong role for
419 ecological processes – both soft allopatry and ecological speciation – in driving speciation and
420 points to multiple mechanisms as explanations for the origin of local endemism and high species
421 diversity in Madagascar.

422

423 **References**

424 Ameijeiras-Alonso, J., R. M. Crujeiras, and A. Rodriguez-Casal. 2021. multimode: An R
425 Package for Mode Assessment. J. Stat. Softw. Vol 1 Issue 9 2021, doi:
426 10.18637/jss.v097.i09.

427 Anacker, B. L., and S. Y. Strauss. 2014. The geography and ecology of plant speciation: range
428 overlap and niche divergence in sister species. *Proc. R. Soc. B Biol. Sci.* 281:20132980.

429 Barve, N. 2011. The crucial role of the accessible area in ecological niche modeling and species
430 distribution modeling. *Ecol. Model.* 10.

431 Berner, D., A.-C. Grandchamp, and A. P. Hendry. 2009. Variable progress toward ecological
432 speciation in parapatry: Stickleback across eight lake-stream transitions. *Evolution*
433 63:1740–1753.

434 Blair, M. E., E. J. Sterling, M. Dusch, C. J. Raxworthy, and R. G. Pearson. 2013. Ecological
435 divergence and speciation between lemur (*Eulemur*) sister species in Madagascar. *J.*
436 *Evol. Biol.* 26:1790–1801.

437 Broennimann, O., M. C. Fitzpatrick, P. B. Pearman, B. Petitpierre, L. Pellissier, N. G. Yoccoz,
438 W. Thuiller, M.-J. Fortin, C. Randin, N. E. Zimmermann, C. H. Graham, and A. Guisan.
439 2012. Measuring ecological niche overlap from occurrence and spatial environmental
440 data: Measuring niche overlap. *Glob. Ecol. Biogeogr.* 21:481–497.

441 Brown, J. H., G. C. Stevens, and D. M. Kaufman. 1996. The geographic range: Size, Shape,
442 Boundaries, and Internal Structure. *Annu. Rev. Ecol. Syst.* 27:597–623.

443 Brown, J. L., A. Cameron, A. D. Yoder, and M. Vences. 2014. A necessarily complex model to
444 explain the biogeography of the amphibians and reptiles of Madagascar. *Nat. Commun.*
445 5:5046.

446 Cardillo, M., R. Dinnage, and W. McAlister. 2019. The relationship between environmental
447 niche breadth and geographic range size across plant species. *J. Biogeogr.* 46:97–109.

448 Caro, L. M., P. C. Caycedo-Rosales, R. C. K. Bowie, H. Slabbekoorn, and C. D. Cadena. 2013.
449 Ecological speciation along an elevational gradient in a tropical passerine bird? *J. Evol.*
450 *Biol.* 26:357–374.

451 Constable, H., R. Guralnick, J. Wieczorek, C. Spencer, A. T. Peterson, and The VertNet Steering
452 Committee. 2010. VertNet: A New Model for Biodiversity Data Sharing. *PLoS Biol.*
453 8:e1000309.

454 Cowen, M. C., J. P. Drury, and G. F. Grether. 2020. Multiple routes to interspecific territoriality
455 in sister species of North American perching birds. *Evolution* 74:2134–2148.

456 Culumber, Z. W., and M. Tobler. 2016. Ecological divergence and conservatism: spatiotemporal
457 patterns of niche evolution in a genus of livebearing fishes (Poeciliidae: Xiphophorus).
458 *BMC Evol. Biol.* 16:44.

459 Doebeli, M., and U. Dieckmann. 2003. Speciation along environmental gradients. *Nature*
460 421:259–264.

461 Elith, J., and C. H. Graham. 2009. Do they? How do they? WHY do they differ? On finding
462 reasons for differing performances of species distribution models. *Ecography* 32:66–77.

463 Fick, S. E., and R. J. Hijmans. 2017. WorldClim 2: new 1-km spatial resolution climate surfaces
464 for global land areas. *Int. J. Climatol.* 37:4302–4315.

465 Fisher, R. A. 1930. *The genetical theory of natural selection*. Clarendon Press, Oxford, England.

466 Florio, A. M., and C. J. Raxworthy. 2016. A Phylogeographic Assessment of the Malagasy Giant
467 Chameleons (*Furcifer verrucosus* and *Furcifer oustaleti*). *PLOS ONE* 11:e0154144.

468 Freeman, B. G., J. A. Tobias, and D. Schluter. 2019. Behavior influences range limits and
469 patterns of coexistence across an elevational gradient in tropical birds. *Ecography*
470 42:1832–1840.

471 Gehara, M., A. A. Garda, F. P. Werneck, E. F. Oliveira, E. M. da Fonseca, F. Camurugi, F. de M.
472 Magalhães, F. M. Lanna, J. W. Sites, R. Marques, R. Silveira-Filho, V. A. São Pedro, G.
473 R. Colli, G. C. Costa, and F. T. Burbrink. 2017. Estimating synchronous demographic
474 changes across populations using hABC and its application for a herpetological
475 community from northeastern Brazil. *Mol. Ecol.* 26:4756–4771.

476 Gehring, P.-S., F. Glaw, M. Gehara, F. M. Ratsavina, and M. Vences. 2013. Northern origin
477 and diversification in the central lowlands? – Complex phylogeography and taxonomy of
478 widespread day geckos (*Phelsuma*) from Madagascar. *Org. Divers. Evol.* 13:605–620.

479 Gehring, P.-S., M. Pabijan, J. E. Randrianirina, F. Glaw, and M. Vences. 2012. The influence of
480 riverine barriers on phylogeographic patterns of Malagasy reed frogs (*Heterixalus*). *Mol.*
481 *Phylogenet. Evol.* 64:618–632.

482 Graham, C. H., J. Elith, R. J. Hijmans, A. Guisan, A. Townsend Peterson, B. A. Loiselle, and
483 The Nceas Predicting Species Distributions Working Group. 2007. The influence of
484 spatial errors in species occurrence data used in distribution models: Spatial error in
485 occurrence data for predictive modelling. *J. Appl. Ecol.* 45:239–247.

486 Graham, C. H., S. R. Ron, J. C. Santos, C. J. Schneider, and C. Moritz. 2004. Integrating
487 phylogenetics and environmental niche models to explore speciation mechanisms in
488 dendrobatid frogs. *Evolution* 58:1781.

489 Guillera-Arroita, G., J. J. Lahoz-Monfort, J. Elith, A. Gordon, H. Kujala, P. E. Lentini, M. A.
490 McCarthy, R. Tingley, and B. A. Wintle. 2015. Is my species distribution model fit for
491 purpose? Matching data and models to applications: Matching distribution models to
492 applications. *Glob. Ecol. Biogeogr.* 24:276–292.

493 Hu, J., O. Broennimann, A. Guisan, B. Wang, Y. Huang, and J. Jiang. 2016. Niche conservatism
494 in Gynandropaa frogs on the southeastern Qinghai-Tibetan Plateau. *Sci. Rep.* 6:32624.

495 Hurlbert, A. H., and W. Jetz. 2007. Species richness, hotspots, and the scale dependence of range
496 maps in ecology and conservation. *Proc. Natl. Acad. Sci.* 104:13384–13389.

497 Jezkova, T., and J. J. Wiens. 2018. Testing the role of climate in speciation: New methods and
498 applications to squamate reptiles (lizards and snakes). *Mol. Ecol.* 27:2754–2769.

499 Jiménez-Valverde, A., J. M. Lobo, and J. Hortal. 2008. Not as good as they seem: the importance
500 of concepts in species distribution modelling. *Divers. Distrib.* 14:885–890.

501 Knouft, J. H., J. B. Losos, R. E. Glor, and J. J. Kolbe. 2006. Phylogenetic analysis of the evolution
502 of the niche in lizards of the *Anolis sagrei* group. *Ecology* 87:S29–S38.

503 Kozak, K. H., and JohnJ. Wiens. 2006. Does niche conservatism promote speciation? A case
504 study in North American salamanders. *Evolution* 60:2604–2621.

505 Lande, R. 1982. Rapid origin of sexual isolation and character divergence in a cline. *Evolution*
506 36:213–223.

507 Lester, S. E., B. I. Ruttenberg, S. D. Gaines, and B. P. Kinlan. 2007. The relationship between
508 dispersal ability and geographic range size. *Ecol. Lett.* 10:745–758.

509 Lobo, J. M., A. Jiménez-Valverde, and R. Real. 2008. AUC: a misleading measure of the
510 performance of predictive distribution models. *Glob. Ecol. Biogeogr.* 17:145–151.

511 Losos, J. B. 2008. Phylogenetic niche conservatism, phylogenetic signal and the relationship
512 between phylogenetic relatedness and ecological similarity among species. *Ecol. Lett.*
513 11:995–1003.

514 Losos, J. B., and R. E. Glor. 2003. Phylogenetic comparative methods and the geography of
515 speciation. *Trends Ecol. Evol.* 18:220–227.

516 Maechler, M. 2016. diptest: Hartigan's Dip Test Statistic for Unimodality - Corrected.

517 Martin, S. H., K. K. Dasmahapatra, N. J. Nadeau, C. Salazar, J. R. Walters, F. Simpson, M.

518 Blaxter, A. Manica, J. Mallet, and C. D. Jiggins. 2013. Genome-wide evidence for

519 speciation with gene flow in *Heliconius* butterflies. *Genome Res.* 23:1817–1828.

520 McCormack, J. E., A. J. Zellmer, and L. L. Knowles. 2009. Does niche divergence accompany

521 allopatric divergence in *Aphelocoma* jays as predicted under ecological speciation:

522 Insights from tests with niche models. *Evolution*, doi: 10.1111/j.1558-

523 5646.2009.00900.x.

524 Niemiller, M. L., B. M. Fitzpatrick, and B. T. Miller. 2008. Recent divergence with gene flow in

525 Tennessee cave salamanders (Plethodontidae: *Gyrinophilus*) inferred from gene

526 genealogies. *Mol. Ecol.* 17:2258–2275.

527 Nosil, P. 2008. Speciation with gene flow could be common. *Mol. Ecol.* 17:2103–2106.

528 Nunes, L. A., and R. G. Pearson. 2017. A null biogeographical test for assessing ecological niche

529 evolution. *J. Biogeogr.* 44:1331–1343.

530 Owens, H. L., V. Ribeiro, E. E. Saupe, M. E. Cobos, P. A. Hosner, J. C. Cooper, A. M. Samy, V.

531 Barve, N. Barve, C. J. Muñoz-R., and A. T. Peterson. 2020. Acknowledging uncertainty

532 in evolutionary reconstructions of ecological niches. *Ecol. Evol.* 10:6967–6977.

533 Papadopulos, A. S. T., M. Kaye, C. Devaux, H. Hipperson, J. Lighten, L. T. Dunning, I. Hutton,

534 W. J. Baker, R. K. Butlin, and V. Savolainen. 2014. Evaluation of genetic isolation

535 within an island flora reveals unusually widespread local adaptation and supports

536 sympatric speciation. *Philos. Trans. R. Soc. B Biol. Sci.* 369:20130342.

537 Pastorini, J., U. Thalmann, and R. D. Martin. 2003. A molecular approach to comparative

538 phylogeography of extant Malagasy lemurs. *Proc. Natl. Acad. Sci.* 100:5879–5884.

539 Pearson, R. G., and C. J. Raxworthy. 2009. The evolution of local endemism in Madagascar:
540 Watershed versus climatic gradient hypotheses evaluated by null biogeographic models.
541 *Evolution* 63:959–967.

542 Pearson, R. G., J. C. Stanton, K. T. Shoemaker, M. E. Aiello-Lammens, P. J. Ersts, N. Horning,
543 D. A. Fordham, C. J. Raxworthy, H. Y. Ryu, J. McNees, and H. R. Akçakaya. 2014. Life
544 history and spatial traits predict extinction risk due to climate change. *Nat. Clim. Change*
545 4:217–221.

546 Pebesma, E., and R. S. Bivand. 2005. S Classes and Methods for Spatial Data: the sp Package.
547 22.

548 Peterson, A. T. 2011. Ecological niche conservatism: a time-structured review of evidence:
549 Ecological niche conservatism. *J. Biogeogr.* 38:817–827.

550 Peterson, A. T., J. Soberón, and V. Sánchez-Cordero. 1999. Conservatism of Ecological Niches
551 in Evolutionary Time. *Science* 285:1265–1267.

552 Prosdij, A. S. J., M. S. M. Sosef, J. J. Wieringa, and N. Raes. 2016. Minimum required number
553 of specimen records to develop accurate species distribution models. *Ecography* 39:542–
554 552.

555 Pyron, R. A., and F. T. Burbrink. 2010. Hard and soft allopatry: physically and ecologically
556 mediated modes of geographic speciation: Modes of allopatric speciation. *J. Biogeogr.*
557 37:2005–2015.

558 Pyron, R. A., G. C. Costa, M. A. Patten, and F. T. Burbrink. 2015. Phylogenetic niche
559 conservatism and the evolutionary basis of ecological speciation: Niche conservatism and
560 speciation. *Biol. Rev.* 90:1248–1262.

561 Pyron, R., F. T. Burbrink, and J. J. Wiens. 2013. A phylogeny and revised classification of
562 Squamata, including 4161 species of lizards and snakes. *BMC Evol. Biol.* 13:93.

563 R Core Team. 2020. R: A Language and Environment for Statistical Computing. R Foundation
564 for Statistical Computing, Vienna, Austria.

565 Rakotoarisoa, J.-E., M. Raheriarisena, and S. M. Goodman. 2013a. A Phylogeographic Study of
566 the Endemic Rodent *Eliurus carletoni* (Rodentia: Nesomyinae) in an Ecological
567 Transition Zone of Northern Madagascar. *J. Hered.* 104:23–35.

568 Rakotoarisoa, J.-E., M. Raheriarisena, and S. M. Goodman. 2013b. Late Quaternary climatic
569 vegetational shifts in an ecological transition zone of northern Madagascar: insights from
570 genetic analyses of two endemic rodent species. *J. Evol. Biol.* 26:1019–1034.

571 Raxworthy, C. 2003. Introduction to the reptiles. *Nat. Hist. Madag.* 934–949. University of
572 Chicago Press Chicago.

573 Raxworthy, C. J., C. M. Ingram, N. Rabibisoa, and R. G. Pearson. 2007. Applications of
574 Ecological Niche Modeling for Species Delimitation: A Review and Empirical
575 Evaluation Using Day Geckos (*Phelsuma*) from Madagascar. *Syst. Biol.* 56:907–923.

576 Raxworthy, C. J., and R. A. Nussbaum. 1995. Systematics, speciation and biogeography of the
577 dwarf chameleons (*Brookesia*; Reptilia, Squamata, Chamaeleontidae) of northern
578 Madagascar. *J. Zool.* 235:525–558.

579 Raxworthy, C. J., R. G. Pearson, B. M. Zimkus, S. Reddy, A. J. Deo, R. A. Nussbaum, and C. M.
580 Ingram. 2008. Continental speciation in the tropics: contrasting biogeographic patterns of
581 divergence in the *Uroplatus* leaf-tailed gecko radiation of Madagascar. *J. Zool.* 275:423–
582 440.

583 Ricklefs, R. E., and E. Bermingham. 2007. The Causes of Evolutionary Radiations in
584 Archipelagoes: Passerine Birds in the Lesser Antilles. *Am. Nat.* 169:285–297.

585 Roubicek, A. J., J. VanDerWal, L. J. Beaumont, A. J. Pitman, P. Wilson, and L. Hughes. 2010.
586 Does the choice of climate baseline matter in ecological niche modelling? *Ecol. Model.*
587 221:2280–2286.

588 Ruane, S., E. A. Myers, K. Lo, S. Yuen, R. S. Welt, M. Juman, I. Futterman, R. A. Nussbaum, G.
589 Schneider, F. T. Burbrink, and C. J. Raxworthy. 2018. Unrecognized species diversity
590 and new insights into colour pattern polymorphism within the widespread Malagasy
591 snake *Mimophis* (Serpentes: Lamprophiidae). *Syst. Biodivers.* 16:229–244. Taylor &
592 Francis.

593 Rundle, H. D., and P. Nosil. 2005. Ecological speciation: Ecological speciation. *Ecol. Lett.*
594 8:336–352.

595 Saupe, E. E., N. Barve, H. L. Owens, J. C. Cooper, P. A. Hosner, and A. T. Peterson. 2018.
596 Reconstructing Ecological Niche Evolution When Niches Are Incompletely
597 Characterized. *Syst. Biol.* 67:428–438.

598 Schluter, D. 2001. Ecology and the origin of species. *Trends Ecol. Evol.* 16:372–380.

599 Seehausen, O., R. K. Butlin, I. Keller, C. E. Wagner, J. W. Boughman, P. A. Hohenlohe, C. L.
600 Peichel, G.-P. Saetre, C. Bank, Å. Brännström, A. Brelsford, C. S. Clarkson, F.
601 Eroukhmanoff, J. L. Feder, M. C. Fischer, A. D. Foote, P. Franchini, C. D. Jiggins, F. C.
602 Jones, A. K. Lindholm, K. Lucek, M. E. Maan, D. A. Marques, S. H. Martin, B.
603 Matthews, J. I. Meier, M. Möst, M. W. Nachman, E. Nonaka, D. J. Rennison, J.
604 Schwarzer, E. T. Watson, A. M. Westram, and A. Widmer. 2014. Genomics and the
605 origin of species. *Nat. Rev. Genet.* 15:176–192.

606 Soultan, A., and K. Safi. 2017. The interplay of various sources of noise on reliability of species
607 distribution models hinges on ecological specialisation. *PLOS ONE* 12:e0187906.

608 Sousa, V., and J. Hey. 2013. Understanding the origin of species with genome-scale data:
609 modelling gene flow. *Nat. Rev. Genet.* 14:404–414.

610 Title, P. O., and D. L. Rabosky. 2016. Do Macrophylogenies Yield Stable Macroevolutionary
611 Inferences? An Example from Squamate Reptiles. *Syst. Biol.* syw102.

612 Townsend, T. M., D. R. Vieites, F. Glaw, and M. Vences. 2009. Testing Species-Level
613 Diversification Hypotheses in Madagascar: The Case of Microendemic *Brookesia* Leaf
614 Chameleons. *Syst. Biol.* 58:641–656.

615 Vences, M., K. C. Wollenberg, D. R. Vieites, and D. C. Lees. 2009. Madagascar as a model
616 region of species diversification. *Trends Ecol. Evol.* 24:456–465.

617 Warren, D. L., R. E. Glor, and M. Turelli. 2008. Environmental niche equivalency versus
618 conservatism: Quantitative approaches to niche evolution. *Evolution* 62:2868–2883.

619 Warren, D. L., N. J. Matzke, and T. L. Iglesias. 2020. Evaluating presence-only species
620 distribution models with discrimination accuracy is uninformative for many applications.
621 *J. Biogeogr.* 47:167–180.

622 Wiens, J. J. 2004. Speciation and ecology revisited: Phylogenetic niche conservatism and the
623 origin of species. *Evolution* 58:193.

624 Wiens, J. J., and C. H. Graham. 2005. Niche Conservatism: Integrating Evolution, Ecology, and
625 Conservation Biology. *Annu. Rev. Ecol. Evol. Syst.* 36:519–539.

626 Wilme, L. 2006. Biogeographic Evolution of Madagascar’s Microendemic Biota. *Science*
627 312:1063–1065.

628 Wollenberg Valero, Marshall, Bastiaans, Caccone, Camargo, Morando, Niemiller, Pabijan,
629 Russello, Sinervo, Werneck, Sites, Jr., Wiens, and Steinfartz. 2019. Patterns, Mechanisms
630 and Genetics of Speciation in Reptiles and Amphibians. *Genes* 10:646.

631 Yoder, A. D., and M. D. Nowak. 2006. Has Vicariance or Dispersal Been the Predominant
632 Biogeographic Force in Madagascar? Only Time Will Tell. *Annu. Rev. Ecol. Evol. Syst.*
633 37:405–431.

634 Zheng, Y., and J. J. Wiens. 2016. Combining phylogenomic and supermatrix approaches, and a
635 time-calibrated phylogeny for squamate reptiles (lizards and snakes) based on 52 genes
636 and 4162 species. *Mol. Phylogenet. Evol.* 94:537–547.

637

638 **Competing interests**

639 The authors declare no competing interests.

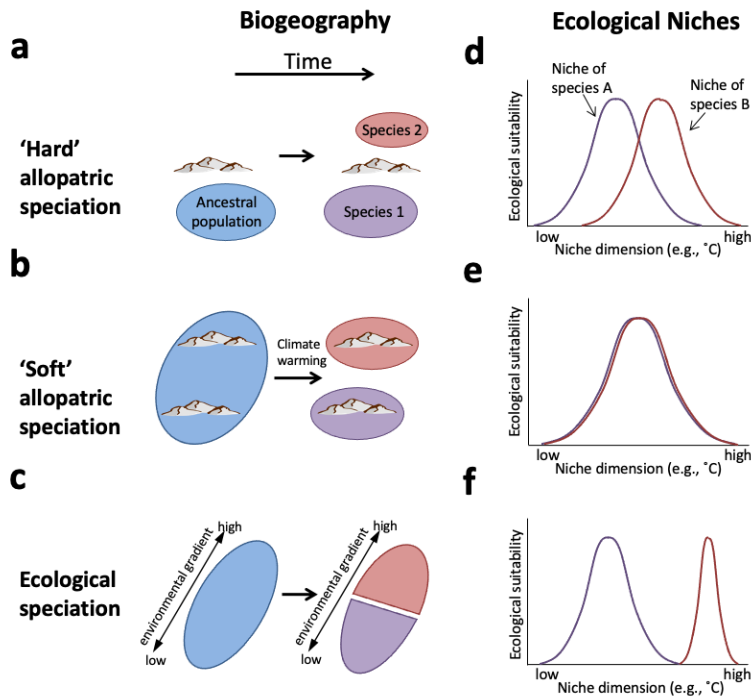
640

641 **Additional information**

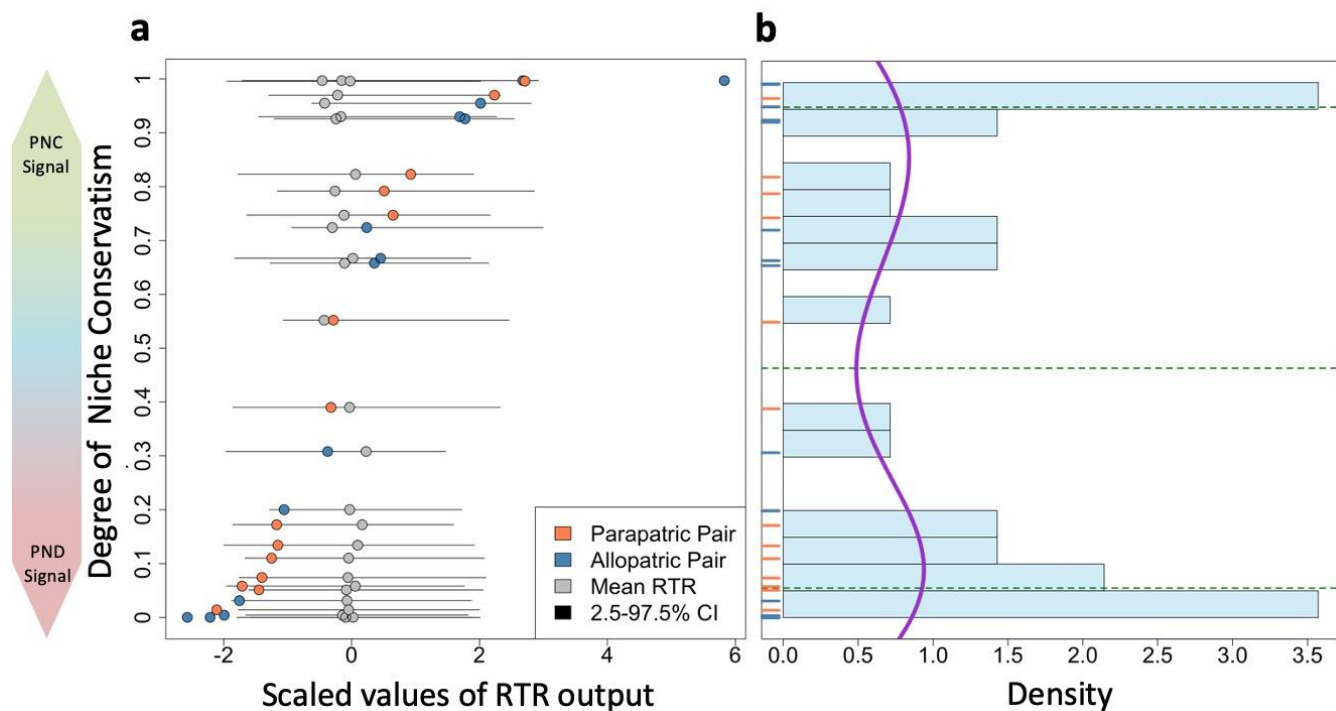
642 Supporting information includes Table 1 showing full set of results for 28 pairs, including scaled
643 CI from RTR model, Table 2 showing the full set of results for the background similarity test,
644 Table 3-5 showing results for generalized linear models for range overlap, degree of niche
645 conservation and speciation age, Table 6-7 showing gene data used for phylogenetic placement
646 of 56 sampled taxa from Zheng and Wiens (2016), Figures S1-S5 showing the analysis based on
647 the background similarity test and a spreadsheet with occurrences for all pairs used in this
648 analysis.

649

650 **Figure Legends**



651
 652 **Fig. 1: Alternative mechanisms of speciation illustrated in terms of biogeography and realized ecological**
 653 **niches. a,** Hard allopatry occurs when a population disperses to a new area that has been isolated by a physical
 654 barrier (e.g., a mountain range) which leads to the isolation of populations from the same ancestral species, resulting
 655 in descendent species with non-overlapping geographic ranges or gene flow. **b,** Soft allopatry occurs when an
 656 environmental disturbance (e.g., climate warming) causes incipient species to become geographically and
 657 genetically isolated due to failure to adapt to novel intervening conditions. **c,** Ecological speciation occurs when
 658 populations adapt to novel environmental features that are distinct from their close relatives, at first with gene flow
 659 but this ceases with time. **d,** In hard allopatry, there may be some ecological niche differences among sister species,
 660 but this differentiation is associated with differences in the availability in ecological features within landscapes (or
 661 random independent evolution) rather than due to species selecting particular features, resulting in no significant
 662 signal of either niche conservation or divergence (i.e., non-ecological signal). **e,** In soft allopatry, lack of adaptation
 663 to novel conditions by descendent species results in ecological niches that are conserved over evolutionary
 664 timescales. **f,** In ecological speciation, species adapt to ecological features that are distinct from their ancestors and
 665 close relatives, leading to ecological niche differentiation between two descendent species.



667

668 **Fig. 2: Bimodal distribution for the degree of niche conservatism for 28 sister pairs of endemic Malagasy**

669 **lizards. a**, Degree of niche conservatism against scaled values of RTR outputs, ordered by decreasing scaled niche

670 overlap values for each sister pair. To illustrate the results for all sister pairs under one common scale, each set of

671 RTR outputs were scaled using their corresponding means and standard deviations using the scale function in R. The

672 degree of niche conservatism of each sister pair is measured as the proportion of niche overlap replicates that are

673 lower than the observed niche overlap (circles), so that higher values reflect a higher degree of phylogenetic niche

674 conservatism (PNC) and lower values are associated with a lower degree of phylogenetic niche conservatism (PND).

675 Cases where the observed niche overlap does not overlap with the 2.5-97.5% intervals of the replicate distributions

676 have a significant ecological signal for either PNC (higher than 97.5%; 3 pairs) or PND (lower than 2.5%, 4 pairs).

677 **b**, Density histogram showing the degree of niche conservatism for 28 pairs fitted with a kernel density line curve

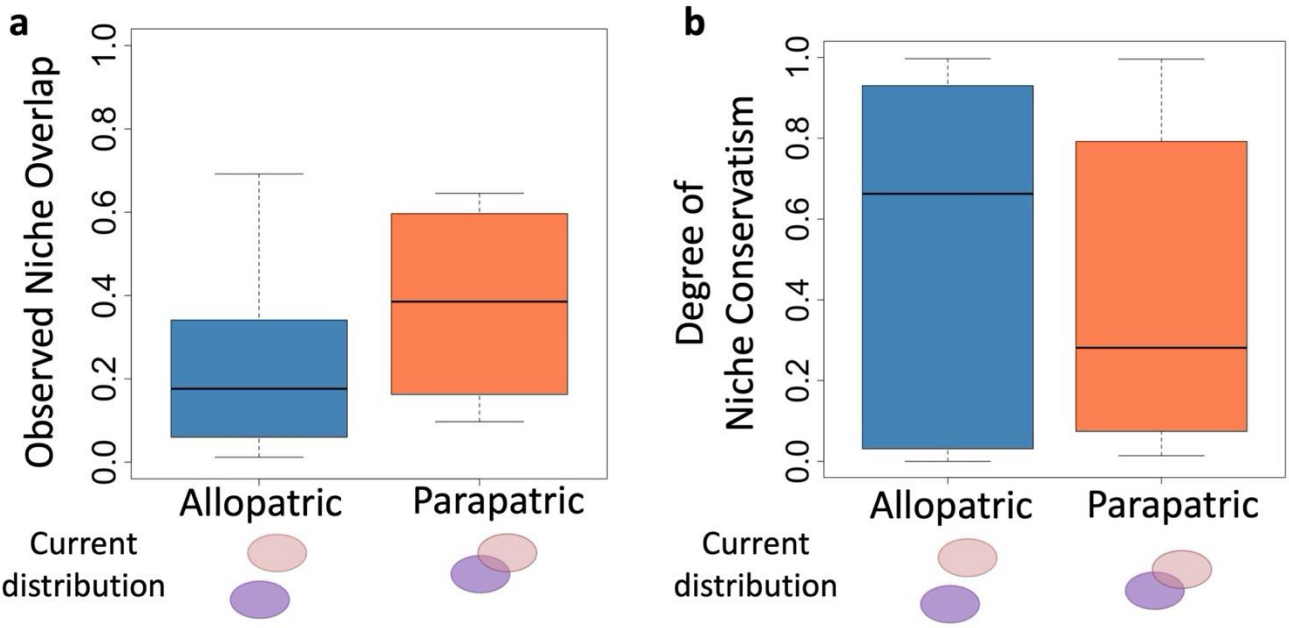
678 (purple). Hartigan's dip test supports a bimodal distribution ($D = 0.11062$, $P < 0.005$). Tick marks represent

679 individual allopatric (blue) and parapatric (orange) sister pairs and dotted lines represent the upper and lower modes

680 (0.954 and 0.055 respectively) and the antimode (0.466) of the distribution. Full table of results in Supplementary

681 Table 1.

682



683

684

Fig. 3: Comparison of observed niche overlap and degree of niche conservatism between allopatric (n=14)

685

and parapatric sister pairs (n=14) of endemic Malagasy lizards. a, Allopatric sister pairs have significantly

686

lower observed niche overlap compared to parapatric sister pairs ($P = 0.055$).

687

b, The degree of niche conservatism in allopatric pairs tends to be higher than in parapatric pairs but differences are not statistically significantly ($P =$

688

0.223).

689

Table 1. Geographic range, biogeographic pattern, niche overlap and degree of niche conservatism among 28 sister pairs of endemic lizards of Madagascar, ranked by genus and degree of niche conservatism based on the RTR model. Data on age of speciation was obtained from Zheng and Wiens (2016). Relative occurrence area refers to the proportion of the total island that is occupied by the minimum convex polygon of the sister pair. Results on niche overlap and degree of niche conservatism based on the ‘background similarity test’ can be found in the Supporting Information Table 2.

Genus	Species #1	Species #2	Age of Speciation (MYBP)	Relative occurrence area	Isolation distance (km)	Range overlap	Biogeography	Climatic niche overlap	Degree of Niche Conservatism
<i>Amphiglossus</i>	<i>A. mandokava</i>	<i>A. tanysona</i>	7.88	0.067	0.000	0.018	Parapatry	0.132	0.074
<i>Amphiglossus</i>	<i>A. punctatus</i>	<i>A. frontoparietalis</i>	9.64	0.153	0.000	0.819	Parapatry	0.645	0.058
<i>Brookesia</i>	<i>B. dentata</i>	<i>B. exarmata</i>	27.55	0.013	290.840	0.000	Allopatry	0.134	0.955
<i>Brookesia</i>	<i>B. minima</i>	<i>B. tuberculata</i>	32.92	0.012	96.134	0.000	Allopatry	0.466	0.930
<i>Brookesia</i>	<i>B. thieli</i>	<i>B. vadoni</i>	10.47	0.130	0.000	0.034	Parapatry	0.617	0.823
<i>Brookesia</i>	<i>B. therezieni</i>	<i>B. superciliaris</i>	19.04	0.174	0.000	0.983	Parapatry	0.460	0.134
<i>Brookesia</i>	<i>B. betschi</i>	<i>B. lineata</i>	11.53	0.008	0.000	0.153	Parapatry	0.162	0.110
<i>Calumma</i>	<i>C. hilleniusi</i>	<i>C. guibei</i>	13.21	0.034	613.034	0.000	Allopatry	0.164	0.997
<i>Calumma</i>	<i>C. furcifer</i>	<i>C. gastrotaenia</i>	16.48	0.093	0.000	0.091	Parapatry	0.418	0.747
<i>Calumma</i>	<i>C. tsaratananense</i>	<i>C. brevicorne</i>	4.48	0.097	39.492	0.000	Allopatry	0.189	0.658
<i>Calumma</i>	<i>C. boettgeri</i>	<i>C. nasutum</i>	4.32	0.283	5.500	0.000	Allopatry	0.312	0.004
<i>Furcifer</i>	<i>F. willsii</i>	<i>F. petteri</i>	19.41	0.358	141.509	0.000	Allopatry	0.409	0.308
<i>Furcifer</i>	<i>F. labordi</i>	<i>F. antimena</i>	4.31	0.107	0.000	0.244	Parapatry	0.210	0.051
<i>Lygodactylus</i>	<i>L. guibei</i>	<i>L. miops</i>	19.62	0.255	0.000	0.546	Parapatry	0.597	0.996
<i>Lygodactylus</i>	<i>L. pictus</i>	<i>L. mirabilis</i>	25.71	0.024	60.206	0.000	Allopatry	0.040	0.724
<i>Madascincus</i>	<i>M. stumpffi</i>	<i>M. polleni</i>	4.76	0.343	304.155	0.000	Allopatry	0.303	0.926
<i>Oplurus</i>	<i>O. fierinensis</i>	<i>O. grandidieri</i>	8.78	0.068	185.458	0.000	Allopatry	0.051	0.000
<i>Paroedura</i>	<i>P. androyensis</i>	<i>P. picta</i>	43.75	0.220	0.000	0.796	Parapatry	0.604	0.390
<i>Paroedura</i>	<i>P. oviceps</i>	<i>P. karstophila</i>	27.98	0.039	0.000	0.085	Parapatry	0.118	0.014
<i>Phelsuma</i>	<i>P. quadriocellata</i>	<i>P. antanosy</i>	16.14	0.185	0.000	0.548	Parapatry	0.174	0.552
<i>Phelsuma</i>	<i>P. dubia</i>	<i>P. ravenala</i>	10.73	0.376	274.942	0.000	Allopatry	0.012	0.031
<i>Tracheloptychus</i>	<i>T. madagascariensis</i>	<i>T. petersi</i>	17.04	0.097	0.000	0.047	Parapatry	0.353	0.970
<i>Trachylepis</i>	<i>T. dumasi</i>	<i>T. aureopunctata</i>	7.55	0.284	0.000	0.804	Parapatry	0.571	0.172
<i>Uroplatus</i>	<i>U. fimbriatus</i>	<i>U. giganteus</i>	14.14	0.246	35.756	0.000	Allopatry	0.692	0.997
<i>Uroplatus</i>	<i>U. malahelo</i>	<i>U. guentheri</i>	56.83	0.366	276.172	0.000	Allopatry	0.123	0.200
<i>Uroplatus</i>	<i>U. pietschmanni</i>	<i>U. alluaudi</i>	41.43	0.052	428.676	0.000	Allopatry	0.060	0.000
<i>Voeltzkowia</i>	<i>V. rubrocaudata</i>	<i>V. lineata</i>	0.01	0.165	34.615	0.000	Allopatry	0.341	0.667
<i>Zonosaurus</i>	<i>Z. trilineatus</i>	<i>Z. quadrilineatus</i>	4.57	0.046	0.000	0.122	Parapatry	0.097	0.792